

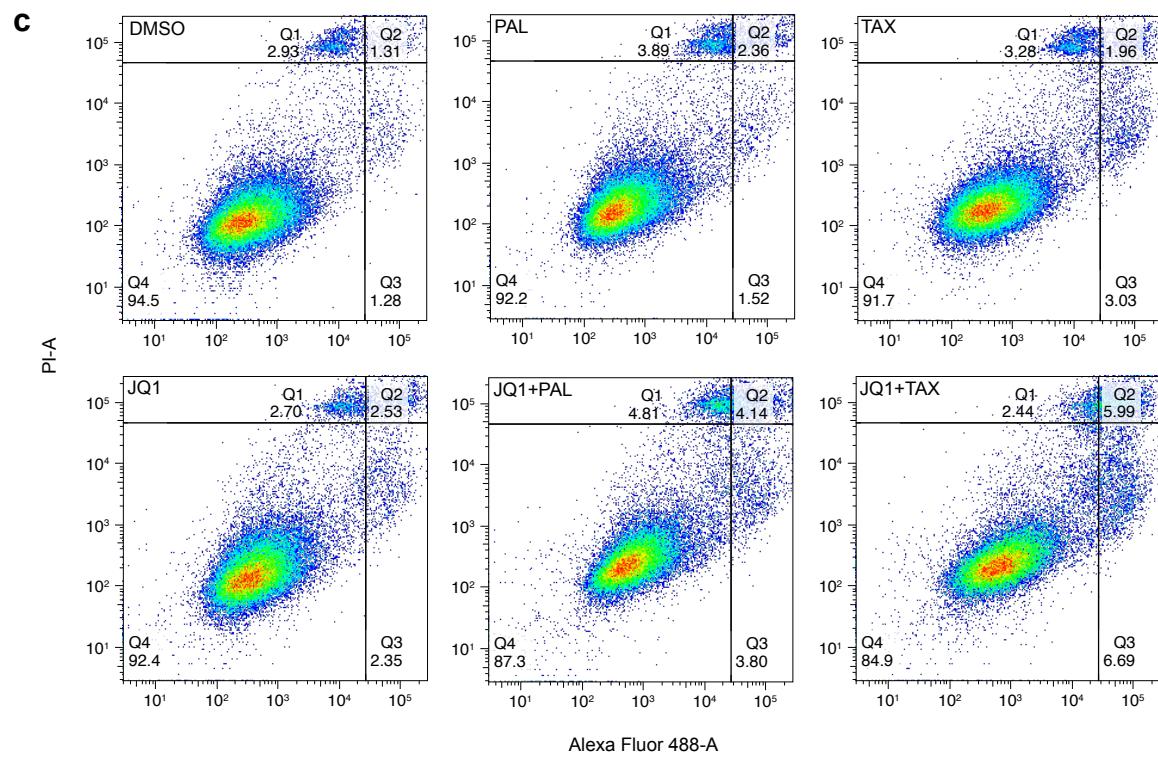
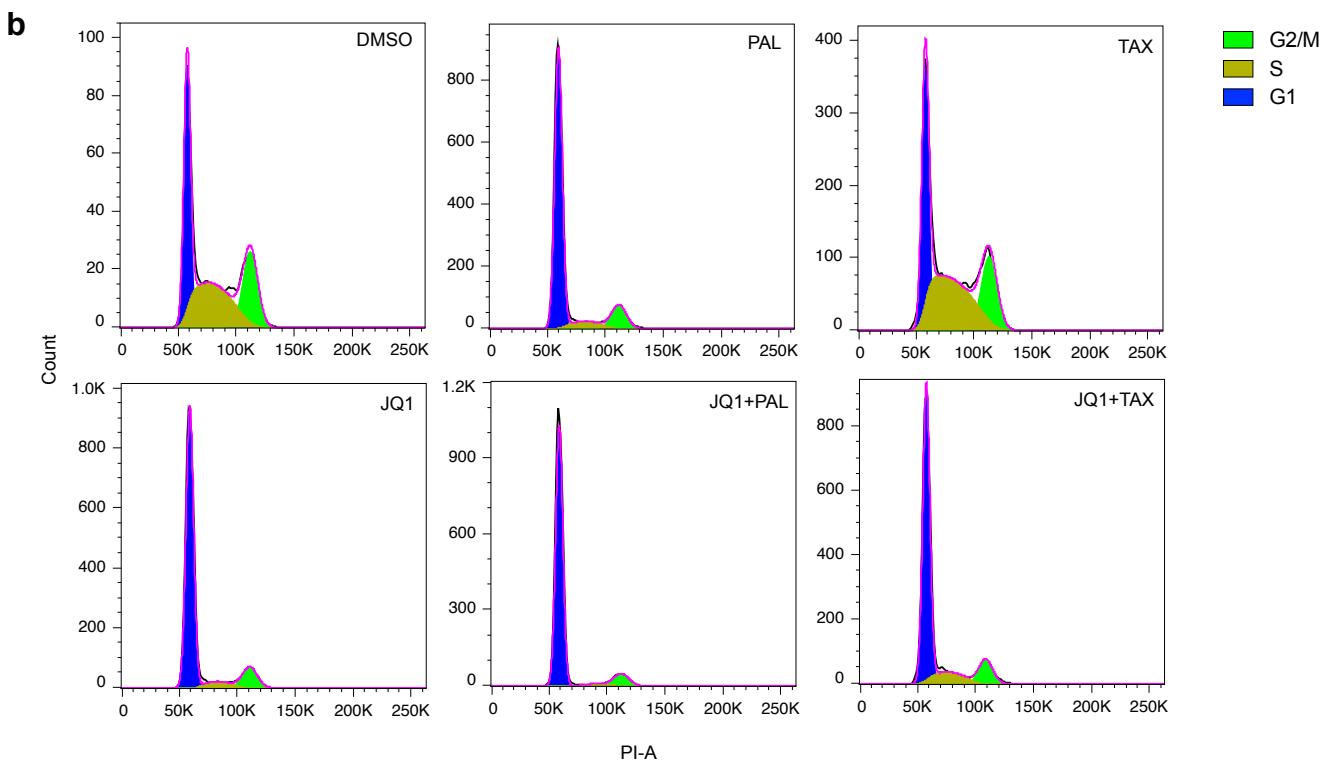
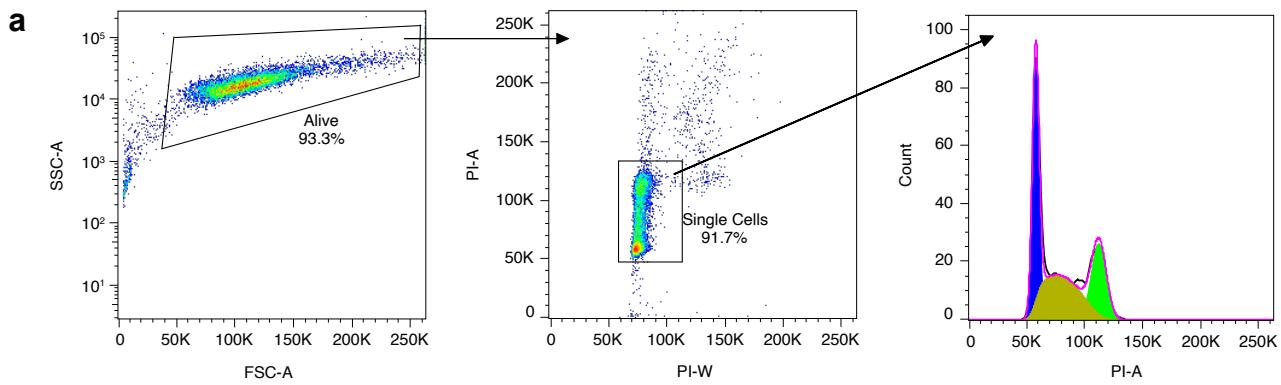
Supplementary Information

**Acquired resistance to combined BET and CDK4/6 inhibition in triple-negative breast
cancer**

Ge et al.

