

Supplementary Materials

Histone methyltransferase G9a protects against acute liver injury through GSTP1

Short title: G9a-GSTP1/2 axis in acute liver injury

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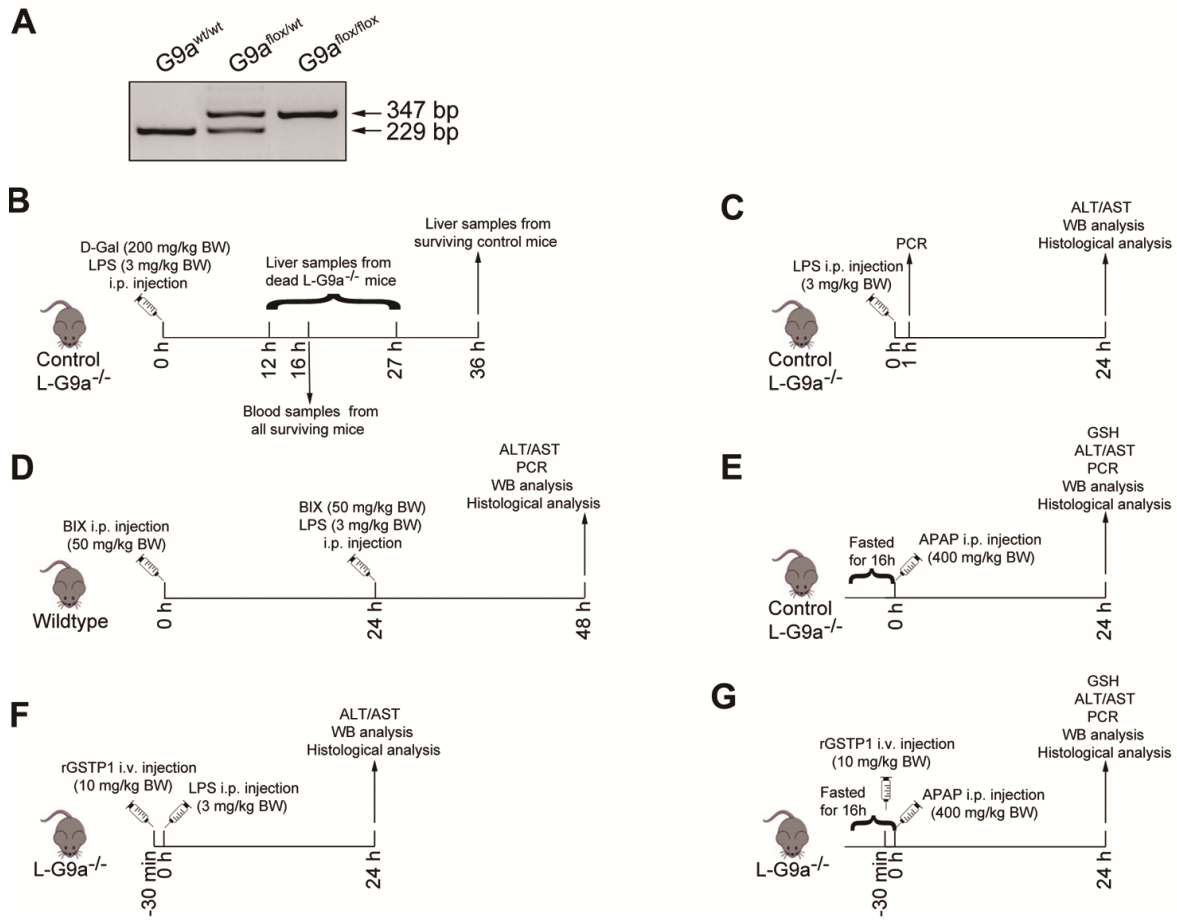
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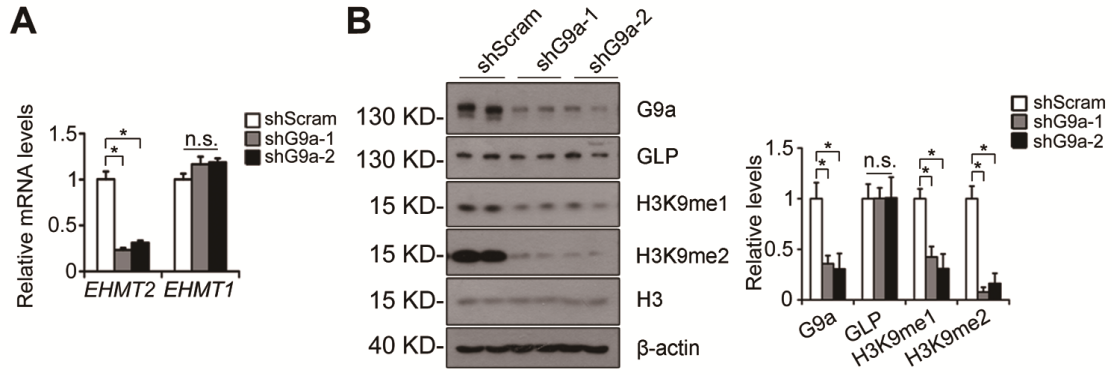
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This supplementary information contains thirteen figures, one table and three videos.



23

24 **Fig. S1. Experimental design.** (A) Genotype analysis by using genomic DNA extracted from the tails
 25 to demonstrated three genotypes of mouse. (B-G) Experimental design flowcharts of LPS/D-Gal-
 26 induced acute liver injury (B), LPS-induced acute liver injury (C), BIX and LPS co-treatment (D),
 27 APAP-induced acute liver injury (E), rGSTP1 treatment for L-G9a^{-/-} mice under LPS injection (F) and
 28 APAP injection (G). BW, body weight.



