

1 **Supplemental Material for: Applied and Environmental Microbiology**

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4 **Diazotrophs show sign of restoration in Amazon rainforest soils with ecosystem**
5 **rehabilitation**

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41 **Supplemental material: S1**

42 **Supplementary Figures:** Figures 1-8

43 **Supplementary Table:** Table S1&2

44 **Identification of various diazotrophic groups that responded to change in land-**

45 **use.** Overall, across all three land use systems, we observed *nifH* sequences from NifH

46 cluster I–V; however, sequences related to clusters I & II were the most dominant (**Fig.**

47 **S6**). The differences among various land use systems were obvious. NifH–I cluster,

48 dominated by the *Alphaproteobacteria*–related *nifH* gene sequences, showed that the

49 *Proteobacteria*–related diazotrophs are susceptible to changes in land use.

50 *Alphaproteobacteria*–related diazotrophs were abundant in the primary and secondary

51 forests (35–41%), whereas *Gammaproteobacteria* (88%) and *Deltaproteobacteria* (55%)

52 were dominated in the pasture soils. Similar to our results, Dörr *et al.* [1] observed the

53 dominance of methanotrophic *Alphaproteobacteria* in the primary forest soils and

54 *Gammaproteobacteria* species in the farmland created after deforestation of the

55 Amazon. Differences among other, less abundant subgroups across different land use

56 systems were also evident (**Fig. S6**), such as in the NifH–II cluster,

57 *Verrucomicrobium*–related *nifH* gene sequences were retrieved in a relatively high

58 abundance (72%) in the pasture lands as compared to primary and secondary forest

59 sites. Similarly, Ranjan *et al.* [2] observed an increase in abundance of overall

60 *Verrucomicrobium*, based on 16S rRNA gene sequences, in response to the conversion

61 of forest to pasture land. The presence of *nifH* gene and N-fixation by *Verrucomicrobium*

62 spp. have been reported previously [3]. Several of the *nifH* genes OPU (operational

63 protein unit) in the NifH–II cluster lacked any cultured representative that suggest the

64 presence of diazotrophs containing novel lineage of *nifH* genes in these different

65 ecosystems (**Fig. S6; Table S1**).

66 The *nifH* gene has been extensively used as a biomarker gene to assess functional
67 diversity across different environments [4-9], and phylogenetic clustering based on the
68 *nifH* gene has been corroborated with the 16S rRNA gene-based clustering [10, 11].
69 However, the taxonomic inference drawn based on the *nifH* gene could possibly be
70 influenced by the horizontal transfer of this gene [12]. It is encouraging that some of the
71 diazotrophs related to *Proteobacteria* (Alpha, Beta, Gamma), *Firmicutes*, and
72 *Actinobacteria* that were detected in this study, have been previously isolated and
73 characterized by 16S rRNA gene sequencing from the primary forest sites [13]. Future
74 ongoing culturing and identification of the diazotrophs across these three different land
75 use systems could further highlight this issue.

76 **MATERIALS AND METHODS:**

77 **Identification of diazotrophs.** Major diazotrophic groups susceptible to the change in
78 land use were identified by analysis of the representative protein sequences at 88%
79 similarity. Translated *nifH* protein sequences (394,920 sequences, 131,640 per land use
80 system, subsampled based on the land use system with the least number of DNA
81 sequences) were aligned with the *nifH* protein sequences of the reference database
82 from Wang *et al.* [7], and a maximum likelihood (ML) phylogenetic tree was generated
83 using FastTree. Operational protein units (OPUs) were collapsed at 88% amino acid
84 similarity level that was corresponding approximately to the class level differences [14].
85 Distribution of sequences within each OPU were presented as bar graphs.

86 **Quantitative PCR for the *nifH* gene.** The abundances of the *nifH* gene sequences
87 across different land use systems were quantified through an absolute quantitative PCR
88 approach. A 25 μ l qPCR reaction contained 12.5 μ l of iTaq™ Fast SYBR Green Super
89 mix (Bio-Rad Inc., Hercules, CA), 100 nM of the *nifH* primers (same as used for

90 high-throughput sequencing) and 10 ng of the template DNA. The *nifH* gene from
91 *Herbaspirillum seropedicae* (American Type Culture Collection 35892) was cloned into
92 TOP 10 *Escherichia coli* competent cells and used for the generation of standard curves.
93 PCR-grade water without template was used as a negative control that did not show any
94 amplification. The real-time PCR amplification efficiency of *nifH* gene was 98%, and the
95 R^2 of 0.99.

96 Although, we followed the absolute quantification procedure the main purpose was to
97 assess the relative abundance of diazotrophs across three different land-use systems. In
98 general, the extracted DNA samples were relatively pure and the quality of the extracted
99 DNA was very good (as indicated by 260/280 ratios >1.75). Overall, the DNA extraction
100 yield across three land-use systems was very similar (Pasture 25 ± 7.5 ng/ μ l; Forest
101 31 ± 3.9 ; Secondary forest 29 ± 8.5). We did evaluate the presence of potential PCR
102 inhibitors in 15 randomly selected DNA samples (five per land-use system) by
103 comparing the *nifH* gene copies quantified from concentrated and 1:10 diluted DNA
104 samples. We did not observe any differences between concentrated vs diluted samples
105 suggesting the absence of PCR inhibition (data not shown).

106 **Supplementary Figures:** Figures S1-S8

107 **Supplementary Figures:**

108 **Fig. S1:** Taxonomic (black) and phylogenetic (grey) α -diversity of the *nifH* genes in
109 response to land use change in the Amazon. Taxonomic diversity was calculated using:
110 A) Margalef species richness index and B) Shannon diversity index. Phylogenetic
111 diversity was calculated as Faith's phylogenetic diversity (Faith, 1992). Overall, 135 000
112 *nifH* gene sequences (45,000 per land use and 1,500 per sample) were used to
113 calculate α -diversity. Error bars represent 95% CI.

114 **Fig. S2 A-C:** Diazotrophic community turnover in space, or β -diversity at different DNA
115 identity levels: A) 95% DNA identity, B) 90% DNA identity, C) 97% protein similarity
116 (amino acid based) in response to land use change in the Amazon. Differences in the
117 slopes across different land use systems were compared by power model. Upper panel
118 represent the variations in the taxonomic β -diversity and lower panel represent the
119 phylogenetic β -diversity across different land use systems. β -diversity was also calculated
120 from subsampled data i.e., total 135,000 *nifH* gene sequences (45,000 per land use and
121 1,500 per sample).

122 **Fig. S3:** Nonmetric multidimensional scaling representation of the *nifH* gene sequence
123 composition at 97% DNA identity for three different land use systems in the Amazon:
124 Subsampled data set containing total 135,000 *nifH* gene sequences (45,000 per land
125 use and 1,500 per sample) were for this comparison.

126 **Fig. S4:** Nonmetric multidimensional scaling representation of the *nifH* gene sequence
127 composition at different DNA identity (95%, 90%) and protein (97%, 95%) similarity levels
128 for three different land use systems in the Amazon: Primary forest samples (black
129 triangle); Pasture samples (white circles); Secondary forest samples (gray squares).
130 Complete data set containing total 443,600 *nifH* gene sequences and distribution of
131 number of sequences retrieved from different treatment were: primary forest samples
132 (158,261); pasture samples (143,700); secondary forest samples (131,640).

133 **Fig. S5:** Log-scale abundance of *nifH* gene copies (g^{-1} dry soil) for the three different land
134 use systems. Each bar represents about 90 samples (30 biological replicates per site and
135 3 technical replicates) and the error bar represents 95% CI.

136 **Fig. S6:** Maximum likelihood phylogenetic tree based on translated amino acid
137 sequences of the *nifH* gene from three different land use systems in the Amazon

138 rainforest. The size of each circle is proportionate to the log number of sequences within
139 an OPU. Total number of sequences in an OPU are also shown next to the taxon
140 names. Overall, a total of 394,920 *nifH* gene sequences were used and the comparison
141 was normalized to 131,640 sequences from each treatment based on the least number
142 of sequences retrieved from secondary forests (131,640 *nifH* gene sequences were
143 randomly sampled from primary forest and pasture samples to match equal numbers).
144 Within an operational protein unit (OPU) translated amino acid *nifH* gene sequences
145 were collapsed at 88% similarity. Numbers at the nodes reflect bootstrap support values
146 above 70% with branches within clusters being collapsed. Operational protein units
147 (OPUs) and the closest matched sequences within each OPU that were obtained
148 through the *nifH* gene reference database were reported in (**SI Table 1**).

