

Supporting Information

Crystal structures of cyanobacterial light-dependent protochlorophyllide oxidoreductase

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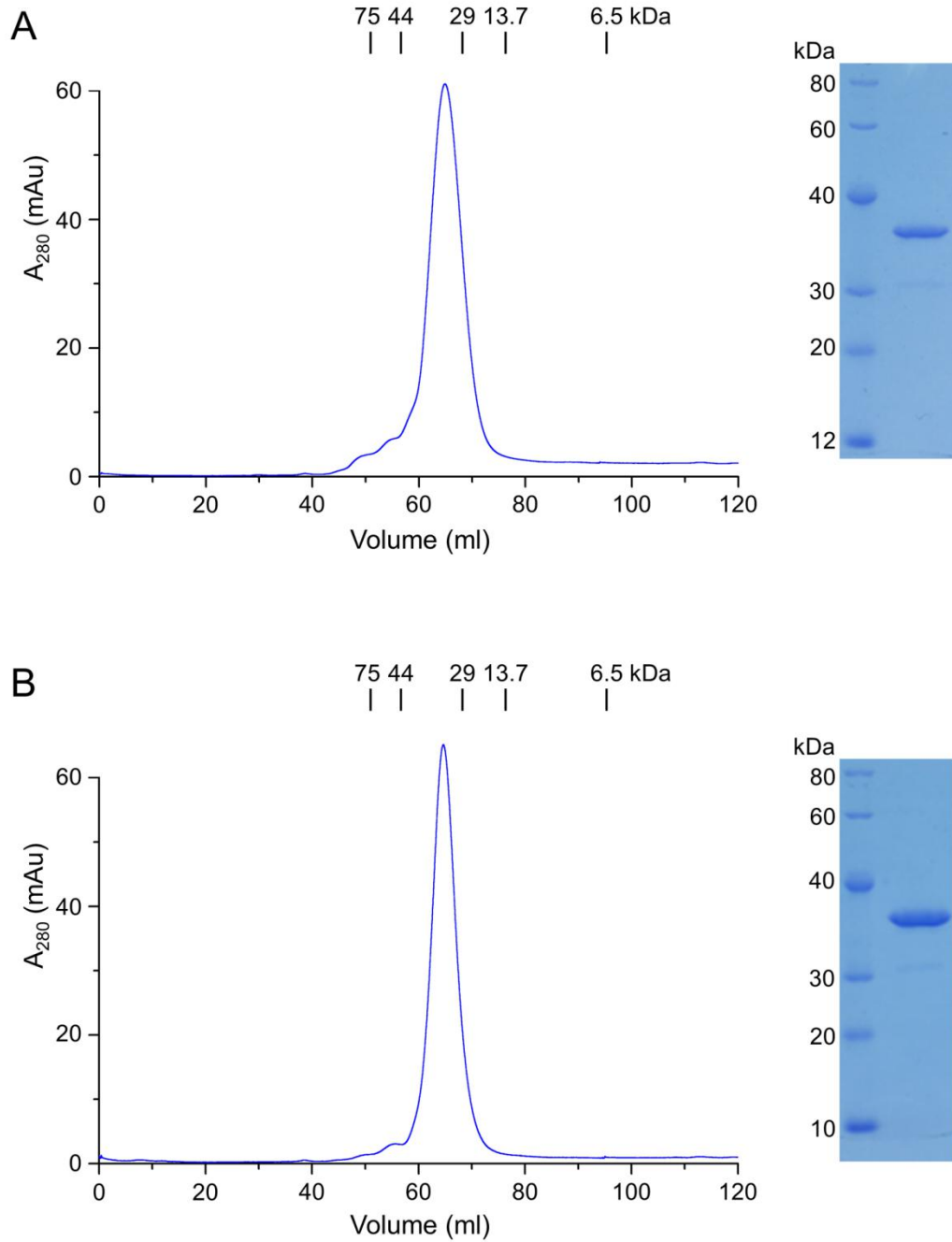


Fig. S1. Purification of *Sy*LPOR (A) and *Te*LPOR (B). The SEC profiles on a HiLoad Superdex 75 pg column (GE Healthcare) were recorded as absorbance at 280 nm. The flow rate was 1 ml min⁻¹. The molecular weight of protein standards (GE Healthcare) was labeled. The purity of the purified protein (peak fraction) was shown by the SDS-PAGE.