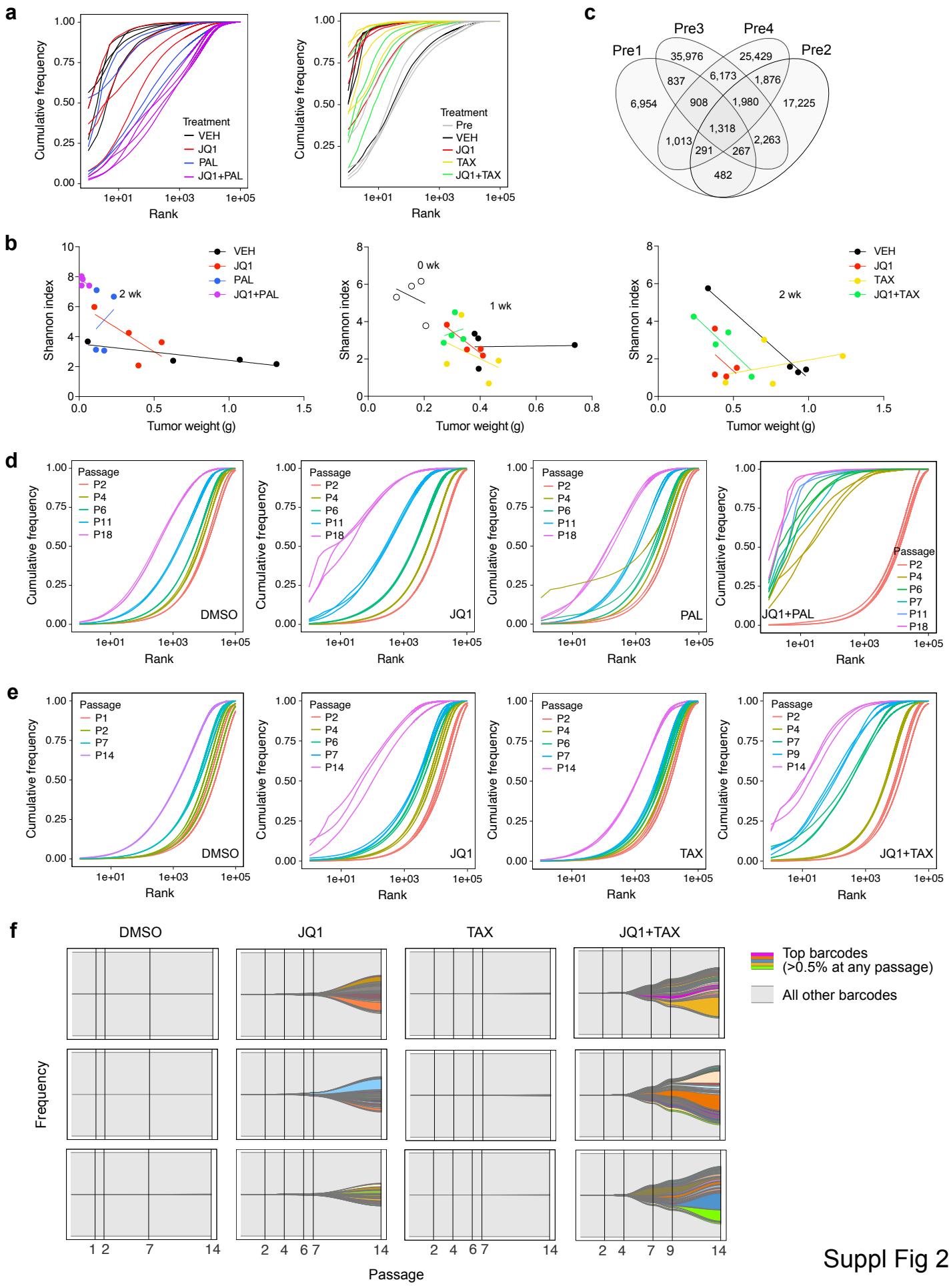
SUPPLEMENTARY FIGURES

Supplementary Figure 1. Flow cytometry analysis of cell cycle and apoptosis. **a**, Gating strategy for flow cytometry analysis of cell cycle in PI-stained cells (shown are DMSO-treated cells). **b**, Representative histograms of PI-stained SUM159 cells in each cell cycle phase, following 24 hours of treatment with the indicated compounds (PAL, palbociclib, and TAX, paclitaxel), by flow cytometry fitted to the Watson cell cycle model. **c**, Representative scatterplots of SUM159 cells that are viable (annexin V-/PI-), early apoptotic (annexin V+/PI-), and late apoptotic (annexin V+/PI+), using flow cytometry following three days of treatment.