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30 **Fig. S2. Downregulation of G9a does not affect the expression level of GLP in HepG2 cells and in**

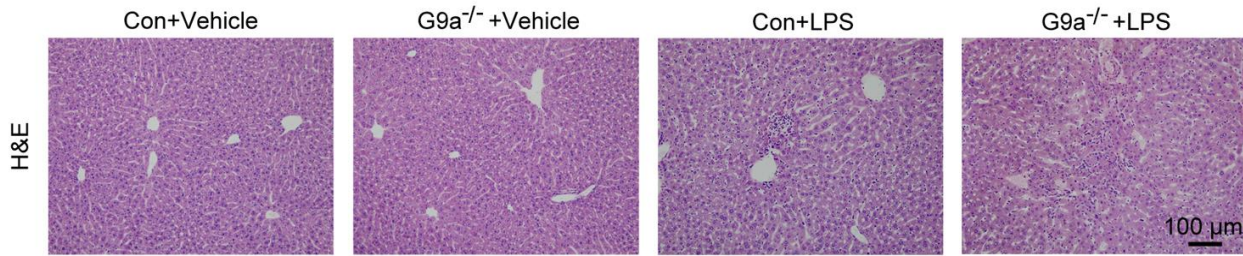
31 **the livers after LPS treatment. (A) Relative mRNA level of *EHMT1/2* in control or two different**

32 **shG9a stable HepG2 cell lines. (B) Representative western blots (*left*) and the quantitative results (*right*)**

33 **for G9a, GLP and H3K9me1/me2 in the control or two different shG9a stable HepG2 cells. H3 and β -**

34 **actin serve as the loading control. n=6; * $p < 0.05$.**

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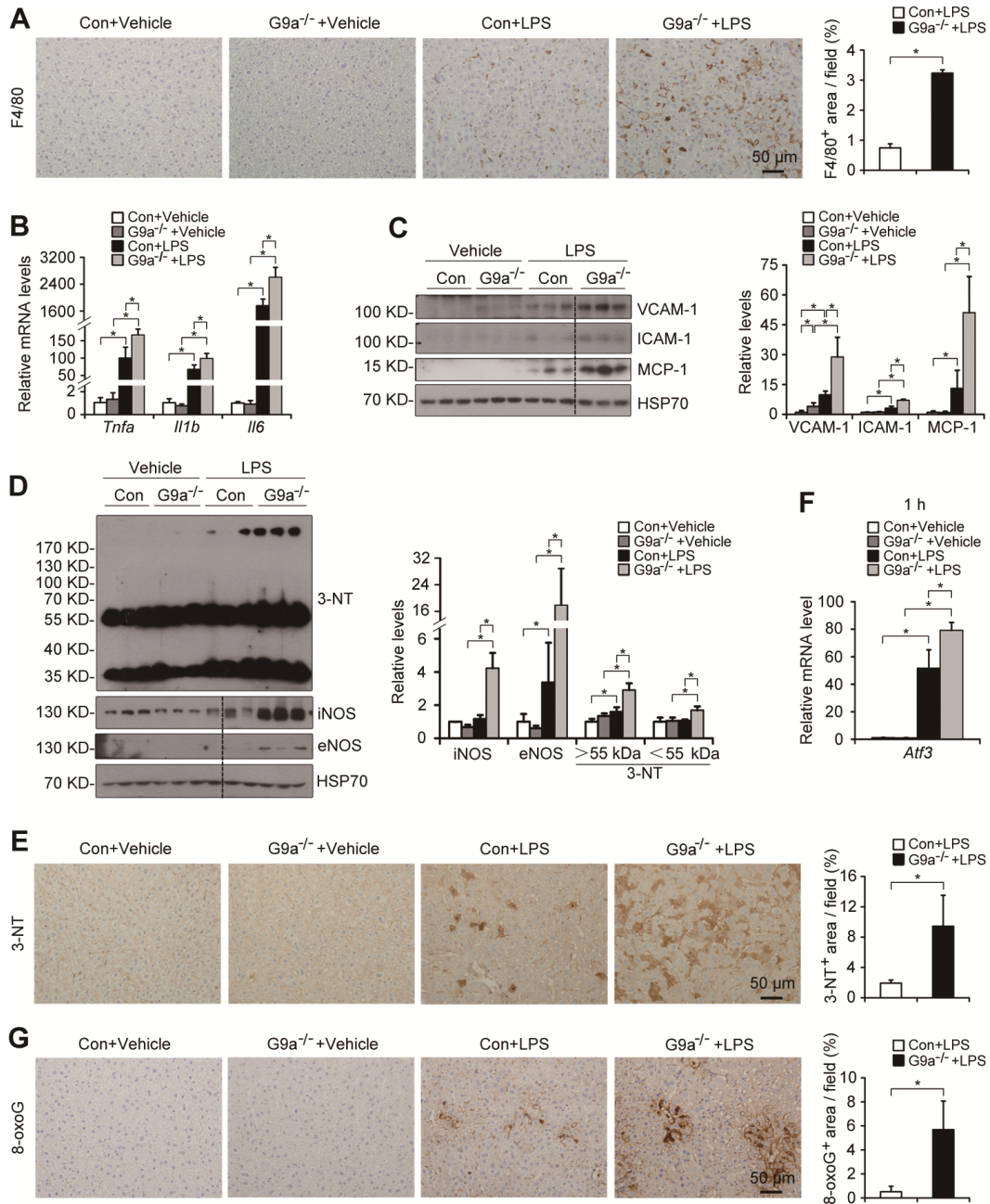


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37 **Fig. S3. Hepatic G9a deletion aggravates LPS-induced liver damage in male mice.** H&E staining in

38 livers after LPS injection. n=3-4 per group; scale bar, 100 μm.