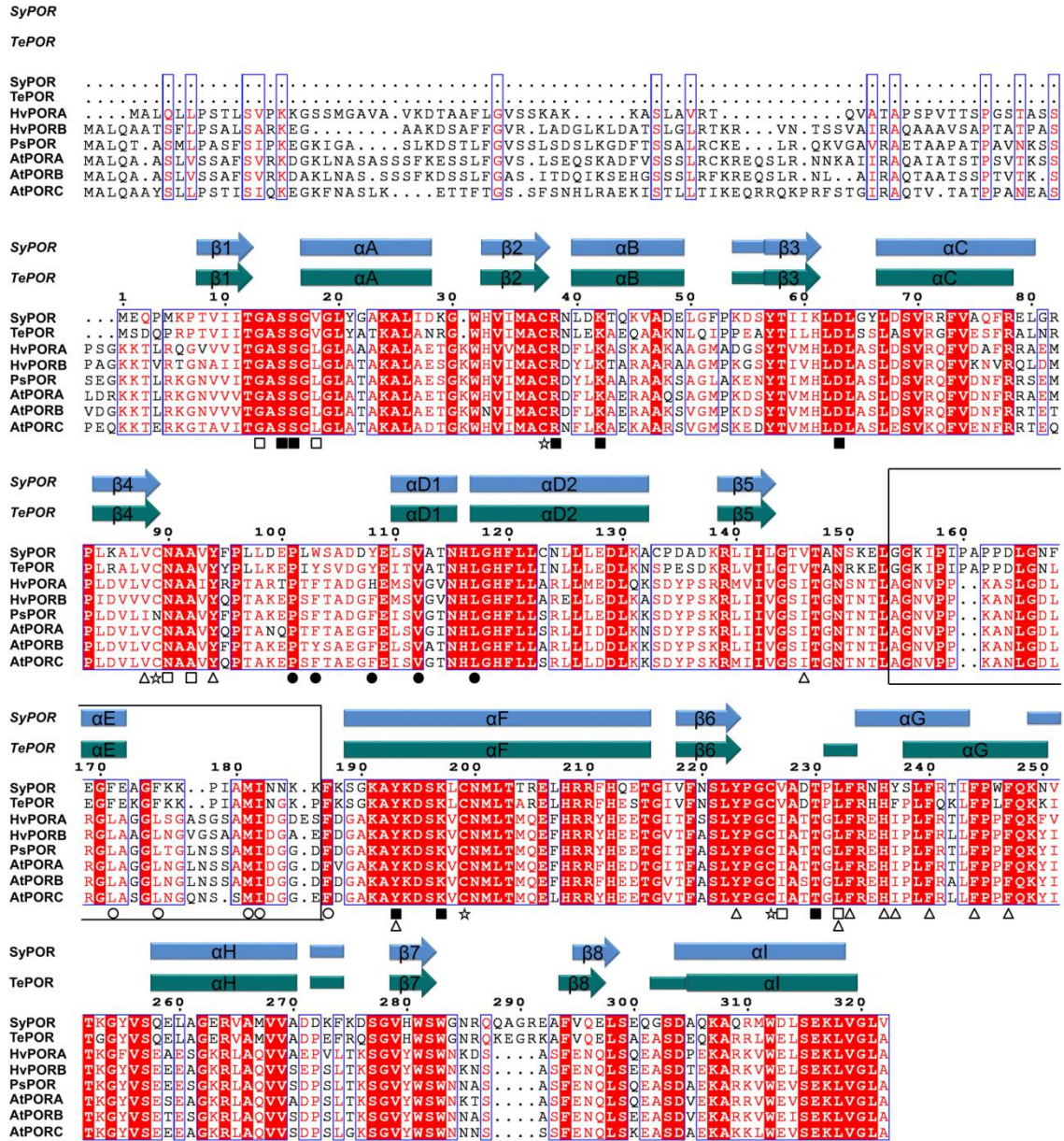


Fig. S2. Multiple LPOR sequences alignment. Identical residues are in white on a red background; similar residues are in red and boxed. Dots indicate gaps introduced during alignment. The secondary structure is calculated by DSSP, and labeled as cylinders for α -helices and arrows for β -strands. The stars denote the four conserved cysteine residues; the rectangles denote residues that interact with NADPH via the side-chain atoms (black) or the backbone atoms (white); the triangles denote residues that possibly participate in Pchlide binding; the circles denote residues that stabilize the LPOR-specific insertion by hydrophobic interactions, with the insertion residues in white and the core residues in black. The 33-residue LPOR-specific insertion is boxed in black rectangle. The abbreviations are: *Sy*, *Synechocystis* sp. PCC 6803; *Te*, *Thermosynechococcus elongates*; *At*, *Arabidopsis thaliana*; *Ps*, *Pisum sativum* L.; *Hv*, *Hordeum vulgare*. The alignment is generated with Clustal Omega (<https://www.ebi.ac.uk/Tools/msa/clustalo/>) and colored with ESPript 3.0 (<http://esript.ibcp.fr/ESPript/ESPript/index.php>).

