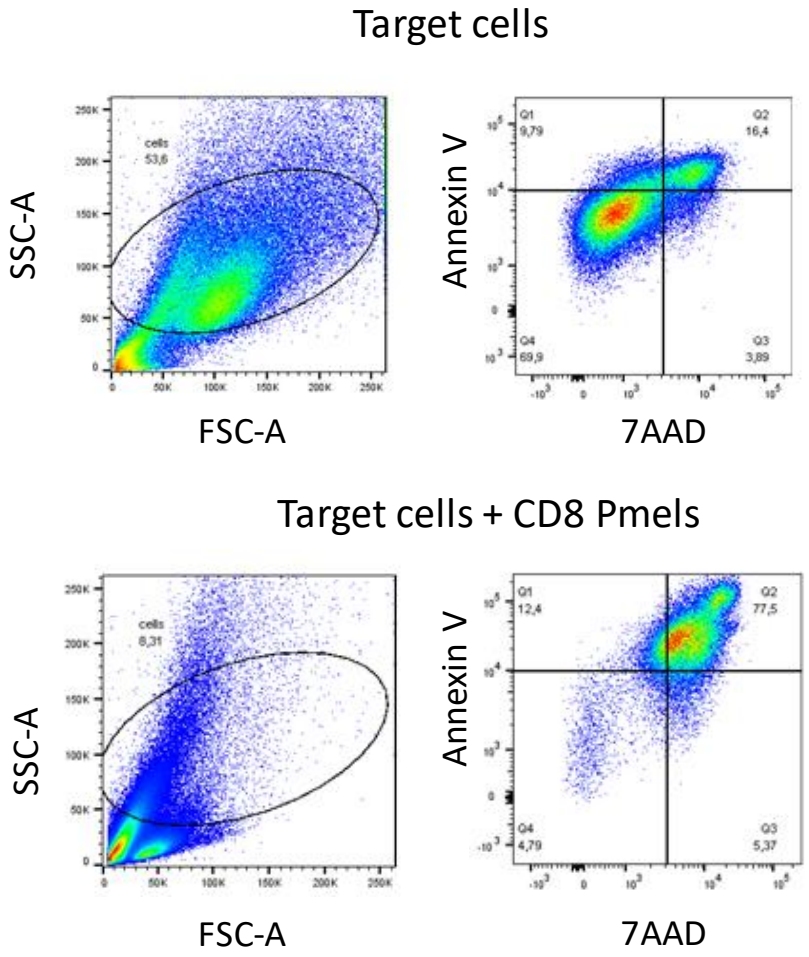
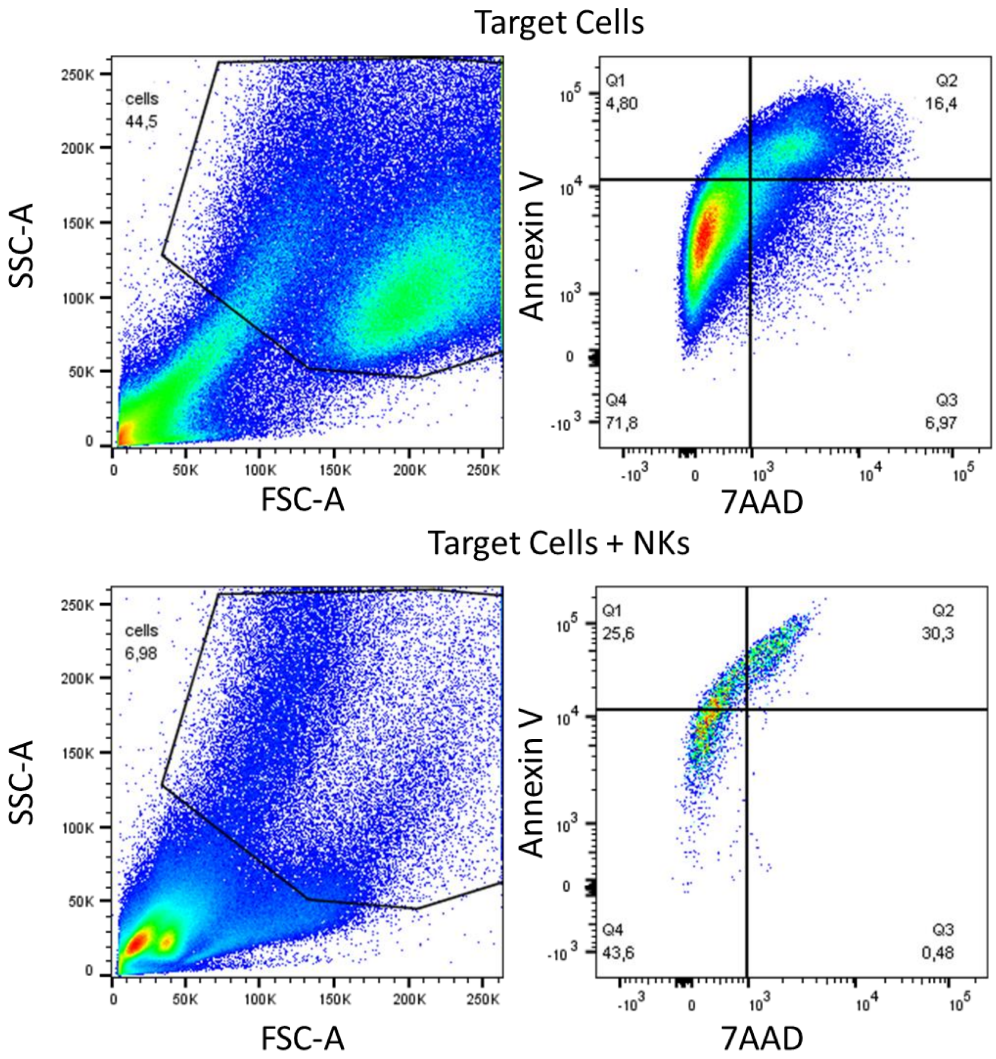