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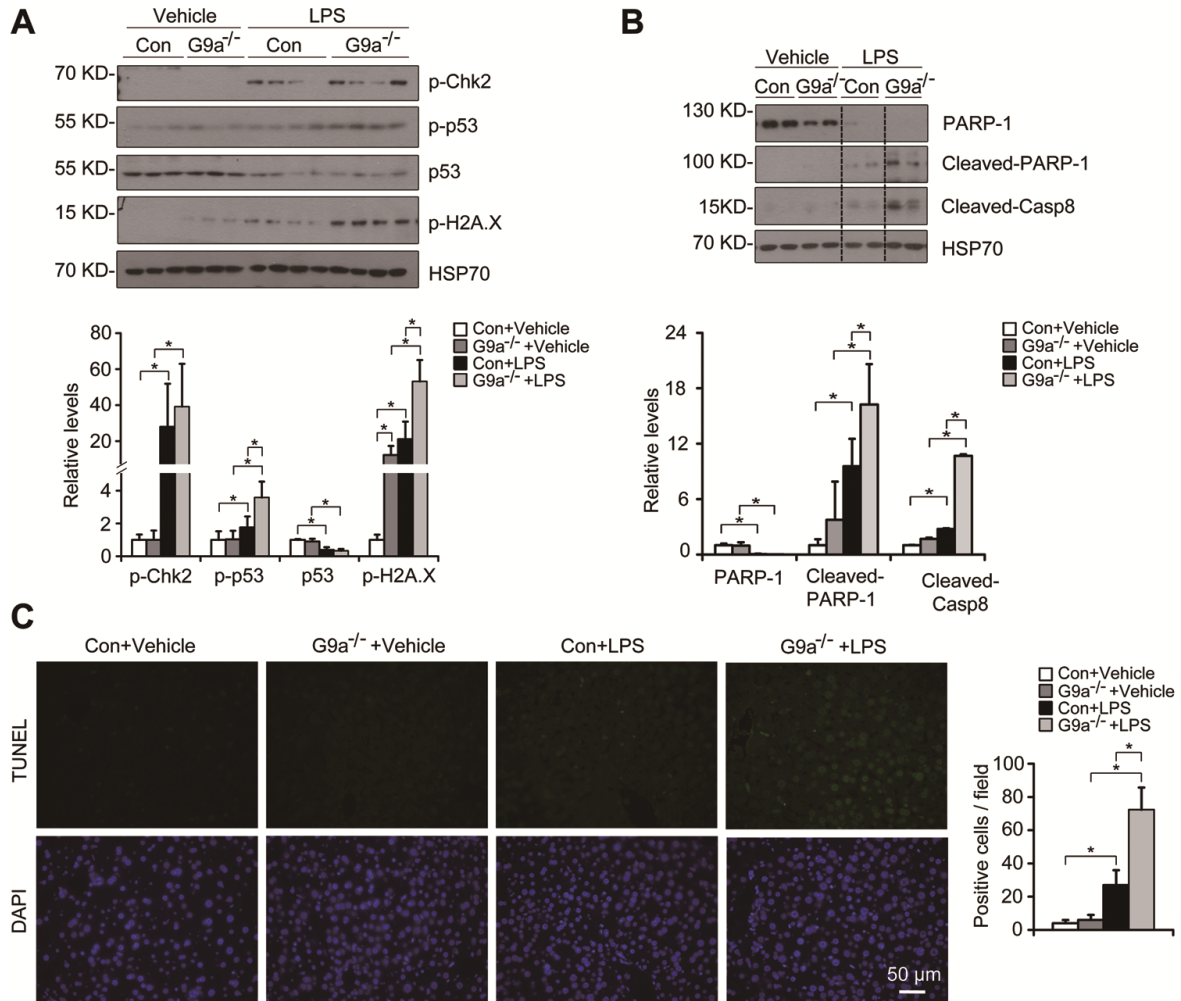


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41 **Fig. S4. Hepatic G9a deletion augments LPS-induced inflammation and RNS/ROS production in**
 42 **female mice.** (A) Representative pictures for F4/80 staining in livers (*left*) and quantitative results
 43 (*right*). (B) qPCR results of the indicated genes in livers of different groups. (C-D) Representative
 44 western blots (*left*) and quantitative results (*right*) for VCAM-1, ICAM-1 and MCP-1 (C), iNOS, eNOS
 45 and 3-NT (D) in livers of different groups. HSP70 serves as the loading control. (E) Representative

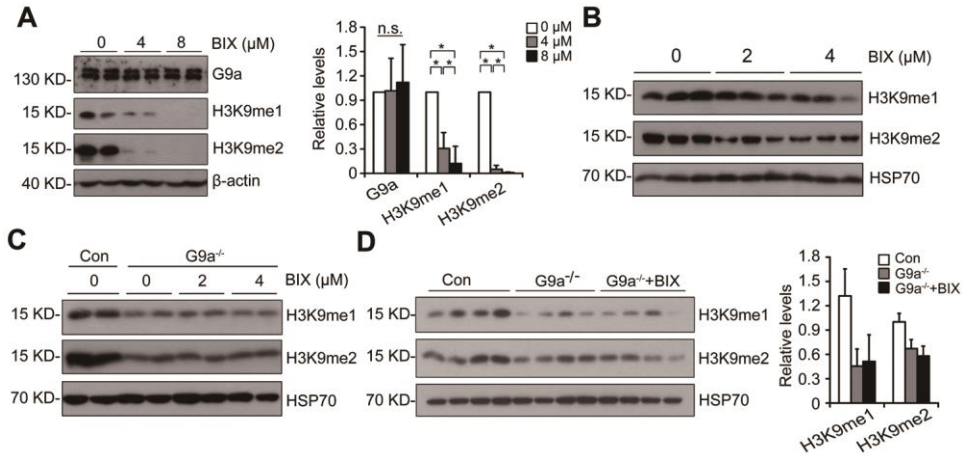
46 pictures for 3-NT staining in livers (*left*) and quantitative results (*right*). (F) Relative mRNA level of
47 *Atf3* in the livers at 1 hour after LPS injection. (G) Representative pictures for 8-oxoG staining in livers
48 (*left*) and quantitative results (*right*). n=3-4 per group; 3-NT, 3-Nitrotyrosine; * $p < 0.05$; scale bar, 50
49 μm .

