

Supplements

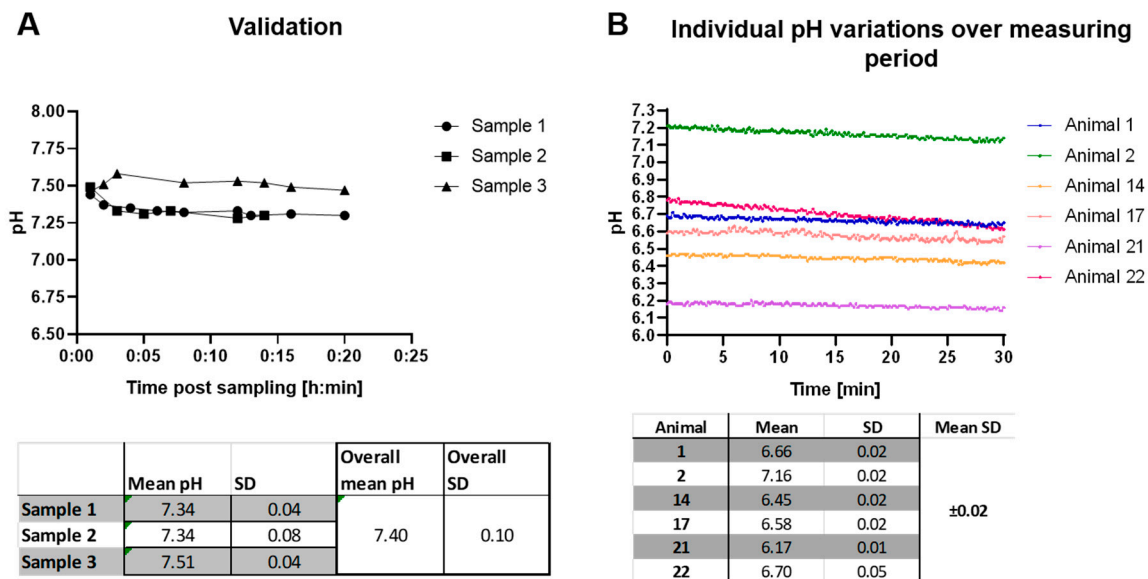


Figure S1. Validation of the PreSens microsensor by measurement of human non-coagulated blood samples (n=3) (A) and by confirming the consistency of measurement in vivo in the rat osteotomy hematoma (B). (A) measured mean pH of three human blood samples of 7.40 ± 0.1 is similar to the known physiological pH of human blood ranging from 7.35- 7.45. This validates the correct measurements by the pH sensor. (B) continuous recording (measurement interval: 10 s) over 30 min indicating a mean SD of individual pH per animal of ± 0.02 .

