SUPPORTING INFORMATION

Structure of an Ancestral Mammalian Family 1B Cytochrome P450 with Increased Thermostability

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List of Supporting Information Included:

Supporting Table 1. Comparison of electrostatic interactions in extant human CYP1B1 vs. the ancestral CYP1B1 enzyme. Supporting Table 2. Comparison of aromatic and pi-cation stacking interactions in extant human CYP1B1 vs. the ancestral CYP1B1 enzyme. Supporting Table 3. MRM transitions for quantification of α -naphthoflavone

Supporting Figure 1. Mass fragmentation of α -naphthoflavone metabolites Supporting Figure 2. Known metabolic pathways of α -naphthoflavone

Table S1. Comparison of salt bridge interactions in extant human CYP1B1 *vs.* the ancestral CYP1B1 enzyme. Red text indicates interactions that are unique to that protein structure. These correspond to the residues show in Figures 6A and 6B, with the exception of partials, which are defined as one are two unique residues that interact with a shared network. Bold indicates that an interaction is between two distinct secondary structure elements. An asterisk indicates that the residue varies between the extant and ancestor enzymes.

Extant Human CYP1B1			Ancestral N98_CYP1B1_Mammal				
2-residue salt	Secondary	Salt-bridge	Secondary	2-residue salt	Secondary	Salt-bridge	Secondary
bridges	Structure	networks	Structure	bridges	Structure	networks	Structure
	Location		Location		Location		Location
D116*:H136*	B/B' loop :	E173 : R213:	D helix : E/F	R348: E499	I helix : β_3	R145 : HEM :	C helix : K"/L
	B'/C loop	E176 : R175/	loop : D helix		loop	R468/R117):	loop : β/β' loop
		H216: D217	: E/F loop		-	HEM: H401	: K/B ₂₋₁
E139 : K142	C helix	E223*: H227:	F helix	R368 : D374	J/J' loop : J'	R444 : E387 :	K'/K" loop : K
		E230*			helix	R390	helix
D192 : R194*	D/E loop : E	R523 : D361 :	C-term : J	E473 : K477	L helix	E474:H149:	L helix : C helix
	helix	R366	helix : J/J'			R469	: K"/L loop
			loop	E220 : K275	F helix : G	D373*:R163	J' helix : D
R233 : E260*	F helix : G	R390 : E387 :	K helix : K'/		helix	: E167	helix
	helix	R444	K" loop	D274 : R278	G helix	H216 : E176 :	E/F loop : D
E229 : K514	F helix : B4-2	R469 : E473 :	K"/L loop : L			R179*	helix
	/β ₃₋₂ loop	K477	helix	R290 : D294	G/H loop :	R523 : D361 :	C-term : J helix
D274 : R278	G helix	R145 : HEM :	C helix :		H helix	R366	: J/J' loop
		R468/R117:	HEM: K"/L	E100 : K433	B helix :	K416:D417:	β ₂₋₂ /β ₁₋₃ loop :
		HEM: H401	loop: B/B'		K'/K" loop	R136*	B'/C loop
			loop : K/β ₂₋₁	D291 : H279	G/H loop :	E318: R130:	I helix : B'/C
D291 · H279	G/H loon : G	Tot	al · 6		G helix	D316 loop : H/I loop	
15251.11275	helix	Unique: 1	+ 2 nartial	D333 : K512	I helix : β ₄₋₂	To	otal:8
D316 · K303*	H/Lloon · H	Cinque. 1	· 2 partial			Unique:	3 + 2 partial
2010.1000	helix			E359 : R362*	J helix		
E318 : R130	I helix : B'/C			D406* : H413	β2		
	loop			E438 : H429	K'-K" loop		
D333 : K512	I helix : B ₄₋₂			R213 : E173	E/F loop : D		
D351*:R355	J helix				helix		
D374 : R368	J' helix : J-J'			D504* : K514	β ₃ /β ₄ loop		
	loop				region		
E438 : H429 K'-K" loop				Total	Total: 14		
Total: 13				Unique: 6 +	- 1 partial	L	
Unique: 7							

Table S2. Comparison of aromatic and pi-cation stacking interactions in extant human CYP1B1 *vs.* the ancestral CYP1B1 enzyme. Red text indicates interactions that are unique to that protein structure. These correspond to the residues show in Figures 6C and 6D, with the exception of partials, which are defined as one are two unique residues that interact with a shared network. Bold indicates that an interaction is between two distinct secondary structure elements. An asterisk indicates that the residue varies between the extant and ancestor enzymes.