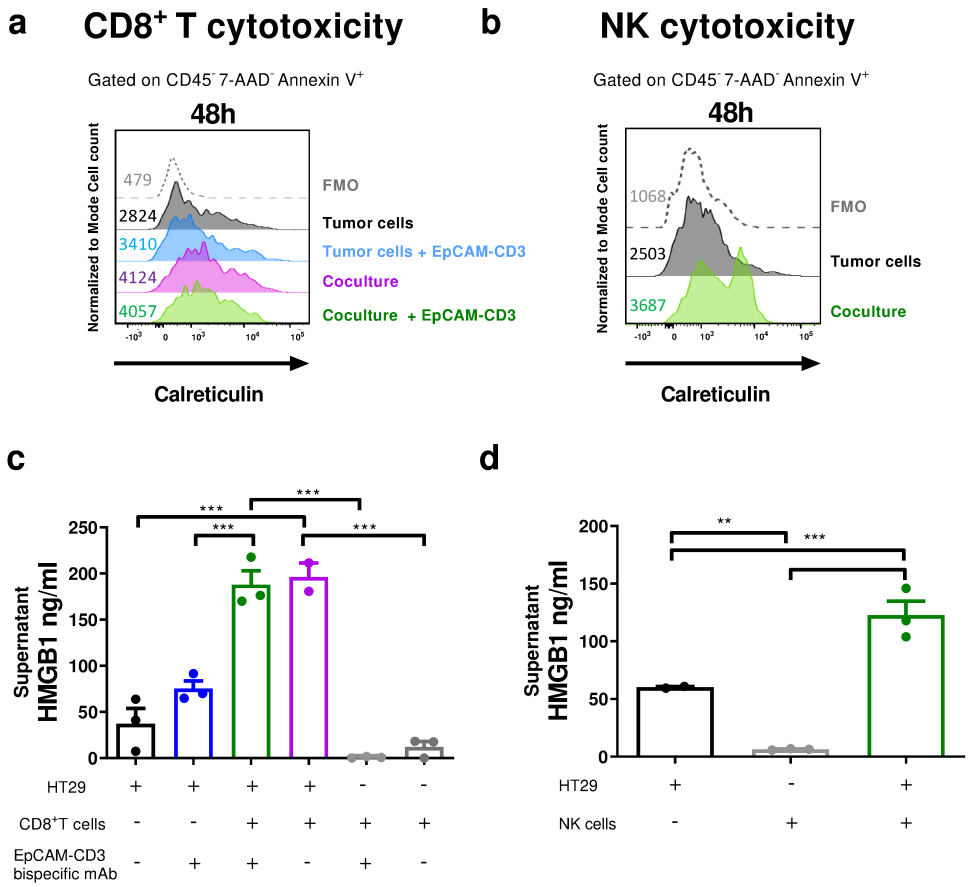
Supplementary Figures



