

## Supplementary Information

**Jasmonic acid is not a biosynthetic intermediate to produce the pyrethrolone moiety in pyrethrin II.**

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### List of Supplementary Information

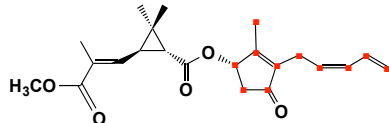
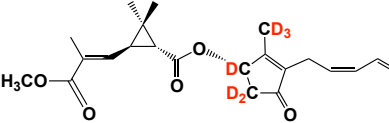
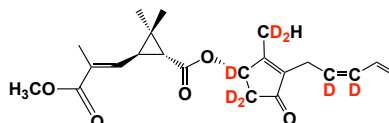
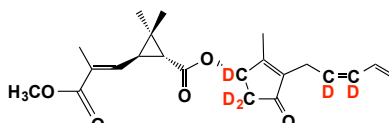
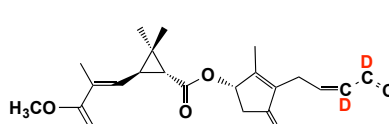
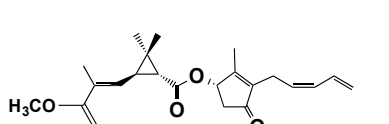
- Table S1.** UPLC MS/MS parameters for measurements of pyrethrins I and II, jasmolins I and II, and cinerins I and II in positive ionization mode.
- Table S2.** UPLC MS/MS parameters for measurements of pyrethrin II and its labeled compounds in positive ionization mode.
- Table S3.** UPLC-MS/MS parameters for measurements of JA, 12-OH-JA and their labeled compounds in negative ionization mode.
- Table S4.** UPLC MS/MS parameters for measurements of JA-L-Ile and their labeled compounds in negative ionization mode.
- Figure S1.** Proposed biosynthetic pathway of pyrethrin II.
- Figure S2.** Reported labeled patterns of *cis*-jasmane derived from [<sup>2</sup>H<sub>1</sub>-7,<sup>2</sup>H<sub>2</sub>-5, <sup>2</sup>H<sub>2</sub>-2] JA (a), and [<sup>2</sup>H<sub>2</sub>-11, <sup>2</sup>H<sub>2</sub>-10, <sup>2</sup>H<sub>2</sub>-8, <sup>2</sup>H<sub>2</sub>-2] methyl *iso*-OPDA (b).
- Figure S3.** UPLC MS/MS chromatograms for analyzing authentic pyrethrin II (a), jasmolin I (b), and pyrethrin I (c) in positive ionization mode.
- Figure S4.** UPLC MS/MS chromatograms for analyzing whole plant of pyrethrum (*T. cinerariifolium*) in positive ionization modes.
- Figure S5.** UPLC MS/MS chromatograms for analyzing authentic JA-L-Ile (a), JA (b), and 12-OH-JA (c) in negative ionization modes.
- Figure S6.** Feeding experiment using airborne [<sup>2</sup>H<sub>3</sub>-12,<sup>2</sup>H<sub>2</sub>-11,<sup>2</sup>H<sub>1</sub>-10] (±)-MeJA.
- Figure S7.** Feeding experiment using [<sup>2</sup>H<sub>3</sub>-16,<sup>2</sup>H<sub>2</sub>-15,<sup>2</sup>H<sub>1</sub>-14] (±)-OPC 6:0.
- Figure S8.** Expected patterns of deuterium labeled pyrethrin II derived from [<sup>2</sup>H<sub>1</sub>-16,<sup>2</sup>H<sub>1</sub>-15,<sup>2</sup>H<sub>2</sub>-11, <sup>2</sup>H<sub>2</sub>-10, <sup>2</sup>H<sub>2</sub>-8, <sup>2</sup>H<sub>2</sub>-2] methyl *iso*-OPDA.
- Figure S9.** Feeding experiment using deuterium-labeled methyl *iso*-OPDA.
- Figure S10.** Feeding experiment [U-<sup>13</sup>C] (+)-OPDA.
- Figure S11.** Proposed conversion pathway to afford deuterium labeled pyrethrin II from deuterium labeled methyl *iso*-OPDA.

**Supporting information Table S1:** UPLC MS/MS parameters for measurements of pyrethrins I and II, jasmolins I and II, and cinerins I and II in positive ionization mode.

Target compound	Parent ion ( <i>m/z</i> )	Transit ion ( <i>m/z</i> )	Cone Voltage (V)	Collision Energy (eV)
pyrethrin I	330.76	161.54	10	15
pyrethrin II	373.50	161.01	15	12
jasmolin I	332.76	163.54	10	15
jasmolin II a)	375.50	163.01	15	12
cinerin I a)	317.76	149.54	10	15
cinerin II a)	361.50	149.01	15	12

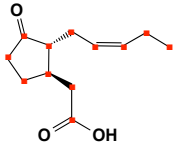
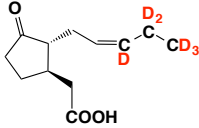
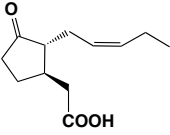
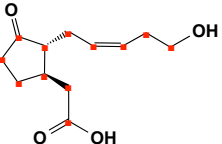
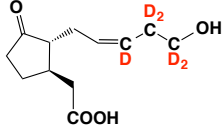
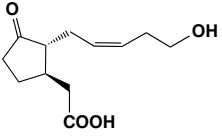
a) parameters were set based on those of authentic pyrethrins I and II, and jasmolin I.

**Supporting information Table S2:** UPLC MS/MS parameters for measurements of pyrethrin II and its labeled compounds in positive ionization mode.

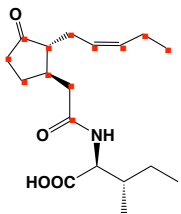
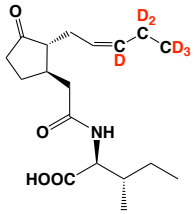
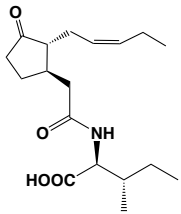
Target compound	Pseudo molecular ion ( $m/z$ )	Transit ion ( $m/z$ )	Cone Voltage (V)	Collision Energy (eV)
	384.50	172.01	15	12
	379.50	167.01	15	12
	380.50	168.01	15	12
	378.50	166.01	15	12
	377.50	165.01	15	12
	373.50	161.01	15	12

■ : indicating  $^{13}\text{C}$

**Supporting information Table S3:** UPLC-MS/MS parameters for measurements of JA, 12-OH-JA and their labeled compounds in negative ionization mode.

