

Supporting Information

Silicon-Free SuFEx Reactions of Sulfonimidoyl Fluorides: Scope, Enantioselectivity, and Mechanism

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2 General information and methods

All commercial chemicals were used as received and stored under argon. Reagents were used without further purification unless otherwise noted. Unless otherwise noted, all reactions were performed using glassware without further preparation. Certain reactions were carried out in anhydrous conditions. This utilized oven-dried glassware which was heated in an oven above 100 °C for at least 5 h or which was dried under vacuum with a heat gun (T > 200 °C), under oxygen-free and water-free conditions. After weighing any solids, the glassware was connected to a Schlenk line, and then placed under vacuum and flushed with nitrogen gas (3x purged). Liquids were added via syringe, through a rubber stopper. Solvent abbreviations are: tetrahydrofuran (THF), ethyl acetate (EtOAc) and *N*-dimethylformamide (DMF), petroleum ether 40-60 (P.E.). Solvents were used as received without any distillation.

Flash column chromatography was performed using a Biotage® system and SiliCycle® precast silica columns (200–300 mesh or 300–400 mesh). TLC analysis was performed on pre-coated, alumina-backed silica gel plates. TLC plates were analyzed by UV fluorescence (254 nm) or I₂ stain.

¹H NMR, ¹³C NMR, ¹⁹F NMR spectra were recorded on a Bruker Avance 400 MHz spectrometer at 298 K. The chemical shifts are listed in ppm on the δ = scale and coupling constants were recorded in Hertz (Hz). Chemical shifts are calibrated relative to the signals corresponding of the non-deuterated solvents (CHCl₃: δ = 7.26 ppm for ¹H and 77.16 ppm for ¹³C, CH₃CN: δ = 1.94 ppm for ¹H and 1.32 and 118.26 ppm for ¹³C, C₆H₆: δ = 7.16 ppm for ¹H and 128.06 ppm for ¹³C). Abbreviations are used in the description of NMR data as follows: chemical shift (δ = ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, dt = doublet of triplets, m = multiplet), coupling constant (*J*, Hz).

High-resolution mass spectra (HR-MS) were measured on a QTOF micro spectrometer using electrospray ionization (ESI) in positive mode (ESI+) or in negative mode (ESI-).

Enantioselectivity was monitored using an HPLC Chiralpak IA column. A reference, racemic product was synthesized to determine the column conditions for baseline separation. Unless specifically mentioned, these were found to be *n*-hexane propan-2-ol 90:10, flow rate 0.5 mL/min, oven temperature 40 °C, detector wavelength 240 nm. Conditions were the same for the sulfonimidoyl starting material, also giving baseline separation.

3 Synthesis of starting materials and special phenols



Scheme S1. The synthesis of sulfonimidoyl fluoride starting material 1.



4-methylbenzenesulfinamide 1b

Compound 1b was synthesized according to literature.1

Oxalyl chloride (9.1 mL, 106 mmol) was added dropwise to sodium *p*-toluenesulfonic acid **1a** (17.8 g, 100 mmol) in anhydrous toluene (100 mL) at 0 °C in a 500 mL 3-necked round bottom flask. The mixture was stirred for 1 h at rt. Thereafter, 100 mL ethyl acetate and 100 mL NH₄OH were added, while at 0 °C. The reaction mixture was then stirred for another hour. The mixture was diluted with 100 mL ethyl acetate, and the organic and aqueous phases separated. The aqueous phase was then extracted with ethyl acetate (2 x 100 mL). The combined organic layers were washed with brine (100 mL) and dried over Na₂SO₄. The volatiles were removed under vacuum evaporation yielding **1b** as a white powder (9.57 g, 61.7 mmol, 62%). ¹H NMR (400 MHz, CDCl₃) δ = 7.63 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 4.31 (s, 2H), 2.41 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₇H₁₀NOS: 156.0483, found: 156.0477. Spectral data are in accordance with literature.¹



N-(p-tolylsulfinyl)benzamide 1c

Compound 1c was synthesized according to literature.²

Compound **1b** (5.0 g, 32.2 mmol) was dissolved in dry THF (100 mL) and cooled to -78 °C. *n*-Butyl lithium (32 mL, 32.5 mmol) was then added dropwise over 10 min. The reaction was stirred at -78 °C for 10 min and warmed to room temperature and stirred for 3 h. The reaction was quenched with saturated NaHCO₃ (100 mL) and diluted with CH₂Cl₂ (150 mL). The organic layer was separated, and the aqueous phase was extracted with CH₂Cl₂ (5 x 50 mL). The combined organic layers were dried with MgSO₄, filtered and concentrated under vacuum. The product was recrystallized from hexane and ethyl acetate (2:1), yielding a white powder (5.6 g, 21.6 mmol, 67%). ¹H NMR (400 MHz, CDCl₃) δ = 8.24 (s, 1H), 7.79 (d, *J* = 8.4 Hz, 2H), 7.68 (d, *J* = 7.6 Hz, 2H), 7.57 (t, *J* = 7.4 Hz, 1H), 7.45 (t, *J* = 7.4 Hz, 2H), 7.36 (d, *J* = 8.0 Hz, 2H), 2.44 (s, 3H). Spectral data are in accordance with literature.²

→ o CI

tert-butyl hypochlorite

tert-butyl hypochlorite was synthesized according to literature.³

NaOCI (421 mL, 5% active chlorine, 275 mmol) was added to a 1 L round bottom flask, cooled to 0 °C and placed in the dark. Acetic acid (16.8 mL, 294 mmol) and *tert*-butyl alcohol (26.1 mL, 275 mmol) were added in one portion. The solution was stirred for 10 min. The aqueous and organic phases were then separated. The organic phase was washed with 10% NaHCO₃, water (5 mL) and dried over CaCl₂ to yield a yellow liquid (17.9 g, 165 mmol, 60%). ¹H NMR (400 MHz, CDCl₃) δ = 1.32 (s, H). Spectral data are in accordance with literature.³