149 **Fig. S7: A).** Operational taxonomic unit (OTU) accumulation curve for three land use
150 systems in the Amazon. Total 443,600 *nifH* gene sequences retrieved from different
151 treatments (primary forest samples (158,261); pasture samples (143 700); secondary
152 forest samples (131,640) were used to generate accumulation curve. The observed
153 number of OTUs were calculated at 97% DNA similarity.

154 **B).** The accumulation curve for each sampling location. Total nine sites, three per land
155 use system (Forest FI, II, III; Pasture P72I, II, III, and Secondary forest SCI, 93, 98).

156 **Fig. S8:** A spatially explicit sampling scheme that indicates the different sampling
157 locations within three land use systems. A square box represents 12 sampling points at
158 each location. Overall 36 soil samples per land use system.

159 **Supplemental Table:**

160 **Table S1:** Operational protein units (OPUs) and the closest matched sequences within each
161 OPU that were obtained through the *nifH* gene reference database by Wang et al.,
162 (2013). **Table S2:** Soil physiochemical characteristics of three land-use systems in the
163 Amazon.

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207 rainforest. *Appl Environ Microbiol* 80:281–288.

209

Fig. S1

97% DNA identity Alpha diversity

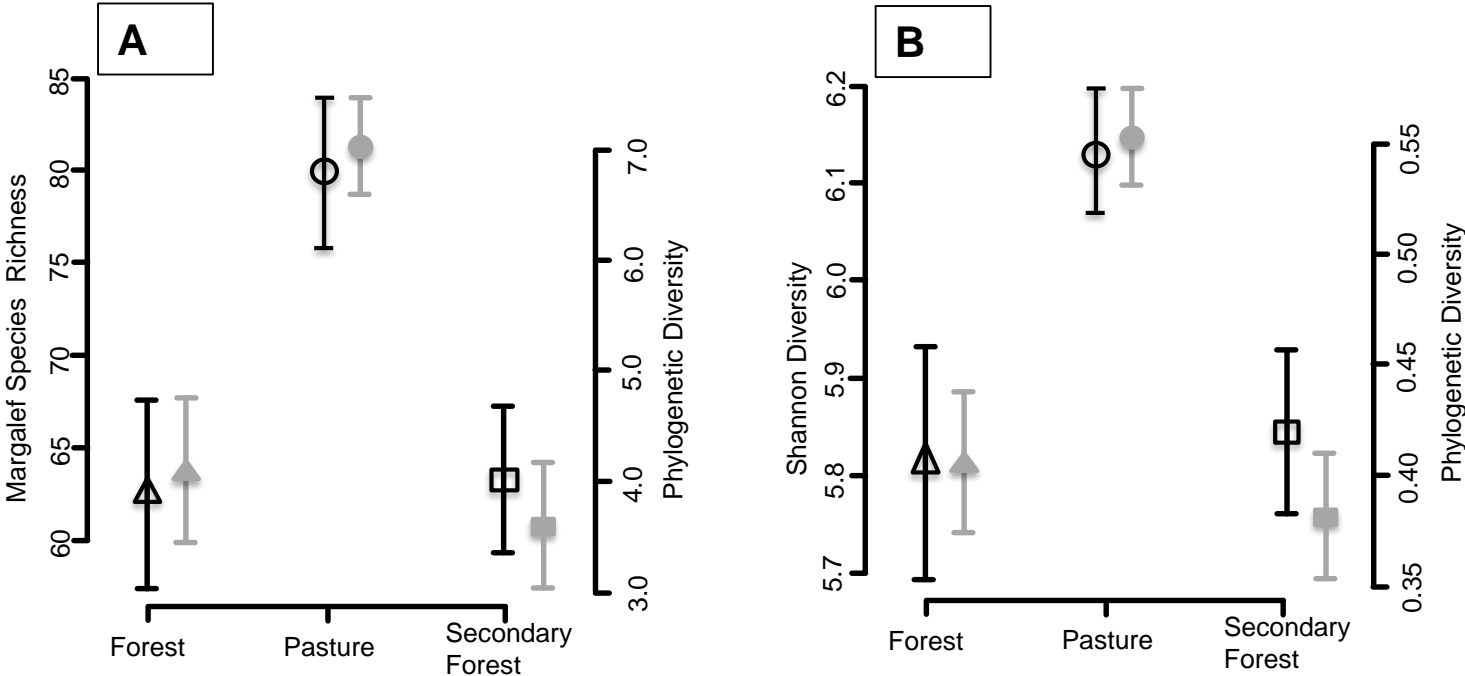
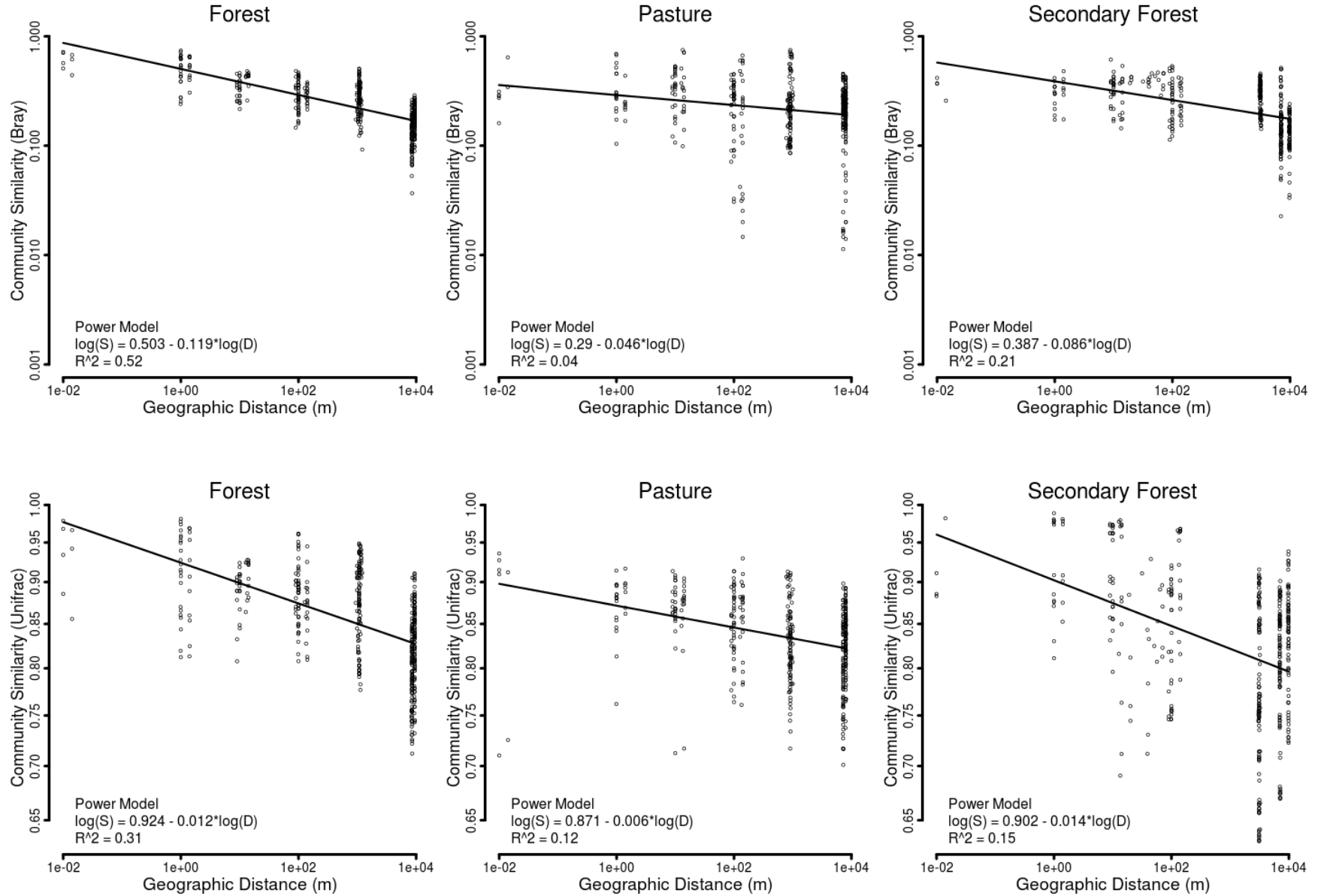


