

Figure S1

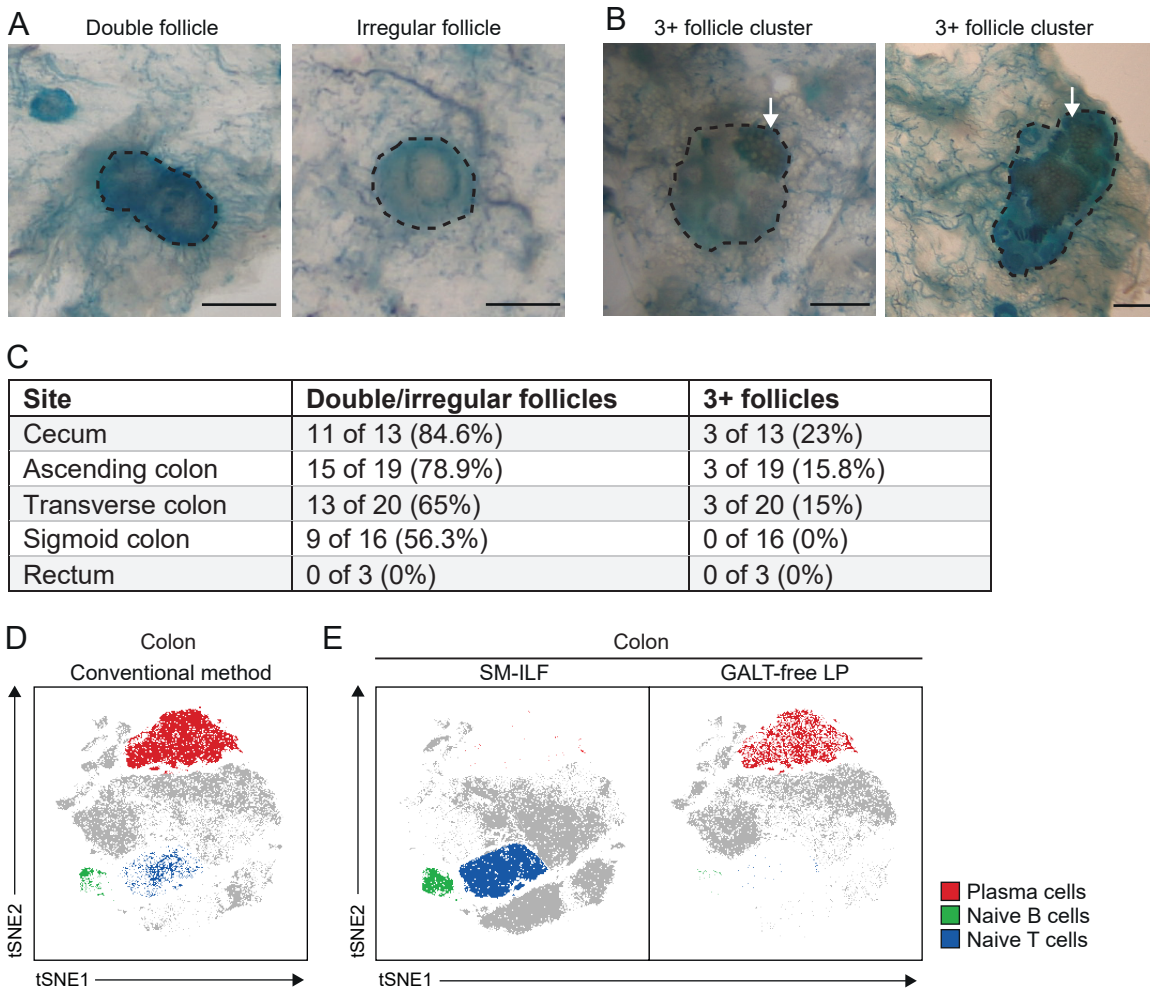


Figure S1. Related to Figure 1.