N-benzoyl-4-methylbenzenesulfonimidoyl fluoride 1

Compound 1 was synthesized according to literature.⁴

tert-Butyl hypochlorite (3.05 g, 28 mmol) was added dropwise to a solution of compound **1c** (5.6 g, 21.6 mmol) in anhydrous CH₂Cl₂ (100 mL) at 0 °C, under argon atmosphere. The mixture was stirred for 2 hours at 0 °C. Subsequently, the volatiles were removed under vacuum. The residue was flushed with argon and dissolved in anhydrous CH₃CN (100 mL). KF (5.02 g, 85 mmol) and 18-crown-6 ether (265 mg, 1 mmol) were subsequently added and the mixture was stirred for 16 h at rt. The mixture was filtered, and the solvent was evaporated under vacuum. The remaining oil was purified by column chromatography (P.E.: EtOAc – 1:1), yielding **1** as an off-white powder (4.40 g, 15.9 mmol, 73%). ¹H NMR (400 MHz, CDCl₃) δ = 8.15 (d, *J* = 7.6 Hz, 2H), 8.06 (d, *J* = 7.6 Hz, 2H), 7.56 (t, *J* = 7.4 Hz, 1H), 7.45 (d, *J* = 7.6 Hz, 2H), 2.50 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): δ = 170.2, 147.3, 134.2, 133.3, 131.5 (d, *J* = 2.0 Hz), 130.4, 130.1, 128.4, 128.1, 22.0. ¹⁹F NMR (376 MHz, CDCl₃) δ = 65.41 (s). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₁₄H₁₃FNO₂S: 278.0651, found: 278.0638. Spectral data are in accordance with literature.⁴



TBDMS-phenol

To a flame-dried flask, under Ar flow, phenol (2.00 g, 21.25 mmol, 1 equiv) was added and dissolved in 30 mL anhydrous CH₂Cl₂. Imidazole (3.18 g, 46.75 mmol, 2.2 equiv) and TBDMS-Cl (3.36 g, 22.31 mmol, 1.05 equiv) were added and a precipitate formed immediately. Stirring was continued for 5 h. The mixture was transferred to a separatory funnel and washed twice with water and twice with brine. After drying over Na₂SO₄, the volatiles were removed under reduced pressure, yielding a colorless oil. This oil was further purified via flash column chromatography (P.E. 40-60), and the product TBDMS-protected phenol was obtained as a colorless oil (3.74 g, 17.98 mmol, 85%). ¹H NMR (400 MHz, CDCl₃) δ = 7.26 – 7.20 (m, 2H), 6.96 (t, *J* = 7.4 Hz, 1H), 6.86 (dd, *J* = 8.5, 1.0 Hz, 2H), 1.00 (s, 10H), 0.21 (s, 6H). Spectral data are in concurrence with literature.⁵



TBDMS-p-cresol

TBDMS-p-cresol was synthesized according to literature.6

p-Cresol (3.16 g, 29.22 mmol) and imidazole (2.98 g, 43.83 mmol) were dissolved in DMF (10 mL) in a 25 mL, 3necked, roundbottom flask. After this, *tert*-butylchlorodimethylsilane (5.73 g, 38 mmol) was added, and the reaction mixture was stirred overnight. The solution was then diluted with diethyl ether (50 mL) and washed with water (3 x 50 mL). The organic layer was then dried over Na₂SO₄ and concentrated under reduced pressure. The residual product was purified with flash column chromatography (P.E.: EtOAc – 20:1) to TBDMS-*p*-cresol a colorless liquid (5.58 g, 27 mmol, 92%). ¹H NMR (400 MHz, CDCl₃) δ = 7.02 (d, *J* = 7.6 Hz, 2H), 6.73 (d, *J* = 7.2 Hz, 2H), 2.27 (s, 3H), 0.98 (s, 9H), 0.18 (s, 6H). Spectral data are in concurrence with literature.⁶



TBDMS-2-tert-butyl-phenol 6

To a flame-dried flask, under Ar flow, 2-tBu-phenol (1.7 mL, 11.07 mmol, 1 equiv) was added and dissolved in 15 mL anhydrous CH₂Cl₂. Imidazole (1.60 g, 24.35 mmol, 2.2 equiv) and TBDMS-CI (1.75 g, 11.62 mmol, 1.05 equiv) were added and a precipitate formed immediately. Stirring was continued for 5 h, after which the reaction had reached about 50% conversion based on TLC analysis. Additional imizadole (828 mg, 12.17 mmol, 1.1 equiv) and TBDMS-CI (876 mg, 5.81 mmol, 0.52 equiv) were added and the reaction was stirred for 12 h. After this time, full conversion was not reached (approx. 90% conversion had occurred). The mixture was transferred to a separatory funnel and washed twice with water and twice with brine. After drying over Na₂SO₄, the volatiles were removed under reduced pressure, yielding a colorless oil. This oil was further purified via flash column chromatography (P.E. 40-60), and the product TBDMS-protected phenol was obtained as a colorless oil (2.33 g, 8.82 mmol, 80%). ¹H NMR (400 MHz, CDCl₃) δ = 7.31 (dd, *J* = 7.8, 1.5 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.89 (t, *J* = 8.0 Hz, 1H), 6.84 (d, *J* = 8.0 Hz, 1H), 1.41 (s, 9H), 1.06 (s, 9H), 0.35 (s, 6H). Spectral data are in concurrence with literature.⁷