Fig. S2A

95% DNA Bray-Curtis and unifrac metrics



90% DNA Bray-Curtis and unifracs metrics

Fig. S2B

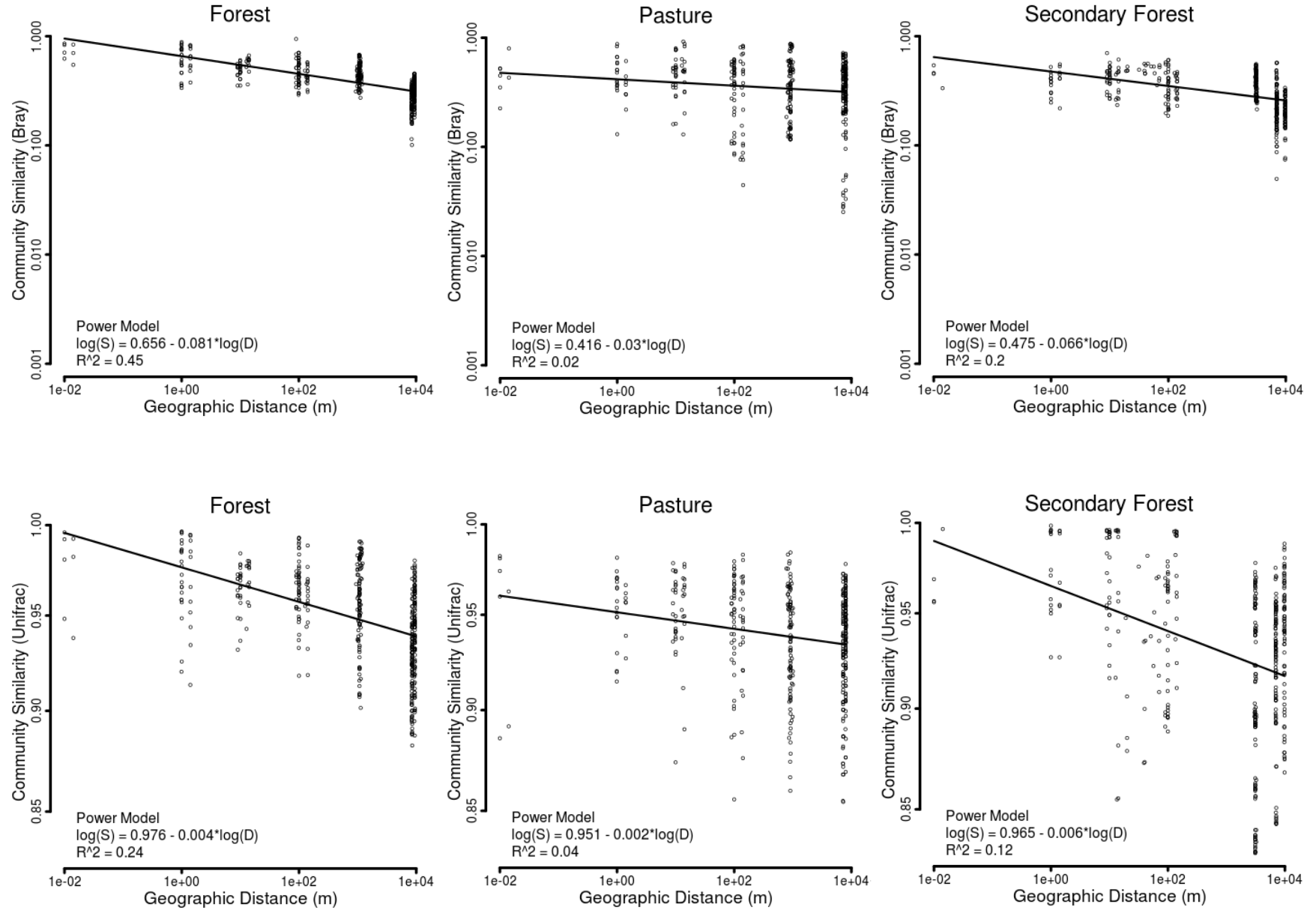


Fig. S2C

97% protein Bray-Curtis and unifracs metrics

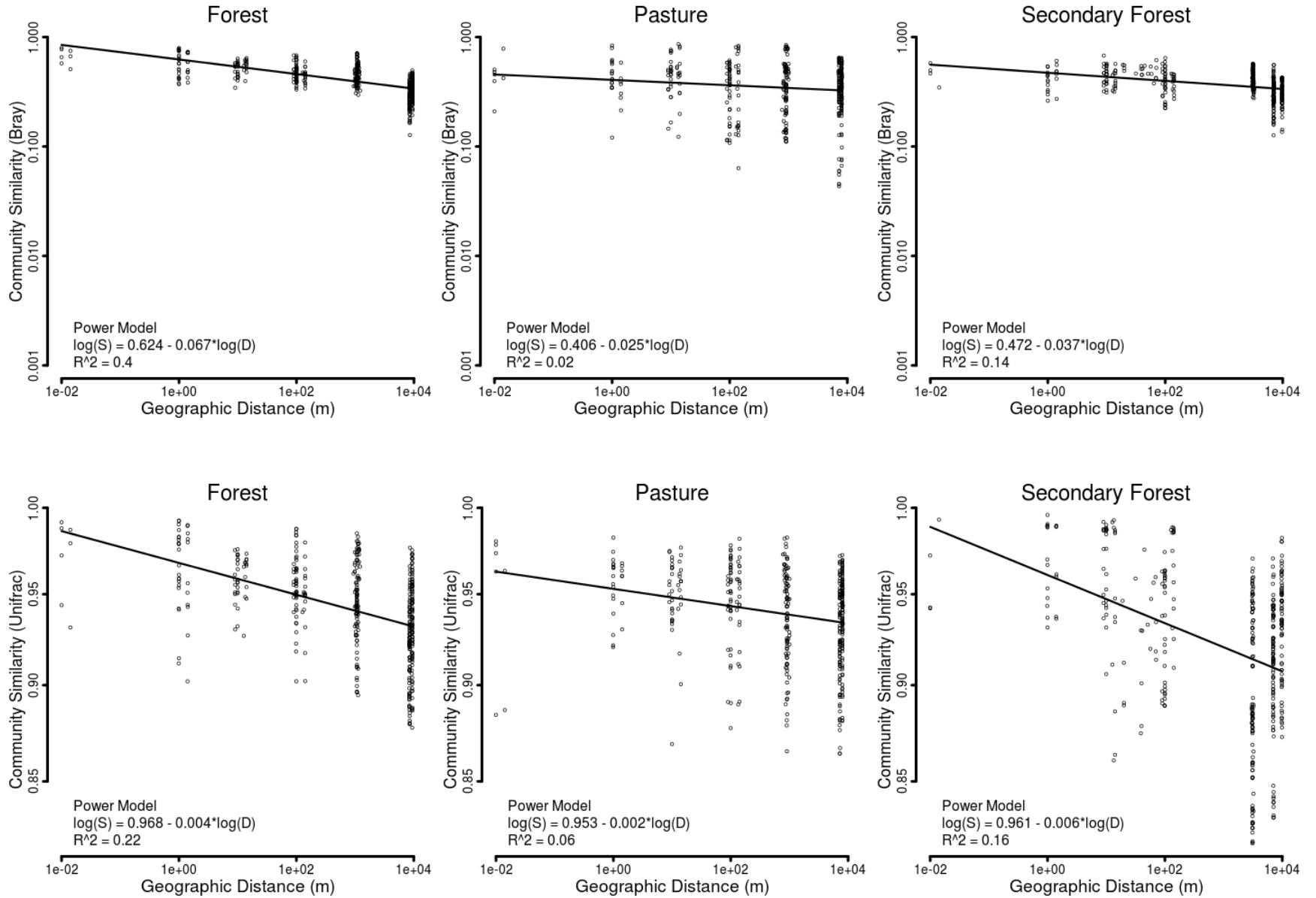
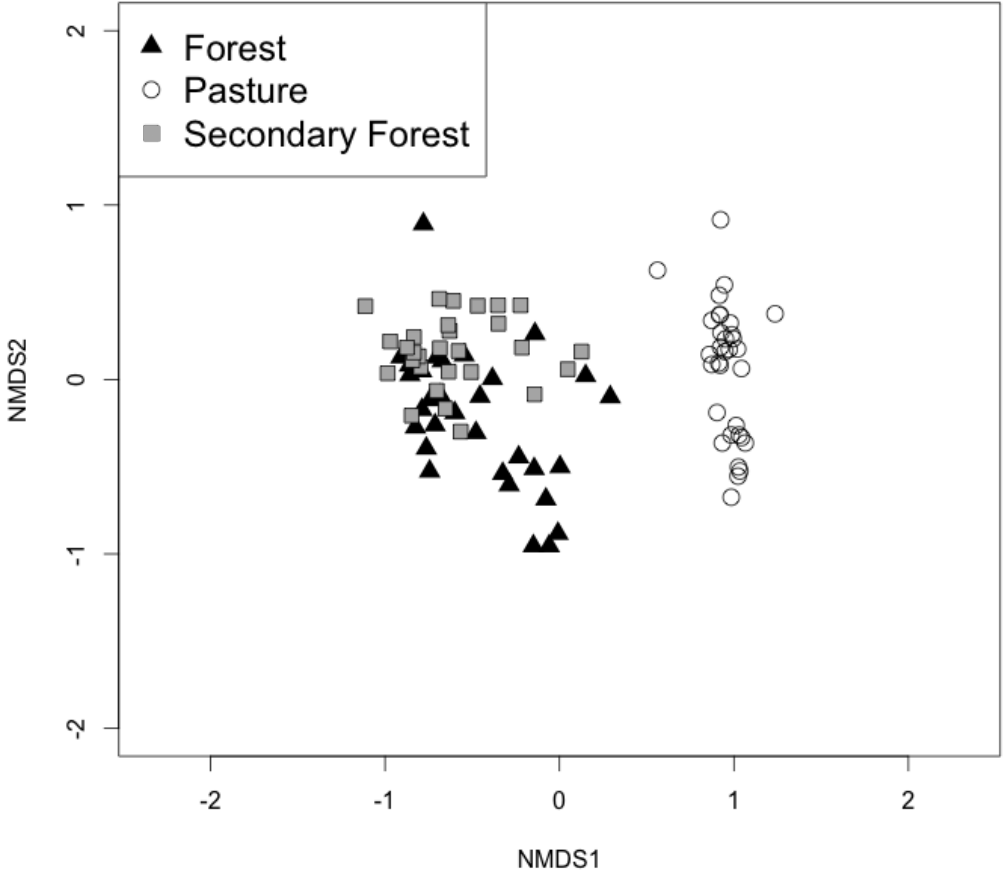