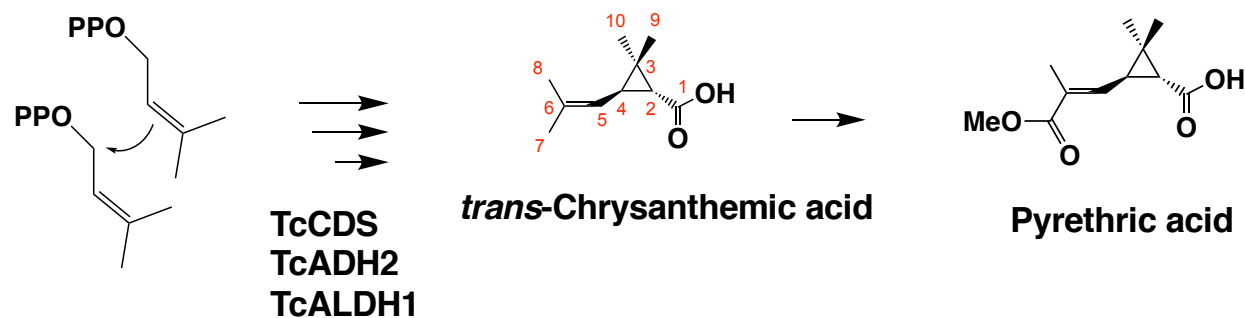
Target compound	Pseudo molecular ion ( $m/z$ )	Transit ion ( $m/z$ )	Cone Voltage (V)	Collision Energy (eV)
	221.00	60.71	24	16
	215.00	58.71	24	16
	209.00	58.71	24	16
	236.97	60.70	32	28
	229.97	58.70	32	28
	224.97	58.70	32	28

**Supporting information Table S4:** UPLC MS/MS parameters for measurements of JA-L-Ile and their labeled compounds in negative ionization mode.

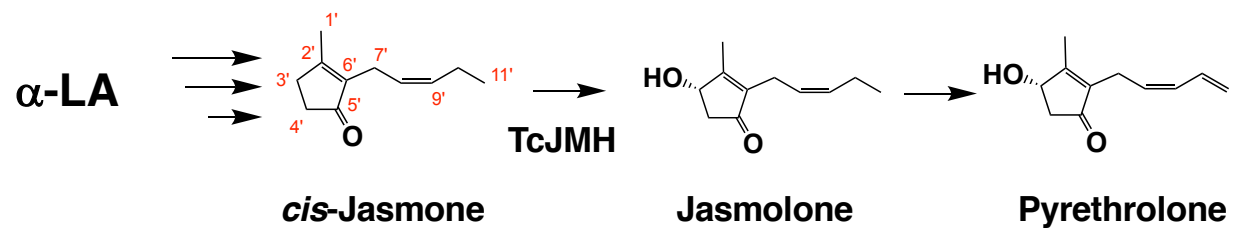
Target compound	Pseudo molecular ion ( <i>m/z</i> )	Transit ion ( <i>m/z</i> )	Cone Voltage (V)	Collision Energy (eV)
	334.03	129.68	45	24
	328.03	129.68	45	24
	322.03	129.68	45	24

■ : indicating <sup>13</sup>C

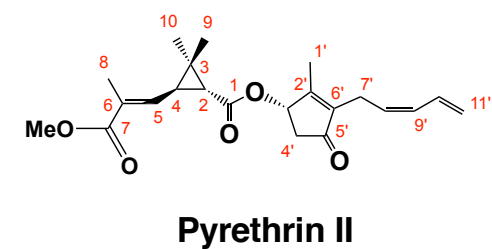
**acid moiety**



**alcohol moiety**



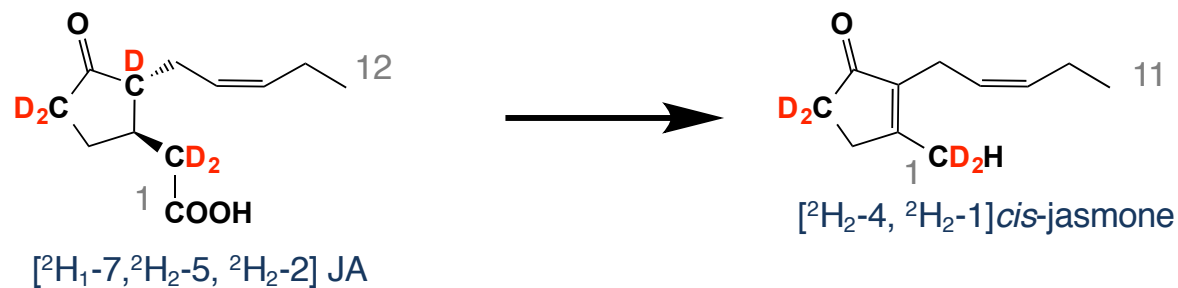
TcGLIP



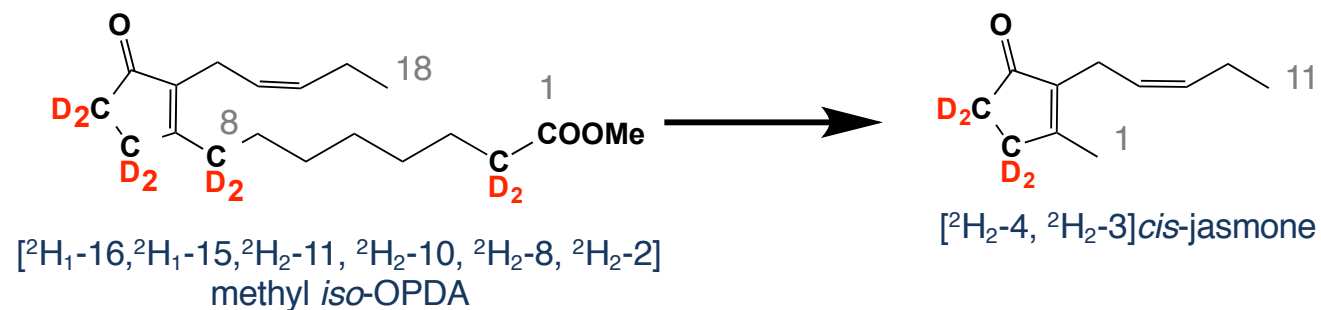
**Supporting information Figure S1:** Proposed biosynthetic pathway of pyrethrin II.