trimethyl(p-tolyloxy)silane

TMS-protected *p*-cresol was synthesized according to literature.⁸ Hexamethyldisilazane (439.8mg, 2.72 mmol) and KAI(SO₄)₂*12H₂O (Alum) (66 mg, 0.141 mmol) were dissolved in anhydrous CH₃CN (2.7 mL). *p*-cresol (294 mg, 2.77 mmol) was added and the mixture was stirred for 2.5 h at rt. The mixture was quenched with water (~20 mL) and diluted with CH₂Cl₂ (20 mL). The organic and aqueous layers were separated and the aqueous layer was washed twice with CH₂Cl₂ (20 mL). The combined organic layers were dried over Na₂SO₄ and concentrated *in vacuo* to yield the title compound. The title compound was obtained as a light yellow oil, and dried overnight under nitrogen flow and used without further purification (36.2mg, 0.200 mmol, 7%). The yield was diminished due to drying under nitrogen flow and subsequent evaporation due to its low boiling point. ¹H NMR (400 MHz, CDCl₃) δ = 7.06-6.98 (d, 2H), 6.78-6.69 (d, 2H), 2.32-2.24 (s, 3H), 0.30-0.21 (s, 9H). Spectral data are in concurrence with literature.⁹

4 Competition study between a phenol (phenolate) and Si-protected phenol in the SuFEx reaction

In our efforts to investigate the Si-free SuFEx reaction, we made a comparison with a traditional SuFEx reaction. Under pseudo-first-order conditions, a Si-protected phenol and free *p*-cresol were reacted with sulfonimidoyl fluoride **1**. Using these conditions, we can determine which phenol derivative reacts faster. This provides information on both the reaction rate, as well as on the relative reactivity of the traditional and Si-free SuFEx substrates.



Scheme S2. The competition reaction between a Si-protected and unprotected *p*-cresol and phenol

Procedures

p-Cresol vs TBDMS-phenol

p-Cresol (31.1 mg, 0.288 mmol, 10 equiv) and TBDMS-phenol (60.0 mg, 0.288 mmol, 10 equiv) were dissolved in CD₃CN (0.55 mL). Mesitylene (4.1 μ L, internal reference) was added and the mixture was shaken well. DBU (43.1 μ L, 0.288 mmol, 10 equiv) was added and the mixture was shaken well for 10 seconds. The mixture was transferred to a vial containing the sulfonimidoyl fluoride **1** (8.0 mg, 0.029 mmol, 1 equiv). The mixture was shaken well and transferred to an NMR tube. ¹H NMR was measured immediately, and again after approximately 15 min. The reaction was executed in duplicate, and measured twice. The ratios were determined using mesitylene as the internal reference, and the presented ratio is averaged over the four measurements.

TBDMS-p-cresol vs phenol

TDBMS-*p*-cresol (64.1 mg, 0.288 mmol, 10 equiv) and phenol (27.1 mg, 0.288 mmol, 10 equiv) were dissolved in CD₃CN (0.55 mL). Mesitylene (4.1 μ L, internal reference) was added and the mixture was shaken well. DBU (43.1 μ L, 0.288 mmol, 10 equiv) was added and the mixture was shaken well for 10 seconds. The mixture was transferred to a vial containing the sulfonimidoyl fluoride **1** (8.0 mg, 0.029 mmol, 1 equiv). The mixture was shaken well and transferred to an NMR tube. ¹H NMR was measured immediately, and again after approximately 15 min. The reaction was executed in duplicate and measured twice. The ratios were determined using mesitylene as the internal reference, and the presented ratio is averaged over the four measurements.

 Table S1. Product distribution in competition experiments comparing the reactivity of SuFEx substrates with Si-protecting group and the Si-free SuFEx substrate.

Entry	Cresol	Phenol	Reaction time (min) #	Product ratio <i>p-</i> cresol : phenol [*]
1	TBDMS-p-cresol	Phenol	< 2	9: 91
2	<i>p</i> -cresol	TBDMS-Phenol	< 2	95: 5

[#]During the reaction time, the transfer of TBDMS-groups was negligible. Interconversion of products was not found, even after letting the reaction stand for 3 days at rt. ^{*}product distribution ratio was determined via ¹H-NMR spectroscopy, at rt, using mesitylene as an internal reference. Standard deviation: < 2%. Experimental error, given relatively small amounts of the minor fraction: ca. 3-4%.

Under these pseudo first-order conditions, the product formation was clearly in favor of the phenolate/cresolate. The traditional TBDMS-protected SuFEx substrates reacted much more slowly under these conditions. Although a small substituent effect (*para*-H vs *p*-Me) could be playing a role, the overwhelming preference for the phenolate shows the power of the Si-free SuFEx reaction. This shows that a roughly 10:1 ratio was found for the reaction rate of unprotected phenols versus Si-protected phenols under these reaction condition.

Pictured below are the ¹H NMR spectra after the completion of the reaction for both reactions, including the peaks of interest used to determine the final product distribution.







Figure S2. The ¹H NMR spectra of the phenol reaction mixture, the *p*-cresol reaction mixture and this pseudo first-order competition experiment in the aromatic region (δ 8.3-6.3) and aliphatic region (δ 2.35-2.10). Product ratios were determined using the integrals of the peaks of interest, belonging to the specific products. Peaks of interest belonging to the phenol product are highlighted in red. Peaks of interest belonging to the *p*-cresol product are highlighted in blue.



Figure S3. ¹H NMR spectrum for the competition reaction between TBDMS-phenol and *p*-cresol (table entry 2).



Figure S4. The ¹H NMR spectra of the phenol reaction mixture, the *p*-cresol reaction mixture and this pseudo firstorder competition experiment in the aromatic region (δ 8.3-6.3) and aliphatic region (δ 2.35-2.10). Product ratios were determined using the integrals of the peaks of interest, belonging to the specific products. Peaks of interest belonging to the phenol product are highlighted in red. Peaks of interest belonging to the *p*-cresol product are highlighted in blue.



