SUPPORTING INFORMATION

Structure of an Ancestral Mammalian Family 1B Cytochrome P450 with Increased Thermostability

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List of Supporting Information Included:

Supporting Table 1. Comparison of electrostatic interactions in extant human CYP1B1 *vs.* the ancestral CYP1B1 enzyme. Supporting Table 2. Comparison of aromatic and pi-cation stacking interactions in extant human CYP1B1 *vs.* the ancestral CYP1B1 enzyme. Supporting Table 3. MRM transitions for quantification of α -naphthoflavone

Supporting Figure 1. Mass fragmentation of α -naphthoflavone metabolites Supporting Figure 2. Known metabolic pathways of α -naphthoflavone

Table S1. Comparison of salt bridge interactions in extant human CYP1B1 *vs.* the ancestral CYP1B1 enzyme. Red text indicates interactions that are unique to that protein structure. These correspond to the residues show in Figures 6A and 6B, with the exception of partials, which are defined as one are two unique residues that interact with a shared network. Bold indicates that an interaction is between two distinct secondary structure elements. An asterisk indicates that the residue varies between the extant and ancestor enzymes.

Table S2. Comparison of aromatic and pi-cation stacking interactions in extant human CYP1B1 *vs.* the ancestral CYP1B1 enzyme. Red text indicates interactions that are unique to that protein structure. **These correspond to the residues show in Figures 6C and 6D, with the exception of partials, which are defined as one are two unique residues that interact with a shared network**. Bold indicates that an interaction is between two distinct secondary structure elements. An asterisk indicates that the residue varies between the extant and ancestor enzymes.

Compound	Q1	Q3	CE	EP
α -naphthoflavone	273.1	115.1	55	10
	273.1	129.1	55	10
	273.1	143.2	55	10
	273.1	171.2	55	10
	273.1	202.3	55	10
	273.1	226.2	55	10
Progesterone (IS)	315.2	97.1	28	8
	315.2	109.1	28	8
	315.2	297.2	47	8
OH- α -naphthoflavone	289.09	215.09	55	10
	289.09	131.05	55	10
	289.09	115.05	55	10
	289.09	103.05	55	10
	289.09	95.05	55	10
	289.09	77.04	55	10

Table S3. MRM transitions used for quantification of α -naphthoflavone, metabolites and internal standard.

Figure S1. Mass fragmentation of α -naphthoflavone metabolites. Tandem mass spectra and predicted fragmentation patterns of (A) α -naphthoflavone, (B) M1, and (C) M2. The fragmentation patterns of M1 and M2 are consistent with the addition of a single oxygen to the two rings indicated.

Figure S2. The known metabolic pathways of α -naphthoflavone (ANF) catalyzed by liver microsomes and recombinant CYP1A forms. The known α -naphthoflavone metabolic pathways catalyzed by CYP1 enzymes and rat liver microsomes (RLM) induced by either 2,3,7,8 tetrachlorodibenzodioxin (66), 3-methylcholanthrene, b-naphthoflavone or phenobarbital (67) are summarized. The two main products of α -naphthoflavone metabolism by rat (r)CYP1A1 are the 5,6-oxide and 7,8-oxide (66,67). These compounds are further metabolized by rat epoxide hydrolase (rEH) to form the respective dihydrodiols. In the absence of rEH, the 5,6-oxide is chiefly observed, as the 7,8-oxide is unstable and will interact with nucleophilic groups on proteins, or spontaneously convert to the 7-hydroxy- α -naphthoflavone product (32,68). Recombinant human (h)CYP1A1, and to a lesser extent hCYP1A2, produced the 5,6-oxide as the main product, which was converted to the 5,6-dihydrodiol in the presence of rEH (32). The 7,8-dihydrodiol was not observed even in the presence of rEH. Other minor metabolites produced by recombinant rCYP1A1 or induced RLM are also shown. Earlier studies (69-71) generally agreed with the metabolite assignments shown here, except that the 7,8-dihydrodiol was originally assigned the structure now defined as the 9,10-dihydrodiol.