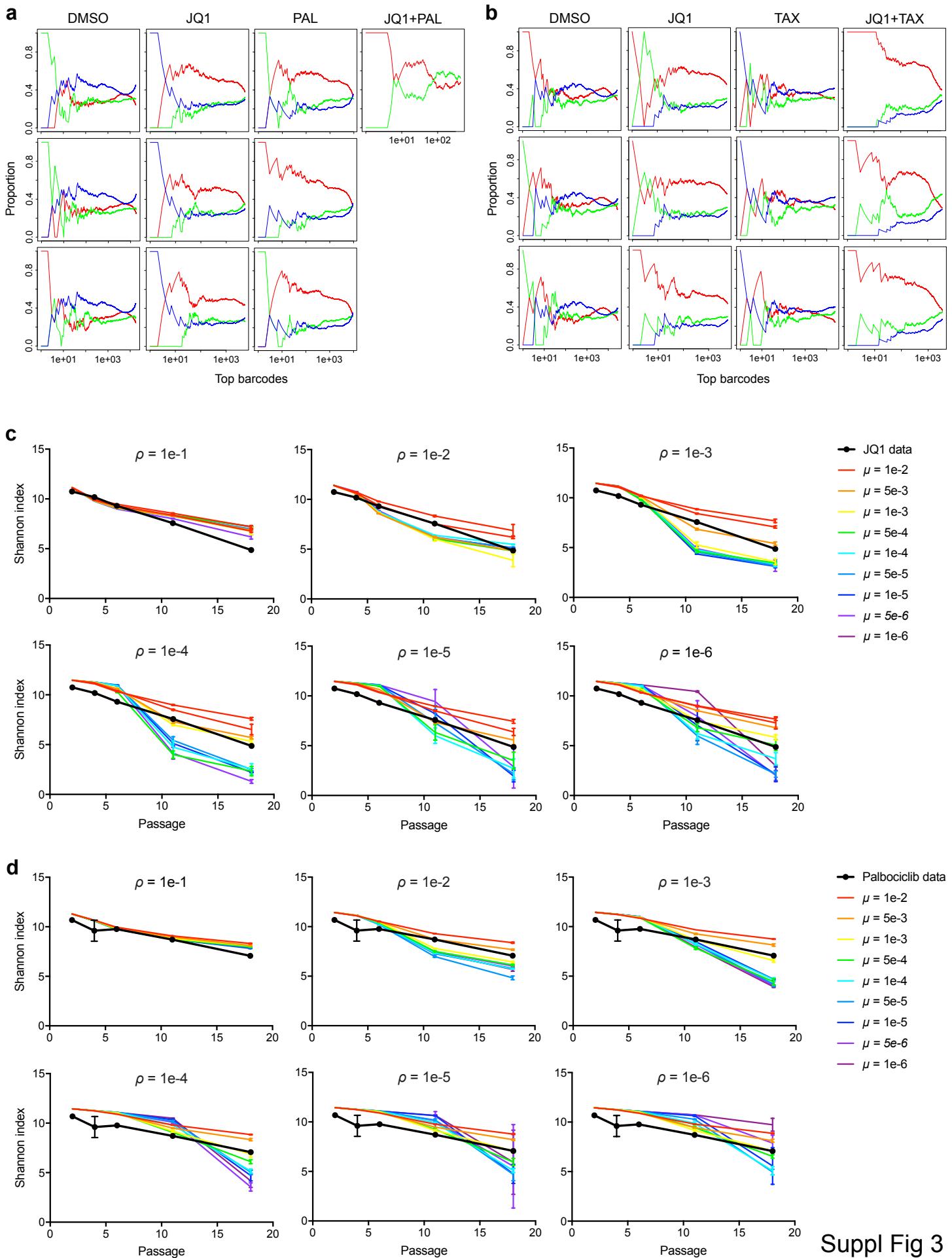
Suppl Fig 1

Supplementary Figure 2. Barcode diversity in xenografts and cells treated with JQ1 combinations. **a**, Cumulative frequencies of ranked barcodes in SUM159 xenografts treated for 2 weeks with JQ1 and palbociclib (left) or JQ1 and paclitaxel (right) alone and in combination. **b**, Scatterplots of Shannon indices vs. tumor weights of xenografts following 2 weeks of treatment with JQ1 and palbociclib (left), before treatment and following 1 week of treatment with JQ1 and paclitaxel (center), and following 2 weeks of treatment with JQ1 and paclitaxel, alone and in combinations. **c**, Venn diagram of the number of shared barcodes between four xenografts prior to treatment. **d**, Cumulative frequencies