5 Mechanism Studies of Bifluoride-Catalyzed SuFEx reactions

Figure S5.. Bifluoride-based (a) traditional SuFEx, (b) traditional SuFEx with 0.2 equiv of catalyst, (c) silicon-free SuFEx, (d) deprotection and (e) catalyst-free, silicon-free SuFEx. TBDMS, *tert*-butyldimethylsilyl

Procedures

Reaction of TBDMS-p-cresol (Figure S5a)

Compound 1 (83 mg, 0.3 mmol) and TBDMS-*p*-cresol (67 mg, 0.3 mmol) were added to a flame-dried 10 mL round bottom flask and dissolved in acetonitrile (3 mL). Then KHF₂ (26 mg, 0.3 mmol) and 18-crown-6 ether (87 mg, 0.33 mmol) were added, and the mixture was stirred for 1 h at rt, during which the reaction reached completion. The reaction mixture was then concentrated and subjected to column chromatography (petroleum ether 4:1 ethyl acetate). The final product was obtained as a white powder (109 mg, 0.3 mmol, >99%). (Full characterization data can be found in 6.4 Experimental data)

Reaction of TBDMS-p-cresol with catalytic KHF2 (Figure S5b)

The procedure was the same as for the previous reaction, except that the amounts of the KHF₂ and 18-crown-6 ether were reduced to 0.2 equiv (5.2 mg, 0.06 mmol) and 0.22 equiv (17 mg, 0.066 mmol) respectively. The mixture

was stirred for 30 h at rt, at which time the reaction had reached completion. The final product was obtained as a white powder (108 mg, 0.3 mmol, >99%) after flash column chromatography (petroleum ether 4:1 ethyl acetate).

Reaction of *p*-cresol (Figure S5c)

Compound **1** (83 mg, 0.3 mmol) and *p*-cresol **2b**-*p* (33 mg, 0.3 mmol) were added to a flame dried 10 mL round bottom flask and dissolved in acetonitrile (3 mL). Then KHF₂(26 mg, 0.3 mmol) and 18-crown-6 ether (87 mg, 0.33 mmol) were added, and the mixture was stirred at rt. The reaction mixture was monitored by TLC analysis, and only trace products were found after 24 h.

Deprotection of TBDMS-p-cresol (Figure S5d)

TBDMS-*p*-cresol (55.6 mg, 0.25 mmol) was added to a flame dried 10 mL round bottom flask and dissolved in acetonitrile (2 mL). Then KHF₂ (20 mg, 0.25 mmol) and 18-crown-6 ether (73 mg, 0.28 mmol) were added, and the mixture was stirred for 1 h at rt. The reaction mixture was then concentrated and subjected to column chromatography (petroleum ether 3:1 ethyl acetate). The final product was obtained as a colorless oil (27 mg, 0.24 mmol, >99%).

Reaction of Phenolate (Figure S5e)

Sodium phenoxide **6** (57.2 mg, 0.5 mmol) was added to a solution of compound **1** (136 mg, 0.5 mmol) in CH₃CN (5 mL) and was monitored until completion with TLC. After 10 min the reaction mixture was concentrated in vacuo. The title compound was obtained as a white powder from flash column chromatography on a silica column (heptane:CH₂Cl₂ = 15:85 to 0:100) (163 mg, 0.46 mmol, 93% yield). (Full characterization data can be found in 6.4 Experimental data)

Discussion

The difluoride is indeed an excellent SuFEx reaction catalyst and performed well with our specific substrate **1** in a traditional SuFEx (Figure S5a). When using catalytic amounts of HF_2 anion (0.2 equiv), the same reaction reached completion, albeit at a much slower pace (30h, Figure S5b).

Nevertheless, no SuFEx product was found in Si-free SuFEx (Figure S5c).

Potassium bifluoride is not a strong base, as the pH of the saturated aqueous solution of potassium bifluoride at room temperature is about 3.¹⁶

Therefore, we believe that the difluoride is used here to remove silvl protecting groups and generate phenolates bearing strong nucleophilicities, *in situ*. In fact, potassium bifluoride (combing with 18-crown-6 ether) is an excellent silvl deprotection reagent (Figure S5d). The reactivity of phenolates has been demonstrated in our experiments (Figure S5e).

The difluoride catalyst is indeed a valuable addition to the SuFEx chemistry. However, our investigations were focused on silicon-free SuFEx reactions, so only preliminary investigations were performed in this direction. As we have shown above, the difluoride catalyst does not play a role in this particular reaction.

So, bifluoride-based SuFEx polymerizations were likely triggered by phenolates resulting from the use of bifluorides. As fluoride ions are released from SuFExable agents during polymerization, the bifluoride catalyst loading can therefore be much lower than stoichiometric.

6 Si-free SuFEx reaction – phenol derivative substrate scope



6.1 Phenol derivatives

Table S2. Substrate scope of the Si-free SuFEx reaction using phenol derivatives.

* In the absence of descriptor, the substituent is H.