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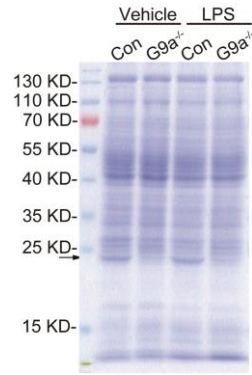
52 **Fig. S5. Hepatic G9a deletion augments DNA damage and hepatocyte apoptosis upon LPS**
 53 **injection in female mice.** (A-B) Representative western blots (*up*) and quantitative results (*down*) for
 54 markers of DNA damage response signaling (A) and apoptosis (B) in livers of different groups. HSP70
 55 serves as the loading control. (C) Representative pictures for TUNEL staining in the livers (*left*) and
 56 quantitative results (*right*). n=3-4 per group; **p* < 0.05; scale bar, 50 μ m.



57

58 **Fig. S6. BIX01294 (BIX) does not further inhibit H3K9me1/me2 levels in the G9a-deficient**
 59 **primary hepatocytes or livers. (A)** Representative western blots (*left*) and quantitative results (*right*)
 60 for G9a, H3K9me1/me2 in HepG2 cells with indicated dosages of BIX. **(B-C)** Representative western
 61 blots for H3K9me1/me2 in primary hepatocytes isolated from the control mice (*B*), as well as the control
 62 and L-G9a^{-/-} mice (*C*) treated with indicated dosages of BIX. **(D)** Representative western blots for
 63 H3K9me1/me2 in livers of the control mice, L-G9a^{-/-} mice and L-G9a^{-/-} mice treated with BIX. β-actin
 64 or HSP70 serves as the loading control. n=6; n.s., not significant; * *p* < 0.05.

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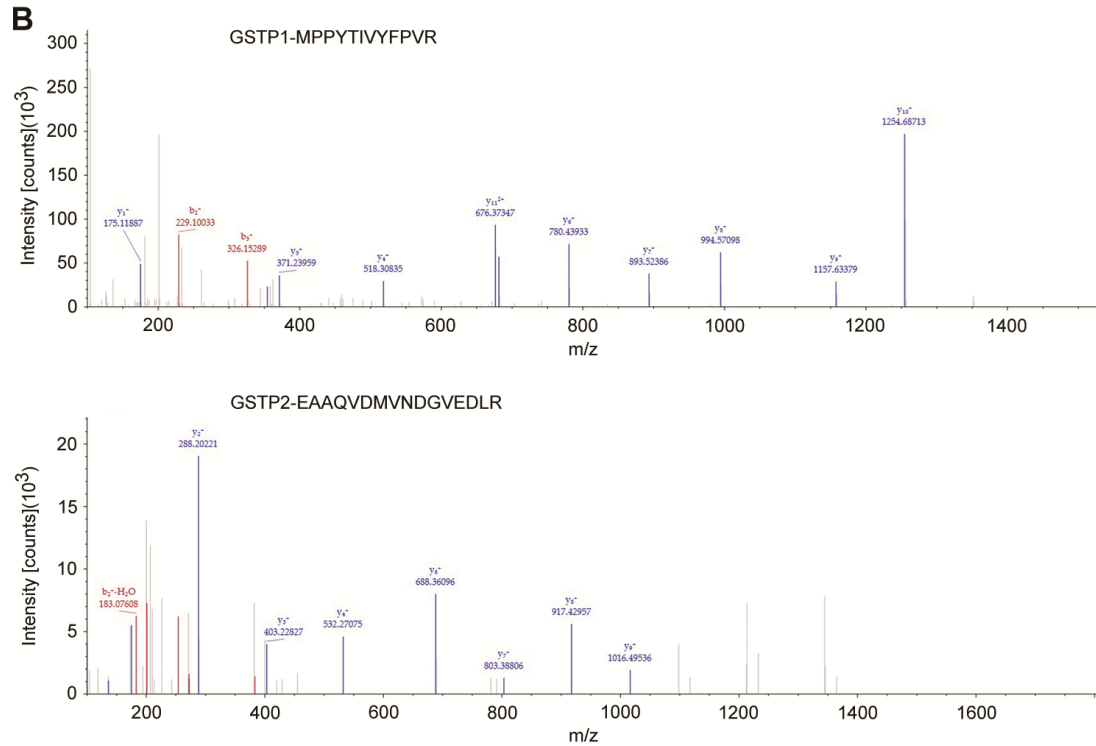


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67 **Fig. S7. The protein expression profile in the liver of different groups.** Coomassie brilliant blue
68 staining of whole liver extracts from control and L-G9a^{-/-} mice with or without LPS i.p. injection. n=4
69 per group.

A

Name	MW (kDa)	Wildtype			L-G9a ^{-/-}			Δ PSMs
		Coverage	Unique Peptides	PSMs	Coverage	Unique Peptides	PSMs	
GSTP1	23.6	62.86	8	884	59.05	8	523	361
GSTP2	23.5	48.57	2	398	44.76	1	273	125
GSTA4	25.5	30.18	8	51	30.18	8	35	16
MGST1	17.5	18.06	2	22	7.74	1	9	13
CATB	37.3	13.57	5	30	15.93	5	20	10
SPRE	27.9	22.99	5	10	3.83	1	1	9



