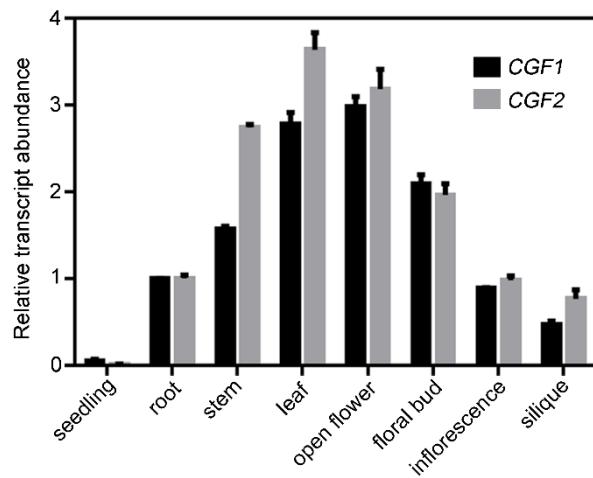


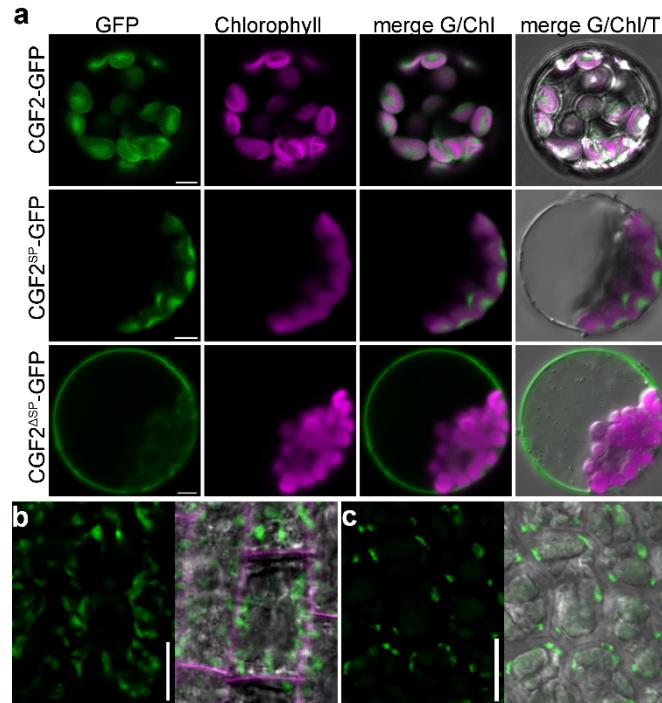
Supplemental Figure 1. CGF1 and CGF2 are homologous with multiple transmembrane domains predicted.

Sequence alignment was performed with VectorNTI. Yellow-highlighted amino acids are identical while green-highlighted amino acids are similar in side chains. Lilac box indicates predicted chloroplast transit peptide (TargetP). Blue boxes indicate TM domains shared by both proteins while empty boxes indicate TM domains only predicted in CGF1 (HMMTOP).

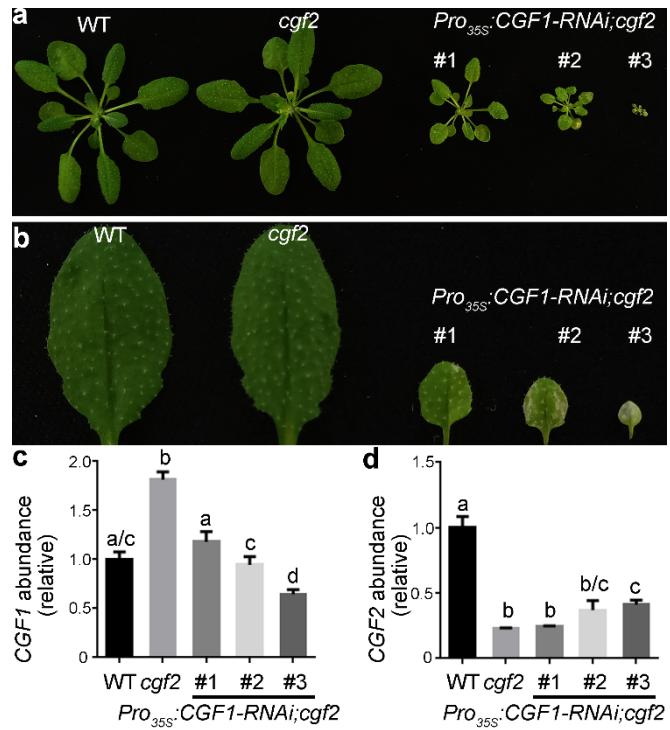


Supplemental Figure 2. *CGF1* and *CGF2* are expressed in diverse tissues and developmental stages.