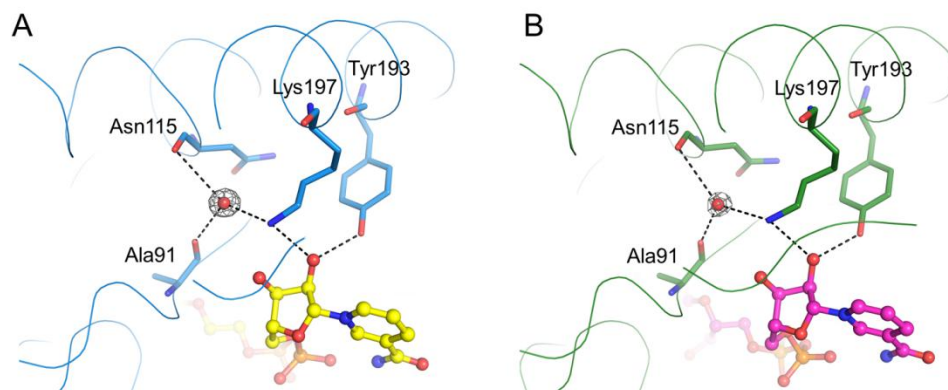


Fig. S3. The proton path within *SyLPOR* (A) and *TeLPOR* (B). The $2F_o - F_c$ electron density map of the water molecule (*red* sphere) bound to Ala91, Asn115, and Lys197 was contoured at 1.0 σ level.

Table S1. Constructs information.

Source organism	<i>Synechocystis</i> sp. PCC 6803	<i>Thermosynechococcus elongatus</i>
DNA source	Synthesized DNA	Synthesized DNA
Forward primer*	GGAATTCC <u>CATATG</u> GAACAGCC GATGAAACC	GGAATTCC <u>CATATG</u> AGTGACCA GCCGCGCCCGA
Reverse primer	GAT <u>CTCGAG</u> TAAACCAGACC AACCAGTTTTTCGC	GAT <u>CTCGAG</u> CGCCAGGCCAA CCAGTTTTTC
Cloning vector	pUC57	pUC57
Expression vector	pET-28a(+)	pET-22b(+)
Expression host	<i>Escherichia coli</i> BL21(DE3)	<i>Escherichia coli</i> BL21(DE3)
Complete amino acid sequence of the construct produced†	<u>MGSSHHHHHSSGLVPRGSH</u> MEQPMKPTVIITGASSGVGLY GAKALIDKGWHVIMACRNLD KTQKVADELGFPKDSYTIKLD LGYLDSVRRFVAQFRELGRPL KALVCNAAVYFPLLDEPLWSA DDYELSVATNHLGHFLLCNLL LEDLKACPDADKRLIILGTVTA NSKELGGKIPIAPPDLGNFEGF EAGFKKPIAMINNKFKSGKA YKDSKLCNMLTTRELHRRFHQ ETGIVFNSLYPGCVADTPLFRN HYSLFRITFPWFQKNVTKGYV SQELAGERVAMVVADDKFKD SGVHWSWGNRQQAGREAFVQ ELSEQGSDAQKAQRMWDLSE KLVGLV	MSDQPRPTVIITGASSGVGLYA TKALANRGWHVIMACRNLEK AEQAAKNLQIPPEAYTILHLDL SSLASVRGFVESFRALNRPLRA LVCNAAVYYPPLLKEPIYSVDG YEITVATNHLGHFLLINLLED LKNSPESDKRLVILGTVTANRK ELGGKIPIAPPDLGNLEGFEK GFKKPIAMINGKPFKSGKAYK DSKLCNMLTARELHRRFHST GIVFNSLYPGCVADTPLFRHHF PLFQKLFPLFQKKITGGYVSQE LAGERVAMVVADPEFRQSGV HWSWGNRQKEGRKAFVQELS AEASDEQKARRLWELSEKLVG <u>LALEHHHHHH</u>

*The restriction site is italicized and underlined.

†Extra amino acids generated during construction are underlined.

Table S2. Data collection and refinement statistics.