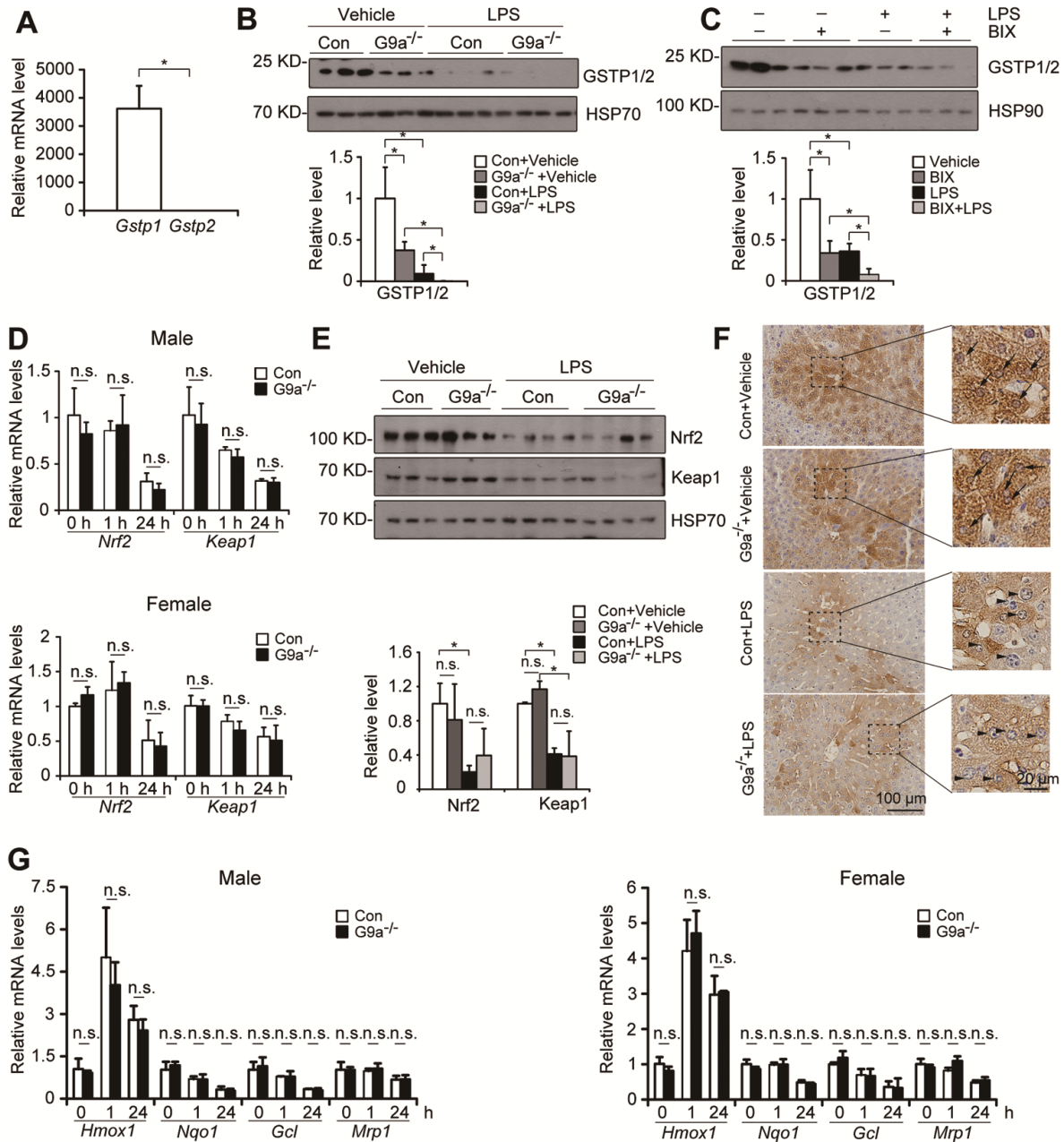
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71 **Fig. S8. Identification of GSTP1 and GSTP2 by mass spectrometry. (A)** Top 6 candidate proteins by

72 analyzing Δ PSM (wildtype - L-G9a^{-/-}). **(B)** Unique peptides of GSTP1 and GSTP2 identified by MS.

73 GSTA4, glutathione S-transferase A4; MGST1, microsomal glutathione S-transferase 1; CATB,

74 cathepsin B; SPRE, sepiapterin reductase.

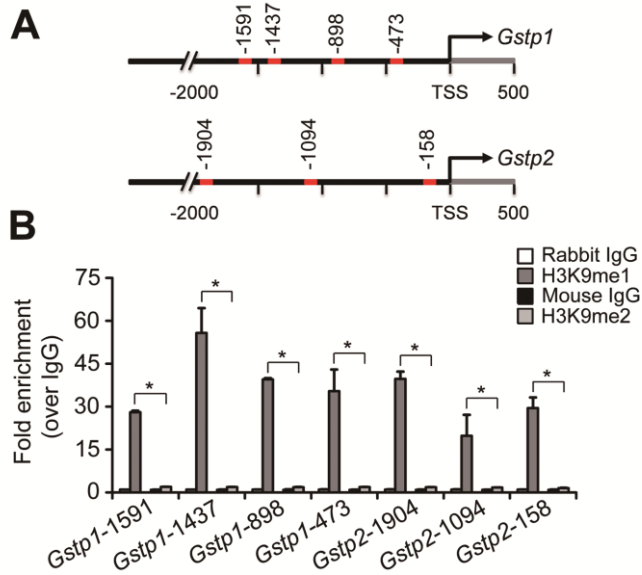


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76 **Fig. S9. G9a-deficiency-caused downregulation of GSTP1/2 is not regulated by Nrf2.** (A) Relative
 77 mRNA levels of *Gstp1* and *Gstp2* in livers of the wildtype male mice. (B-C) Representative western
 78 blots (up) and quantitative results (down) for GSTP1/2 in livers upon LPS injury in female mice (B) and
 79 BIX with LPS co-treatment in male mice (C). (D) Relative mRNA levels of *Nrf2* and *Keap1* in livers of
 80 the male (up) and female (down) control and L-*G9a*^{-/-} mice at 0, 1 or 24 hours after LPS injection. (E)

81 Representative western blots (*up*) and quantitative results (*down*) for Nrf2 and Keap1 in livers of the
82 control and L-G9a^{-/-} male mice at 24 hours after LPS injection. (F) Representative pictures of Nrf2
83 staining in the livers of different groups. Nucleus with or without Nrf2 staining were marked by arrows
84 or arrowheads, respectively. (G) Relative mRNA levels of *Hmox1*, *Nqo1*, *Gcl* and *Mrp1* in livers of the
85 control and L-G9a^{-/-} mice at 0, 1 or 24 hours after LPS injection. HSP70 or HSP90 serves as the loading
86 control. n=3-4 per group; n.s., not significant; **p* < 0.05.