of ranked barcodes in SUM159 cells at various passages of treatment with DMSO, JQ1, palbociclib (PAL), and JQ1+palbociclib (JQ1+PAL). **e**, Cumulative frequencies of ranked barcodes in SUM159 cells at various passages of treatment with DMSO, JQ1, paclitaxel (TAX), and JQ1+paclitaxel. **f**, Frequencies of top barcodes (those representing at least 0.5% of the population at any passage) in cell populations selected with JQ1 and paclitaxel, alone and in combination. Colors represent unique barcodes, gray background represents all other barcodes in the population, and each plot represents one replicate. Frequencies of all barcodes (y-axis) add up to 100%.



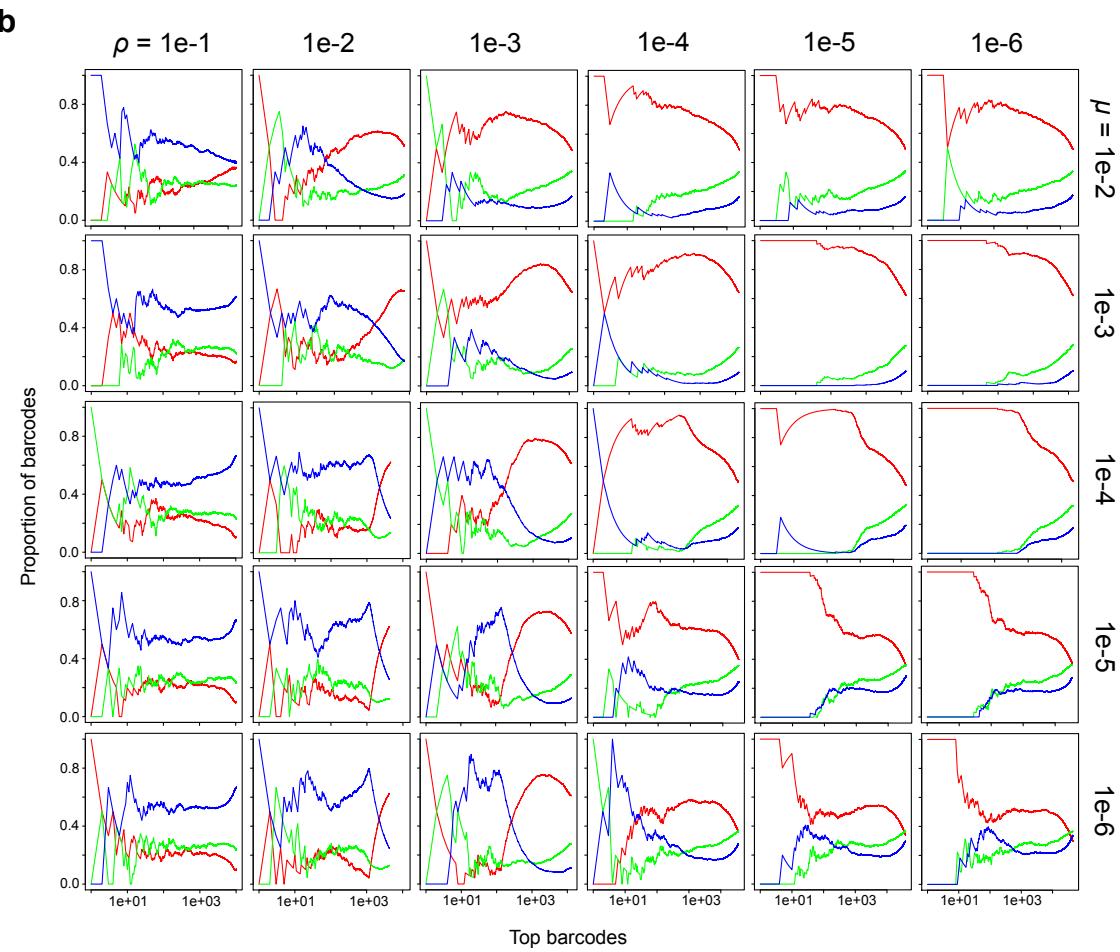
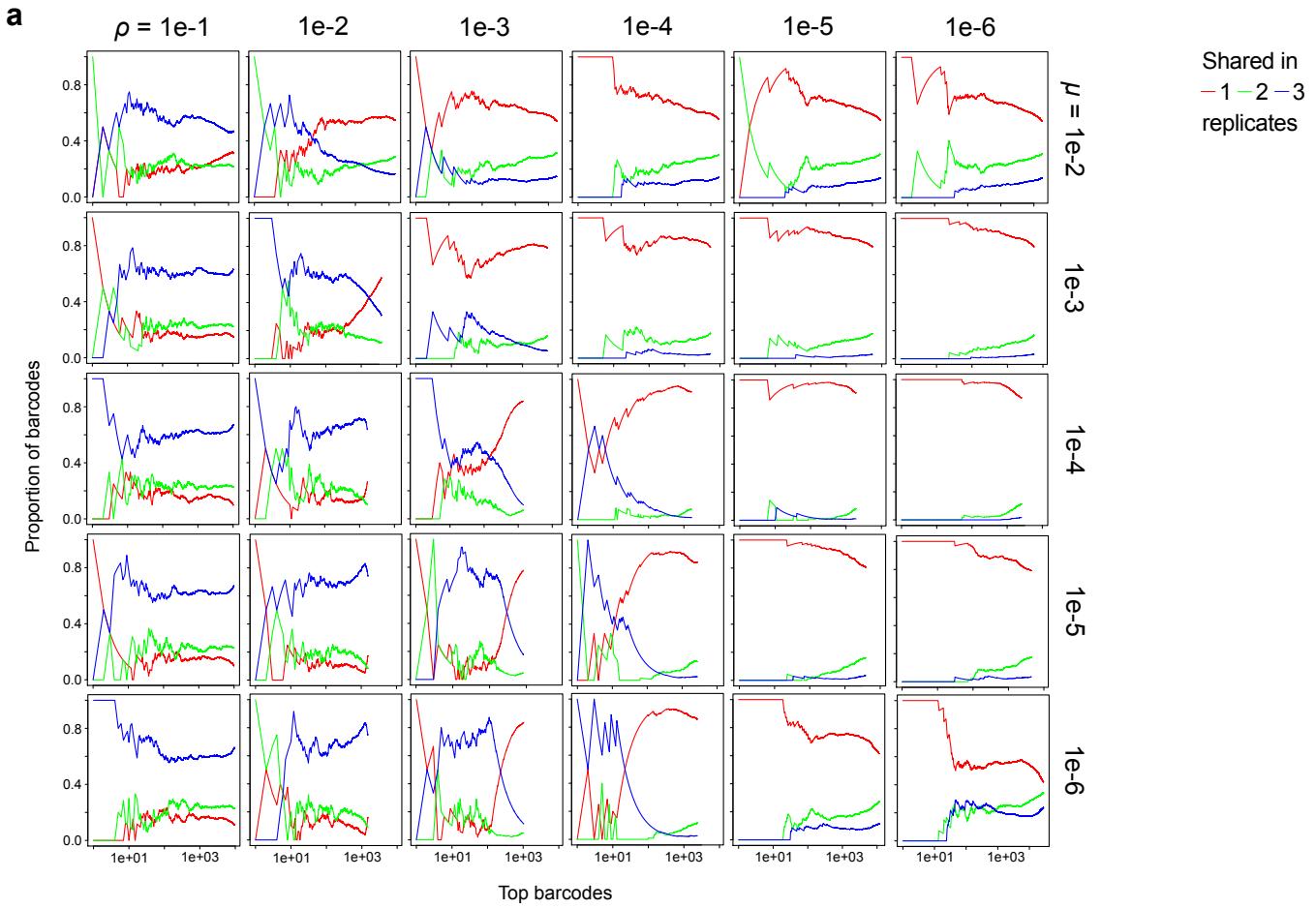
Suppl Fig 2