	SyLPOR–NADPH	TeLPOR–NADPH
Diffraction Data		
Diffraction source	BL18U1, SSRF	BL17U1, SSRF
Detector	Pilatus3 6M	Eiger X 16M
Wavelength (Å)	0.979	0.979
Unit-cell parameters		
<i>a</i> , <i>b</i> , <i>c</i> (Å)	156.0, 57.1, 72.9	52.9, 96.2, 141.7
α , β , γ (°)	90, 90, 90	90, 90, 90
Space group	P 21 21 2	P 21 21 21
Resolution (Å)	50-2.20 (2.28-2.20)*	50-2.40 (2.49-2.40)
Total No. of reflections	163488 (16060)	305028 (22810)
No. of unique reflections	33980 (3338)	27727 (2401)
Average redundancy	4.8 (4.9)	11.0 (9.5)
Mean <i>I</i> / σI	15.2 (2.2)	18.1 (4.5)
Completeness (%)	99.9 (99.9)	96.9 (85.9)
Wilson B-factor (Å ²)	37.26	43.01
<i>R</i> _{merge}	0.107 (0.874)	0.112 (0.547)
<i>R</i> _{pim}	0.052 (0.428)	0.036 (0.183)
CC _{1/2}	0.990 (0.686)	0.988 (0.913)
Refinement		
Resolution range (Å)	46.09-2.20 (2.26-2.20)	49.57-2.40 (2.51-2.40)
No. of reflections used in refinement	33794 (3279)	27674 (2275)
No. of reflections used for <i>R</i> _{free}	1710 (173)	1329 (126)
<i>R</i> _{work}	0.179 (0.256)	0.181 (0.244)
<i>R</i> _{free}	0.207 (0.289)	0.221 (0.292)
No. of non-hydrogen atoms		
protein	4620	4730
ligand	116	96
water	211	186
Average <i>B</i> factor (Å ²)		
protein	43.81	39.78
ligand	39.68	33.04
water	50.58	40.86
Model quality		
RMS bond lengths (Å)	0.004	0.004
RMS bond angles (°)	0.73	0.79
Ramachandran plot		
favored (%)	98.10	97.99
allowed (%)	1.90	2.01
PDB code	6L1G	6L1H

*Values in parentheses are for highest resolution shell.

Table S3. NADPH environment in *SyLPOR* and *TeLPOR*.

Bonding atom	NADPH atom		Distance (Å) in chain A/B	
	PDB V3 (IUPAC)	PDB V2	<i>SyLPOR</i>	<i>TeLPOR</i>
Water1 O	O7N	NO7	2.63/2.61	3.01/2.41
Water2 O	O2N	NO2	2.44/2.44	2.61/2.50
	O5B	AO5*	2.85/3.01	2.94/2.78
Water3 O	O1A	AO1	2.84/3.30	2.83/2.52
Water4 O	O1A	AO1	2.92/3.58	–
	O3X	AOP3	3.11/3.02	–
Water5 O	O3X	AOP3	3.06/3.99	2.72/2.73
Water6 O	O2X	AOP2	2.61/2.54	2.69/2.72
Water7 O	O2X	AOP2	2.78/2.81	2.47/2.69
Water8 O	O2B	AO2*	2.99/2.95	3.19/–
	N3A	AN3	2.92/2.82	2.76/–
Water9 O	N6A	AN6	2.95/2.99	2.79/3.15
Gly13 backbone C=O	O3B	AO3*	3.21/3.23	3.05/2.96
Ser15 γ O	O3B	AO3*	3.08/3.05	3.08/2.82
Ser15 γ O	O1X	AOP1	2.52/2.92	–
Ser16 γ O	O2A	AO2	2.89/2.96	2.85/2.75
Val18 backbone N	O2N	NO2	2.93/2.81	2.89/2.84
Arg38 η N	O3X	AOP3	3.00/2.62	2.71/2.75
Arg38 ϵ N	O2X	AOP2	2.80/4.07	2.90/4.12
Lys42 ζ N	O1X	AOP1	2.96/2.77	2.79/2.77
Asp63 δ O	N6A	AN6	3.43/3.29	3.22/3.10
Asn90 backbone C=O	O3D	NO3*	2.68/2.72	2.56/2.56
Ala92 backbone N	O4B	AO4*	3.43/3.29	3.15/3.07
Tyr193 η O	O2D	NO2*	2.68/2.72	2.70/2.64
Lys197 ζ N	O2D	NO2*	3.05/3.05	3.18/3.03
Lys197 ζ N	O3D	NO3*	2.95/3.01	3.12/3.03
Val227 backbone N	O7N	NO7	2.99/2.93	2.75/3.08
Thr230 γ O	O1N	NO1	2.64/2.52	2.74/2.77
Leu232 backbone N	O3	O3	–	3.13/3.15
Arg234 η N ₁	O1X	AOP1	–	2.86/2.96
Arg234 η N ₂	O1A	AO1	–	2.87/2.93