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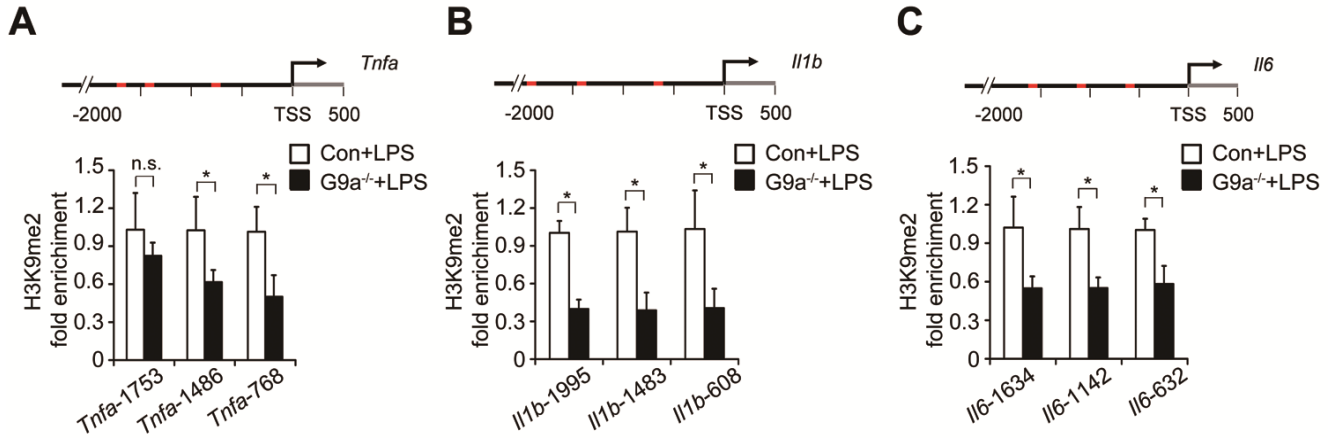
89 **Fig. S10. H3K9me1/me2 levels at promoters of *Gstp1/2* in the livers of wildtype mice. (A)** Locations

90 of primers used for ChIP-qPCR. **(B)** ChIP-qPCR analysis of H3K9me1 and H3K9me2 levels at the

91 various regions of promoters of *Gstp1* and *Gstp2* in livers of control mice. n=3 per group; * $p < 0.05$.

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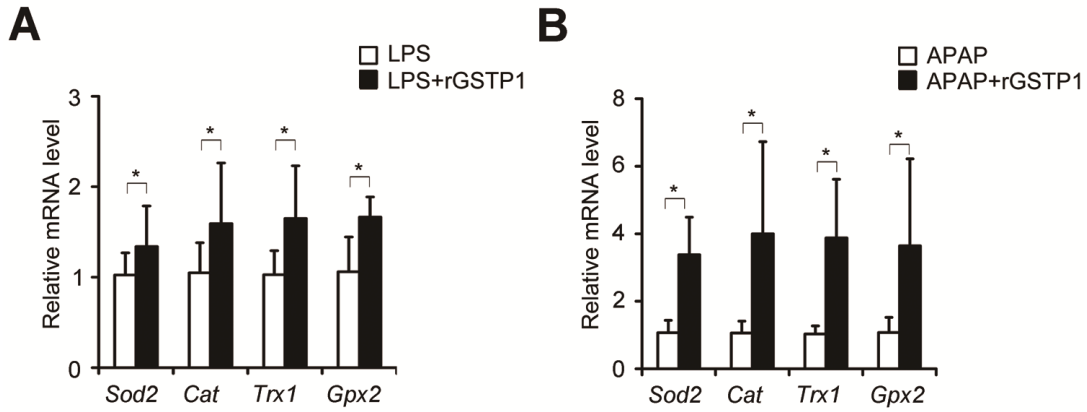
95 **Fig. S11. H3K9me2 levels on the promoters of inflammatory genes in the livers of the control and**

96 **L-G9a^{-/-} mice upon LPS injection.** (A-C) Locations of primers used for ChIP-qPCR (*up*) and the ChIP-

97 qPCR analysis of the H3K9me2 at various promoter regions of inflammatory genes (*down*) such as *Tnfa*

98 (*A*), *Il1b* (*B*) and *Il6* (*C*). n=4 per group; n.s., not significant; **p* < 0.05.