Supplementary Figure 3. Proportions of shared barcodes between replicates selected with JQ1 combinations and simulated barcode diversities. **a**, Proportion of top barcodes (x-axis) in each replicate at the last passage that are unique to that replicate, shared between 2 replicates, or shared between all 3 replicates passaged in DMSO, JQ1, palbociclib (PAL), and JQ1+palbociclib. One JQ1+palbociclib treated replicate died out during the treatment course, so barcodes are only shared between 2 replicates. **b**, Proportion of top barcodes (x-axis) in each replicate at the last passage that are unique to that replicate, shared between 2 replicates, or shared between all 3 replicates passaged in DMSO, JQ1, paclitaxel (TAX), and JQ1+paclitaxel. **c**, Comparison between simulated data and experimental data of Shannon indices for cells passaged in JQ1, over a range of values for ρ and μ . Data are represented as mean \pm SD, $n = 3$. **d**, Comparison between simulated data and experimental data of Shannon indices for cells passaged in palbociclib, over a range of values for ρ and μ . Data are represented as mean \pm SD, $n = 3$.



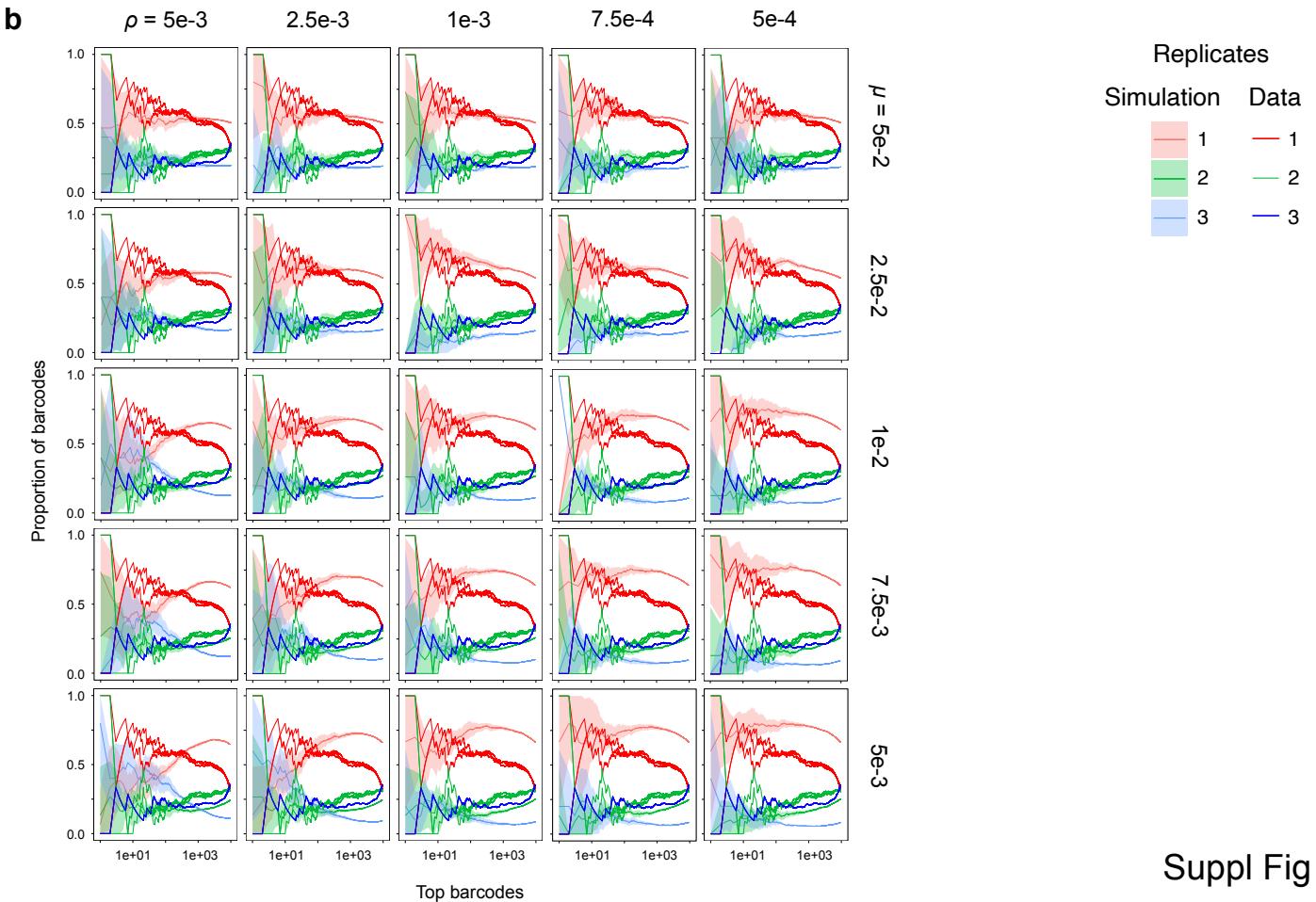
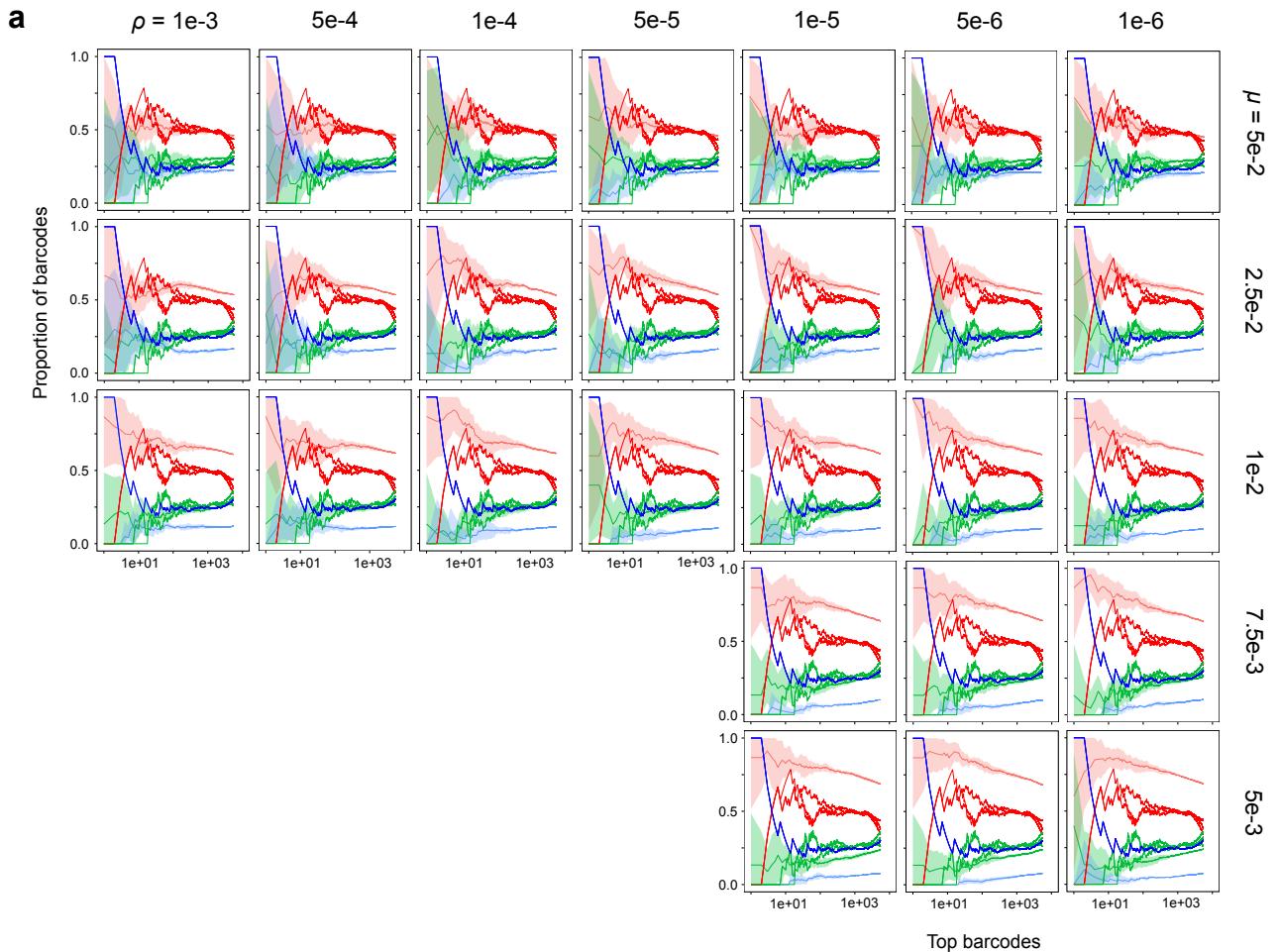
Suppl Fig 3

Supplementary Figure 4. Simulated proportions of shared barcodes. **a**, Simulated proportions of top barcodes (x-axis) that are unique, shared between 2 replicates, or shared between all 3 replicates at the last passage of JQ1, over a range of values for ρ and μ . **b**, Simulated proportions of top barcodes (x-axis) that are unique, shared between 2 replicates, or shared between all 3 replicates at the last passage of palbociclib, over a range of values for ρ and μ .



