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## **Supplemental Information**

# **Oncolytic Adenovirus Armed with BiTE, Cytokine,**

### and Checkpoint Inhibitor Enables CAR T Cells

## to Control the Growth of Heterogeneous Tumors

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**Supplemental Fig. 1**. *CRISPR/Cas9 system effectively knocks out target genes in FaDu cells*. We used the CRISPR/Cas9 system to knock either CD44 or HER2 out of FaDu cells. After sorting knockout lines, we confirmed CD44 or HER2 receptor expression on these cells.



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**Supplemental Fig. 2**. *HDAd-derived CD44v6.BiTE, IL-12p70 and PD-L1 blocking antibody increase the antitumor efficacy of HER2.CAR T-cells in vitro.* (**A**) FaDu and FaDuHER2-/- expressing *ffLuc* cells were infected with 100 vp/cell of HDAd0 (no transgene), HDAd*CD44v6.BiTE*, HDAd*Duo* or HDAd*Trio* (n=5 per group). HER2.CARTs were added at 24 hr post-infection (E:T=1:20). Cells were harvested 72 hours post-coculture, and viable cancer cells were analyzed by luciferase assay. Data are presented as means  $\pm$  SD. \**P* < 0.001. (**B**) HER2.CARTs were harvested 72 hours post-coculture, and CD25, PD-1, 4-1BB and OX40 expression were analyzed by flow cytometry.



**Supplemental Fig. 3**. *Co-infection of Onc.Ad with HDAd expressing BiTE, IL-12p70 and PD-L1 mini-antibody amplifies IL-12p70 and PD-L1 blocking antibody expression with oncolysis in vitro.* (**A**) FaDu was infected with a total 20 Vp/cell of HDAd*Trio*, Onc.Ad or with an CAd*Trio* (Onc.Ad:HDAd=1:10) (n=4 per group). Medium samples were collected 48 hours post-infection. Levels of IL-12p70 and PD-L1 mini-antibody in media samples were quantified by IL-12p70 ELISA assay and western blotting for PD-L1 mini-antibody, respectively. Data are presented as means  $\pm$  SD. \**P* < 0.001. (**B**) FaDu was infected with increasing doses of HDAd*s*, Onc.Ad or with CAds (Onc.Ad: HDAd=1:10) (n=6 per group). Viable cells were analyzed at 96 hours by MTS assay. Data are presented as means  $\pm$  SD.



**Supplemental Fig. 4.** *Combinatorial treatment can control both primary and metastasized tumors in an orthotopic HNSCC model.* (A) FaDu cells were transplanted into the tongues of NSG mice (n=5 per group). A total of 1x10<sup>8</sup> vp of CAds (Onc:HD=1:20) were injected into the tongue. A total of 0.2 x10<sup>6</sup> HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAd. Bioluminescence of HER2.CARTs at the tumor area was monitored at different time points. Data are presented as means  $\pm$  SD. (B) Body weight was measured at different time points. Data are presented as means  $\pm$  SD. (C) FaDu cells expressing *ffLuc* were transplanted into the tongues of NSG mice. 1x10<sup>8</sup> vp of CAd*BiTE* (Onc:HD=1:20) were injected into the tongue. 0.2x10<sup>6</sup> HER2.CARTs were systemically administered 3 days post-injection of CAds. IFN $\gamma$  levels in were measured by ELISA from serum samples collected at 0, 3, 10, 24, 45, and 66 days post-injection of CAds (n=10). Data are presented as means  $\pm$  SD. **Table** shows median survival of animals treated with CART alone or CAd*BiTE* plus CART.



Supplemental Fig. 5. *CAd-derived CD44v6.BiTE improves HER2.CAR T-cell anti-tumor effects in an orthotopic HER2<sup>-/-</sup> HNSCC model.* FaDuHER2-/- cells expressing *ffLuc* were transplanted into the tongues of NSG mice. A total of  $1 \times 10^8$  vp of CAd*BiTE* (Onc:HD=1:20) were injected into the tongue. A total of  $1 \times 10^6$  HER2.CARTs were systemically administered 3 days post-injection of CAd*BiTE*. Bioluminescence of FaDuHER2-/- cells was monitored at different time points. Serum samples were collected at 0, 3, 10, 24, 45, 66 days post-injection of CAd*BiTE* (n=5), and IFN $\gamma$  levels in serum were measured by ELISA. Data are presented as means  $\pm$  SD. Kaplan-Meier survival curve after CAd*BiTE* administration (n=5). Data are presented as means  $\pm$  SD. \**P* < 0.02.