Fig. S3

Analysis of similarity using subsampled data (135,000 *nifH* gene sequences) at 97% DNA similarity



ANOSIM statistic R: 0.7451

Significance: 0.001 Dissimilarity: bray

Fig. S4 Analysis of similarity using complete data (443,000 *nifH* gene sequences) at different DNA and protein similarity levels

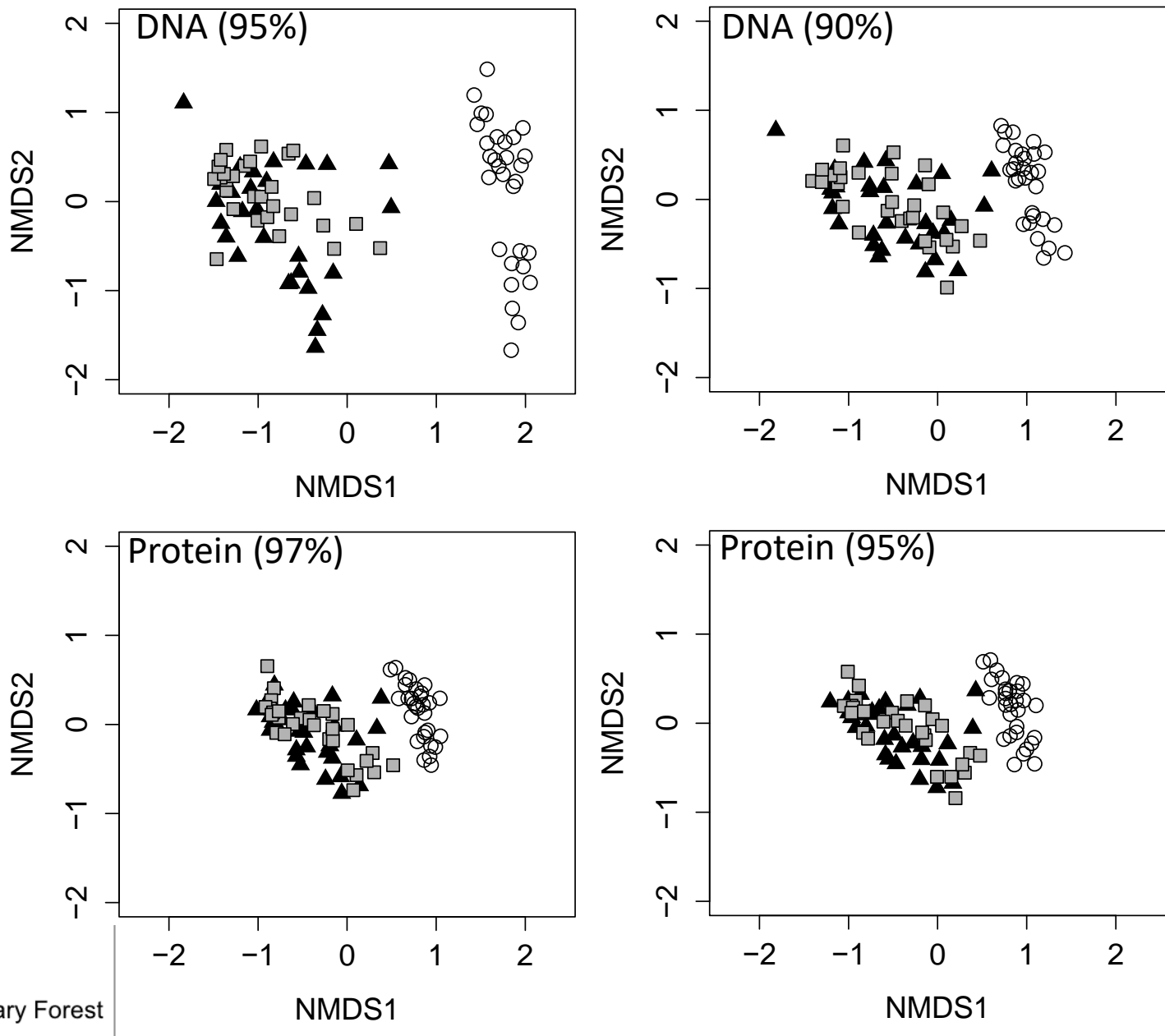


Fig S5: *NifH* gene abundance

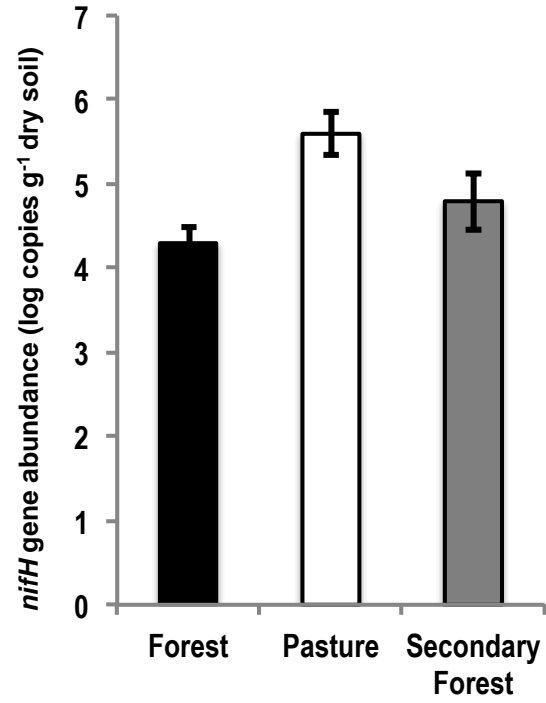
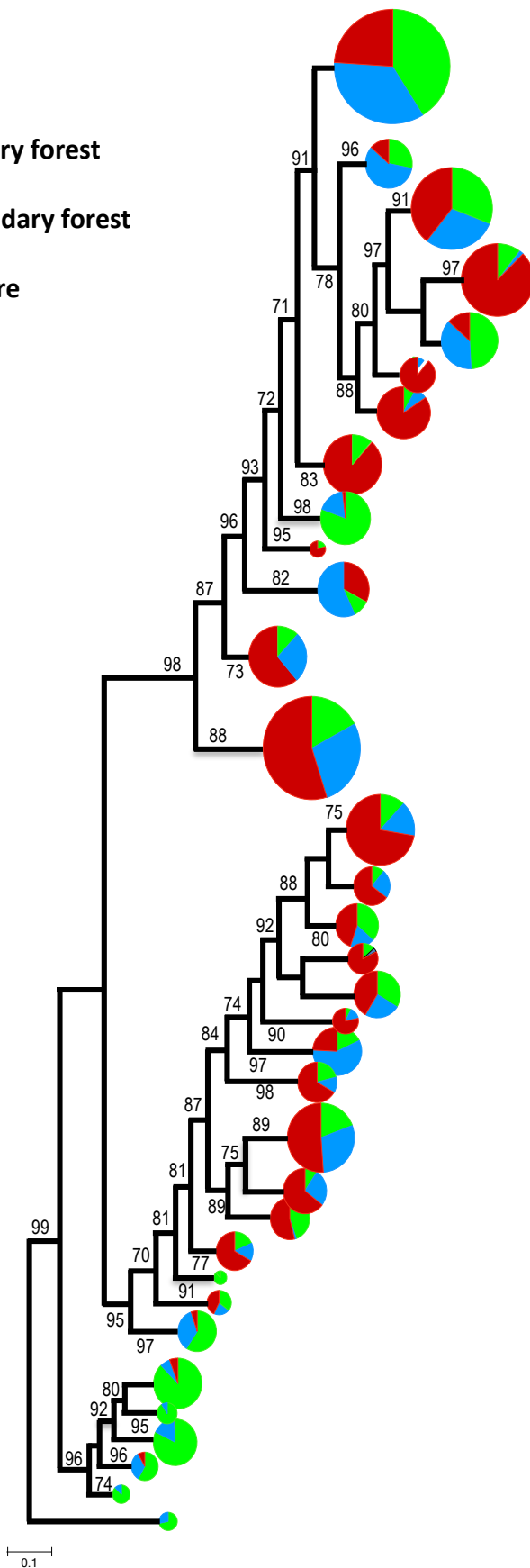