Supplementary Figure 1. Assessment of CTL-induced tumor cell death by flow cytometry. MC38hEGFROVA cells were incubated 48 hours with IFNγ (15UI/ml) and gp100 peptide (100ng/ml) and Pmel-1-derived splenocytes were activated for 48 hours with gp100 peptide (100ng/ml) and 48 hours with IL-2 (30 UI/ml). At day -3, tumor cells and Pme1-1 cells were co-cultured. At day 0, the result of the co-culture was analyzed by flow cytometry after staining with 7-AAD, and Annexin V. Dot plots represent the gating strategy and 7-AAD and Annexin V double staining in a representative co-culture

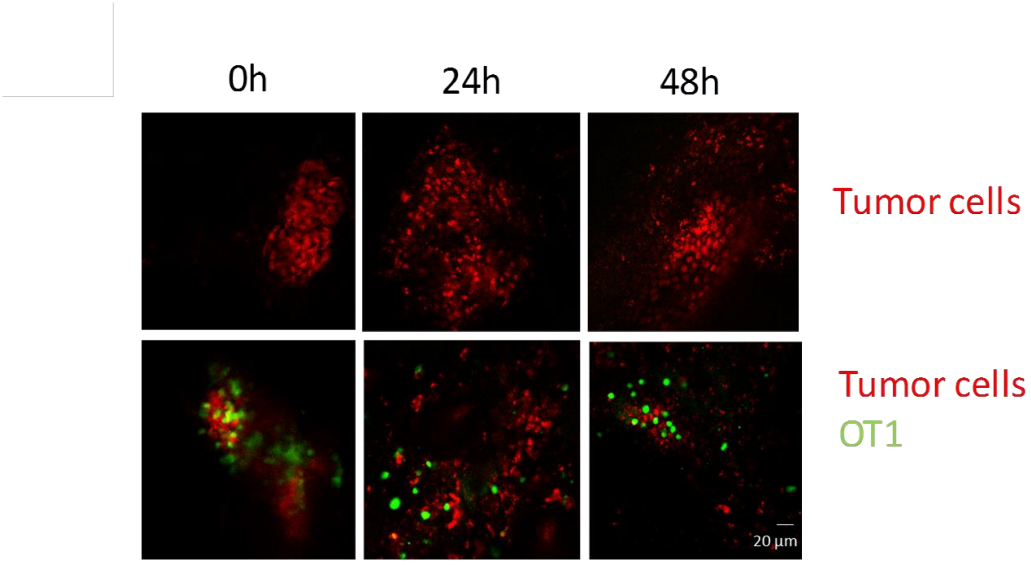


Supplementary Figure 2. Assessment of NK-induced tumor cell death by flow cytometry. RAG1 mice were hydrodynamically injected with a plasmid coding for IL-15. After three days, spleens were harvested and NK isolated by immunomagnetic negative selection. MC38hEGFROVA cells were incubated with such IL-15 *in vivo* activated NK cells for 72 hours. The result of the co-culture was analyzed by flow cytometry after staining with 7-AAD, and Annexin V. Dot plots represent the gating strategy and 7-AAD and Annexin V double staining in a representative co-culture.