Suppl Fig 4

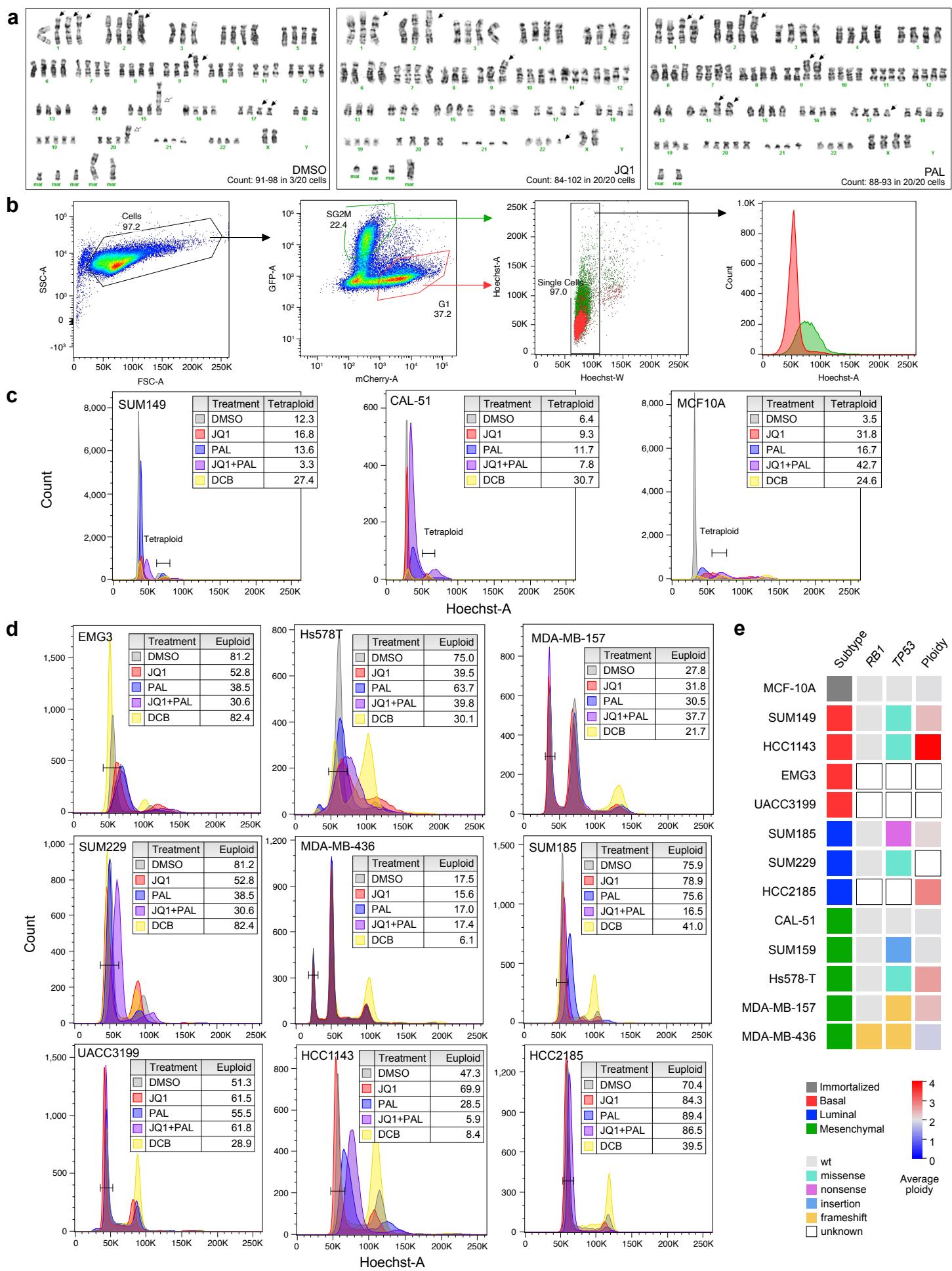
Supplementary Figure 5. Comparison between simulated and experimental proportions of shared barcodes. **a**, Comparison between simulated data and experimental data of proportions of top barcodes (x-axis) that are unique, shared between 2 replicates, or shared between all 3 replicates at the last passage of JQ1, over a narrowed down range of values for ρ and μ . Simulated distributions are represented as mean \pm SD, $n = 5$ independent simulations with 3 replicates each. **b**, Comparison between simulated data and experimental data of proportions of top barcodes (x-axis) that are unique, shared between 2 replicates, or shared between all 3 replicates at the last passage of palbociclib, over a narrowed down range of values for ρ and μ . Simulated distributions are represented as mean \pm SD, $n = 5$ independent simulations with 3 replicates each.