$\alpha$ -LA:  $\alpha$ -linolenic acid, CDS: chrysanthemyl diphosphate synthase, ADH: alcohol dehydrogenase, ALDH: aldehyde dehydrogenases, JMH: jasmone hydroxylase.

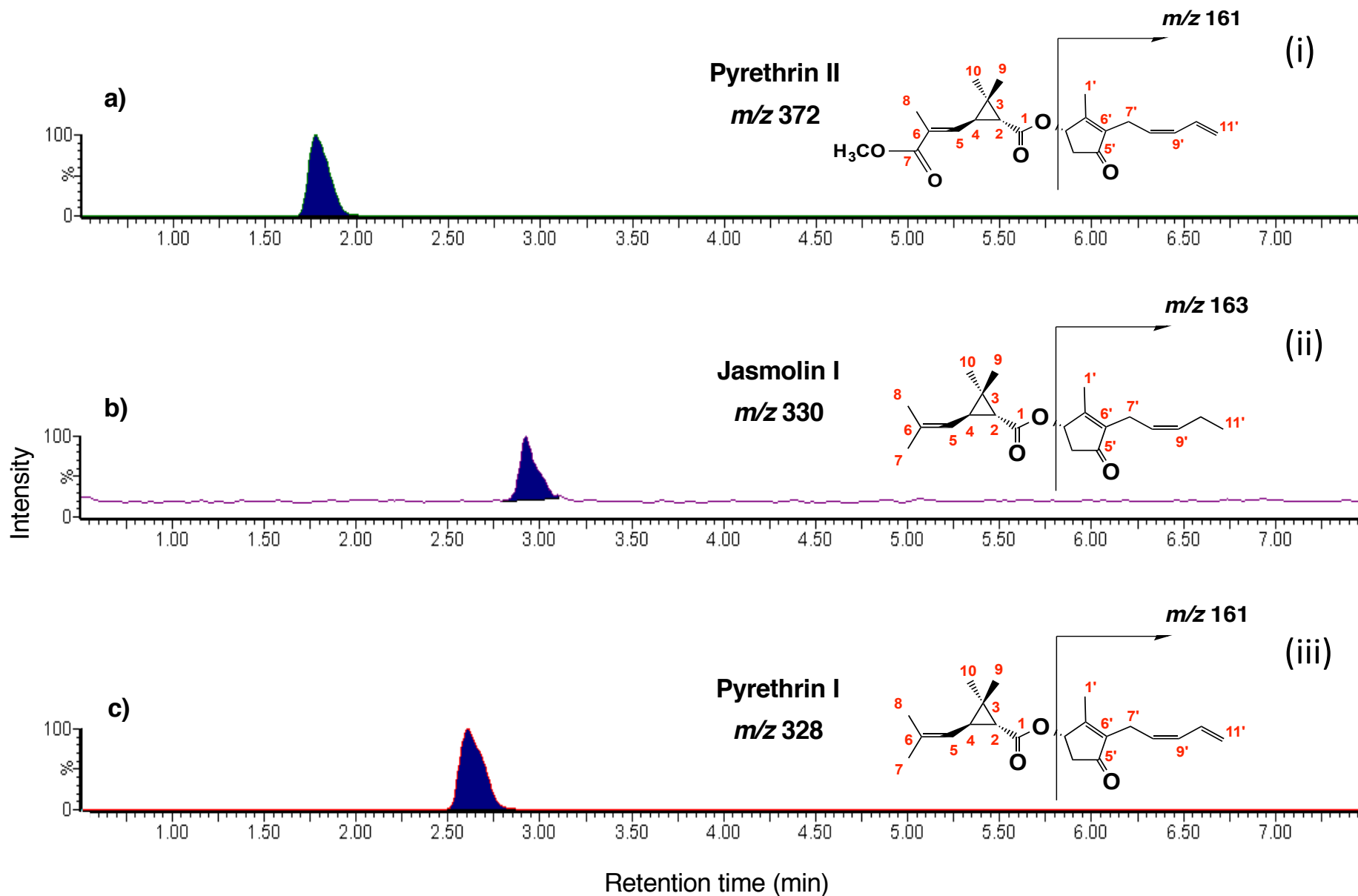
a) Koch et. al. (1997) using *Jasrinurn rincospernum* (1).



b) Matusi et. al. (2017, 2019) using *Lsiodiplodia theobromae* (2).