#			Phenol*			Meth.	Conv/%	Time	lsol vield	Hydroly- sis	Notes
	R ²	R ³	R⁴	R⁵	R ⁶						
3a						а	>95	5 min	93		Phenol
3b- <i>p</i>			Me			а	>95	6 min	87		
3b- <i>m</i>		Me				а	>95	6 min			
3b-o	Me					а	>95	6 min			
3c	Me		Me		Me	а	>95	10 min	91		
3d			OMe			а	>95	2 min	93		
3e		Pyr.				а	>95	3 min	87		3-hydroxy pyridine
3f			NH ₂			а	>95	3 min		Y (2%)	
3g			OCF ₃			а	>95	3 min			
3ĥ			CONH ₂			а	>95	10 min			
3i			COOH			С	>95	10 min			
3j			NHCOMe			а	>95	3 min			
3k		OH				d	>95	5 min	89		Resorcinol
31			[/] Pr-C ₆ H₅OH			d	>95	3 min			BPA
3m- <i>p</i>			F			а	>95	3 min			
3m- <i>m</i>		F				а	>95	5 min			
3m-o	F					а	>95	3 min			
3n		F	F	F		а	93	11 min		Y (5%)	
30			CI			а	>95	3 min	87		
Зр-р			Br			а	>95	3 min			
3p-m		Br				а	>95	3 min			
3р-о	Br					а	>95	30 min	79	Y (3%)	
3q						а	>95	3 min			
3r-p			<i>t</i> -Bu			а	>95	15 min			
3r-o	<i>t</i> -Bu					а	>95	40 min			
3r-o	<i>t</i> -Bu					е	>95	2 min	92		
3s			COOMe			а	>95	25 min			
3t			CH₂OH			а	>95	5 min		Y (18%)	
3u			B(OH) ₂			а	>95	30 min		Y (8%)	
3v			CF₃			а	>95	30 min			
3w			CHO			а	89	30 min		Y (4%)	
3x			SO ₂ -C ₆ H ₆ -O [/] Pr			а	89	30 min		Y (5%)	
3y-p			CN			а	87	30 min		Y (3%)	
3y-m		CN				а	>95	15 min	33 ^c		
3z		CF ₃		CF ₃		а	88	21 h		Y (26%)	
3aa	<i>t</i> -Bu		Me		<i>t</i> -Bu	а	66	21 h		Y (>99%)	
3aa	<i>t</i> -Bu		Me		<i>t</i> -Bu	е	>95	42 min		Y (>99%)	
3ab	CI				CI	а	44	22 h		Y (6%)	
3ac	NO ₂		Me			а	56	70 min	71 ^C	Y (21%)	
3ad			NO ₂			а	77	45 min		Y (53%)	
3zy	CI	CI	CI	CI	CI	а	< 5	27 h		Y (>99%)	

6.2 Natural phenol derivatives

#	Phenolic derivative	Cond.	Conv. /%	Time	Hydrolysis
4a	Eugenol	а	>95	3 min	
4b	Vanillic acid	С	>95	12 min	
4c	Raspberry ketone	а	>95	3 min	
4d	Tyrosol	а	>95	3 min	Y (7%)
4e	Sesamol	а	>95	5 min	
4f	Ferrulic acid	С	>95	6 min	
4g	Thymol	а	>95	15 min	
4h	Salicyl aldehyde	а	96	60 min	Y (2%)
4i	Vanillin	а	93	4h	Y (5%)
4z	Salicyclic acid	С	65	24h	Y (100%)
	Salicyclic acid	е	>95	45 min	Y (100%)

Table S3. Substrate scope of the Si-free SuFEx reaction using natural phenol derivatives.

6.3 Experimental conditions

6.3.1 General observations and methods for determining conversion via ¹H-NMR

The SuFEx reactions were generally fast, clean, high-yielding reactions. The reactions were monitored via ¹H-NMR (in MeCN-*d*₃), whereby the disappearance of starting material (δ = 8.14 (t, *J* = 7.2 Hz, 4H)) coincided with the appearance of product (generally two doublets, with a typical shift around 8.10 and 7.98, both 2H). Furthermore, the starting material methyl peak also shifts upfield upon product formation, from 2.51 ppm to 2.46-2.49 ppm. Hydrolysis of the product can be observed by the formation of a singlet at 2.35/2.36 ppm corresponding to the Mesignal of the –OH hydrolysis product.

6.3.2 General procedures for substrate screening via ¹H-NMR

a) In an NMR tube, fluoride (1 equiv) and the phenol (1.05 equiv) were dissolved in CD₃CN (0.1M). DBU was added (1.0 equiv) and the reaction progress was monitored via ¹H NMR.

b) (used for determination isolated yield) in a flame-dried flask, under Ar flow, SF (1 equiv) and the phenol (1.05 equiv) were dissolved in anhydrous MeCN (0.1 M). DBU was added (1.0 equiv) and the reaction progress was monitored via TLC. Once full conversion was reached, the product was isolated via flash column chromatography. c) in an NMR tube, SF (1 equiv) and the phenol (1.05 equiv) were dissolved in CD₃CN (0.1 M). DBU was added (2.0 equiv) and the reaction progress was monitored via ¹H NMR.

d) in an NMR tube, SF (2.01 equiv) and the phenol (1 equiv) were dissolved in CD₃CN (0.1 M). DBU was added (1.0 equiv) and the reaction progress was monitored via ¹H NMR.

e) in an NMR tube, SF (1 equiv) and the phenol (10 equiv) were dissolved in CD₃CN (0.1 M). DBU was added (10 equiv) and the reaction progress was monitored via ¹H NMR.

6.3.3 Examples of procedures

- a) Sulfonimidoyl fluoride 1 (14.2 mg, 0.051 mmol, 1 equiv) was dissolved in 0.55 mL CD₃CN. Phenol (6.3 mg, 0.067 mmol, 1.3 equiv) was added and the reaction mixture was analyzed via ¹H NMR. DBU (8.1 μL, 0.054 mmol, 1.05 equiv) was added and the mixture was shaken well. Immediately after, ¹H NMR measurements were continued until the starting material had disappeared fully. Full conversion of the starting material towards the desired product was reached after 5 min.
- b) Sulfonimidoyl fluoride 1 (138 mg, 0.50 mmol) was dissolved in anhydrous acetonitrile (5 mL, 0.1 M) in a 25 mL round bottom flask at room temperature. The unprotected phenol was added (0.50 mmol), followed by the addition of DBU (75 µL, 0.50 mmol). The reaction was monitored with TLC until the starting materials

were not observed anymore, or until no change in the TLC was observed. After this, the reaction mixture was concentrated and purified by flash column chromatography, yielding the desired product. Eluents for the purification and the obtained yield are detailed in the data section 5.4.