(A and B) GALT in the submucosa were occasionally found to consist of (A) two follicles or have irregular shapes or (B) clusters of three or more follicles. Scale bars represent 1 mm. (C) Proportion of intestinal site that contained irregular or multi-follicular SM-GALT. (D) Representative tSNE plot derived from flow cytometric analysis of CD45⁺ cells isolated from digested colon mucosa with attached submucosa using conventional protocols. Surface markers used for tSNE: CD45, CD3, CD8, CD20, CD38, CD138, CD45RA, TCR $\alpha\beta$, IgD, TCR $\alpha\beta$, CD117, CCR6, IgA. (E) Representative tSNE plots derived from flow cytometric analysis of CD45⁺ cells isolated from colon SM-ILF and GALT-free colon LP. GALT, gut-associated lymphoid tissue; SM-ILF, submucosal follicle; LP, lamina propria.

Figure S2

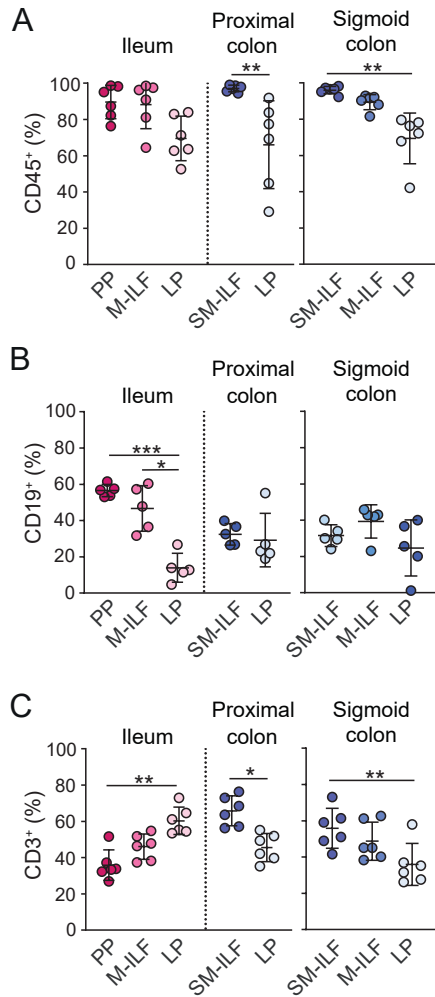


Figure S2. Related to Figure 2.

(A-C) Flow cytometric analysis of indicated immune niches. (A) Proportion of viable cells expressing CD45, and proportion of CD45⁺ cells expressing (B) CD19 and (C) CD3. Each circle represents an individual donor with paired samples from 4 patients. SM-ILF and M-ILF data are from at least 5 pooled follicles. Bars represent mean \pm 1SD and statistical significance between paired samples was determined using the paired Friedman test with Dunn's multiple comparisons, *p<0.05, **p<0.01, ***p<0.001. PP, Peyer's patch; SM-ILF, submucosal follicle; M-ILF, mucosal follicle; LP, lamina propria.