Fig S6: Variation in diazotrophic species



Number of sequences	Taxonomic description	Cluster
267,204	<i>α-proteobacteria</i>	nifH cluster I
551	<i>α-proteobacteria</i>	
17,800	<i>β-proteobacteria</i>	
4,272	<i>γ-proteobacteria</i>	
876	<i>γ-proteobacteria</i>	
112	<i>Proteobacteria</i> (UC)	
638	<i>Proteobacteria</i> (UC)	
1,097	<i>Proteobacteria</i> (UC)	
906	<i>Paenibacillus</i>	
15	<i>Nostoc</i>	
720	<i>Frankia</i>	nifH cluster II
1,379	<i>Firmicutes</i>	
88,108	<i>δ-proteobacteria</i>	
3,800	<i>Verrucomicrobium</i>	
159	<i>Desulfobulbus</i> (δ)	
288	Uncultured	
68	<i>Desulfotomaculum</i>	
432	<i>Spirochetes</i>	
41	Uncultured	
1,164	Uncultured	
168	<i>Spirochetes</i>	nifH cluster III
2,958	Uncultured	
315	<i>δ-proteobacteria</i> (UC)	
185	<i>Bacteroidetes</i>	
159	<i>Spirochetes</i> (<i>treponema</i>)	
15	<i>Clostridia</i>	
41	Uncultured	
232	<i>Clostridia</i>	
647	<i>γ-proteobacteria</i>	
32	<i>α-proteobacteria</i>	
445	<i>Clostridia</i>	
71	<i>Clostridia</i>	
23	<i>Methanomicrobia</i>	
21	<i>Firmicutes</i> (<i>clostridium</i>)	nifH cluster IIV&V

Fig. S7

97% DNA similarity

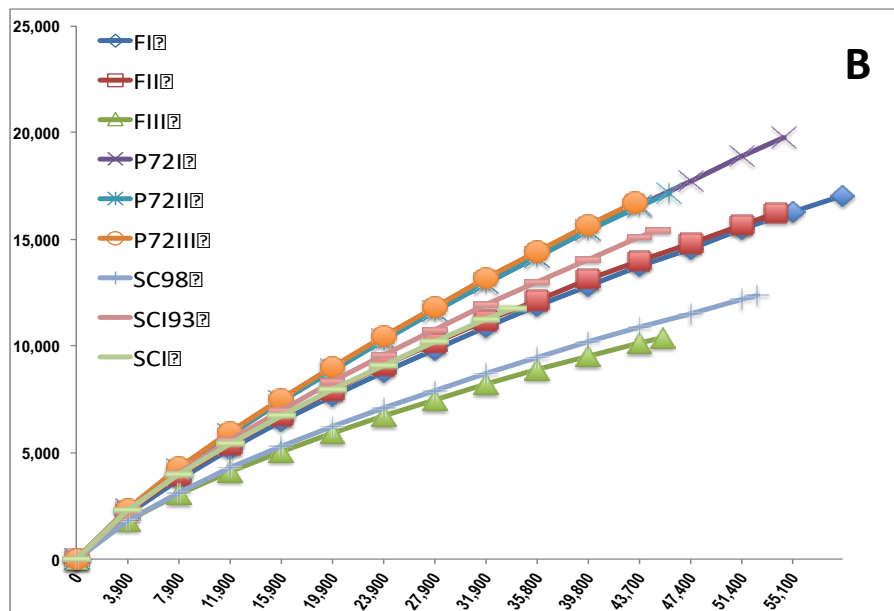
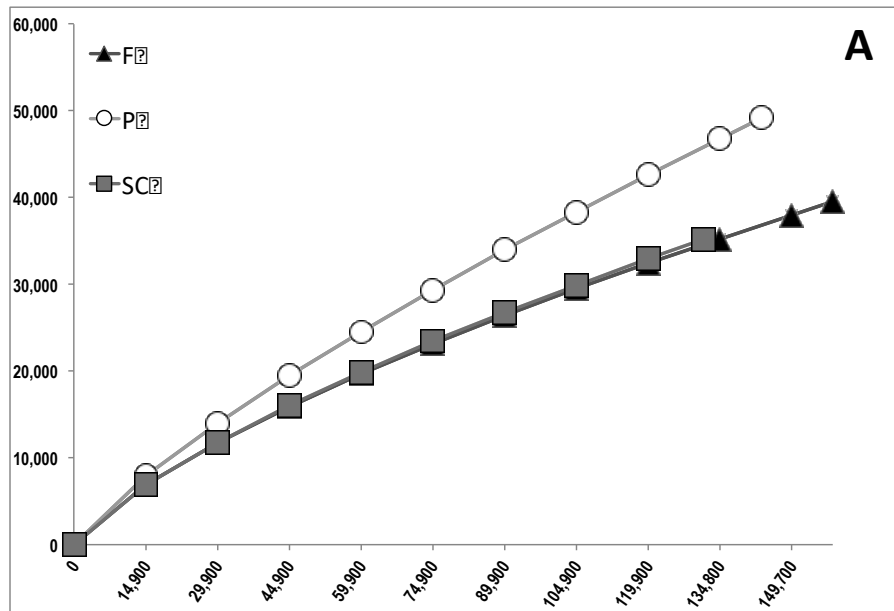