Suppl Fig 5

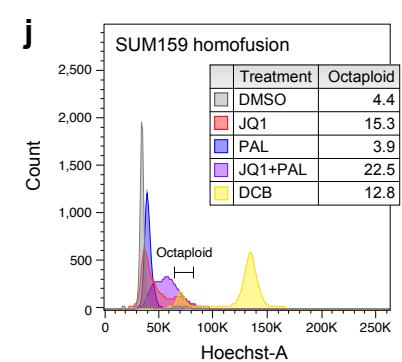
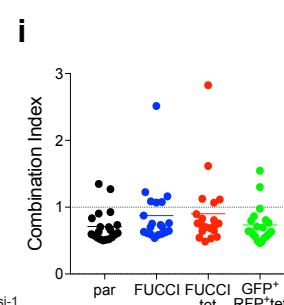
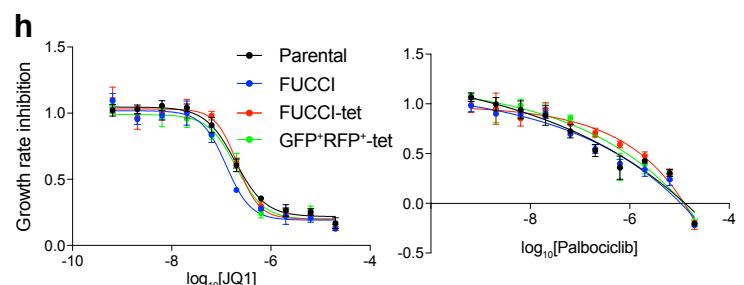
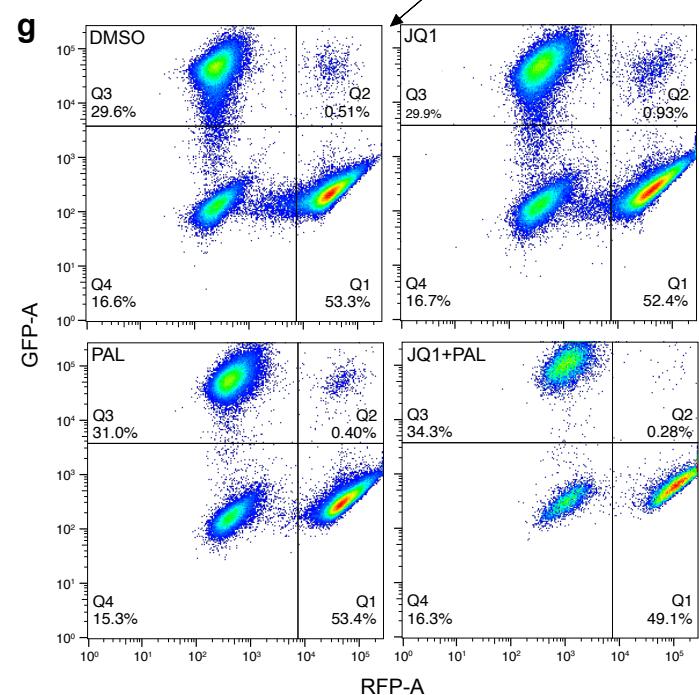
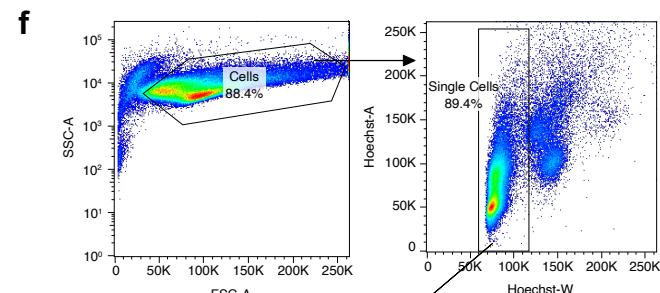
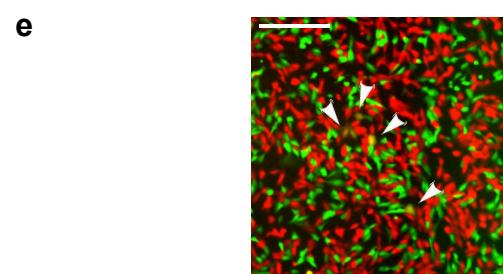
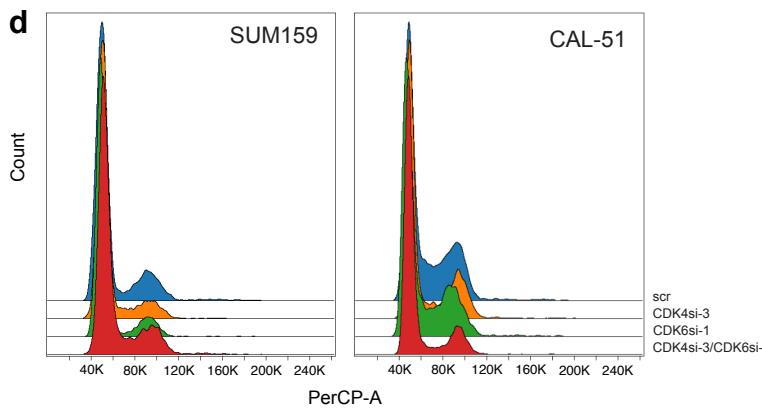
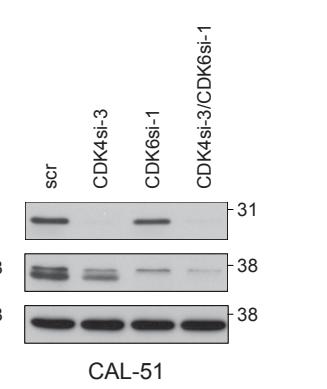
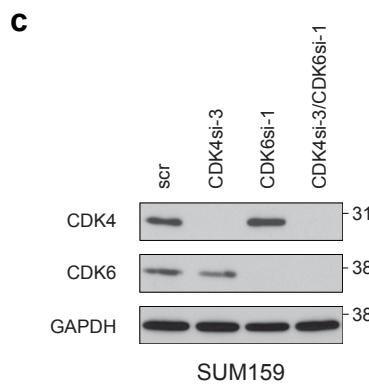
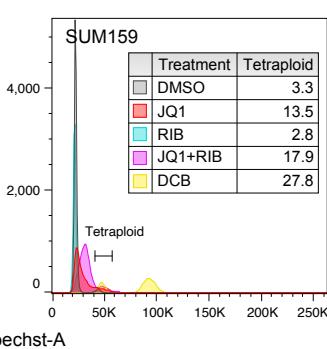
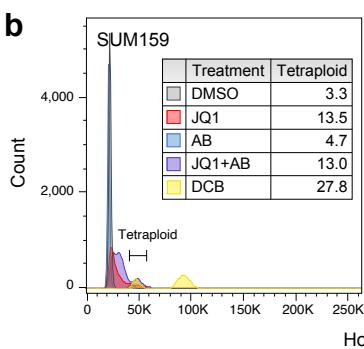
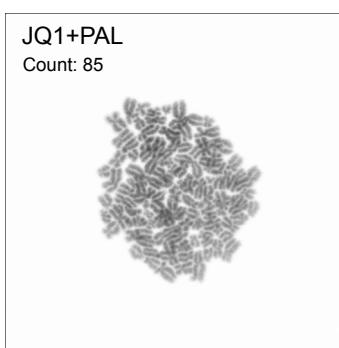
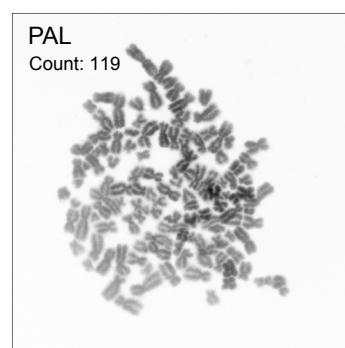
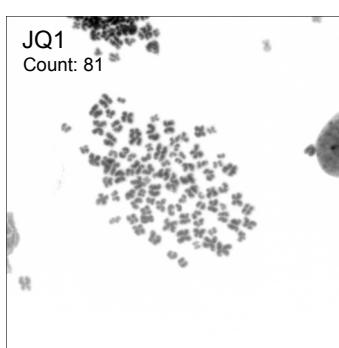
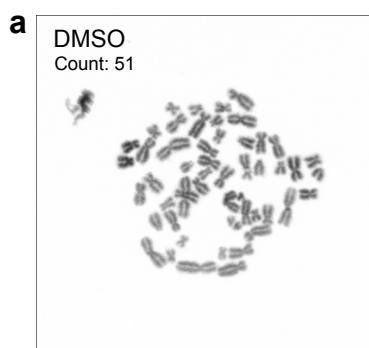
Supplementary Figure 6. JQ1 and palbociclib induce increased ploidy through cell division failure. **a**, Representative karyotypes of a minority of cells that was tetraploid after passaging in DMSO and of cells after passaging in JQ1 and palbociclib (PAL). Ranges of chromosome numbers and numbers of metaphase spreads counted are shown. Arrows indicate abnormal chromosomes. **b**, Gating strategy for flow cytometry analysis of DNA content in G1 and S/G2M fractions of Hoechst-stained FUCCI-labeled cells (shown are DMSO-treated cells). **c**, Histograms of DNA content by flow cytometry in Hoechst-stained FUCCI-labeled G1 SUM149, CAL-51, and MCF10A cells treated with DMSO, JQ1, palbociclib, and JQ1+palbociclib for 7 days. Dihydrocytochalasin B (DCB)-induced tetraploid cells were used as positive controls. Table indicates proportion of cells in the tetraploid gate. **d**, Histograms of DNA content by flow cytometry in a panel of TNBC cells treated with DMSO, JQ1, palbociclib, and JQ1+palbociclib for 7 days and stained with Hoechst. Dihydrocytochalasin B (DCB)-induced tetraploid cells were used as positive controls. Table indicates proportion of cells in the euploid gate, i.e. those with the baseline ploidy for each cell line. Note: these cell lines were not FUCCI-labeled; thus, the analysis includes cells at any cell cycle phase. **e**, TNBC subtype, *RB1* and p53 status, and average ploidy of cell lines tested in this study.