Figure S3

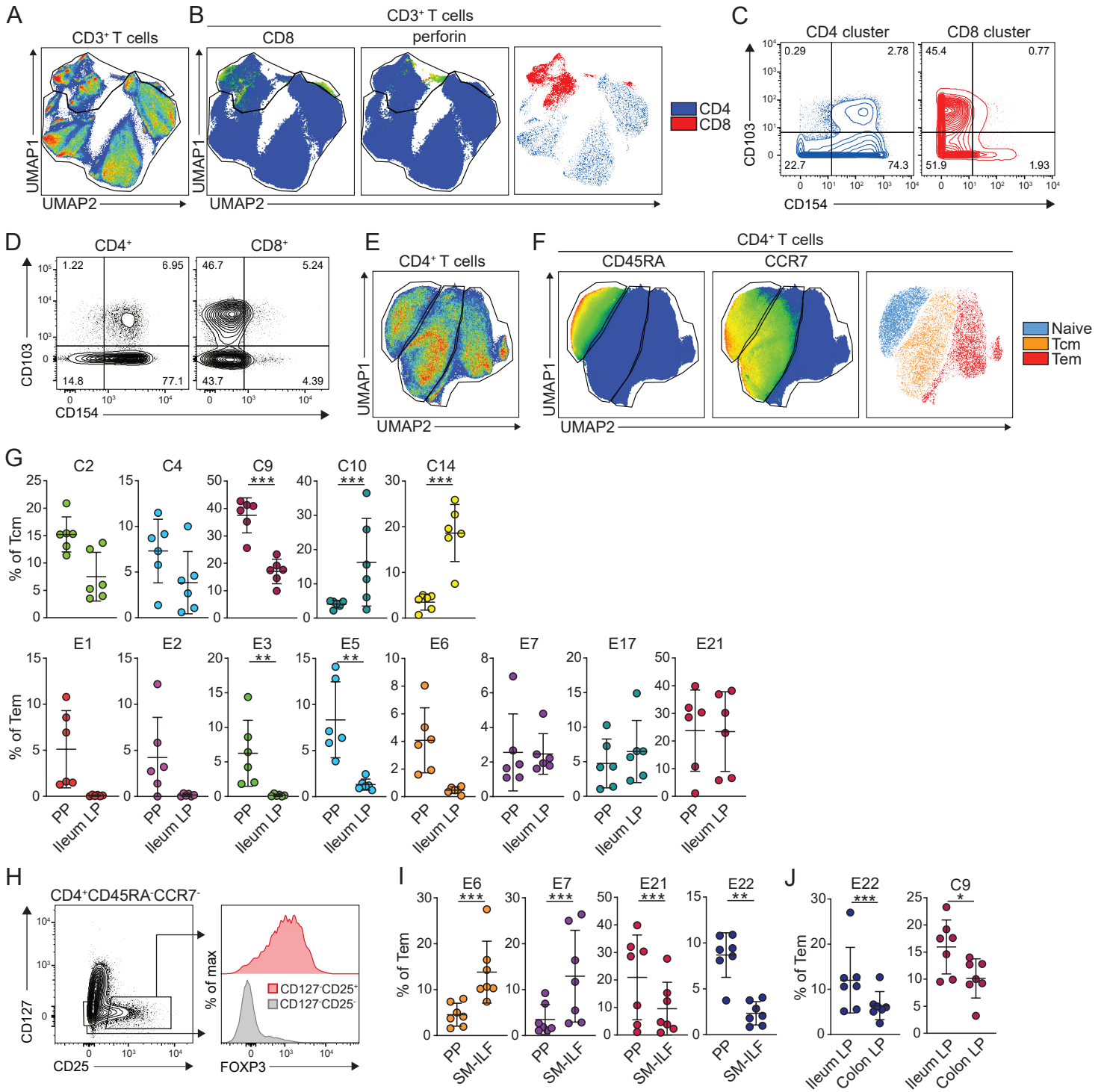


Figure S3. Related to Figure 3.

(A-H) Single-cell suspensions of PP, proximal colon SM-ILF and ileum and colon LP were stimulated with PMA and ionomycin and the T cell populations were analyzed by CyTOF. (A) Pseudocolour density plot of total T cell UMAP used to distinguish CD4⁺ from CD8⁺ T cells. (B) CD8 and perforin expression, and putative CD4 (blue) and CD8 (red) gates overlaid on the total T cell UMAP. (C) CD103 and CD154 expression overlaid on the CD4⁺ (blue) and CD8⁺ (red) T cell clusters defined in (B). (D) CD103 and CD154 expression by CD4⁺ and CD8⁺ T cells in colon LP as assessed by flow cytometry. (E) Pseudocolour density plot of CD4⁺ T cell UMAP (see STAR methods for details) including manual gates demarcating CD4⁺ naïve, Tcm, and Tem. (F) CD45RA and CCR7 expression overlaid on CD4⁺ T cell UMAP plot. (G) Frequency of memory CD4⁺ T cell clusters in PP and ileum LP. Each circle represents a paired sample from 6 individual patients. Bars represent mean \pm 1SD. (H) FOXP3 expression by CD25⁺CD127⁻CD4⁺ Tem in SM-ILF. Results are from 1 representative patient of 3, with at least 10 pooled SM-ILF per patient. (I and J) Frequency of memory CD4⁺ T cell clusters in (I) Peyer's patch and SM-ILF and (J) ileum LP and colon LP. Each circle represents a paired sample from 7 individual patients. Bars represent mean \pm 1SD. PP, Peyer's patch; SM-ILF, submucosal isolated lymphoid follicle; LP, lamina propria. Statistical significance was determined using paired 2-way ANOVA, false discovery rate was corrected with the two-stage step-up method of Benjamini, Krieger and Yekutieli, *p<0.05, **p<0.01, ***p<0.001. See also Figure S3.

