#### Appendix

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# TMEM135 regulates primary ciliogenesis through modulation of

#### intracellular cholesterol distribution

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#### Appendix Figure S1. TMEM135 localizes to peroxisomes.

(A) Huh7 cells were transfected with TMEM135-myc plasmid (Upper and lower panel), followed by immunostaining with antibodies against PMP70 (red) and Myc (green) in the upper panel, Myc (red) and Tomm20 (green) in lower panel. Scale bar, 10 μm. Celle were co-transfected with TMEM135-Myc and RFP-SKL plasmid (middle panel).

(B) RPE1 cells were transfected with TMEM135-Myc plasmids (Upper panel and lower panel) and immunostained with antibodies against PMP70 (red) and Myc (green) in Upper panel and Myc (red) and Tomm20 (green) in lower panel. Scale bar, 10 μm. Endogenous TMEM135 and PMP70 was stained by TMEM135 antibody (green) and PMP70 antibody (red) middle panel. Scale bar, 10 μm.



# Appendix Figure S2. TMEM135 depletion is not associated with impaired peroxisomeal matrix protein import or accumulation of VLCFA.

(A) Huh7 cells were transfected with siRNAs as indicated followed by Western blot analysis for TMEM135 knock down efficiency by using TMEM135 antibody.

**(B)** Huh cells were transfected with siRNAs as indicated followed by Western blot analysis of different peroxisome protein (PMP70, catalase, Pex5, Pex14).

(C) Huh cells were transfected with siRNAs as indicated and immunostained with antibodies against catalase (red) and PMP70 (green). Scale bar, 10 µm.

(D) Huh7 cells were transfected with siRNAs as indicated followed by GC-MS analysis of

VLCFA. Data represent mean  $\pm$  SD (n =3 experiments). NS, not significant. Student's T-test.

(E) RPE1 cells were transfected with siRNAs as indicated followed by Western blot analysis for TMEM135 antibody.

(F) RPE1 cells were transfected with siRNAs as indicated and immunostained with antibodies against catalase (red) and PMP70 (green). Scale bar, 10 µm.



**Appendix Figure S3: TMEM135 siRNA efficiently reduces TMEM135 fluorescent signal from peroxisomes.** Representative image showing that all three siRNA efficiently reduces TMEM135 protein from peroxisomes stained with PMP70 in RPE1 cell. Scale bar, 10 μm.



Serum starvation

Rab8

#### Appendix Figure S4. Few cells were detected with ciliary endogenous ciliary Rab8.

RPE1 cells were serum-starved for 24 h and immunostained with Rab8 antibody. Representative image showing ciliary Rab8. Scale bar, 10  $\mu$ m. Arrow indicate the endogenous Rab8 localizes in few cell populations during serum-starvation.





(A, B) RPE1 cells were transfected with siRNAs as indicated followed by Western blot analysis for Rab11 and Rabin8 knockdown efficiency respectively.

(C) RPE1 cells were transfected with siRNAs as indicated followed methanol fixation and immunostaining with EHD1 (red) and  $\gamma$ -tubulin (green) antibody. Scale bar, 10  $\mu$ m.

(**D**) Quantitative data of (**C**) showing percentage of cells with EHD1 localization in the cilium or basal body. Data represents average (n=2 experiments), 100 cells were scored per condition per experiment.

(E) Cells were transfected as shown and the cell lysates were incubated with purified GST-JCF1 (RBD) fusion protein. The amount of GTP-Rab8 bound to GFT-JCF1(RBD) was analyzed by Western blot with Rab8 antibody.



#### Appendix Figure S6. TMEM135 depletion increases peroxisome-Rab8 overlap.

(A) RPE1 cells were transfected with siRNAs as indicated for 48 h, incubated with/without HP $\beta$ CD in serum starved media for additional 18 h, and immunostained with antibodies against Rab8 (red) and PMP70 (green). Scale bar, 10  $\mu$ m.

(B) Quantification of peroxisome-Rab8 overlap in (A) Data represent mean  $\pm$  SD (n = 3 experiments). 50 cells were scored per condition per experiment). \*P < 0.05, Student's t-test.





(B) Quantification of percentage of cells with IFT88 localized to cilium in (A) Data represent mean  $\pm$  SD (n = 3 experiments). 100 cells were scored per condition per experiment \*P < 0.05, Student's t-test.