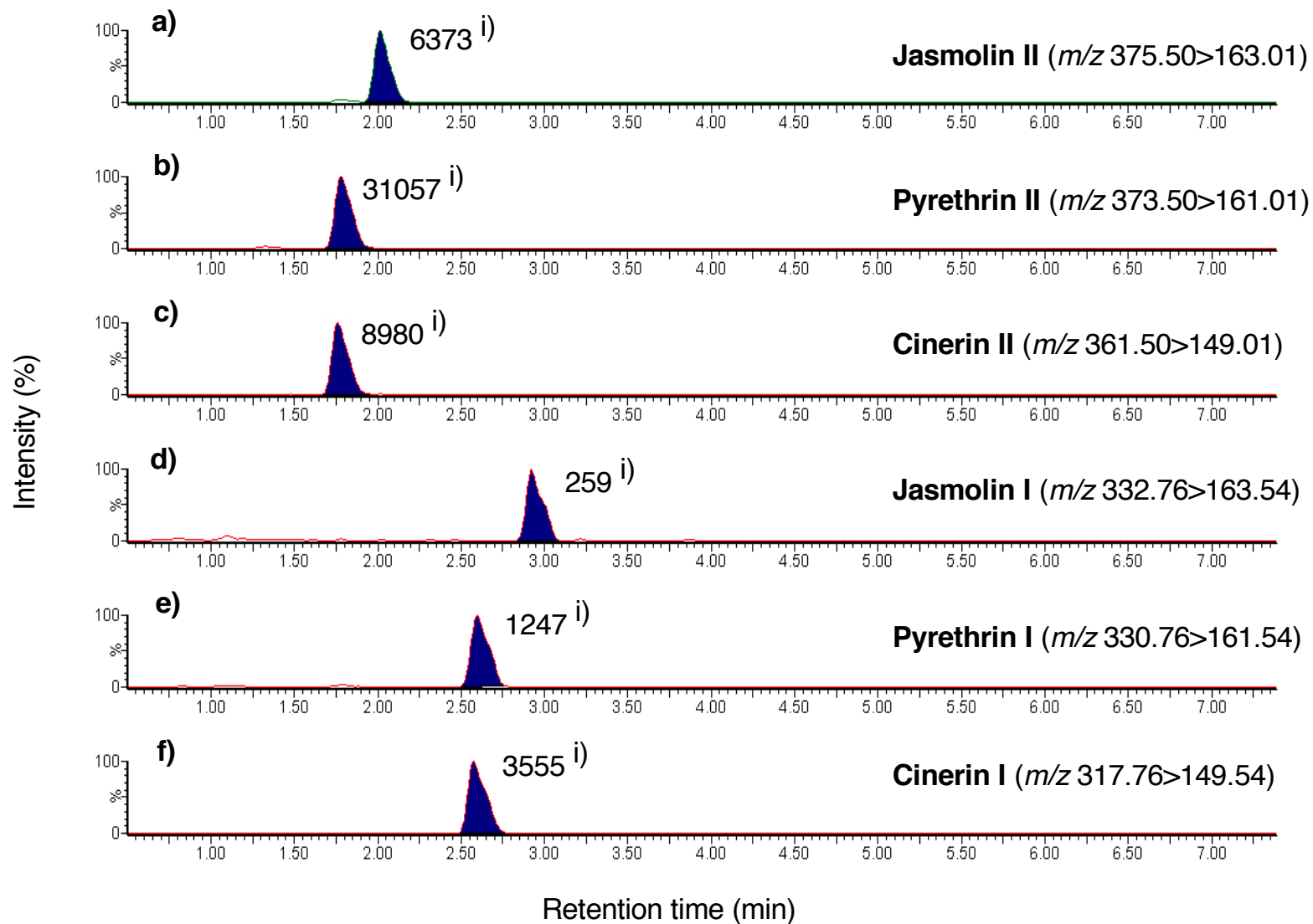


**Supporting information Figure S2:** Reported labeled patterns of *cis*-jasmone derived from  $[^2\text{H}_1-7, ^2\text{H}_2-5, ^2\text{H}_2-2]$  JA (a), and  $[^2\text{H}_2-11, ^2\text{H}_2-10, ^2\text{H}_2-8, ^2\text{H}_2-2]$  methyl *iso*-OPDA (b).



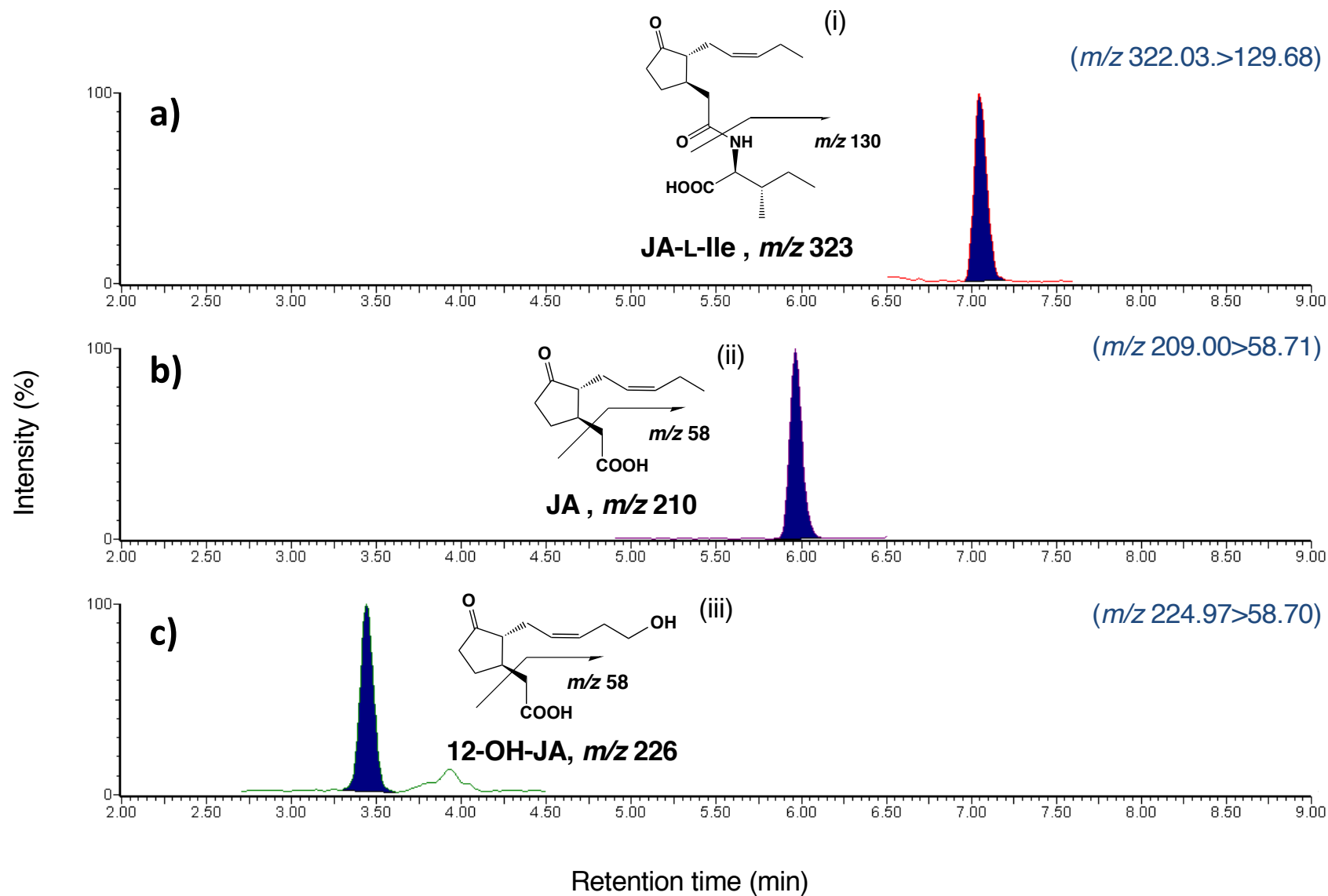
**Supporting information Figure S3:** UPLC MS/MS chromatograms for analyzing authentic pyrethrin II (a), jasmolin I (b), and pyrethrin I (c) in positive ionization mode. Insets indicate the fragment patterns of pyrethrin II (i), jasmolin I (ii), and pyrethrin I (iii).