Figure S4

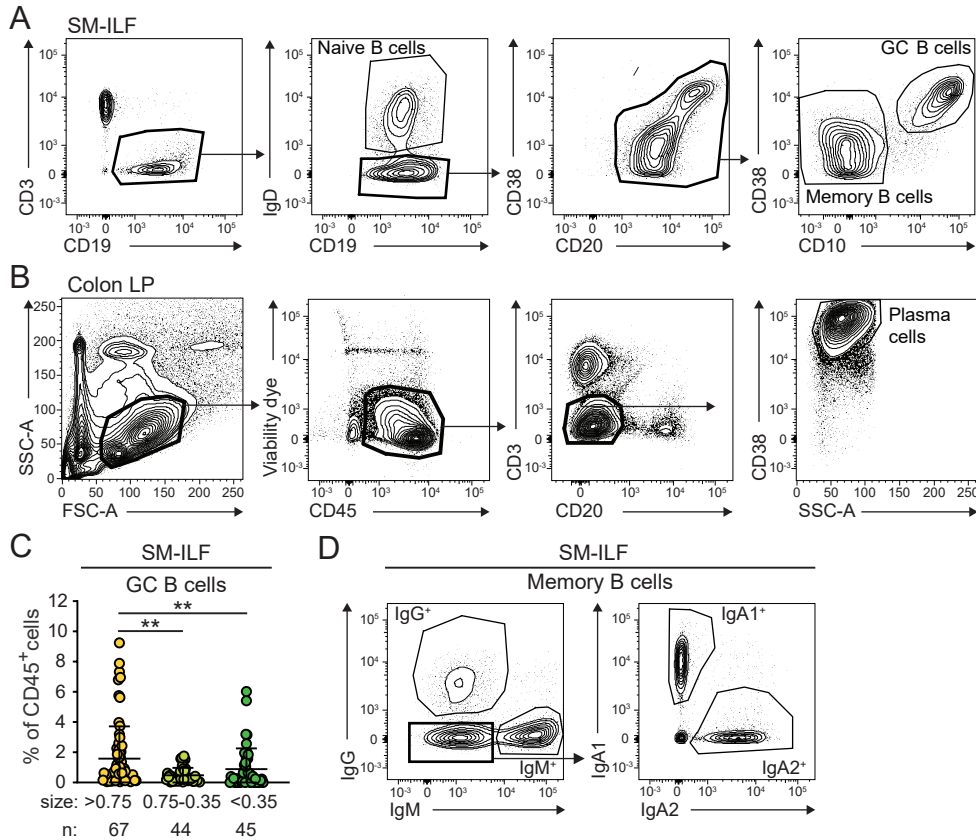


Figure S4. Related to Figure 4.

(A-B) Gating strategy to identify naïve, memory and GC B cells and CD45⁺ PC by flow cytometry in (A) colon SM-ILF and (B) colon LP. (C) Proportion of CD10⁺CD20^{hi} GC B cells amongst total CD45⁺ cells in colon SM-ILF of the indicated size. Each circle represents a single SM-ILF. Samples are from 6 proximal colon and 5 distal colon resections. Bars represent mean \pm 1SD. (D) Gating strategy to identify IgM⁺, IgG⁺, IgA1⁺ and IgA2⁺ memory and GC B cells by flow cytometry. Statistical significance between samples was determined using the Kruskal-Wallis test with Dunn's multiple comparisons, ** $p < 0.01$. GC, germinal center; SM-ILF, submucosal follicle; LP, lamina propria.