(C) Quantification of percentage of cells with IFT88 localized to centrioles in (A) Data represent mean  $\pm$  SD (n = 3 experiments). 100 cells were scored per condition per experiment. NS, not significant, Student's t-test

# Appendix Table S1: List of siRNA sequences used in this study

siRNA	Sense strand sequence	Reference
TMEM135 (1)	5'-GCUCAAAUUAUGCAGUAGA-3'	Bioneer
TMEM135 (2)	5'-AGUUCAGGACAGCUGUGAU-3'	Bioneer
TMEM135 (3)	5'-CUGCAUUUAGGCAUCUGUU-3'	Bioneer
IFT20	5'-GUAAAGUAGAAGCAGAACA-3'	Bioneer
Rab8a	5'-GUAGUGCCAUGAAGCGAUU-3'	Bioneer
Rab8b	5'-CUGUCAUGUUCACUCUGAA-3'	Bioneer
Rab11a (1)	5'-CGAGCUAUAACAUCAGCAU-3'	Bioneer
Rabin8 (1)	5'-GUCAUCAGGACCUCAGUGU-3'	Bioneer

## Appendix Table S2: List of antibodies used in this study

Antibody	Application	Dilution	Source	Reference	
c-Myc	IF	1:1000	Mouse	sc-40, Santa Cruz Biotech	
c-Myc	IF	1:1000	Rabbit	A190-205A, Bethyl	
PMP70	IF and WB	1:500	Rabbit	ab3421, Abcam	
PMP70	IF	1:500	Mouse	SAB4200181, Sigma	
Tomm20	IF	1:500	Mouse	sc-17764, Santa Cruz Biotech	
SREBP2	WB	1:1000	Rabbit	Lee JN, et al. (2004)	
UBXD8	WB	1:2000	Rabbit	Lee JN, et al. (2010)	
CREB	WB	1:2000	Rabbit	#9192, Cell Signaling	
CP110	IF	1:500	Rabbit	12780-I-AP, Proteintech	
IFT20	IF, and WB	1:500	Rabbit	13615-1-AP, Proteintech	
IFT88	IF	1:500	Rabbit	13967-1-AP, Proteintech	
Rab8a	IF and WB	1:500 and 1:2000	Rabbit	ab188574, Abcam	
ARL13B	IF	1:500	Rabbit	17711-1-AP, Proteintech	
Acetelated-tubbulin	IF	1:1000	Mouse	T7451, Sigma	
γ-tubulin	IF	1:500	Mouse	T5326, Sigma	
GM130	WB	1:1000	Mouse	610822, BD Transduction Laboratories	
Rab11	IF	1:200 and 1:2000	Mouse	610656, BD Transduction Laboratoreis	
GFP (B-2) antibody	WB	1:1000	Mouse	SC-9996, Santa Cruz Biotech	
Rabin8	IF	1:500	Rabbit	12321-I-AP, Proteintech	
TCTN1	IF	1:500	Rabbit	15004-I-AP, Proteintech	
TMEM135	IF and WB	1:500 and 1:1000	Rabbit	SAB3500832, Sigma	
TMEM135	IF and WB	1:500 and 1:1000	Rabbit	Ab222237, Abcam	
LAMP1	IF and WB	1:500 and 1:1000	Mouse	sc-2011, Santa Cruz Biotech	
Catalase	IF and WB	1:500 and 1:2000	Rabbit	Ab4877, Abcam	
Pex5	WB	1:2000	Rabbit	GTX109798	
Pex14	WB	1:1000	Rabbit	A303-0858, Bethyl	
EEA1	WB	1:1000	Rabbit	C45B10, Cell signalling	
Na <sup>+</sup> /K <sup>+</sup> ATPase	WB	1:2000	Rabbit	Sc-21712, Santa Cruz Biotech	
EHD1	IF	1:250	Rabbit	HPA067747, ATLAS ANTIBODIES	

Gene	Primer sequence	Reference	
TMEM135	F: 5'-CCAGCTTGGAGCTTTTCTTG-3'	Bioneer	
	R: 5'AGTTCATCATCTAAGTTTCTGATCCA- 3'		
HMGCoA synthase	F: 5'-GACTTGTGCATTCAAACATAGCAA-3'	Bioneer	
	R: 5'-GCTGTAGCAGGGAGTCTTGGTACT-3'		
HMGCoAreductase	F: 5'-CAAGGAGCATGCAAAGATAATCC-3'	Bioneer	
	R: 5'-GCCATTACGGTCCCACAC-3'		
LDLR	F: 5'-GGCTGCGTTAATGTGACACTCT-3'	Bioneer	
	R: 5'-CTCTAGCCATGTTGCAGACCTTGT-3'		
Insig1	F: 5'-CCCAGATTTCCTCTATATTCGTTCTT-3'	Bioneer	
	R: 5'-CACCCATAGCTAACTGTCGTCCTA-3'		
Rab8a	F: 5'-GCCTACTACAGGGGTGCAAT-3'	Bioneer	
	R: 5'-GGATGTTGTCGAAGGACTTC-3'		
Rab8b	F: 5'-ACGAGATATAATGACAAAAC-3'		
	R: 5'-GATCGGTTTTCTGTTATTT-3'		

### Appendix Table S3: List of primer sequence used for RT-QPCR

#### References

Lee, J.N., and Ye, J. (2004). Proteolytic activation of sterol regulatory element-binding protein induced by cellular stress through depletion of Insig-1. *J Biol Chem.* **279**, 45257-65.

Lee, J. N., Kim, H., Yao, H., Chen, Y., Weng, K., Ye, J. (2010). Identification of Ubxd8 protein as a sensor for unsaturated fatty acids and regulator of triglyceride synthesis. *Proc Natl Acad Sci USA*. **107**, 21424-9.