Extant Human CYP1B1				
Aromatic or cation-pi	Secondary Structure			
stacking interactions	Location			
H71 : W425 : H429	A helix : K' helix :			
	K'-K" loop			
F120 : F123 : F134	B-B' loop : B' helix :			
	B'-C loop			
H149 : R153	C helix			
F155 : R158*/F156*	C-D loop			
F346 : F384	I helix : K helix			
Y349 : F440	I/J loop : K'-K" loop			
Total : 6				
Unique: 3 + 1 partial				

Ancestral N98 CYP1B1 Mammal					
Aromatic or cation-pi	Secondary Structure				
stacking interactions	Location				
W57 : F74	N-term-β: A helix				
H71 : W425 : H429	A helix : K' helix :				
	K'-K" loop				
R117 : H401	B-B' loop : K-β2-1				
	loop				
F123 : F134	B' helix : B'-C loop				
W141 : R468	C helix : K"-L loop				
H149 : R153	C helix				
F231 : F261	F helix : G helix				
R366 : H489	J-J' loop				
Total : 8					
Unique: 5					

Compound	Q1	Q3	СЕ	EP
α -naphthoflavone	273.1	115.1	55	10
	273.1	129.1	55	10
	273.1	143.2	55	10
	273.1	171.2	55	10
	273.1	202.3	55	10
	273.1	226.2	55	10
Progesterone (IS)	315.2	97.1	28	8
	315.2	109.1	28	8
	315.2	297.2	47	8
OH-α-naphthoflavone	289.09	215.09	55	10
	289.09	131.05	55	10
	289.09	115.05	55	10
	289.09	103.05	55	10
	289.09	95.05	55	10
	289.09	77.04	55	10

Table S3. MRM transitions used for quantification of α -naphthoflavone, metabolites and internal standard.



Figure S1. Mass fragmentation of α -naphthoflavone metabolites. Tandem mass spectra and predicted fragmentation patterns of (A) α -naphthoflavone, (B) M1, and (C) M2. The fragmentation patterns of M1 and M2 are consistent with the addition of a single oxygen to the two rings indicated.



The known metabolic pathways of α -naphthoflavone (ANF) catalyzed by liver Figure S2. microsomes and recombinant CYP1A forms. The known α -naphthoflavone metabolic pathways catalyzed by CYP1 enzymes and rat liver microsomes (RLM) induced by either 2,3,7,8tetrachlorodibenzodioxin (66), 3-methylcholanthrene, β -naphthoflavone or phenobarbital (67) are summarized. The two main products of α -naphthoflavone metabolism by rat (r)CYP1A1 are the 5,6-oxide and 7,8-oxide (66,67). These compounds are further metabolized by rat epoxide hydrolase (rEH) to form the respective dihydrodiols. In the absence of rEH, the 5,6-oxide is chiefly observed, as the 7,8-oxide is unstable and will interact with nucleophilic groups on proteins, or spontaneously convert to the 7-hydroxy- α -naphthoflavone product (32,68). Recombinant human (h)CYP1A1, and to a lesser extent hCYP1A2, produced the 5,6-oxide as the main product, which was converted to the 5,6-dihydrodiol in the presence of rEH (32). The 7,8-dihydrodiol was not observed even in the presence of rEH. Other minor metabolites produced by recombinant rCYP1A1 or induced RLM are also shown. Earlier studies (69-71) generally agreed with the metabolite assignments shown here, except that the 7,8-dihydrodiol was originally assigned the structure now defined as the 9,10-dihydrodiol.