Variation of 4 MV X-ray dose rate strongly impacts biological response both

in vitro and in vivo

M Ben Kacem, MA Benadjaoud, M Dos Santos, F Soysouvanh, V Buard, G Tarlet, B Le

Guen, A François, O Guipaud, F Milliat and V Paget

Supplementary Information

SF	0.63 Gy.min ⁻¹ vs 2.5 Gy.min ⁻¹	95% CI*
0.01	1.11	[1.06;1.16]
0.02	1.14	[1.09;1.19]
0.05	1.21	[1.15;1.27]
0.10	1.21	[1.16;1.27]
0.15	1.20	[1.14;1.27]
0.20	1.19	[1.12;1.27]
0.25	1.18	[1.10;1.27]
0.30	1.17	[1.08;1.27]
0.35	1.16	[1.06;1.28]
0.40	1.16	[1.05;1.28]
0.45	1.15	[1.03;1.28]
0.50	1.15	[1.02;1.29]
0.55	1.14	[1.00;1.29]
0.60	1.14	[0.99;1.29]#
0.65	1.13	[0.98;1.30] #
0.70	1.13	[0.97;1.30] #
0.75	1.12	[0.96;1.30] #
0.80	1.12	[0.95;1.30] #
0.85	1.11	[0.94;1.31] #
0.90	1.11	[0.93;1.31] #

Supplementary Table S1: Clonogenic assay RBE values (Ratio of doses for a given SF).

* 95% CI = 95% confidence interval. # Not statistically significant (CI including 1).

Dose (Gy)	RBE	95% CI*
0.5	1.01	[0.87 ; 1.18] [#]
1	0.97	[0.78 ; 1.23] #
1.5	0.91	[0.70 ; 1.20] #
2	0.83	[0.62 ; 1.10] #
2.5	0.75	[0.57 ; 0.98]
3	0.68	[0.51;0.88]
3.5	0.62	[0.47 ; 0.79]
4	0.58	[0.44 ; 0.74]
4.5	0.56	[0.43 ; 0.71]
5	0.54	[0.42 ; 0.69]
5.5	0.53	[0.41 ; 0.68]
6	0.53	[0.41 ; 0.68]
6.5	0.53	[0.40 ; 0.67]
7	0.53	[0.40 ; 0.67]
7.5	0.53	[0.40 ; 0.68]
8	0.53	[0.40 ; 0.68]
8.5	0.53	[0.40 ; 0.68]
9	0.53	[0.40 ; 0.69]
9.5	0.53	[0.40 ; 0.69]
10	0.53	[0.40 ; 0.70]
10.5	0.54	[0.40 ; 0.70]
11	0.54	[0.40 ; 0.71]
11.5	0.54	[0.40 ; 0.72]
12	0.55	[0.40 ; 0.72]
12.5	0.55	[0.41 ; 0.72]
13	0.55	[0.41 ; 0.72]
13.5	0.56	[0.41 ; 0.73]
14	0.56	[0.41 ; 0.74]
14.5	0.56	[0.42 ; 0.75]
15	0.57	[0.42 ; 0.75]
15.5	0.57	[0.42 ; 0.75]
16	0.57	[0.42 ; 0.76]
16.5	0.58	[0.42 ; 0.76]
17	0.58	[0.43 ; 0.77]
17.5	0.58	[0.43 ; 0.78]
18	0.59	[0.43 ; 0.79]
18.5	0.59	[0.43 ; 0.79]
19	0.59	[0.43 ; 0.80]
19.5	0.59	[0.43 ; 0.81]
20	0.60	[0.43 ; 0.82]

Supplementary Table S2: Senescence RBE (ratio for a given dose). * 95% CI = 95%

confidence interval. # Not statistically significant (Cl including 1).

	Estimated RBE	95% CI*
Severe damage	0.5117	[0.4487 ; 0.5834]

Supplementary Table S3: Severe damage RBE (ratio of severe damage percentages for a given dose) * 95% CI = 95% confidence interval. # Not statistically significant (CI including 1).



Supplementary Figure S1. "RBE" based on cell viability (ratios for a given dose and time). (A) "RBE" representation at day 1 (green curve), day 4 (orange curve), day 7 (purple curve), day 14 (red curve) and day 21 (black curve) as a function of dose (Gy) (B) Numerical values of RBE extracted from experimental data. *95% CI = 95% confidence interval. # Not statistically significant (CI includes 1).



Supplementary Figure S2. Cell cycle analysis performed on controls just before irradiation. (A) Example of one representative cell cycle analysis performed using the FlowJo cell cycle tool on single cells (set up to remove debris first, then by using size (APC-W)/intensity (APC-A) bi-parametric analysis to remove doublets) (B) Mean of cell cycle proportions for controls at T0 (n=14 in total, left panel); n=7 for 0.63 Gy.min⁻¹ irradiation (purple plots) plus n=7 for 2.5 Gy.min⁻¹ (blue plots) (right panel); 5x104 cells were analyzed per n.





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Supplementary Figure S3. Example of analysis set-up for C₁₂FDG experiments for 0.63 Gy.min⁻¹ irradiation. (A) First analysis on size (FSC: forward scatter)/granulometry (SSC: side scatter) parameters, to collect cells (gate G1) and to remove fragmented cells and debris. Triton 0.06X final was used as positive control to ensure good detection of living cells (Gate 2) and dead cells. (B) DMSO (vehicle of C₁₂FDG) used as negative control to ensure good detection of C₁₂FDG in controls (C) Detection of C₁₂FDG in controls. (D) DMSO used as negative control to ensure good detection of C₁₂FDG in 2 Gy. (F) Detection of C₁₂FDG in 10 Gy. (G) Bi-parametric (size/FITC) overlay of control versus 2 Gy irradiation (upper panel) and control vs 10 Gy irradiation (bottom panel).



Supplementary Figure S4. RT-qPCR TLDA custom array at D21. Assay heat map illustrating differentially expressed genes among 44 genes measured at day 21 post-irradiation. For one gene considered, red dots represent overexpression and green dots represent downregulation. Corrected p-values using the Benjamini-Hochberg method, p < 0.05. (n = 4 independent experiments per condition). Purple rounds correspond to 0.63 Gy.min⁻¹ irradiation and blue rounds to 2.5 Gy.min⁻¹ irradiation, the color gradient indicating the dose.