- c) Sulfonimidoyl fluoride 1 (15.0 mg, 0.054 mmol, 1 equiv) was dissolved in 0.55 mL CD₃CN. 4hydroxybenzoic acid (8.22 mg, 0.060 mmol, 1.1 equiv) was added and the reaction mixture was analyzed via ¹H NMR. DBU (16 μL, 0.108 mmol, 2.0 equiv) was added and the mixture was shaken well. Immediately after, ¹H NMR measurements were continued until the starting material had disappeared fully. Full conversion of the starting material towards the desired product was reached after 10 min.
- d) Sulfonimidoyl fluoride **1** (14.6 mg, 0.053 mmol, 2.2 equiv) was dissolved in 0.55 mL CD₃CN. Resorcinol (2.5 mg, 0.023 mmol, 1 equiv) was added and the reaction mixture was analyzed via ¹H NMR. DBU (8.0 μ L, 0.055 mmol, 1.0 equiv) was added and the mixture was shaken well. Immediately after, ¹H NMR measurements were continued until the free phenol had disappeared fully (characteristic peak, a triplet at 7.01 (t, *J* = 8.1 Hz, 1H). The sulfonimidoyl fluoride starting material was added in excess and the usual peaks to determine full conversion could therefore not be used. Full conversion of the phenol towards the desired product was reached after 5 min.
- e) Sulfonimidoyl fluoride 1 (8.0 mg, 0.029 mmol, 1 equiv) was dissolved in 0.55 mL CD₃CN. 2-*tert*-butyl-phenol (43.4 mg, 0.30 mmol, 10 equiv) was added and the reaction mixture was analyzed via ¹H NMR. DBU (43 μL, 0.29 mmol, 10 equiv) was added and the mixture was shaken well. Immediately after, ¹H NMR measurements were continued until the sulfonimidoyl fluoride starting material had disappeared fully. Full conversion to the desired SuFEx product was obtained after 3 min.

6.4 Experimental data



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.0 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.50 – 7.42 (m, 4H), 7.38–7.27 (m, 3H), 7.11–7.06 (m, 2H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₈NO₃S: 352.1002, found: 352.0999.

Isolated product

The title compound was obtained as a grey powder from flash column chromatography on a silica gel (CH₂Cl₂:P.E. - 5:1) (163 mg, 93%). ¹H NMR (400 MHz, CDCl₃) δ = 8.14 (d, *J* = 7.1 Hz, 2H), 7.96 (d, *J* = 8.4 Hz, 2H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.45-7,35 (m, 4H), 7.34–7.22 (m, 3H), 7.12 (d, *J* = 7.8 Hz, 2H), 2.47 (s, 3H). ¹³C NMR (100 MHz, C₆D₆) δ = 171.4, 150.0, 145.0, 136.3, 134.6, 132.4, 130.2, 129.86, 129.82, 128.5, 128.4, 127.3, 123.4, 21.2. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₈NO₃S: 352.1002, found: 352.0999. IR (neat): 3062, 1640, 1585, 1267 cm⁻¹.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.0 Hz, 2H), 7.92 (d, *J* = 8.0 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.48-7.42 (m, 4H), 7.14 (d, *J* = 8.3 Hz, 2H), 6.96 (d, *J* = 8.5 Hz, 2H), 2.46 (s, 3H), 2.28 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₃S: 366.1158, found: 366.1148.

Isolated product

The title compound was obtained as a white powder from flash column chromatography on silica gel (CH₂Cl₂: P.E.-5:1) (160 mg, 87%). ¹H NMR (400 MHz, CDCl₃) δ = 8.14 (d, *J* = 8.3, 2H), 7.95 (d, *J* = 8.4 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.44–7.33 (m, 4H), 7.08 (d, *J* = 8.3 Hz, 2H), 6.99 (d, *J* = 8.6 Hz, 2H), 2.47 (s, 3H), 2.30 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 172.0, 147.1, 145.7, 137.4, 135.5, 133.5, 132.5, 130.3, 130.0, 129.8, 128.5, 128.2, 122.7, 21.9, 21.1. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₃S: 366.1158, found: 366.1148. IR (neat): 3060, 2915, 1643, 1595, 1500, 1251 cm⁻¹.





Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 7.4 Hz, 2H), 7.93 (d, *J* = 8.3 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.46 (t, *J* = 6.8 Hz, 4H), 7.21 (t, *J* = 7.9 Hz, 1H), 7.12 (d, *J* = 7.5 Hz, 1H), 6.92 (s, 1H), 6.87 (d, *J* = 8.1 Hz, 1H), 2.46 (s, 3H), 2.25 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₃S: 366.1158, found: 366.1149.



reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.06 (d, *J* = 7.2 Hz, 2H), 7.95 (d, *J* = 8.4 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.46 (t, *J* = 8.3 Hz, 4H), 7.30–7.12 (m, 4H), 2.46 (s, 3H), 2.16 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₃S: 366.1158, found: 366.1150.

Isolated product

The title compound was obtained as a white powder from flash column chromatography on silica (hexane : CH_2CI_2 = 15 : 85) (169 mg, 92%).