Fig S8: A Spatially Explicit Sampling Scheme

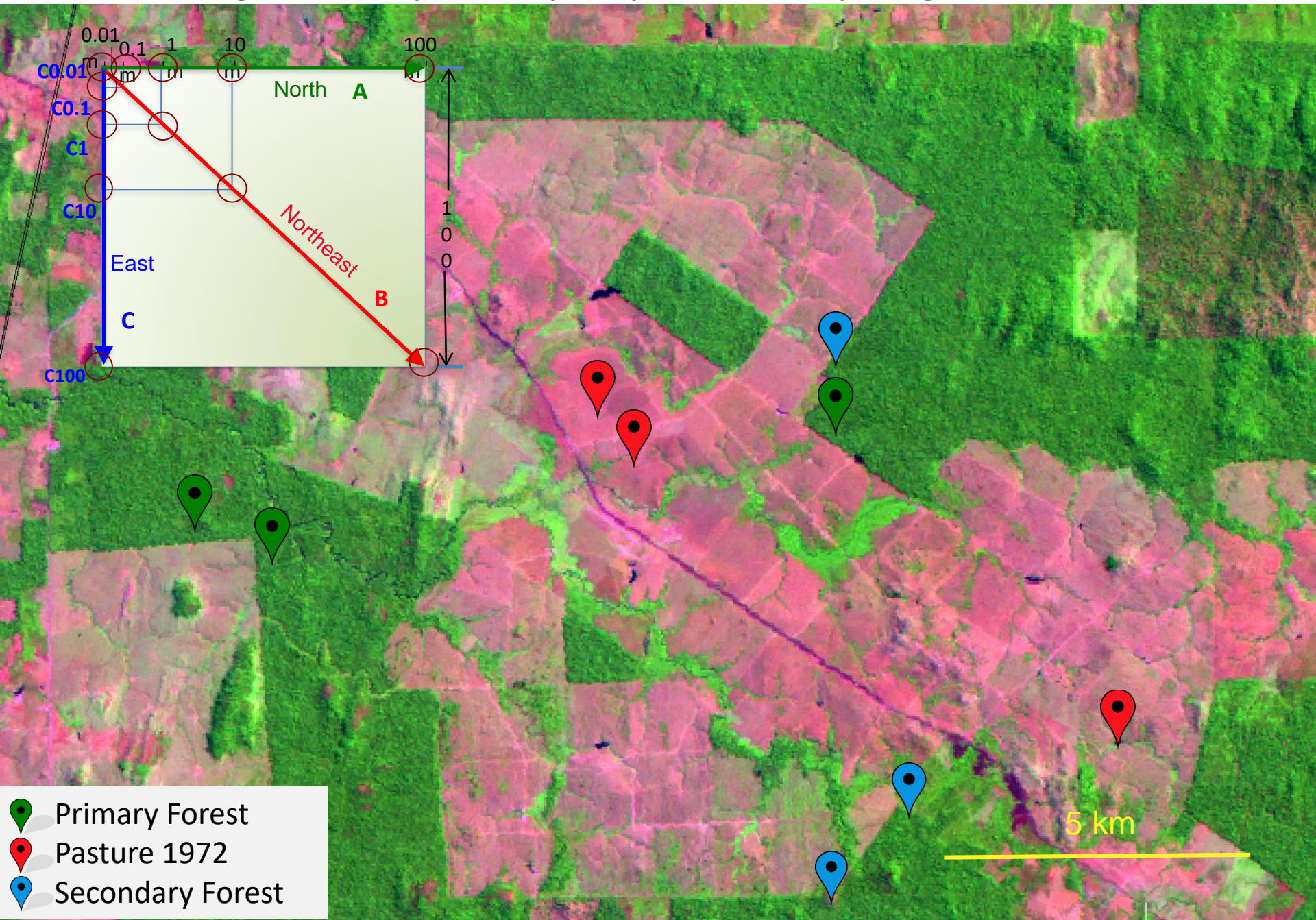


Table S1. Operational protein units (OPUs) and the closest matched sequences obtained through Wang et al., (2013) reference data. OPUs were clustered at 88% similarity.

Cluster	OPU (No sequences)	Accession No.
NifH_typer	<i>Alphaproteobacteria</i> (267,204)	<i>Zymomonas mobilis</i> (ACQU01000014), <i>Magnetospirillum magneticum</i> (AP007255), <i>Methylocystis</i> sp. (AEVM01000015), <i>Novosphingobium nitrogenifigens</i> (326209574), <i>Rhizobium</i> 1 encoding nitrogenase (152287), <i>Bradyrhizobium</i> sp. ORS278 (146195046), <i>Beijerinckia indica</i> (182633352), <i>Azospirillum amazonense</i> Y2 (346917376), <i>Rhizobium</i> sp ANU289 (169750), <i>Hyphomicrobium</i> sp. MC1 (337761014), Unidentified bacterium nif cluster (13435000), <i>Bradyrhizobium japonicum</i> (EF394157), <i>Bradyrhizobium</i> sp. SEMIA (HQ259566), <i>Azospirillum</i> (DQ787332), <i>Methylocapsa aurea</i> (FN433472), <i>Bradyrhizobium yuanmingense</i> (EF394167), uncultured bacterium (EU912866)
	<i>Alphaproteobacteria</i> (551)	<i>Azorhizobium doebereineriae</i> (FJ223129),
	<i>Betaproteobacteria</i> (17,800)	<i>Sideroxydans lithotrophicus</i> ES1 (CP001965), <i>Candidatu Accumulibacter phosphatis</i> (CP001715), <i>Azoarcus</i> sp. BH72 (AF200742), <i>Dechloromonas aromatica</i> RCB (CP000089)
	<i>Gammaproteobacteria</i> (4,272)	<i>Klebsiella pneumoniae</i> 342 (CP000964), <i>Halorhodospira halophila</i> (AB189641)
	<i>Gammaproteobacteria</i> (876)	<i>Thiorhodococcus drewsii</i> (AFWT01000026)
	UC-proteobacteria (111)	Uncultured <i>Proteobacteria</i>
	UC-proteobacteria (636)	Uncultured <i>Proteobacteria</i>
	<i>Alphaproteobacteria</i> (1,099)	<i>Rhodomicrobium vannielii</i> (CP002292)
	<i>Paenibacillus</i> (902)	<i>Paenibacillus graminis</i> (AB485747), <i>Paenibacillus polymyxa</i> (HM146187), <i>Paenibacillus azotofixans</i> (AJ299453), <i>Paenibacillus fujiensis</i> (AB489070), <i>Paenibacillus sabiniae</i> (HM583798)
	Cyanobacteria (15)	<i>Nostoc</i> sp. PCC 7120 (17130219), <i>Nostoc azollae</i> (YP_003720733), <i>Anabaena</i> sp. (NC_003272)
	<i>Frankia</i> (718)	<i>Frankia</i> sp. CcI3 (CP000249), <i>Frankia</i> sp. CeF (AC035776), <i>Frankia</i> sp. Cc1. 17 (ACJ24362)
	<i>Firmicutes</i> (1,374)	<i>Desulfitobacterium hafniense</i> DCB2 (CP001336), <i>Desulfosporosinus orientis</i> DSM765 (CP003108)
	<i>Deltaproteobacteria</i> (88,108)	<i>Geobacter metallireducens</i> GS15 (CP000148), <i>Geobacter bemidjensis</i> (CP001124), <i>Geobacter</i> sp. M18 (ACPJ01000029), <i>Pelobacter propionicus</i> (CP000482), <i>Anaeromyxobacter</i> sp Fw109-5 (CP000769), <i>Anaeromyxobacter</i> sp. K (CP001131), <i>Desulfuromonas acetoxidans</i> DSM 684 (AAEW02000007), <i>Geobacter lovleyi</i> _SZ (CP001089), <i>Geobacter daltonii</i> (YP_002538026), <i>Anaeromyxobacter</i> sp.(YP_001380211), uncultured bacterium (AY601069), uncultured bacterium (AY601072), <i>Geobacter uraniireducens</i> (NC_009483)
	NifH_typerII	<i>Verrucomicrobium</i> (3,800)
<i>Deltaproteobacteria</i> (<i>Desulfobulbus</i>) (158)		<i>Desulfobulbus propionicus</i> DSM 2032 (CP002364)
Uncultured (287)		None
<i>Firmicutes</i> (<i>Desulfotomaculum</i>) (67)		<i>Desulfobacca acetoxidans</i> _DSM_11109 (CP002629)
<i>Spirochetes</i> (431)		<i>Spirochaeta thermophila</i> DSM6192 (CP001698), <i>Syntrophobacter fumaroxidans</i> MPOB (CP000478)
Uncultured (41)		Uncultured methanogenic archaeon_RC-I (AM114193)
Uncultured (1160)		None
<i>Spirochetes</i> (167)		<i>Spirochaeta smaragdinae</i> DSM11293 (CP002116)
Uncultured (2,962)		None
<i>Deltaproteobacteria</i> (314)		<i>Deltaproteobacterium</i> NaphS2 (300447381)
<i>Bacteroidetes</i> (185)		<i>Paludibacter propionis</i> WB4 (CP002345)
<i>Spirochetes</i> (treponema) (158)		<i>Treponema primitia</i> ZAS-2 (CP001843), <i>Treponema azotonutricium</i> ZAS-9 (CP001841)
<i>Clostridia</i> (15)		<i>Clostridium thermocellum</i> (CP000568)
Uncultured 41		None
<i>Clostridia</i> (231)	<i>Clostridium lentocellum</i> DSM 5427 (296249647), <i>Clostridium cellobioparum</i> (U59414), <i>Clostridium papyrosolvans</i> DSM 2782 (ACXX01000015)	
NifH_typerIII	<i>Gammaproteobacteria</i> (645)	<i>Thiorhodococcus drewsii</i> AZ1 (343918744), <i>Dickeya dadantii</i> Ech703 (CP001654)
	<i>Alphaproteobacteria</i> (32)	<i>Zymomonas mobilis</i> subsp pomaceae (CP002865)
	<i>Clostridia</i> (445)	<i>Clostridium pasteurianum</i> (40593), <i>Desulfosporosinus</i> sp OT (344326083), <i>Clostridium</i> sp. (357172598), <i>Chloroherpeton thalassium</i> ATCC 35110 (CP001100)
	<i>Clostridia</i> (70)	<i>Syntrophothermus lipocalidus</i> DSM 12680 (CP002048)
	Methanomicobia (23)	<i>Methanosarcina acetivorans</i> str C2A (19915043), <i>Ethanolins harbinense</i> YUAN-3 (289520023)
NifH_typerIV & V	<i>Firmicutes</i> (<i>clostridia</i>) (21)	<i>Clostridium cellulolyticum</i> H10 (NC_011898), <i>Geobacter</i> sp FRC-32 (CP001390), <i>Syntrophobotulus glycolicus</i> DSM8271 (CP002547), <i>Clostridiales</i> genomosp BVAB3 (CP001850), <i>Clostridium kluyveri</i> DSM 555 (CP000673), <i>Rhodopseudomonas palustris</i> DX1 (283578518)

