Supplementary Information

Structural basis for impairment of DNA methylation by the DNMT3A R882H mutation

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Supplementary Figure 1. Structural details of the DNMT3A^{WT}-CGA and DNMT3A^{WT}-CGT complexes. (a) Electrostatic surface of DNMT3A^{WT}-DNMT3L tetramer bound to CGA DNA. (b) Fo-Fc omit map (violet) of the CGA DNA (sticks) bound to DNMT3A^{WT}, contoured at 2.0 sigma level. (c) Schematic view of the DNMT3A^{WT}-DNA interactions in the DNMT3A^{WT}-CGA complex. The hydrogen-bonding, electrostatic and van der Waals interactions are indicated by red, blue and black arrows, respectively. The water-mediated hydrogen bond is indicated by letter 'W'. (d,e) Close-up views of the DNA interactions of the TRD loop in the DNMT3A^{WT}-CGT (d) and DNMT3A^{WT}-CGA (e) complexes. The hydrogen bonding interactions are indicated by dashed lines. The Fo-Fc omit maps (violet) for the interacting DNAs are contoured at 2.0 sigma level.



Supplementary Figure 2. Structural analysis of the DNMT3A^{R882H}-CGA complex. (a) Fo-Fc omit map (violet) of the CGA DNA (sticks) bound to DNMT3A^{R882H}, contoured at 2.0 sigma level. (b) Schematic view of the DNMT3A^{R882H}-DNA interactions in the DNMT3A^{R882H}-CGA complex. The hydrogen-bonding, electrostatic and van der Waals interactions are indicated by red, blue and black arrows, respectively. The water-mediated hydrogen bond is indicated by letter 'W'. (c) Close-up view of the interactions between the RD interface and DNA in the DNMT3A^{R882H}-CGA complex. The hydrogen-bonding interactions are shown as dashed lines.



DNMT3AR882H-bound CGT DNA



Supplementary Figure 3. Structural details of the DNMT3A^{R882H}-CGT complex. (a) Fo-Fc omit map (violet) of the CGT DNA (sticks) bound to DNMT3A^{R882H}, contoured at 2.0 sigma level. **(b)** Schematic views of the protein-DNA interactions in the DNMT3A^{R882H}-CGT complex. The hydrogen-bonding, electrostatic and van der Waals interactions are indicated by red, blue and black arrows, respectively. **(c)** Close-up view of the interaction between the RD interface and DNA in the DNMT3A^{R882H}-CGT complex. The hydrogen bonding interactions are shown as dashed lines.

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Supplementary Figure 4. Structural details of the DNMT3A^{R882H}-CAG complex. (a) Fo-Fc omit maps (violet) of the CAG DNA (sticks) bound to DNMT3A^{R882H}, contoured at 2.0 sigma level. **(b)** Schematic views of the protein-DNA interactions in the DNMT3A^{R882H}-CAG complex. The hydrogen-bonding, electrostatic and van der Waals interactions are indicated by red, blue and black arrows, respectively. **(c)** Close-up views of the interactions between the catalytic loop and DNA in the DNMT3A^{R882H}-CAG complex. **(d)** Close-up view of the interaction between the TRD loop and DNA in the DNMT3A^{R882H}-CAG complex, with the Fo-Fc omit map (violet) of residues R836-N838 contoured at 2.0 sigma level. The hydrogen bonds in (c) and (d) are shown as dashed lines. The water molecule is shown as a red sphere.





N879'

T862

E820

W860

M674'

H821





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Supplementary Figure 5. Intermolecular interactions at the RD interfaces of DNMT3A-DNA complexes. (a-e) Stick representations of protein residues located at the RD interfaces of the DNMT3A^{WT}-CGT (PDB 5YX2 [https://www.rcsb.org/structure/5YX2]) (a), DNMT3A^{WT}-CGA (b), DNMT3A^{R882H}-CGA (c), DNMT3A^{R882H}-CGT (d) and DNMT3A^{R882H}-CAG (e) complexes. The hydrogen bonding interactions are shown as dashed lines.



Supplementary Figure 6. Biochemical and enzymatic analyses of the DNMT3A^{R882H}- and DNMT3A^{WT}-mediated methylation. (a,b) Representative ITC binding curves of DNMT3A^{WT} (a) and DNMT3A^{R882H} (b) titrated against (GAC)₈ DNA. (c) *In vitro* CpG methylation of DNMT3A^{R882H}-DNMT3L and DNMT3A^{WT}-DNMT3L on a 36-