Figure S5

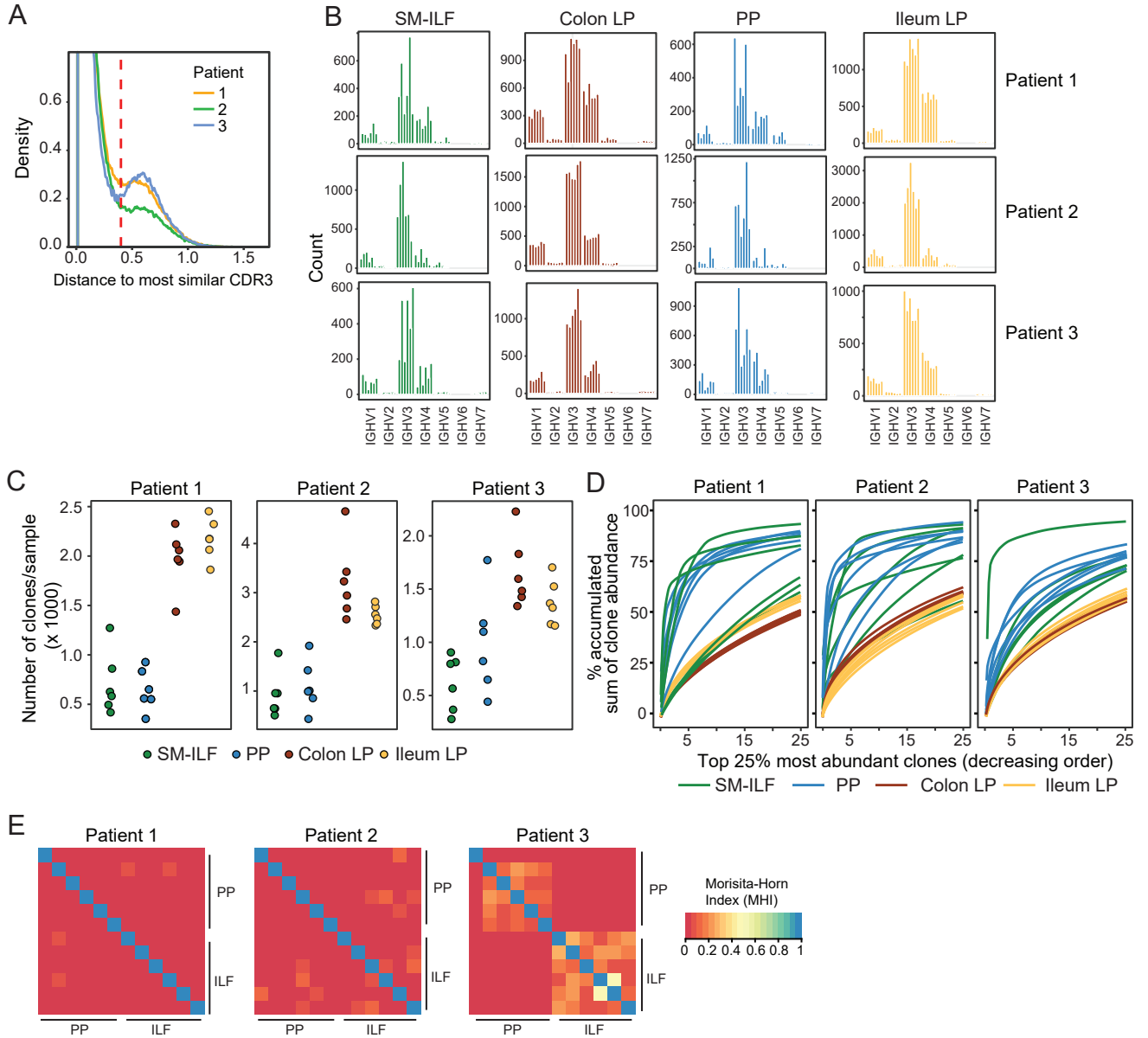


Figure S5. Related to Figure 5.