Quantitative real-time PCRs (qPCRs) of *CGF1* and *CGF2* among different tissues. Results shown are means \pm standard errors (SEM, n=3). Each biological replicates were repeated three times with similar results.

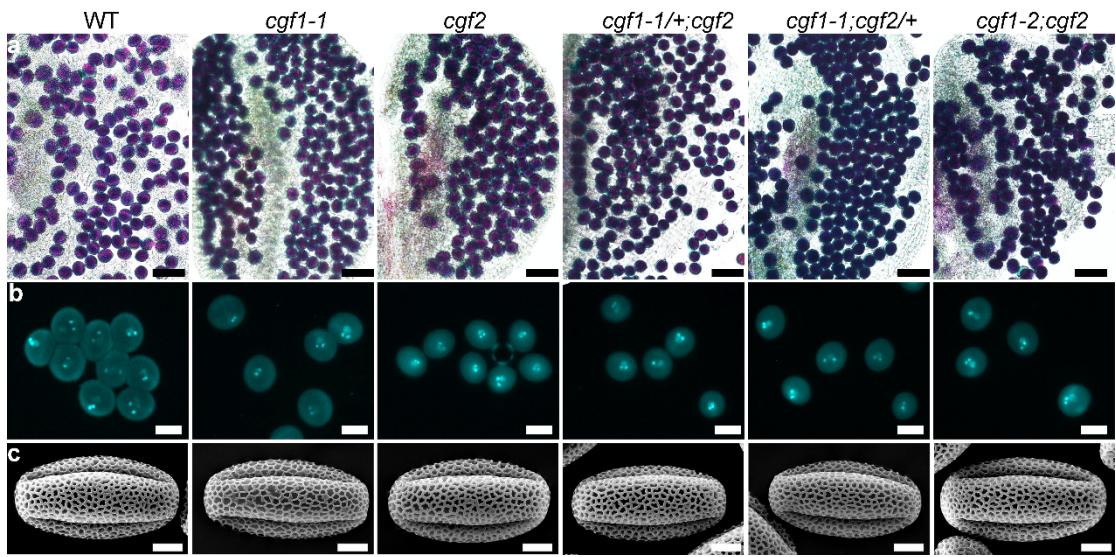


Supplemental Figure 3. CGF2 targets to chloroplasts through its N-terminal sequences. (a) CLSM of protoplasts from *Pro_{UBQ10}:CGF2-GFP*, *Pro_{35S}:CGF2^{SP}-GFP*, or *Pro_{35S}:CGF2^{ΔSP}-GFP* transgenic plants. From left to right: the GFP channel, autofluorescence channel (chlorophyll), merge of the GFP and autofluorescence (Chl) channels, merge of the GFP, autofluorescence, and transmission channels. (b) CLSM of root epidermal cells from *Pro_{UBQ10}:CGF2-GFP*. The right image is the merge of the GFP, RFP (FM4-64), and transmission channels. (c) CLSM of pavement cells from embryonic cotyledons of the *Pro_{UBQ10}:CGF2-GFP* transgenic plants. The right image is the merge of the GFP and transmission channels. Bars = 5 μm for (a); 10 μm for (b-c).