mer DNA containing single CpG site. **(d)** *In vitro* CpA and CpT methylation of DNMT3A^{R882H}-DNMT3L and DNMT3A^{WT}-DNMT3L on a 36-mer DNA containing single CpA or CpT site. **(e-g)** *In vitro* methylation kinetics of DNMT3A^{WT}-DNMT3L and DNMT3A^{R882H}-DNMT3L over (e) CpG, (f) CpA and (g) CpT DNA. The experiment in (e-g) was performed independently from (c,d). **(h)** Biological replicate of the bisulfite sequencing analysis in Figure 6d. (Top) Relative *in vitro* CpG and CpH methylation of WT and R882H-mutated DNMT3A-DNMT3L on a 626-bp DNA. The data are displayed as ratios of the methylation events of each indicated sequence context over total methylation events. (Bottom) Summary of total cytosine sites and methylation events in each indicated context, derived from bisulfite sequencing analyses of 7 clones of the 626-bp DNA incubated with WT or R882H-mutated DNMT3A-DNMT3L. Note that each DNA fragment contains 15 CpG, 38 CpA, 26 CpT and 29 CpC sites. **(i)** Relative ratio of methylation efficiency of DNMT3A^{R882H}-DNMT3L and DNMT3A^{WT}-DNMT3L for indicated sequence contexts on the 626 bp-long DNA. The experiment was repeated once with consistent results. In (c-g), n = 3 biological replicates. Data are mean ± s.d.

Supplementar	y Table 1. X-ra	y data collection	and refinement statistics.
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	DNMT3A ^{R882H} -	DNMT3A ^{R882H} -	DNMT3A ^{R882H} -	DNMT3A ^{WT} -
	DNMT3L-CGT	DNMT3L-CGA	DNMT3L-CAG	DNMT3L-CGA
	DNA	DNA	DNA	DNA
	(PDB: 6W8D)	(PDB: 6W89)	(PDB: 6W8J)	(PDB: 6W8B)
Data collection				
Space group	H 3	P 3 ₂	H 3	P 3 ₂
Cell dimensions				
a, b, c (Å)	205.6, 205.6, 89.5	187.6, 187.6, 82.4	205.9, 205.9, 89.4	186.3, 186.3, 81.7
α, β, γ (°)	90, 90, 120	90, 90, 120	90, 90, 120	90, 90, 120
Resolution (Å)	33.16-2.60(2.69-	37.72-2.50(2.59-	43.35-2.45(2.54-	46.59-2.40(2.49-
	$(2.60)^{a}$	2.50)	2.45)	2.40)
R _{merge}	0.051(1.155)	0.078(0.818)	0.112(1.496)	0.074(1.249)
$I/\sigma(I)$	13.7(0.9)	10.4(1.4)	7.8(1.9)	10.0(0.8)
$CC_{1/2}$	0.998(0.636)	0.999(0.726)	0.992(0.404)	0.999(0.644)
Completeness (%)	98.9(99.5)	98.5(93.0)	100.0(99.9)	99.9(99.9)
Redundancy	5.1(4.9)	6.7(5.9)	4.8(4.6)	5.1(4.7)
Refinement				
No. reflections	43,403	110,518	51,900	123,895
$R_{\rm work} / R_{\rm free}$	0.199/0.253	0.196/0.240	0.183/0.223	0.212/0.233
No. atoms				
Protein and DNA	8,238	16,730	8,330	16,745
SAH	52	104	52	104
Water	96	259	151	288
<i>B</i> factors ($Å^2$)				
DNMT3A	89.9	65.2	77.4	70.9
DNMT3L	140.2	124.0	125.9	130.8
DNA	158.4	120.0	129.8	116.8
SAH	89.1	60.1	80.6	66.6
Water	88.5	68.8	83.2	68.1
r.m.s deviations				
Bond lengths (Å)	0.004	0.006	0.004	0.004
Bond angles (°)	0.703	0.968	0.708	0.675

^aValues in parentheses are for highest-resolution shell. Each structure was determined using the dataset collected from a single crystal.

Supplementary Table 2. Summary of primer sequences.

Name	Sequence	Notes	
DNMT3A(R882H)_F	CAATATGAGCCATCTGGCACGTCAGCGT	DNMT3A R882H mutation	
DNMT3A(R882H)_R	TGCCAGATGGCTCATATTGCTAACATCGG	DNMT3A R882H mutation	
DNMT3A_MTase_F	CGCGGCGGTCTCGGATCCGCAGAAAAACGT AAACCGATTCGTG	Cloning of DNMT3A MTase domain into pRSFDuet-1	
DNMT3A_MTase_R	GGCGGCGCGGCCGCTTAAACGCAGGCAAAA TATTC	Cloning of DNMT3A MTase domain into pRSFDuet-1	
DNMT3L_F	CGCGGCCATATGATGTTCGAAACCGTGCCT GTG TGG	Cloning of DNMT3L C-terminal domain into pRSFDuet-1	
DNMT3L_R	CGCGGCGGTCTCCTCGAGTTATAAAGAGGA AGTGAGTTCTG	Cloning of DNMT3L C-terminal domain into pRSFDuet-1	
GST3-F	TTGAAGAAAAATATGAAGAGGATTTGTATG AG	Bisulfite sequencing analysis	
GST3-R	CCCCTCCAACACAACTTCC	Bisulfite sequencing analysis	