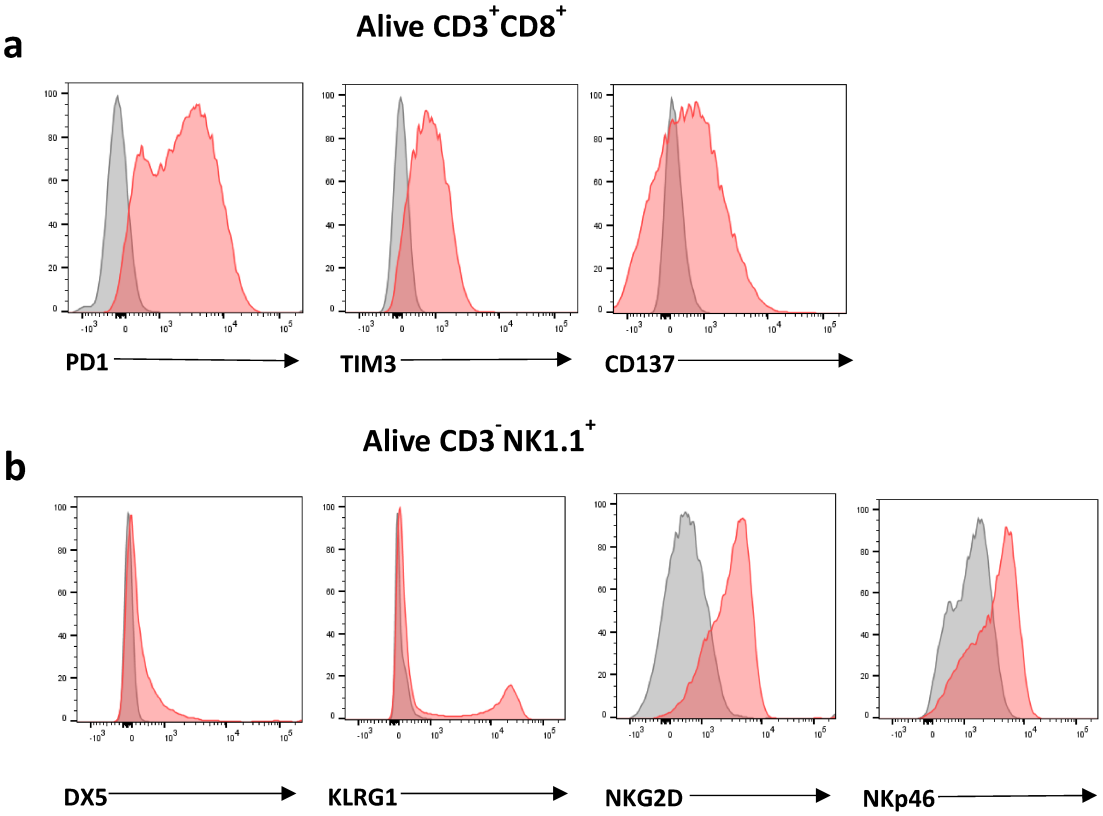


Supplementary Figure 3. Cellular cytotoxicity induces the release of danger-associated molecular patterns by dying human cancer cells in culture. A) HT29 cells were co-cultured with human T cells at a ratio 1:1 with or without EpCAM-CD3ε bispecific antibody (1µg/ml) for 48 hours and calreticulin expression on dying tumor cells ($CD45^-7\text{-}AAD^-Annexin\ V^+$) was analyzed by flow cytometry. Numbers in histograms indicate FMI. B) As in A but HT29 were killed by human $CD3^-CD56^+$ NK cells pre-activated 48 hours before with IL-2 (100 UI/ml) and IL-15 (10ng/ml). Supernatants from the co-cultures with human T cells (C) or human NK cells (D) were analyzed for HMGB1 concentration by ELISA. n=2 for HT29+T, n=3 for other groups (C). n=2 for HT29, n=3 for other groups (D). One-way ANOVA test with Tukey's