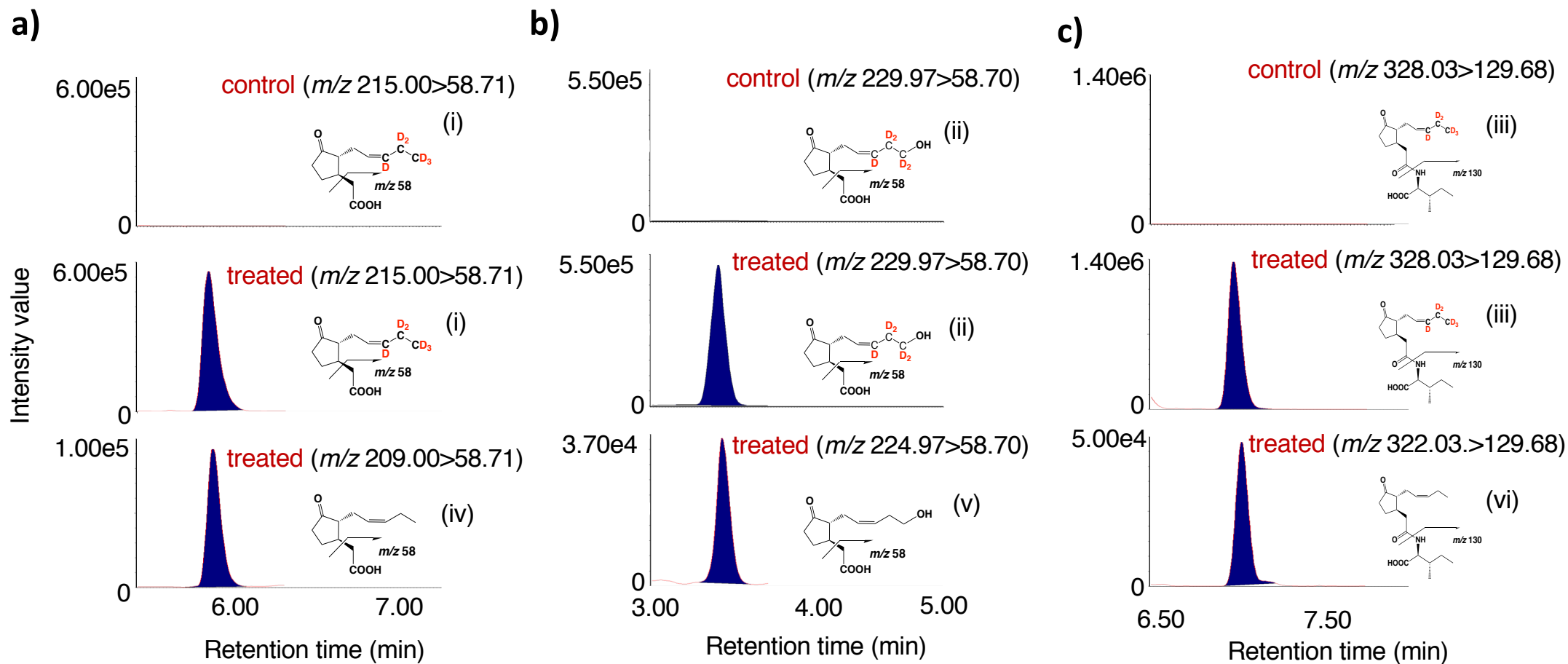


**Supporting information Figure S4:** UPLC MS/MS chromatograms for analyzing whole plant of pyrethrum (*T. cinerariifolium*) in positive ionization modes. Analyzing jasmolin II (a) , pyrethrin II (b), cinerin II (c), jasmolin I (d) , pyrethrin I (e), cinerin I (f).

i): indicating vale of peak area



**Supporting information Figure S5:** UPLC MS/MS chromatograms for analyzing authentic JA-L-Ile (a), JA (b), and 12-OH-JA (c) in negative ionization modes. Insets indicate the fragment patterns of JA-L-Ile (i), JA (ii), and 12-OH-JA (iii).

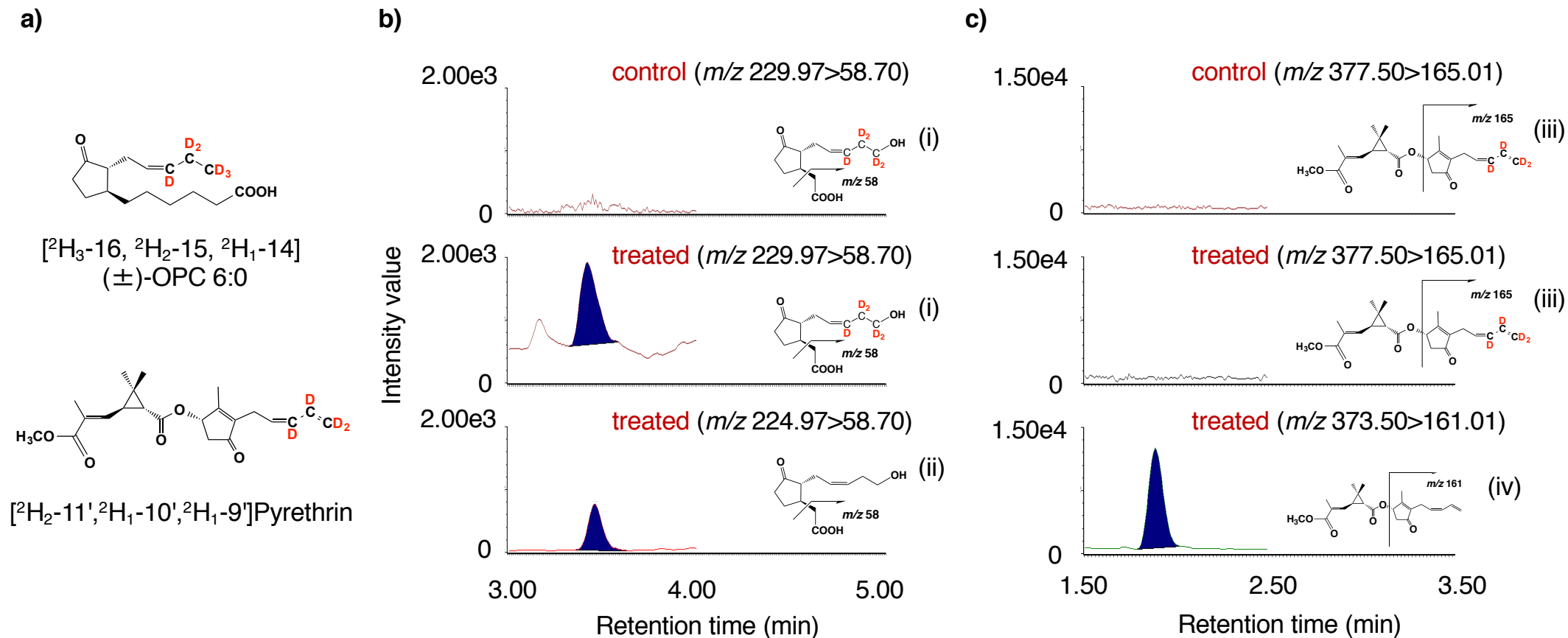


**Supporting information Figure S6:** Feeding experiment using airborne [ $^2\text{H}_3$ -12, $^2\text{H}_2$ -11, $^2\text{H}_1$ -10] ( $\pm$ )-MeJA.