Table S2: Soil physiochemical characteristics of thee land-use systems in the Amazon.

sample	pH	OM	C	N	C/N	P	S	K	Ca	Mg	Al	H+Al	SB	T	V	m	B	Cu	Fe	Mn	Zn
FIA001	4.8	45	1.586	0.159	9.956	17	10	1.6	48	14	1	22	63.6	85.6	74	2	0.22	0.9	6	61.6	13
FIA1	4.8	45	1.586	0.159	9.956	17	10	1.6	48	14	1	22	63.6	85.6	74	2	0.22	0.9	6	61.6	13
FIA10	5.5	28	3.171	0.293	10.84	9	9	1.3	56	20	0.01	15	77.3	92.3	84	0	0.2	1	50	40.5	3.1
FIA100	4.6	33	5.567	0.401	13.882	11	10	2.6	47	12	1	34	61.6	95.6	64	2	0.21	1.5	45	77.7	5.3
FIB1	5.6	34	1.851	0.156	11.839	9	10	1.4	36	11	0.01	13	48.4	61.4	79	0	0.2	1	98	47.9	3.7
FIB10	4.8	28	2.25	0.18	12.53	10	11	1.6	49	15	1	31	65.6	96.6	68	2	0.2	0.7	46	36.9	2.2
FIB100	4.8	30	1.603	0.152	10.543	9	13	1	31	13	2	34	45	79	57	4	0.15	1.8	115	162.8	5.7
FIC01	5.4	25	2.364	0.193	12.271	6	9	1	16	5	2	18	22	40	55	8	0.2	1.1	104	57.3	2.9
FIC1	5.4	25	2.364	0.193	12.271	6	9	1	16	5	2	18	22	40	55	8	0.2	1.1	104	57.3	2.9
FIC100	4.1	9	4.042	0.353	11.457	22	11	3	130	35	2	22	168	190	88	1	0.26	1.6	39	84.5	8
FIIA001	3.9	16	1.105	0.076	14.497	5	9	0.8	10	3	1	31	13.8	44.8	31	7	0.19	0.9	49	42.7	0.8
FIIA01	3.9	16	1.105	0.076	14.497	5	9	0.8	10	3	1	31	13.8	44.8	31	7	0.19	0.9	49	42.7	0.8
FIIA1	3.9	16	1.105	0.076	14.497	5	9	0.8	10	3	1	31	13.8	44.8	31	7	0.19	0.9	49	42.7	0.8
FIIA100	3.6	17	1.145	0.104	11.028	5	11	0.6	7	2	3	31	9.6	40.6	24	24	0.24	1.9	71	94	1.1
FIIB1	4.5	15	0.781	0.068	11.536	5	9	0.4	16	4	6	18	20.4	38.4	53	23	0.2	0.8	32	34	0.5
FIIB10	4.9	29	3.62	0.195	18.588	8	9	6	20	12	1	16	38	54	70	3	0.22	0.8	39	75.2	1
FIIB100	5.2	36	1.275	0.095	13.422	9	12	1.8	43	10	0.01	18	54.8	72.8	75	0	0.32	1.3	45	73.4	1.3
FIIC01	4.1	15	0.946	0.044	21.464	5	8	1.4	10	3	1	25	14.4	39.4	37	6	0.2	0.8	43	43.4	0.4
FIIC1	4.1	15	0.946	0.044	21.464	5	8	1.4	10	3	1	25	14.4	39.4	37	6	0.2	0.8	43	43.4	0.4
FIIC10	4.6	18	1.245	0.116	10.735	6	8	0.9	17	5	2	18	22.9	40.9	56	8	0.21	0.9	39	83.8	0.5
FIIC100	4.2	19	1.905	0.095	20.064	6	9	0.6	20	3	1	25	23.6	48.6	49	4	0.23	1.2	57	29.5	0.6
FIIAA001	3.9	13	0.658	0.044	14.997	4	8	1	4	2	2	20	7	27	26	22	0.18	0.6	36	26	0.5
FIIAA01	3.9	13	0.658	0.044	14.997	4	8	1	4	2	2	20	7	27	26	22	0.18	0.6	36	26	0.5
FIIAA1	3.9	13	0.658	0.044	14.997	4	8	1	4	2	2	20	7	27	26	22	0.18	0.6	36	26	0.5
FIIAA100	4.4	14	0.306	0.015	20.569	7	9	2.8	7	3	1	11	12.8	23.8	54	7	0.18	0.3	26	12.2	0.7
FIIBB1	3.8	12	0.7	0.043	16.475	5	9	0.6	5	1	1	20	6.6	26.6	25	13	0.2	0.9	45	27.8	0.5
FIIBB10	4.2	16	0.513	0.022	23.825	4	8	0.4	8	1	6	13	9.4	22.4	42	39	0.16	0.7	62	23.7	1
FIIBB100	4.2	8	0.486	0.02	24.41	4	8	0.4	1	1	1	15	2.4	17.4	14	29	0.18	0.1	64	2	0.4
FIIBC01	3.7	10	0.642	0.041	15.827	4	9	0.6	2	1	2	11	3.6	14.6	25	36	0.22	0.8	43	24.1	0.3
FIIBC1	3.7	10	0.642	0.041	15.827	4	9	0.6	2	1	2	11	3.6	14.6	25	36	0.22	0.8	43	24.1	0.3
FIIBC10	3.7	19	0.565	0.024	23.852	6	8	1.2	17	4	7	22	22.2	44.2	50	24	0.18	0.6	7	15.2	1
SC93A01	4.5	28	1.09	0.073	14.917	4	9	1.5	14	8	2	31	23.5	54.5	43	8	0.16	1.6	87	50.9	1.7
SC93A10	4	19	2.183	0.133	16.434	4	8	1.4	6	3	4	31	10.4	41.4	25	28	0.2	1	83	11.2	0.8
SC93A20	3.9	35	1.86	0.14	13.285	8	8	1.8	10	7	6	47	18.8	65.8	29	24	0.18	0.8	133	16	1.3
SC93B1	4.2	28	2.835	0.177	15.999	6	8	1	19	7	1	31	27	58	47	4	0.15	0.8	88	41.5	2.4
SC93B40	3.6	51	1.758	0.134	13.102	9	10	1.1	6	4	13	80	11.1	91.1	12	54	0.16	0.8	191	28.2	4.8
SC93C01	4.1	32	2.438	0.158	15.436	6	9	1.3	16	7	3	42	24.3	66.3	37	11	0.12	0.8	96	64	2.8
SC93C1	4.1	32	2.438	0.158	15.436	6	9	1.3	16	7	3	42	24.3	66.3	37	11	0.12	0.8	96	64	2.8
SC93C10	4.3	17	1.179	0.093	12.723	4	9	4.1	13	7	1	22	24.1	46.1	52	4	0.15	1.1	62	25.2	3.4