¹H NMR (400 MHz, CDCl₃) δ = 8.13 (d, *J* = 8.0 Hz, 2H), 7.97 (d, *J* = 8.0 Hz, 2H), 7.52 (t, *J* = 7.0 Hz, 1H), 7.45–7.33 (m, 4H), 7.23–7.08 (m, 4H), 2.48 (s, 3H), 2.22 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 171.9, 148.2, 145.8, 135.5, 134.1, 132.5, 132.1, 131.8, 130.1, 129.8, 128.2, 128.2, 127.4, 127.0, 122.9, 21.9, 16.7. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₃S: 366.1158, found: 366.1150. IR (neat): 3068, 2918, 1642, 1596, 1270 cm⁻¹.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 7.96-8.04 (m, 4H), 7.57 (t, *J* = 7.4 Hz, 1H), 7.49 (d, *J* = 8.3 Hz, 2H), 7.52-7.40 (m, 2H), 6.93 (s, 2H), 2.48 (s, 3H), 2.26 (s, 3H), 2.18 (s, 6H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₃H₂₄NO₃S: 394.1471, found: 394.1471.

Isolated product

The title compound was obtained as a white powder from flash column chromatography on a silica gel (CH₂Cl₂: P.E. - 4:1) (179 mg, 91%). ¹H NMR (400 MHz, CDCl₃) δ = 8.07 (d, *J* = 8.0 Hz, 2H), 8.00 (d, *J* = 8.4 Hz, 2H), 7.50 (t, *J* = 7.4 Hz, 1H), 7.45-7.34 (m, 4H), 6.85 (s, 2H), 2.47 (s, 3H), 2.26 (s, 3H), 2.19 (s, 6H). ¹³C NMR* (100 MHz, CDCl₃) δ = 171.7, 145.4, 145.3, 136.8, 135.6, 135.2, 132.4, 130.1, 129.7, 128.2, 128.1, 21.9, 20.8, 17.8; (100 MHz, C₆D₆) δ = 171.2, 146.0, 144.8, 136.58, 136.55, 136.2, 132.7, 132.3, 130.3, 130.1, 129.9, 21.2, 20.6, 17.9. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₃H₂₄NO₃S: 394.1471, found: 394.1460. IR (neat): 3065, 2924, 1655, 1265 cm⁻¹. (The ¹³C NMR was obtained with two different solvents to account for overlap of carbon peaks. For example, peaks located at 132.4 and 130.1 ppm in CDCl₃ are split into two peaks when C₆D₆ was adopted, respectively 132.7 & 132.3, 130.3 & 130.1. Peaks located at 128.2 and 128.1 ppm in CDCl₃ will overlap with C₆H₆ solvent residue peaks.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.0 Hz, 2H), 7.91 (d, *J* = 8.0 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.50-7.43 (m, 4H), 6.99 (d, *J* = 9.1 Hz, 2H), 6.84 (d, *J* = 9.1 Hz, 2H), 3.73 (s, 3H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₄S: 382.1108, found: 382.1096.

Isolated product

¹H NMR (400 MHz, CDCl₃) δ = 8.15 (d, *J* = 8.0 Hz, 2H), 7.93 (d, *J* = 8.0 Hz, 2H), 7.51 (t, *J* = 7.3 Hz, 1H), 7.45-7.30 (m, 4H), 7.01 (d, *J* = 9.1 Hz, 2H), 6.77 (d, *J* = 9.1 Hz, 2H), 3.74 (s, 3H), 2.45 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 172.0, 158.5, 145.7, 142.7, 135.4, 133.1, 132.5, 130.0, 129.7, 128.4, 128.2, 123.9, 114.6, 55.6, 21.8. HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₁H₂₀NO₄S: 382.1108, found: 382.1096.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.49 (d, *J* = 4.1 Hz, 1H), 8.28 (s, 1H), 8.08 (d, *J* = 7.5 Hz, 2H), 7.95 (d, *J* = 7.8 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.49 (q, J = 9.9, 8.4 Hz, 5H)., 7.35 (dd, *J* = 7.6, 5.1 Hz, 1H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₁₉H₁₇N₂O₃S: 353.0954, found: 353.0949.

Isolated product

The title compound was obtained as a yellow crystal from flash column chromatography on a silica gel (petroleum ether 2:1 ethyl acetate) (0.1534 g, 87%). ¹H NMR (400 MHz, CDCl₃) δ = 8.51 (d, *J* = 4.3 Hz, 1H), 8.23 (d, *J* = 1.4 Hz 1H), 8.14 (d, *J* = 8.0 Hz, 2H), 7.97 (d, *J* = 8.4 Hz, 2H), 7.66 (ddd, *J* = 8.4, 2.6, 1.4 Hz, 1H), 7.54 (t, *J* = 7.4 Hz, 1H), 7.46–7.37 (m, 4H), 7.31 (dd, *J* = 8.4, 4.7 Hz, 1H), 2.49 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 171.7, 148.5, 146.5, 146.4, 144.4, 135.0, 132.8, 132.7, 131.0, 130.3, 129.8, 128.3, 128.3, 124.4, 21.9. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₁₉H₁₇N₂O₃S: 353.0954, found: 353.0949. IR (neat, cm⁻¹): 3063, 2923, 1648, 1254, 1145.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

During the reaction, a trace of hydrolysis product was obtained (<2%).

