

Supplementary Material

Teresa L. Capasso¹, Bijun Li¹, Harry J. Volek², Waqas Khalid², Elizabeth R. Rochon^{1,3}, Arulselvi Anbalagan¹, Chelsea Herdman⁴, H. Joseph Yost⁴, Flordeliza S. Villanueva^{3,5}, Kang Kim^{2,3,5}, Beth L. Roman^{1,3}

¹Department of Human Genetics, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, PA 15261

²Department of Bioengineering, Swanson School of Engineering, University of Pittsburgh, Pittsburgh, PA 15260

³Heart, Lung, Blood and Vascular Medicine Institute, University of Pittsburgh, Pittsburgh, PA 15261

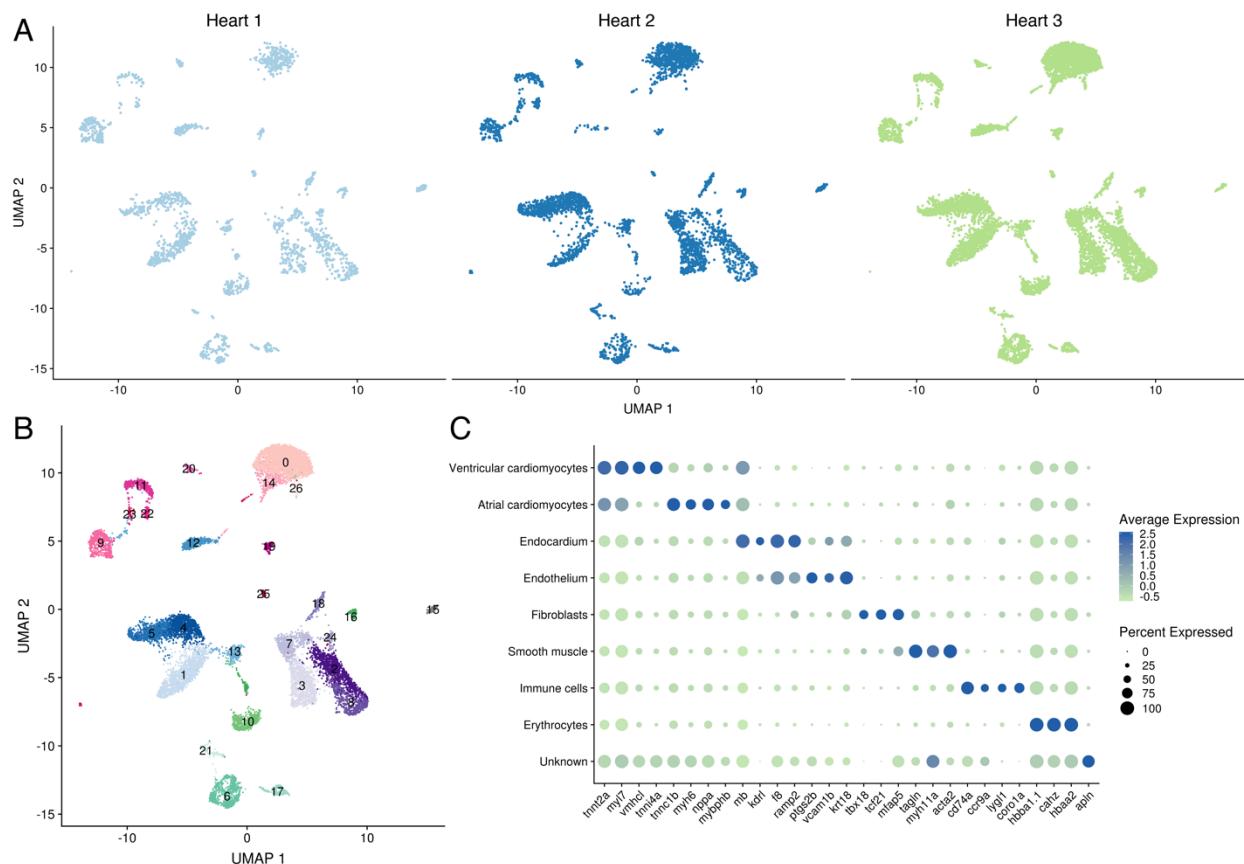
⁴Molecular Medicine Program, University of Utah, Salt Lake City, UT 84112

⁵Center for Ultrasound Molecular Imaging and Therapeutics, Division of Cardiology, Department of Medicine, School of Medicine, University of Pittsburgh, Pittsburgh, PA 15261

Corresponding author: Beth L. Roman, romanb@pitt.edu, 412.624.7006.