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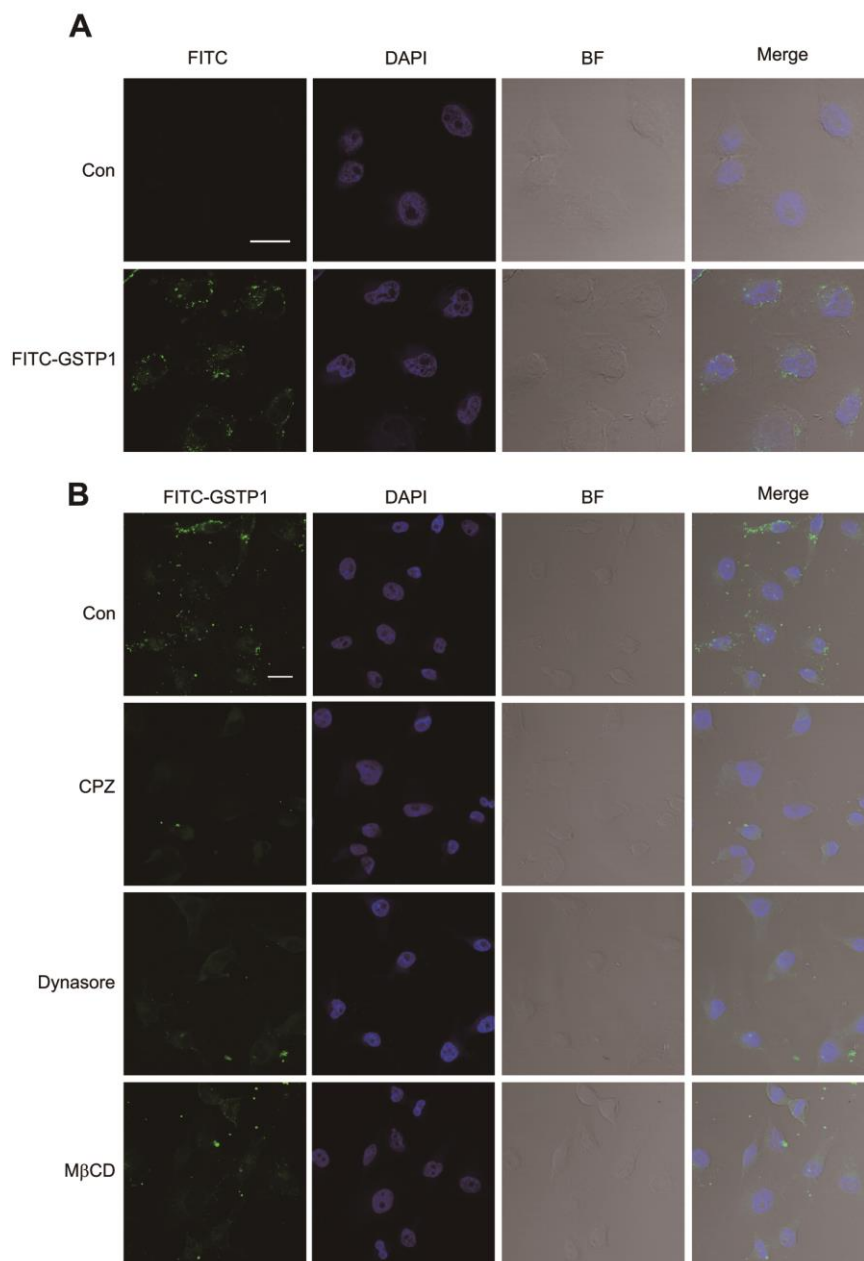
101 **Fig. S12. rGSTP1 upregulates redox and detoxification genes in L-G9a^{-/-} mice upon LPS- or**

102 **APAP- induced liver damage. (A-B) Relative mRNA level of some redox and detoxification genes in**

103 **the livers of indicated groups upon LPS- (A) or APAP- (B) stress. All these data were obtained from**

104 **male L-G9a^{-/-} mice. n=4-6 per group; * $p < 0.05$.**

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106

107 **Fig. S13. rGSTP1 may be transported into cells by clathrin-dependent endocytosis. (A)**

108 Representative pictures of the transport of FITC-labeled rGSTP1 into Hepa1-6 cells. (B) Representative

109 pictures of the intracellular localization of FITC-labeled rGSTP1 in Hepa1-6 cells after the treatment of

110 different endocytosis inhibitors. Scale bar, 20 μ m.

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Table S1 Information of materials used in the present study

Antibodies		
Antibodies	Vendor	Catalog number
G9a	Abcam (for WB)	31874
G9a	Abcam (for ChIP)	185050
GLP	Bioss	16789
H3K9me2	Abcam	1220
H3K9me1	Abcam	9045
Histone H3	Cell Signaling	9715
HSP70	BD Biosciences	03719
HSP90	BD Biosciences	03717
VCAM-1	Santa Cruz	8304
ICAM-1	Santa Cruz	1511
MCP-1	Santa Cruz	1785
iNOS	BD Biosciences	610431
eNOS	BD Biosciences	610298
3-Nitrotyrosine	Millipore	MAB5404
p-Chk2	Cell Signaling	2197
p-Chk1	Cell Signaling	2348
p-p53	Cell Signaling	9284
p53	Santa Cruz Bio	99
p-H2A.X	Cell Signaling	9718
PARP-1	ENZO Life Sciences	ALX-804-211
Caspase3	Cell Signaling	9662
Caspase8	Beyotime	AC056

GSTP1	Santa Cruz	28494
Nrf2	Santa Cruz (for WB)	365949
Nrf2	Cell Signaling (for IHP)	12721
Keap1	Santa Cruz	33569
F4/80	Santa Cruz	52664
Ly-6G	BD Biosciences	551459
CD3	Gene Tech	GA045229
8-oxoG	Santa Cruz	130914

qPCR primers

Genes	Species	Sequences
<i>Ehmt2</i>	<i>Mouse</i>	Forward: TCGGGCAATCAGTCAGACAG Reverse: TGAGGAACCCACACCATTAC
<i>Ehmt1</i>	<i>Mouse</i>	Forward: GAACAGGAGTCTCCCGACAC Reverse: GGGCTGTCAGTCTTCCCTC
<i>Tnfa</i>	<i>Mouse</i>	Forward: GGTGATCGGTCCCCAAAGGGATGA Reverse: TGGTTTGCTACGACGTGGGCT
<i>Il1b</i>	<i>Mouse</i>	Forward: GAAGAAGAGCCCATCCTCTG Reverse: TCATCTCGGAGCCTGTAGTG
<i>Il6</i>	<i>Mouse</i>	Forward: CACTTCACAAGTCGGAGGCT Reverse: CTGCAAGTGCATCATCGTTGT
<i>Mcp-1</i>	<i>Mouse</i>	Forward: GCTCTCTCTCCTCCACCAC Reverse: GCGTTAACTGCATCTGGCT
<i>Atf3</i>	<i>Mouse</i>	Forward: CTCTCACCTCCTGGGTCCT Reverse: TCTGGATGGCGAATCTCAGC
<i>Cox2</i>	<i>Mouse</i>	Forward: CAACACCTGAGCGGTTACCA