UPLC MS/MS chromatograms of the leaf extract of untreated (control) and treated plants.

Upper and middle panels: JA (a), 12-OH-JA (b), JA-L-Ile (c) having the expected labeled pattern in the control and treated plants, respectively. Lower panel: endogenous target compounds in the treated plants.

Insets indicate the fragment patterns of deuterium labeled JA (i), 12-OH-JA (ii), JA-L-Ile (iii) and endogenous JA (iv), 12-OH-JA (v), JA-L-Ile (vi)



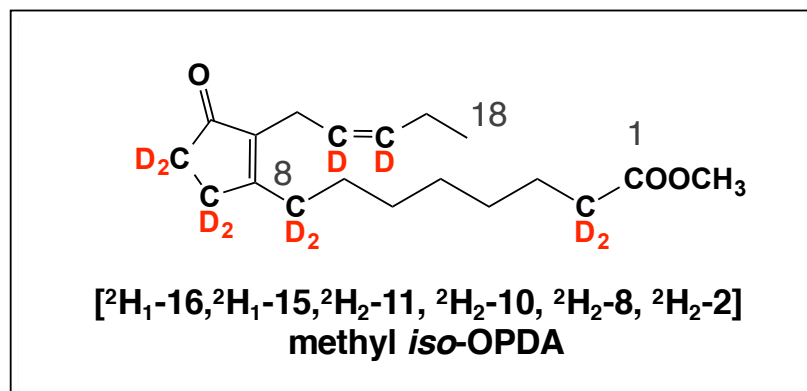
**Supporting information Figure S7:** Feeding experiment using  $[^2\text{H}_3\text{-16}, ^2\text{H}_2\text{-15}, ^2\text{H}_1\text{-14}]$  ( $\pm$ )-OPC 6:0.

a) Chemical structures of  $[^2\text{H}_3\text{-16}, ^2\text{H}_2\text{-15}, ^2\text{H}_1\text{-14}]$  ( $\pm$ )-OPC 6:0 and pyrethrin II having expected labeled pattern.

b), c) UPLC MS/MS chromatographs analyzing leaf extract of non-treated (control) and treated plants.

Upper and middle panels: analyzing 12-hydroxyJA (b) and pyrethrin II (c) having expected labeled pattern for control and treated plants, respectively. Lower panel: analyzing endogenous 12-hydroxyJA (b) and pyrethrin II (c) for treated plants.

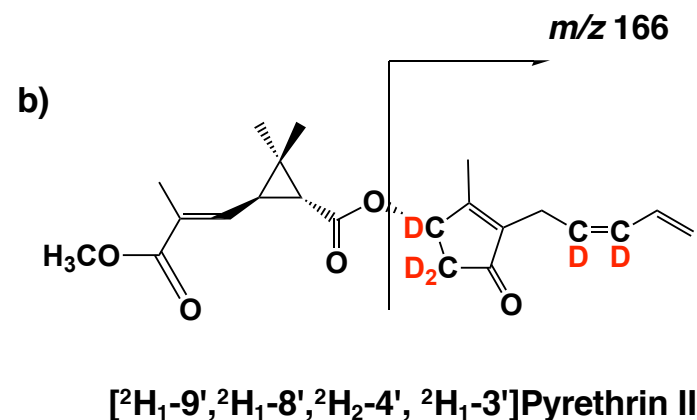
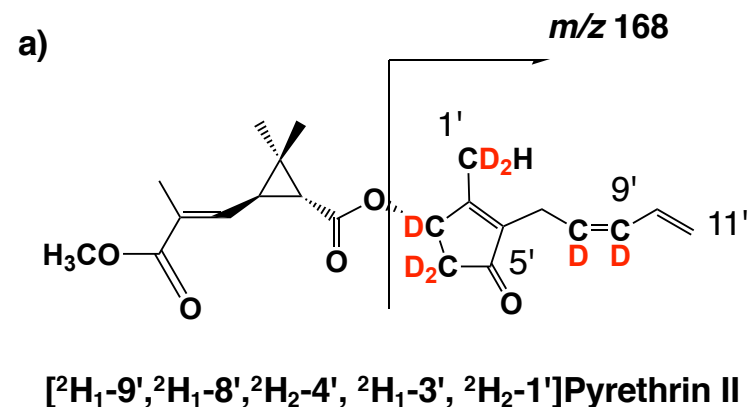
Insets indicate the fragment patterns of deuterium labeled 12-hydroxyJA (i), endogenous 12-hydroxyJA (ii), deuterium labeled pyrethrin II (iii), and endogenous pyrethrin II (iv).



Koch et al. (1)