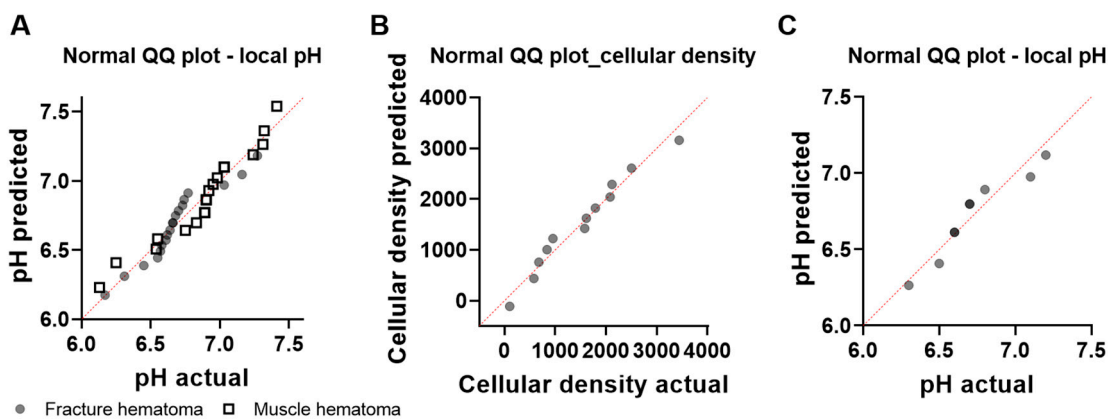


Figure S2. Normal QQ plots for visualization of normality analysis of data sets derived from the rat in vivo study. (A)- (C): Depicted is the fitting of all values belonging to the data set (actual values, x-axis) to the predicted values (y-axis), predicted under the assumption of normal distribution. (A) local pH of all fracture and muscle hematoma samples utilized to perform regression analysis of pH measured within both hematomas (Figure 3), n= 19. (B) local pH of fracture hematoma samples utilized for cellular density (C) calculation, for (B) and (C): n= 12.

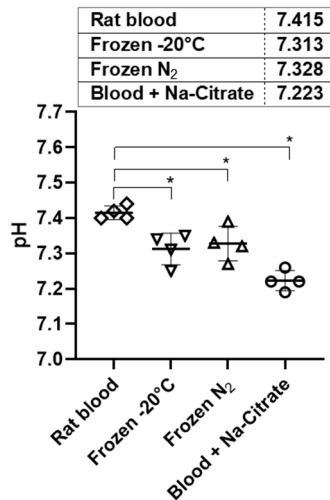


Figure S3. Storage and sample supplementation with anti-coagulant affect pH of sample (rat blood, n=4 independent blood samples from 4 rats). Direct pH measurement of rat blood, of previously frozen blood, either by freezing in -20°C environment or snap-freezing in liquid nitrogen and direct measurement in rat blood supplemented with the anti-coagulant sodium citrate. Two-way Mann-Whitney-U test was conducted to compare storage and supplementation with the directly measured pH.

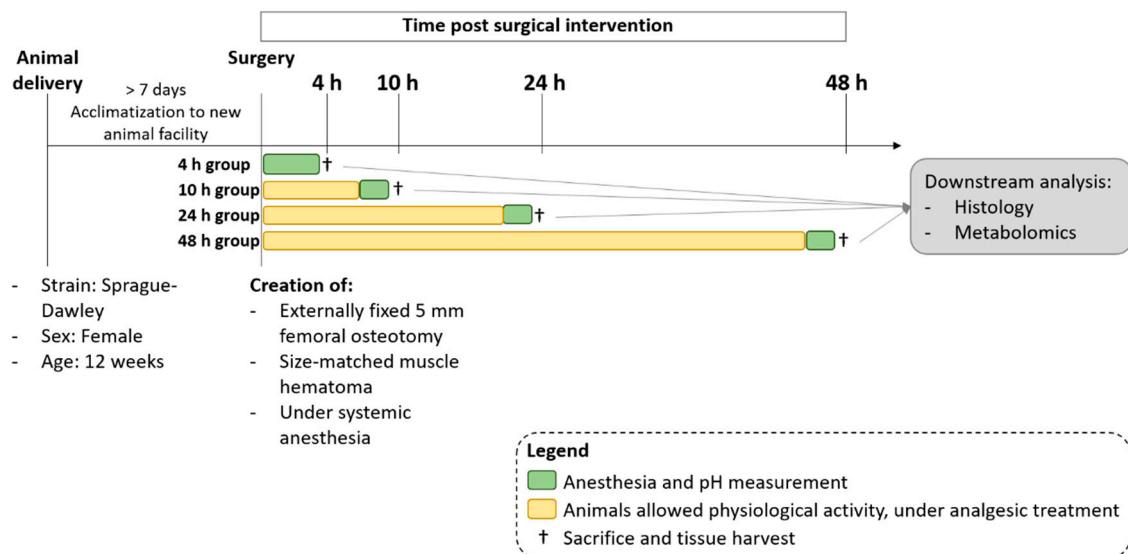


Figure S4. Study design of rat in vivo study.