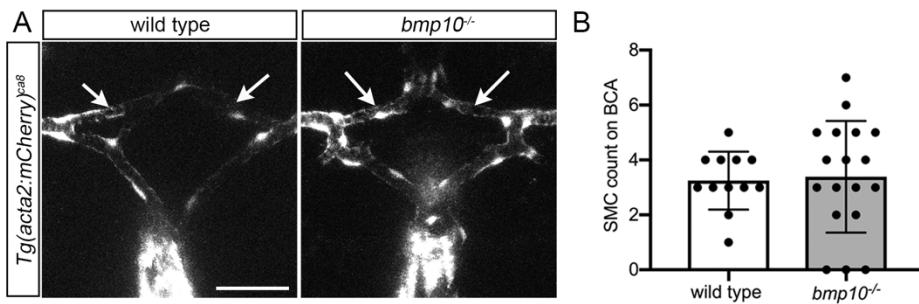
ORCID 0000-0002-1250-1705

Supplementary Material



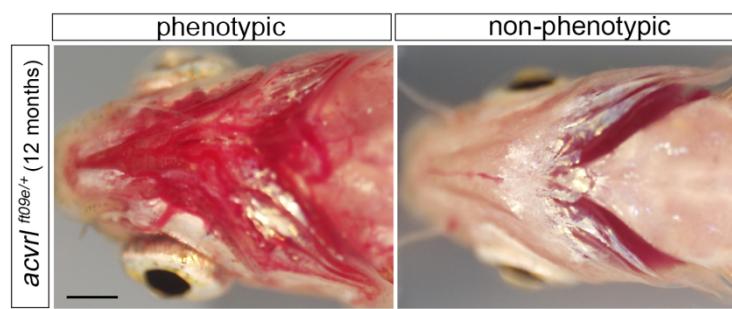
Supplementary Fig. 1 **a** UMAP plots of individual hearts (Heart 1 = 2086 cells, Heart 2 = 3924 cells, Heart 3 = 8462 cells). **b** UMAP plot of the three integrated heart datasets. Cluster numbers correspond to the original cluster IDs from the `FindClusters` function (`res 0.8`) and colors correspond to cell type as annotated in Supplementary Dataset 1. Cluster identities are as follows: 0, erythrocytes; 1, atrial cardiomyocytes; 2, endothelium; 3, endocardium; 4, ventricular cardiomyocytes; 5, ventricular cardiomyocytes; 6, fibroblasts; 7, endocardium; 8, endothelium; 9, immune cells; 10, smooth muscle; 11, immune cells; 12, atrial cardiomyocytes; 13, atrial cardiomyocytes; 14, erythrocytes; 15, unknown; 16, smooth muscle; 17, fibroblasts; 18, endothelium; 19, immune cells; 20, immune cells; 21, fibroblasts; 22, immune cells; 23, immune cells; 24, endothelium; 25, immune cells; 26, erythrocytes. **c** Dotplot demonstrating the scaled expression values of select genes used to characterize the clusters. The size of each dot represents the percentage of cells within the cluster that express the gene.

Supplementary Material



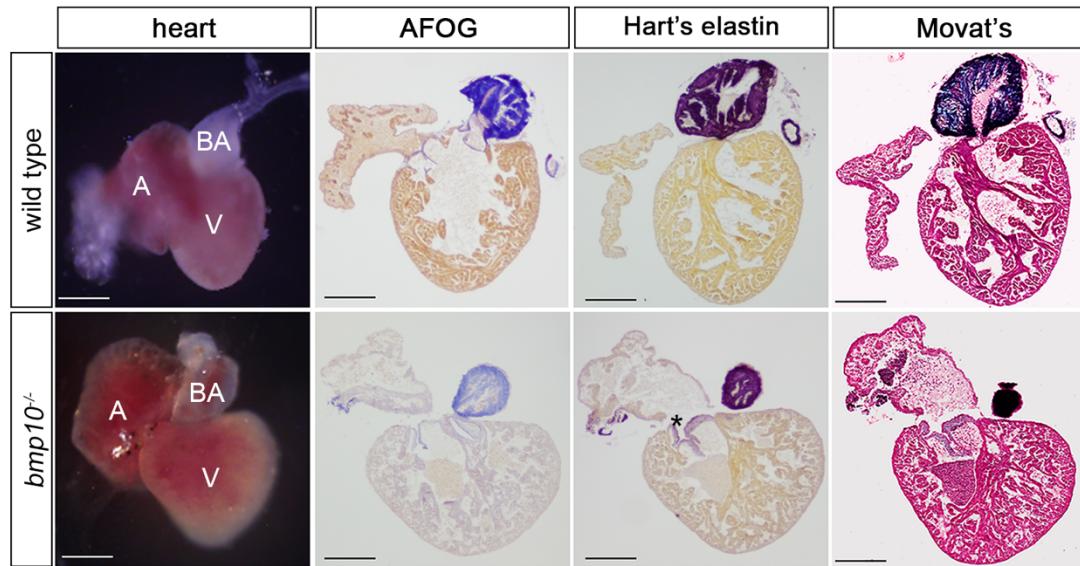
Supplementary Fig. 2 *bmp10* mutant embryos have normal smooth muscle cell coverage on the basal communicating artery (BCA) at 5 dpf. **a** Cranial vasculature in *bmp10^{pt527};Tg(acta2:mcherry)^{ca8}* mutant and wild type siblings at 5 dpf. Arrows, BCA. 2D confocal projections, dorsal views, anterior top. Scale bar: 50 μ m. **b** Quantification of smooth cell number on BCA. N= 12 wild types, 18 mutants over 3 experiments. Bars shown mean \pm SD. Not significant by unpaired Student's *t* test.