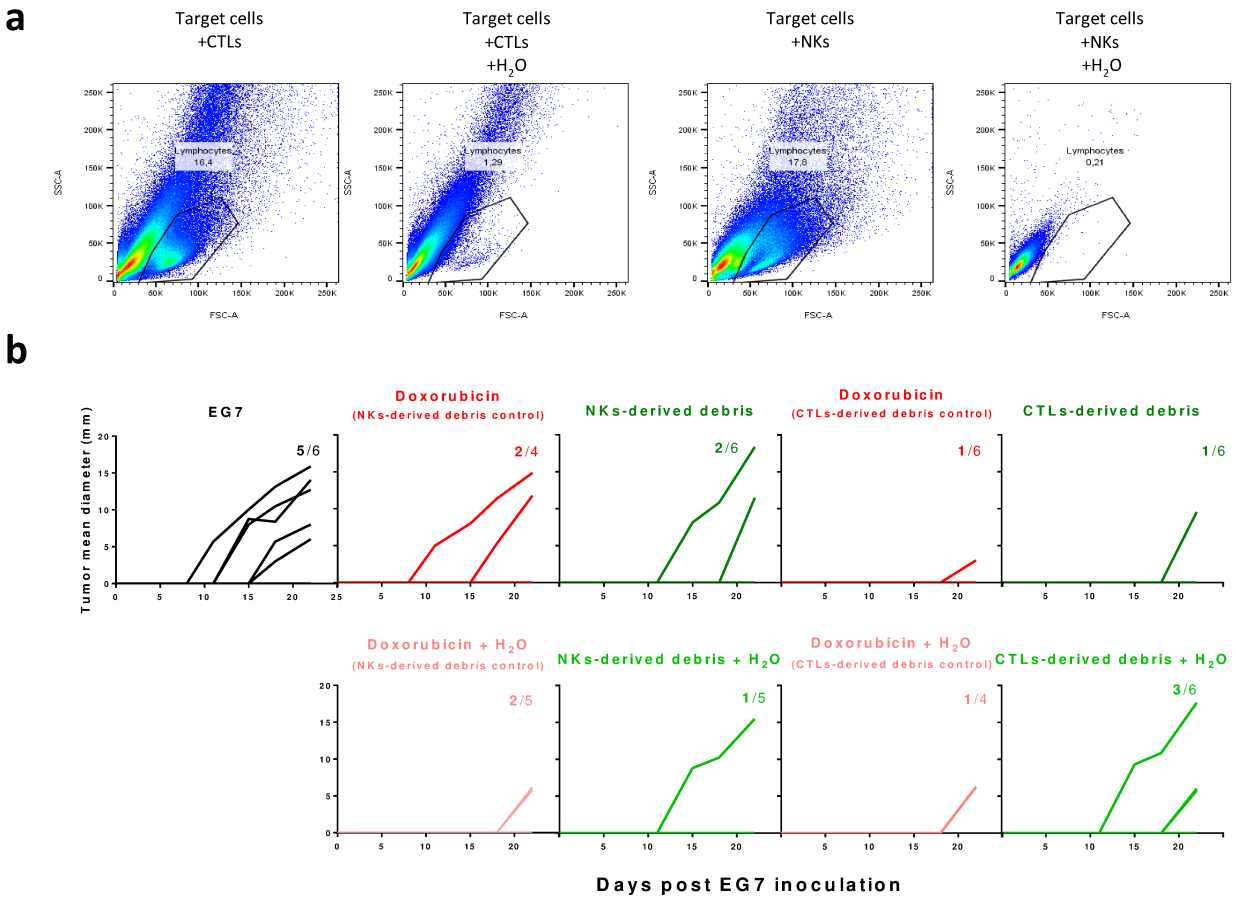
multiple comparisons tests, ***p<0.001. Results are representative of two experiments performed.



Supplementary Figure 4. Visualization of OT-I-mediated killing of tumor cells *in vivo*. *In vitro* activated OT-I-EGFP-derived splenocytes were co-injected with DRAQ 5 pre-labeled MC38hEGFROVA cells at a ratio 20:1 and injected subcutaneously into the ear dermis of C57BL/6 mice. Tumor and OT-I cells were visualized at day 0, 1, and 2 in mice by intravital imaging using an inverted confocal microscope.



Supplementary Figure 5. Surface antigen characterization by flow cytometry of NKs and CTLs used for killing assays gated as living $CD3^+CD8^+$ and $CD3^-NK1.1^+$ cells.



Supplementary Figure 6. Distilled-water elimination of effector cytotoxic cells does not impair immunization. In experiment as those in Fig. 2 and 4 effector cytotoxic T or NK cells (A) were removed by 60 min incubation in distilled water. Such lysates of MC38hEGFROVA deprived of cytotoxic effectors were tested to immunize against

OVA^+ EG7 cells when given 7 days later (B and C). Doxorubicin-killed lysates with or without preincubation with gp100 peptide and IFNγ have been used as positive control for CTLs-derived debris or NKs-derived debris, respectively. Log-rank tests were used for the indicated comparisons.