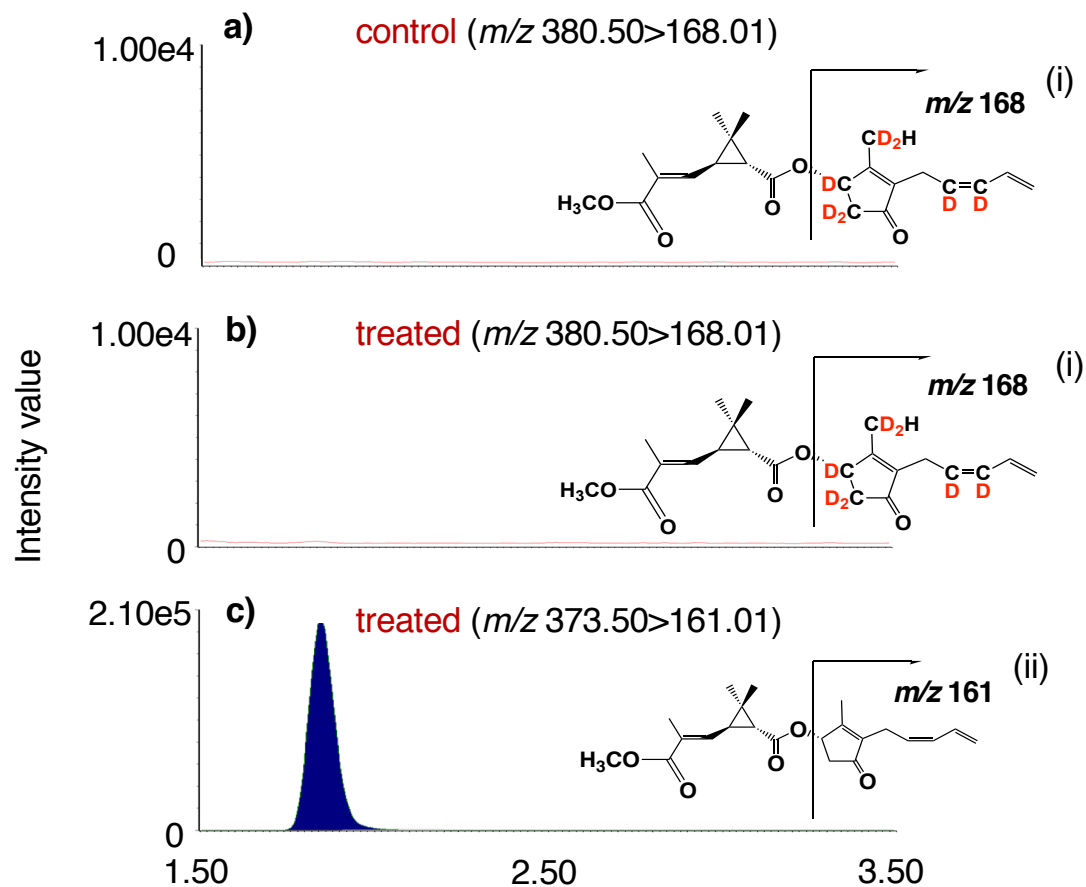


Matsui et al. (1, 2)



**Supporting information Figure S8:** Expected patterns of deuterium labeled pyrethrin II derived from [<sup>2</sup>H<sub>1</sub>-16, <sup>2</sup>H<sub>1</sub>-15, <sup>2</sup>H<sub>2</sub>-11, <sup>2</sup>H<sub>2</sub>-10, <sup>2</sup>H<sub>2</sub>-8, <sup>2</sup>H<sub>2</sub>-2] methyl *iso*-OPDA.

The plants were given a lanolin paste having [<sup>2</sup>H<sub>1</sub>-16, <sup>2</sup>H<sub>1</sub>-15, <sup>2</sup>H<sub>2</sub>-11, <sup>2</sup>H<sub>2</sub>-10, <sup>2</sup>H<sub>2</sub>-8, <sup>2</sup>H<sub>2</sub>-2] methyl *iso*-OPDA, and two kinds of labeled patterns were expected. One (a) is based on the report of Koch et al. (1) and the other (b) is Matsui et al. (2, 3).



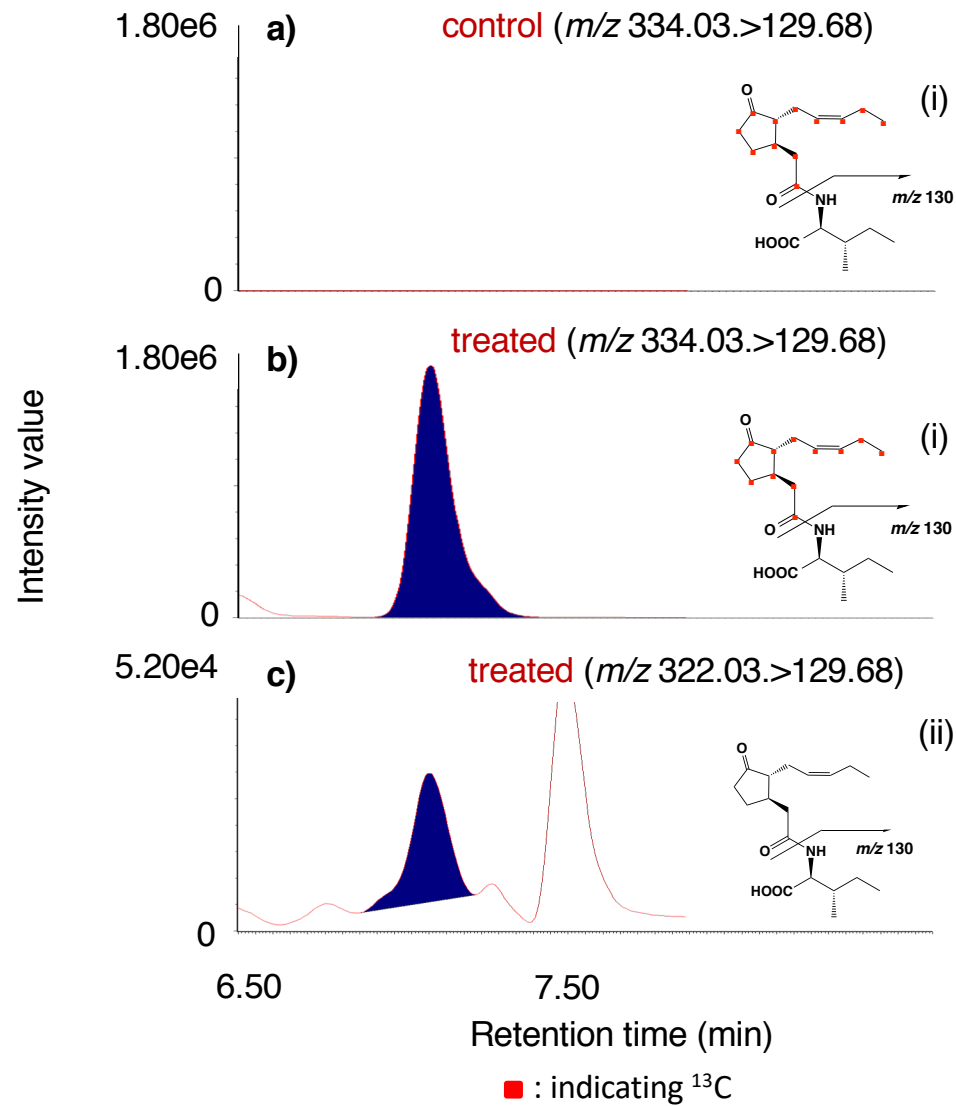
**Supporting information Figure S9:** Feeding experiment using deuterium-labeled methyl *iso*-OPDA.

a) UPLC MS/MS chromatograph analyzing leaf extract of non-treated plants (control) for [ $^2H_1$ -9', $^2H_1$ -8', $^2H_2$ -4',  $^2H_1$ -3',  $^2H_2$ -1']pyrethrin II.

b) analyzing leaf extract of treated plant with the labeled compound for [ $^2H_1$ -9', $^2H_1$ -8', $^2H_2$ -4',  $^2H_1$ -3',  $^2H_2$ -1']pyrethrin II.

c) analyzing leaf extract of treated plants with the labeled compound for endogenous pyrethrin II.