Suppl Fig 6



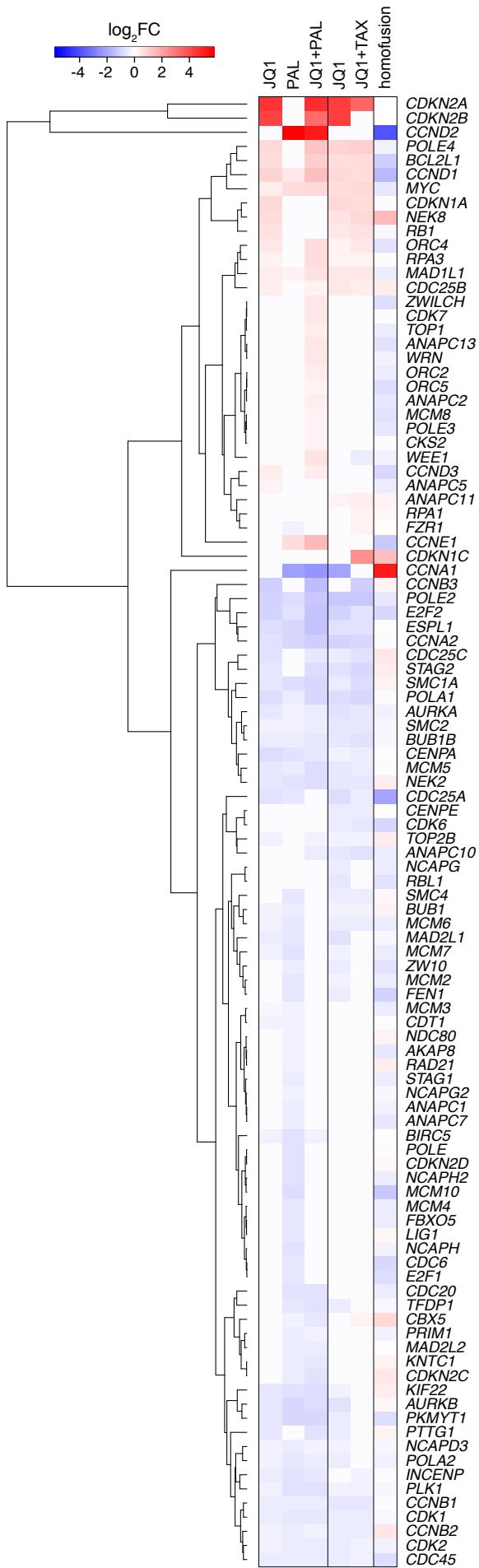
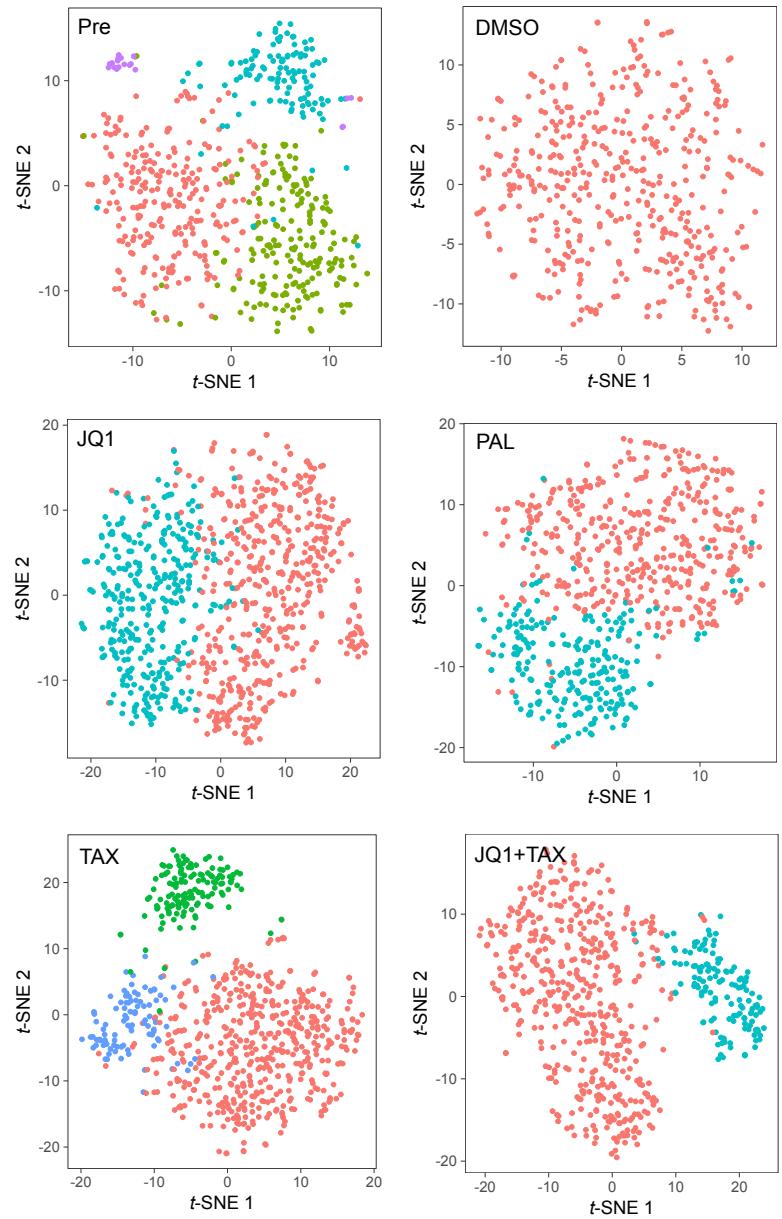
Supplementary Figure 7. JQ1 and CDK4/6 inhibitors induce increased ploidy through cell division failure. **a**, Metaphase spreads of SUM159 cells following treatment with DMSO, JQ1, palbociclib (PAL), and JQ1+palbociclib for 7 days. Counts indicate numbers of chromosomes. Images are representative of one experiment with 20 cells counted per group. **b**, Histograms of DNA content by flow cytometry in Hoechst-stained FUCCI-labeled G1 SUM159 cells treated with DMSO, JQ1, abemaciclib (AB), JQ1+abemaciclib, ribociclib (RIB), and JQ1+ribociclib for 7 days. Dihydrocytochalasin B (DCB)-induced tetraploid cells were used as positive controls. Table indicates proportion of cells in the tetraploid gate. **c**, Western blots show CDK4 and CDK6 levels in SUM159 and CAL-51 cells after CDK4, CDK6, and CDK4/6 siRNA knockdown. **d**, Histograms of DNA content by flow cytometry in 7-AAD-stained SUM159 and CAL-51 cells 6 days post-transfection with scramble, CDK4, CDK6, or CDK4 and 6 siRNA. Note: these cell lines were not FUCCI-labeled; thus, the analysis includes cells at any cell cycle phase. **e**, Image shows co-cultured RFP⁺ and GFP⁺ cells. Arrows indicate fused (GFP⁺RFP⁺, yellow) cells. Image is representative of two independent experiments. Scale bar represents 500 μm. **f**, Gating strategy for flow cytometry analysis of fused cells in co-cultures of GFP⁺ and RFP⁺ cells (shown are DMSO-treated cells). **g**, Scatterplots of flow cytometry analysis of co-cultured GFP⁺ and RFP⁺ cells quantifying the proportion of fused (GFP⁺RFP⁺) cells following 7 days of treatment. **h**, Dose response curves for JQ1 and palbociclib of diploid cells (parental and FUCCI) and sorted cells enriched for tetraploids (FUCCI-tet from FUCCI G1 tetraploids and GFP/RFP-tet from GFP⁺RFP⁺ fusions). Data are represented as mean ± SD, $n = 4$. Source data are provided as a Source Data file. **i**, Levels of synergism between JQ1 and palbociclib in diploid cells (parental and FUCCI) and sorted cells enriched for tetraploids (FUCCI-tet and GFP⁺RFP⁺-tet). Each point represents CI for one pair of concentrations, averaged over 8 replicates. CI = 1 additive, CI < 1 synergistic, and CI > 1 antagonistic. Source data are provided as a Source Data file. **j**, Histograms of DNA content by flow cytometry in Hoechst-stained SUM159 homofusions following treatment with DMSO, JQ1, palbociclib, and JQ1+palbociclib for 7 days. Table indicates proportion of cells in the octaploid gate. Note: these cell lines were not FUCCI-labeled; thus, the analysis includes cells at any cell cycle phase.

Suppl Fig 7



Supplementary Figure 8. RNA-seq and single cell RNA-seq analysis of post-selection cells. a,

Heat map of differentially expressed genes in the core cell cycle process network by bulk RNA-seq of post JQ1, palbociclib (PAL), JQ1+palbociclib (JQ1+PAL), and JQ1+paclitaxel (JQ1+TAX) selected cells compared with post-DMSO selected cells, as well as tetraploid homofusion cells compared with diploid parental cells. **b**, Separate *t*-SNE plots of cells from the pre-treatment population (Pre) and from the post-selection populations of DMSO, JQ1, palbociclib (PAL), paclitaxel (TAX), and JQ1+paclitaxel (JQ1+TAX) by single-cell RNA-seq. Each point represents one single cell.

a**b**

Suppl Fig 8