		Reverse: TTCAGAGGCAATGCGGTTCT
<i>Hmox1</i>	<i>Mouse</i>	Forward: CCTCACAGATGGCGTCACTT
		Reverse: GCTGATCTGGGGTTTCCCTC
<i>Gstp1</i>	<i>Mouse</i>	Forward: GCGGCAAATATGTCACCCTCAT
		Reverse: GCAGGGTCTCAAAGGCTTC
<i>Gstp2</i>	<i>Mouse</i>	Forward: ATATGGCACCATGATCTACAGAAA
		Reverse: TCTGGTCACCCACGATGAAA
<i>Nrf2</i>	<i>Mouse</i>	Forward: CCTCTGTCACCAGCTCAAGG
		Reverse: TTGATGACCAGGACTCACGG
<i>Keap1</i>	<i>Mouse</i>	Forward: GTGGCACCTACAGAGACACC
		Reverse: GCGCTTGGAGAAGGGCA
<i>Nqo1</i>	<i>Mouse</i>	Forward: CATTGCAGTGGTTTGGGGTG
		Reverse: TCTGGAAAGGACCGTTGTCTG
<i>Gcl</i>	<i>Mouse</i>	Forward: AGCTTTGGGTCGCAAGTAGG
		Reverse: GGTCTCGCGCTCCCAC
<i>Mrp1</i>	<i>Mouse</i>	Forward: CAAAGCCGGTGGAAAATGGG
		Reverse: GTGGGAAGACGAGTTGCTGA
<i>Sod2</i>	<i>Mouse</i>	Forward: TAACGCGCAGATCATGCAGCTG
		Reverse: AGGCTGAAGAGCGACCTGAGTT
<i>Cat</i>	<i>Mouse</i>	Forward: TGAGAAGCCTAAGAACGCAATTC
		Reverse: CCCTTCGCAGCCATGTG
<i>Trx1</i>	<i>Mouse</i>	Forward: GAAATGCATGCCGACCTTCCAG
		Reverse: GCTGGTTACACTTTTCAGAGCATG
<i>Gpx2</i>	<i>Mouse</i>	Forward: TGTGACGTCAATGGGCAGAA
		Reverse: GTCATGAGGGAGAACGGGTC

<i>β-actin</i>	<i>Mouse</i>	Forward: GGCTACAGCTTCACCACCAC Reverse: TAGCTCTTCTCCAGGGAGGA
ChIP-qPCR primers		
Name	promoter region (+1 for TSS)	Sequences
<i>Gstp1-1591</i>	-1591 to -1506	Forward: AAGGGGGCCCCATGTTGA Reverse: ATCCAAAGCCAGGGAATTAGCTCTG
<i>Gstp1-1437</i>	-1437 to -1344	Forward: ACCTTGCTGGTGATACGTGAG Reverse: CTCCTCGGTGTCTTACTTACTCTG
<i>Gstp1-898</i>	-898 to -751	Forward: TGGCCTGTTTAAAGGTGGAGAA Reverse: TGTGAGAATAGGGACAAATCTTGAG
<i>Gstp1-473</i>	-473 to -375	Forward: TCCAACCCTGTTAGTCTCGTTT Reverse: CCTTCGCCAGGAGTAATCAG
<i>Gstp2-1904</i>	-1904 to -1832	Forward: GCCCAGAACCTCAGTGATGAA Reverse: TGCACTGTAGCGTCACATTCT
<i>Gstp2-1094</i>	-1094 to -1014	Forward: TGCGGGAGGAAAGAGTGAGA Reverse: GTGTCTCCCTGGGTAATGGG
<i>Gstp2-158</i>	-158 to -40	Forward: ACATGCTAGTGTCTCGGGC Reverse: TATAAGGGGCATTGCGCTC
<i>Tnfa-1753</i>	-1753 to -1669	Forward: TTCTTTGGAGCCTTTGCACT Reverse: ACCCTTGAAACAACGGTCAG
<i>Tnfa-1486</i>	-1486 to -1329	Forward: GGACATGAAGGAGGGACAGA Reverse: CACCTTTGTGTCTGGGACCT
<i>Tnfa-768</i>	-768 to -523	Forward: CGCAGTCAAGATATGGCAGA Reverse: CTTGGAGGAAGTGGCTGAAG

<i>Il1b-1995</i>	-1995 to -1823	Forward: CTCCAAATCCTCCCAGACAA Reverse: AAGGGTAACTAGGGGCCTGA
<i>Il1b-1483</i>	-1483 to -1431	Forward: ATAGCTGGTCAAAGGCAGGA Reverse: GCATCTCGATTTCAGGAAGG
<i>Il1b-608</i>	-608 to -512	Forward: AAGCACCTGGCATTATCAG Reverse: AGGGCACACACAGTATGCAG
<i>Il6-1634</i>	-1634 to -1502	Forward: CACACGGTGAAAGAATGGTG Reverse: AAAGCCGGTTGATTCTTGTG
<i>Il6-1142</i>	-1142 to -1038	Forward: GGTGGACAGAAAACCAGGAA Reverse: TAACCCCTCCAATGCTCAAG
<i>Il6-632</i>	-632 to -527	Forward: TGCACAAAATTTGGAGGTGA Reverse: ACCCAACCTGGACAACAGAC

113 TSS, transcription starting site.