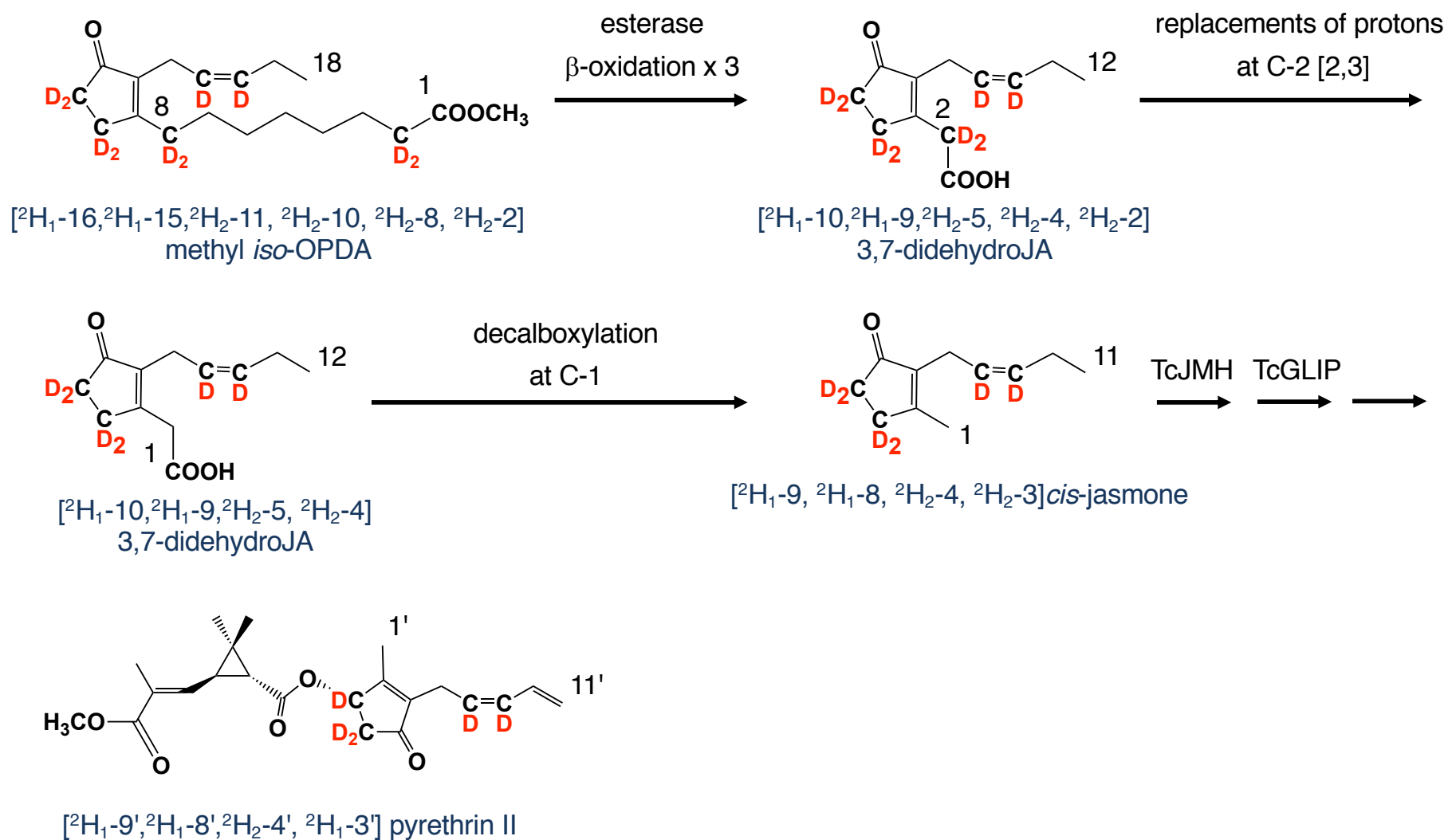
Insets indicate the fragment patterns of [ $^2H_1$ -9', $^2H_1$ -8', $^2H_2$ -4',  $^2H_1$ -3',  $^2H_2$ -1'] pyrethrin II (i) and endogenous pyrethrin II (ii).



**Supporting information Figure S10:** Feeding experiment [ $\text{U-}^{13}\text{C}$ ] (+)-OPDA.

- a) UPLC MS/MS chromatographs analyzing leaf extract of non-treated (control) plant set for [ $\text{U-}^{13}\text{C}$ ]JA-L-Ile.  
 b) analyzing leaf extract of treated plant set for [ $\text{U-}^{13}\text{C}$ ]JA-L-Ile.  
 c) analyzing leaf extract of treated plants set for JA-L-Ile.

Insets indicate the fragment patterns of [ $\text{U-}^{13}\text{C}$ ]JA-L-Ile (i) and endogenous JA-L-Ile (ii).



**Supporting information Figure S11:** Proposed conversion pathway to afford deuterium labeled pyrethrin II from deuterium labeled methyl *iso*-OPDA.



## References

1. Koch, T., Bandemer, K., and Boland, W. (1997) Biosynthesis of cis-jasmone: A pathway for the inactivation and the disposal of the plant stress hormone jasmonic acid to the gas phase? *Helvetica Chimica Acta* 80, 838-850
2. Matsui, R., Amano, N., Takahashi, K., Taguchi, Y., Saburi, W., Mori, H., Kondo, N., Matsuda, K., and Matsuura, H. (2017) Elucidation of the biosynthetic pathway of cis-jasmone in *Lasiodiplodia theobromae*. *Sci Rep* 7
3. Matsui, R., Takiguchi, K., Matsuda, K., Takahashi, K., and Matsuura, H. (2019) Feeding experiment using uniformly (13)C-labeled alpha-linolenic acid supports the involvement of the decarboxylation mechanism to produce cis-jasmone in *Lasiodiplodia theobromae*. *Biosci Biotechnol Biochem* 83, 2190-2193