(A) Distribution of the sequence difference, as given by the SF5 substitution matrix, of each CDR3 to the most similar other CDR3 in the IgA repertoire that have the same V and J-gene combination. (B) Distribution of V gene usage in indicated tissues and patients. (C and D) The IgA repertoire in isolated ileum and proximal colon GALT and LP was determined in 3 individual donors. (C) Total number of clones with ≥ 10 reads in the indicated tissue in each donor and (D) the cumulative abundance of the 25% most abundant clones within each sample. (E) Morisita-Horn similarity index (MHI) between PP and ILF (SM-ILF) samples taking into account only the top 100 clones in each compartment. PP, Peyer's patch; SM-ILF, submucosal follicle; LP, lamina propria.

Table S1

	Total	GALT count	B cells	T cells	CytoTOF	IgA	FOXP3	B cell Migration	IHC
Number of patients	99	68	24	12	11	3	3	4	25
Age	70.7 ± 9.6	70.7 ± 9.4	72.8 ± 7.3	67.5 ± 11.7	69.8 ± 13.1	73 ± 13.2	70.7 ± 2.1	64.8 ± 5.7	77.7 ± 4.7
Sex									
Female	51	37	9	6	4	2	1	1	20
Male	48	31	15	6	7	1	2	3	5
Site									
Terminal ileum	54	14	18 ^c	6 ^c	8 ^c	3 ^c	-	4 ^c	15
Proximal colon	73 ^a	49 ^a	18 ^c	6 ^c	8 ^c	3 ^c	1	4 ^c	14
Sigmoid colon/rectum	26 ^b	20 ^b	6	6	3	-	2	-	4
Proximal colon site									
Cecum	16	13	5	4	2	1	-	-	4
Ascending	25	19	7	1	2	-	-	2	6
Transverse	36	20	6	1	2	2	1	2	4
Combined proximal	2	-	-	-	2	-	-	-	-
Diagnosis									
Volvulus	4	2	1	1	2	-	-	-	-
Diverticulitis	3	2	1	2	-	-	-	-	-
Stenosis	2	1	-	-	-	-	1	-	-
Adenocarcinoma	90	63	22	9	9	3	2	4	25
UICC stage									
0	15	11	4	-	1	1	-	-	5
1	14	12	3	1	1	-	-	1	2
2	33	20	7	5	6	2	-	1	9
3	22	16	8	1	-	-	1	2	7
4	6	4	-	2	1	-	1	-	2

^a Samples taken from multiple proximal sites per patient

^b One patient had samples taken from both sigmoid colon and rectum

^c Samples taken from both ileum and colon per patient allowing paired data analysis

Table S1. Related to experimental model and subject details section of STAR methods

Table S2

Gene name	Primers name	Forward	Reverse
Actin γ 1	ACTG1	CCGAGCCGTGTTTCCTCC	GCCATGCTCAATGGGGTACT
Ribosomal protein S27	RPS27	ATGCCTCTCGCAAAGGATCTC	TGAAGTAGGAATTGGGGCTCT
Activation Induced Cytidine Deaminase	AICDA.1	GAGGCGTGACAGTGCTACATC	CAGGGTCTAGGTCCCAGTCC
Activation Induced Cytidine Deaminase	AICDA.2	AAGGGCTGCATGAAAATTCAGT	AAGGGCTGCATGAAAATTCAGT
Syndecan-1 (CD138)	SDC1.1	CTGCCGCAAATTGTGGCTAC	TGAGCCGGAGAAGTTGTCAGA
Syndecan-1 (CD138)	SDC1.2	TTGTGGCTACTAATTTGCC	TATCTTGCAAAGCACCTGC

Table S2. Related to Figure 4.