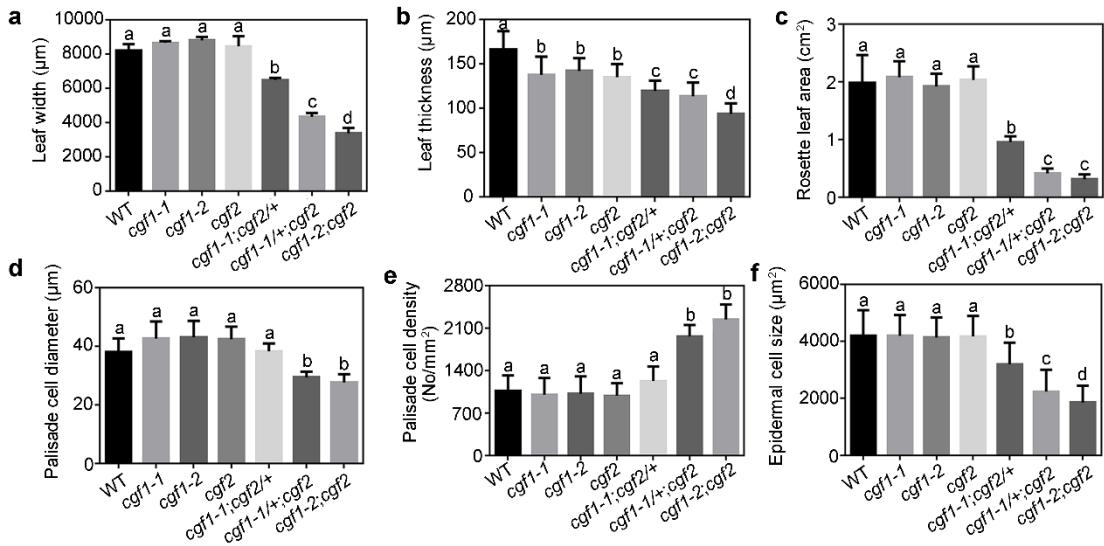
Supplemental Figure 4. Downregulating *CGF1* in *cgf2* mimicked defects of the double mutants.

(a-b) A representative 4 WAG plant (a) or the 6th true leaf (b) from 4 WAG wild-type, *cgf2*, or three lines of the *Pro_{35S}:CGF1-RNAi;cgf2* transgenic plants. (c-d) Relative *CGF1* (c) or *CGF2* (d) transcript abundance by qPCRs. Results are means \pm SEM ($n=3$). Each biological replicates were repeated three times with similar results. Different letters indicate significant different groups (OneWay ANOVA, Tukey's multiple comparisons test, $P<0.05$).



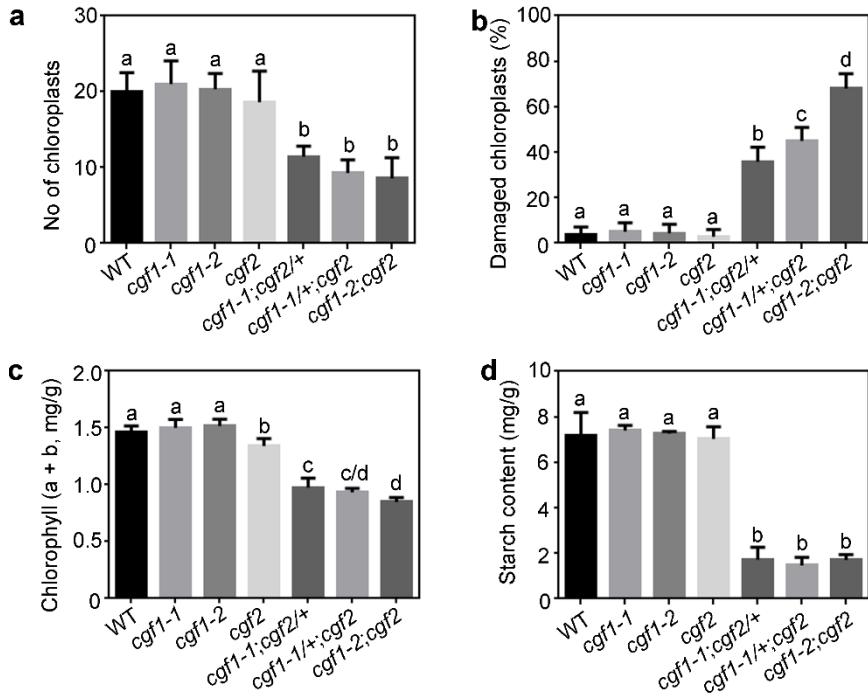
Supplemental Figure 5. Mutations of *CGF1* and *CGF2* did not affect pollen development.

(a-c) Alexander staining of a dehiscing anther (a), DAPI staining of mature pollen grains (b), or scanning electron micrographs (SEM, c) from wild-type, *cgf1-1*, *cgf1-2*, *cgf2*, *cgf1-1+/cgf2*, *cgf1-1;cgf2/+*, or *cgf1-2;cgf2* plants. Bars = 50 µm for (a); 20 µm for (b); 5 µm for (c).



Supplemental Figure 6. Mutations of both *CGF1* and *CGF2* affected leaf development.