SC93C50	4.3	25	2.471	0.139	17.78	5	8	1.5	14	7	2	31	22.5	53.5	42	8	0.18	0.9	109	14.8	26.3
SC98A001	4	19	0.795	0.069	11.5	6	13	0.9	7	3	4	34	10.9	44.9	24	27	0.22	0.7	41	259.8	0.8
SC98A01	4	19	0.795	0.069	11.5	6	13	0.9	7	3	4	34	10.9	44.9	24	27	0.22	0.7	41	259.8	0.8
SC98A1	4	19	0.795	0.069	11.5	6	13	0.9	7	3	4	34	10.9	44.9	24	27	0.22	0.7	41	259.8	0.8
SC98A10	3.7	14	1.005	0.081	12.474	5	14	0.8	3	2	7	34	5.8	39.8	15	55	0.24	0.8	42	205	0.5
SC98A100	4.4	21	0.797	0.056	14.353	5	9	0.8	13	4	1	28	17.8	45.8	39	5	0.16	0.4	44	134.4	0.5
SC98B1	3.9	14	0.766	0.064	12.025	5	17	0.7	4	2	4	31	6.7	37.7	18	37	0.2	0.6	35	223.2	0.4
SC98B10	3.9	15	0.847	0.065	13.087	5	19	0.7	1	1	5	31	2.7	33.7	8	65	0.21	0.5	35	207.4	0.4
SC98B100	4.4	15	1.261	0.083	15.137	3	9	0.8	9	2	1	31	11.8	42.8	28	8	0.16	0.5	41	55	0.3
SC98C01	3.9	11	0.919	0.077	11.872	4	21	0.6	3	1	6	31	4.6	35.6	13	57	0.21	0.6	34	182.4	10.9
SC98C1	3.9	11	0.919	0.077	11.872	4	21	0.6	3	1	6	31	4.6	35.6	13	57	0.21	0.6	34	182.4	10.9
SC98C10	4.3	15	0.843	0.069	12.207	6	25	2.3	8	3	3	25	13.3	38.3	35	18	0.23	0.9	33	189.2	4.7
SC98C100	3.7	11	0.725	0.052	14.062	5	13	0.8	2	1	7	25	3.8	28.8	13	65	0.21	0.7	36	151.4	0.4
SCIA1	4.7	33	1.21	0.11	11.03	6	12	1.3	15	9	1	31	25.3	56.3	45	4	0.17	1.3	69	59.8	6.3
SCIA10	4.4	33	1.68	0.15	11.1	6	12	1.6	20	8	1	34	29.6	63.6	47	3	0.19	1.6	95	82.8	1
SCIA100	3.8	17	1.38	0.12	11.79	5	11	0.5	3	2	5	31	5.5	36.5	15	48	0.2	1.3	62	7.8	0.3
SCIB1	4.6	37	3.6	0.22	16.61	6	11	1.8	25	15	1	25	41.8	66.8	63	2	0.18	1.4	69	34.9	1
SCIB10	4.4	26	2.16	0.16	13.39	6	9	1.1	12	8	1	25	21.1	46.1	46	5	0.18	1.4	59	41.3	1.5
SCIB100	4.1	17	1.62	0.12	13.24	4	12	0.8	7	3	3	42	10.8	52.8	20	22	0.2	1.2	64	13.3	0.3
SCIC1	4.6	43	3.33	0.21	15.98	6	13	1.5	20	13	1	28	34.5	62.5	55	3	0.15	1.3	48	34.2	4.3
SCIC100	4.3	19	1.13	0.1	11.8	5	9	1.7	8	5	1	28	14.7	42.7	34	6	0.2	1	64	65	0.5
P72IA001	4	25	1.2	0.07	17.91	5	8	1.3	9	2	2	15	12.3	27.3	45	14	0.17	1	104	26.2	1
P72IA01	4	25	1.2	0.07	17.91	5	8	1.3	9	2	2	15	12.3	27.3	45	14	0.17	1	104	26.2	1
P72IA1	4	25	1.2	0.07	17.91	5	8	1.3	9	2	2	15	12.3	27.3	45	14	0.17	1	104	26.2	1
P72IA10	4.6	28	1.68	0.11	15.67	4	8	1.5	9	2	2	20	12.5	32.5	38	14	0.17	1.4	71	26.8	1.6
P72IA100	4.6	28	2.21	0.14	16.17	3	8	0.5	16	2	2	22	18.5	40.5	46	10	0.14	1.8	95	44	1.8
P72IB1	4.6	29	1.63	0.11	14.98	3	8	1.4	11	3	1	22	15.4	37.4	41	6	0.14	1.3	105	26.4	1.5
P72IB10	4.6	29	2.16	0.113	19.113	3	8	1.7	10	3	1	22	14.7	36.7	40	6	0.19	1.5	65	30.9	2
P72IB100	4.6	23	1.25	0.072	17.45	3	8	1	9	3	1	22	13	35	37	7	0.21	1	67	33	1.1
P72IC01	4.8	30	1.645	0.107	15.398	4	6	1.9	10	4	1	22	15.9	37.9	42	6	0.18	1	112	26	1.5
P72IC1	4.8	30	1.645	0.107	15.398	4	6	1.9	10	4	1	22	15.9	37.9	42	6	0.18	1	112	26	1.5
P72IC10	4.7	29	1.63	0.096	16.91	6	8	1.3	13	4	1	25	18.3	43.3	42	5	0.18	1.3	87	33.3	2.2
P72IC100	4.5	29	1.755	0.11	16.011	4	8	1	10	3	1	25	14	39	36	7	0.15	0.8	98	26.1	1.6
P72IIA001	4.5	25	2.071	0.135	15.382	3	8	0.7	7	2	2	28	9.7	37.7	26	17	0.14	0.7	47	23.9	0.9
P72IIA01	4.5	25	2.071	0.135	15.382	3	8	0.7	7	2	2	28	9.7	37.7	26	17	0.14	0.7	47	23.9	0.9
P72IIA1	4.5	25	2.071	0.135	15.382	3	8	0.7	7	2	2	28	9.7	37.7	26	17	0.14	0.7	47	23.9	0.9
P72IIB10	4.3	26	1.885	0.141	13.395	3	8	0.8	6	3	2	25	9.8	34.8	28	17	0.17	0.8	65	24.3	1.2
P72IIB100	5	33	2.038	0.128	15.928	4	12	7	13	8	0.01	18	28	46	61	0	0.18	1	96	48	2.1
P72IIC01	4.3	30	1.98	0.133	14.927	3	8	1.2	8	3	3	28	12.2	40.2	30	20	0.18	0.8	57	24.9	1.3
P72IIC10	4.8	44	3.532	0.203	17.387	6	8	1.2	23	5	1	31	29.2	60.2	49	3	0.22	1.5	88	51.2	4
P72IIC100	4.4	33	2.544	0.172	14.834	4	7	1.7	9	4	3	31	14.7	45.7	32	17	0.2	1.3	77	55.8	1.5

P72IIIA01	5.1	44	5.342	0.409	13.061	18	9	3.2	28	7	0.01	31	38.2	69.2	55	0	0.2	5.8	198	222	14
P72IIIA1	5.1	44	5.342	0.409	13.061	18	9	3.2	28	7	0.01	31	38.2	69.2	55	0	0.2	5.8	198	222	14
P72IIIA10	5.3	54	5.245	0.424	12.365	16	9	4.3	39	12	0.01	28	55.3	83.3	66	0	0.22	5.8	142	203	16.4
P72IIIA100	5.1	49	5.186	0.452	11.474	10	9	3.5	27	7	0.01	31	37.5	68.5	55	0	0.22	5.6	70	193.4	9.4
P72IIIB1	5.2	41	2.495	0.219	11.4	8	9	1.7	25	4	0.01	28	30.7	58.7	52	0	0.2	5.1	103	184.6	9.6
P72IIIB100	5.3	50	7.476	0.581	12.873	29	15	4.6	33	14	0.01	28	51.6	79.6	65	0	0.22	5.4	128	171.6	12.3
P72IIIC01	5.2	62	6.446	0.47	13.724	15	9	2.4	30	9	0.01	34	41.4	75.4	55	0	0.2	5.7	131	163.4	15.5
P72IIIC1	5.2	62	6.446	0.47	13.724	15	9	2.4	30	9	0.01	34	41.4	75.4	55	0	0.2	5.7	131	163.4	15.5
P72IIIC10	5.5	43	3.604	0.268	13.43	19	9	1.4	35	9	0.01	20	45.4	65.4	69	0	0.18	4.2	59	165.8	12.2
P72IIIC100	5.3	50	3.043	0.276	11.045	11	13	1.7	23	8	0.01	31	32.7	63.7	51	0	0.21	5	255	174.8	7.9

Measured soil variables: OM, organic matter; C, carbon; N, nitrogen; pH, potential activity of hydrogen; P, phosphorus; S, sulfur; K, potassium; Ca, calcium; Mg, magnesium; Al, aluminum; H⁺, hydrogen; CEC, potential cation exchange capacity; V, base saturation; Al⁺ sat, aluminum saturation. C and N are expressed in g.Kg⁻¹ soil; P and S are expressed as mg dm⁻³; K, Ca, Mg, Al and CEC are expressed in mmol c.dm⁻³.

List of the 19 soil samples produced <1500 nifH gene sequences were not included in this study. Overall 89 soil samples were analyzed across three different land-use systems.

FIA01, FIC10, FIA10, FIIIA10, FIIC100
P72IB1, P72IIA10, P72IIA100, P72IIB1, P72C1, P72IIIA001, P72IIIB10
SC98B1, SCI93A001, SCI93B10, SCIC10,
SCI93A40, SCIA001, SCIA01