Supplementary Material



Supplementary Fig. 3 *acvr1*^{flo9e/+} fish phenocopy *bmp10* mutants with low penetrance. 12-month fish. Scale bar: 1 mm.

Supplementary Material



Supplementary Fig. 4 *bmp10* mutants have abnormally shaped hearts at 3 months. Hearts from 3-month wild type and *bmp10^{pt527}* mutant siblings, whole mount or sections stained with acid fuchsin orange G (AFOG) for collagen (blue) and fibrin (red); Hart's elastin (purple); or Movat's pentachrome for muscle (red), collagen (yellow), elastin (blue-black), and ground substance (light blue). A, atrium; V, ventricle; BA, bulbus arteriosus; asterisks, valves. Scale bars, 200 μ m. Images representative of N = 6 hearts per genotype from 3 independent lines.

Supplementary Material

Supplementary Table 1: TALENs and gRNAs

Gene	TALEN (Addgene #)	Target sequence (5'-3')
<i>bmp9/gdf2</i>	TAL3010 (36002) TAL3011 (36003)	TTGAACAAGGTGGAGAGTttcttaggcttatgaAGGAAGATTTTGAGGA
<i>bmp10</i>	TAL3032 (41206) TAL 3033 (41207)	TCAGCTCCCCGGAGAGGCaccgcactgctccagggTTGGATGATGGACATGGA
Gene	gRNA name	Target sequence
<i>bmp10-</i> <i>like</i>	gRNA <i>bmp10l.3</i>	AGTGGAGGACTGCAGAATAG

Supplementary table 2: Genotyping assays

Gene	Allele	Forward primer	Reverse primer	Enzyme	Result
<i>bmp9</i>	pt533	CACTTAAGGAACCC CGATTTC	ACTCACCCCTGAAC GACAAAGC	Ddel	WT 80+95+255 bp M 95+327 bp
<i>bmp9</i>	pt536	CACTTAAGGAACCC CGATTTC	ACTCACCCCTGAAC GACAAAGC	Ddel	WT 80+95+255 bp M 95+328 bp
<i>bmp10</i>	pt527	CAAAGTAGCCCCAT CAGCTC	CTTCAGGGTCTCC ATCAAGC	NA	WT 138 bp M 130 bp*
<i>bmp10</i>	pt543	CAAAGTAGCCCCAT CAGCTC	CTTCAGGGTCTCC ATCAAGC	BstNI	WT 44+94 bp M 131 bp
<i>bmp10</i> <i>-like</i>	pt544	GAGTCGGCGCAGC GCTAAAGTGAAG	TGGGGACTCTTCA GATTGAGCAGCG	MboII	WT 208 bp M 155+35 bp
<i>bmp10</i> <i>-like</i>	pt545	CGCAGCGCTAAAGT GGAGGACTGCTGA	TGGGGACTCTTCA GATTGAGCAGCG	Ddel	WT 200 bp M 168+24 bp
<i>bmp10</i> <i>-like</i>	sa11654	CAGAACTGCGCATT CACATGTTTC	GATGCCGACTTTT CTCCAGTCTC	Ddel	WT 197 bp M 176+21 bp

*Separation requires high-resolution gel e.g. 4% Metaphor agarose (Lonza, Walkersville MD USA).

Supplementary Material

Supplementary Table 3: PCR primers

Gene	Forward primer	Reverse primer
<i>bmp9</i>	CGCAGGAACACAGAAAGGTT	GTTGGGTTTGTGCCTTGT
<i>bmp10</i>	ATGCCTTCGGCAAACATCATACGC	TTGAAGAGAAGTGGGTGTCGTCTCAC
<i>bmp10-like</i>	GGCAGCTAACATCATCAGGAGCTC	AGATGTTGAACTGGAGACGCTGC
<i>acvrl1</i>	GGGTCTCGTCTTGTGGGAGA	GTCAGAGGGCACCATGTCAA
<i>actb2</i>	CGTGCTGTCTTCCCATCCA	TCACCAACGTAGCTGTCTTCTG

Supplementary Table 4: Primers used to generate templates for riboprobes

Gene	Forward primer	Reverse primer (with T7 site)
<i>Itbp3</i>	CCTGGGGCCAAAATAATGCTACA	TAATACGACTCACTATAGGGAGAAATGGG GACTTCCGGAGGGCTTGAC
<i>nkx2.5</i>	CATTAACCCTCACTAAAGGGAAGTG CGGGACATACTGAACCT	TAATACGACTCACTATAGGGTGCCTCTTG CACTTGATCG
<i>tbx20</i>	AGCCGCTCATCCCGACGACTC	TAATACGACTCACTATAGGGAGAGACGCG GTGTGATCTTCTTCTTG