(a-f) Quantification of leaf width (a), leaf thickness (b), rosette area (c), palisade cell diameter (d), palisade cell density (e), or pavement cell size (f) of the 4th true leaf from 3 WAG wild-type, *cgf1-1*, *cgf1-2*, *cgf2*, *cgf1-1;cgf2*, *cgf1-1;cgf2/+*, or *cgf1-2;cgf2* plants. Results are means \pm SE for (a) ($n > 4$); for (d) and (e) ($n > 3$). Results are means \pm SD for (b) ($n > 75$); for (c) ($n > 8$); for (f) ($n > 65$). Different letters indicate significant different groups (OneWay ANOVA, Tukey's multiple comparisons test, $P < 0.05$).



Supplemental Figure 7. Mutations of both *CGF1* and *CGF2* compromised chloroplast integrity.

(a-d) Number of chloroplasts per cross-section (a), the percentage of damaged chloroplasts among total chloroplasts (b), chlorophyll content (c), or starch content (d) of the 4th true leaf from 3 WAG wild-type, *cgf1-1*, *cgf1-2*, *cgf2*, *cgf1-1+/cgf2*, *cgf1-1;cgf2/+*, or *cgf1-2;cgf2* plants. Results are means \pm SE ($n > 7$). Different letters indicate significant different groups (OneWay ANOVA, Tukey's multiple comparisons test, $P < 0.05$).

Supplemental Table 1. Segregation ratio.

Parents	Progenies		
Female X Male	Genotype	Expected ratio	Observed
AaBb X AaBb	A(A/a)B(B/b): aaB(B/b): A(B/b)bb: aabb	9: 3: 3: 1	397: 101: 125: 0 ^a
Aabb X Aabb	AAbb: Aabb: aabb	1: 2: 1	68: 73: 0 ^b
aaBb X aaBb	aaBB: aaBb: aabb	1: 2: 1	76: 89: 0 ^b

^a Significantly different from 9: 3: 3: 1 (χ^2 , P< 0.01).
^b Significantly different from 1: 2: 1 (χ^2 , P< 0.01).
A: *CGF1*; B: *CGF2*; a: *cgf1-1*; b: *cgf2*.

Supplemental Table 2. Oligos used in this study.

Application	No.	5'-3' sequences
Cloning	<i>CGF1g</i>	ZP5837 CACCGTGGGACTCACTGGATTGC
		ZP4998 GTAGAGGCTAAAACCAAAGAAAGGG
	<i>CGF2g</i>	ZP5838 CACCACTCTCCACAACACCACCAATG
		ZP5739 ATACAGGCTGAATCCAAAGAAC
	<i>CGF2g-</i> 3'UTR	ZP5838 CACCACTCTCCACAACACCACCAATG
		ZP9022 GAGCAAATCAACCCCTAAGAGAGGAAC
	<i>CGF1</i>	ZP4997 CACCATGGAAAGGCTTCTCAACCATC
		ZP4998 GTAGAGGCTAAAACCAAAGAAAGGG
	<i>CGF1^{SP}</i>	ZP4997 CACCATGGAAAGGCTTCTCAACCATC
		ZP9786 TTTGGGAGACCCATTGAAGGAATTG
	<i>CGF2^{SP}</i>	ZP5738 CACCATGGATAGGCTTTGCAACCTC
		ZP9784 AGAAGCAATCAAGAAATTACTACTCTG
	<i>CGF1^{ΔSP}</i>	ZP9787 CACCTCGGATGAATCGAAACCTAATCCC
		ZP4998 GTAGAGGCTAAAACCAAAGAAAGGG
	<i>CGF2^{ΔSP}</i>	ZP9785 CACCTCTAAACCGATGCGTCGAAAC
		ZP5739 ATACAGGCTGAATCCAAAGAAC
qPCR	<i>CGF1</i>	ZP5839 ATATATGGTCTCGATTGAAGTCAGCTAACCGGTGTT
		ZP5840 TGAAGTCAGCTAAATCCGGTGTAGAGCTAGAAATAGC
	<i>CGF1&CGF2-</i> <i>CRISPR</i>	ZP5841 AACCAGGACTGAAGTATTGACACAATCTTAGTCGACTCTAC
		ZP5842 ATTATTGGTCTCGAAACCAGGACTGAAGTATTGACACAA
	<i>CGF2</i>	ZP9333 CGTCACTGTATA CGCCTATAGG
		ZP9334 AATCCGGGATTAGGTTCGATT
	<i>GAPDH</i>	ZP5013 GCTTCTCTCAAACCGATGCG
		ZP5014 GAAAGCTAGCGAAAGCAGGC
		ZP687 TGAAATCAAAAGCTATCAAGG
		ZP688 CATCATCCTCGGTGTATCAA