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.0 Hz, 2H), 7.89 (d, *J* = 8.0 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.50-7.42 (m, 4H), 6.74 (d, *J* = 8.0 Hz, 2H), 6.50 (d, *J* = 8.0 Hz, 2H), 4.41 (br s, 2H*), 2.45 (s, 3H). *also starting material OH and NH. HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₀H₁₉N₂O₃S: 367.1111, found: 367.1098.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.3 Hz, 2H), 7.95 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.51–7.42 (m, 4H), 7.27 (d, *J* = 8.0 Hz, 2H), 7.19 (d, *J* = 8.0 Hz, 2H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₇F₃NO₄S: 436.0825, found: 436.0812.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ 8.19 (br s, 1H), 8.08 (d, J = 7.1 Hz, 2H), 7.98-7.90 (m, 4H), 7.58 (t, J = 7.4 Hz, 1H), 7.50–7.42 (m, 4H), 7.13 (d, J = 8.8 Hz, 2H), 6.42 (br s, 1H), 2.45 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₉N₂O₄S: 395.1060, found: 395.1049.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.0 Hz, 2H), 7.92 (d, *J* = 7.9 Hz, 2H), 7.86 (d, *J* = 8.1 Hz, 2H), 7.58 (t, *J* = 7.2 Hz, 1H), 7.49-7.43 (m, 4H), 6.97 (d, *J* = 8.2 Hz, 2H), 2.45 (s, 3H). HRMS (ESI) *m/z* [*M*-H]⁻ calcd. for C₂₁H₁₆NO₅S: 394.0755, found: 394.0749.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.3 Hz, 2H), 7.90 (d, *J* = 8.3 Hz, 2H), 7.63 (d, *J* = 9.0 Hz, 2H), 7.57 (t, *J* = 7.4 Hz, 1H), 7.48-7.41 (m, 4H), 6.93 (d, *J* = 9.1 Hz, 2H), 5.34 (br s, 1H)*, 2.44 (s, 3H), 2.02 (s, 3H). *NH overlaps with starting material. HRMS (ESI) *m/z* [*M*+H]* calcd. for C₂₂H₂₁N₂O₄S: 409.1217, found: 409.1212.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess sulfonimidoyl fluoride)

¹H NMR (400 MHz, CD₃CN) δ = 8.07 (d, *J* = 8.0 Hz, 4H), 7.90-7.84 (m, 4H), 7.62–7.57 (m, 2H), 7.50–7.42 (m, 8H), 7.32(td, *J* = 8.4, 1.6 Hz, 1H), 7.09-7.02 (m, 2H), 6.97 (t, *J* = 2.2 Hz, 1H), 2.46 (s, 6H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₃₄H₂₉N₂O₆S₂: 625.1462, found: 625.1446.

Isolated product

The title compound was obtained as a purple liquid by flash column chromatography on silica gel (P.E.: EtOAc – 3:2, 277 mg, 89%*). *both diastereoisomers.

¹H NMR (400 MHz, CDCl₃) δ = 8.15–8.07 (m, 8H), 7.92-7.85 (m, 8H), 7.57–7.48 (m, 4H), 7.46-7.38 (m, 8H), 7.38-7.31 (m, 8H), 7.28 (d, 3.2 Hz, 1H), 7.24 (d, 2.8 Hz, 1H), 7.16-7.09 (m, 4H), 6.95 (t, *J* = 4 Hz, 1H), 6.90 (t, *J* = 4 Hz, 1H), 2.47 (s, 6H), 2.46 (s, 6H). Two sets of peaks can be found in a 1:1 ratio. The ratio was determined by comparing the integral of two triplets found at δ = 6.90-6.95. ¹³C NMR (100 MHz, CDCl3) δ = 171.7, 157.5, 150.0, 149.6, 146.2, 135.1, 133.3, 132.9, 132.8, 132.7, 132.5, 130.3, 130.0, 130.1, 130.0, 129.82, 129.79, 128.4, 128.25, 128.18, 128.0, 122.2, 122.1, 118.0, 114.9, 114.4, 110.5, 29.71, 21.9, 21.8. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₃₄H₂₉N₂O₆S₂: 625.1462, found: 625.1446. **IR** (neat): 3026, 2924, 1647, 1595, 1260 cm⁻¹.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess sulfonimidoyl fluoride)

¹H NMR (400 MHz, CD₃CN) δ = 8.04 (d, *J* = 8.0 Hz, 4H), 7.91 (d, *J* = 8.4 Hz, 4H), 7.57 (t, *J* = 7.4 Hz, 2H), 7.47-7.40 (m, 8H), 7.11 (d, *J* = 8.8 Hz, 4H), 6.98 (d, *J* = 8.8 Hz, 4H), 2.44 (s, 6H), 1.55 (s, 6H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₄₃H₃₉N₂O₆S₂: 743.2244, found: 743.2224.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.3, 2H), 7.93 (d, *J* = 8.5 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.52-7.40 (m, 4H), 7.14–7.03 (m, 4H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇FNO₃: 370.0908, found: 370.0898.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.10 (d, *J* = 8.2 Hz, 2H), 7.97 (d, *J* = 8.2 Hz, 2H), 7.61 (t, *J* = 6.9 Hz, 1H), 7.52-7.45 (m, 4H), 7.42-7.33 (m, 1H), 7.10 (td, *J* = 8.5, 2.3 Hz, 1H), 6.98-6.89 (m, 2H), 2.48 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇FNO₃: 370.0908, found: 370.0897.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.07 (d, *J* = 7.9 Hz, 2H), 7.96 (d, *J* = 8.0 Hz, 2H), 7.59 (t, *J* = 7.3 Hz, 1H), 7.50-7.43 (m, 4H), 7.39-7.28 (m, 2H), 7.23–7.14 (m, 2H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇FNO₃: 370.0908, found: 370.0896.



3n

The reaction with 3,4,5-trifluorophenol was rather fast, yielding a high conversion after 2 min. Conversion of the starting material increased, but after 11 min hydrolysis of the product started to diminish the yield of product.

t = 2 min		%	t = 11 min	%
	Conversion	85	Conversion	93
Of which	Starting material	15	Starting material	7
	Product	82	Product	87
	Hydrolysis of product	2	Hydrolysis of product	5

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.11 (d, *J* = 7.9 Hz, 2H), 7.98 (d, *J* = 8.0 Hz, 2H), 7.62 (t, *J* = 7.1 Hz, 1H), 7.55 – 7.46 (m, 4H), 7.01–6.90 (m, 2H), 2.49 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₅F₃NO₃S: 406.0719, found: 406.0710.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 7.4 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.52-7.43 (m, 4H),), 7.34 (d, *J* = 9.0 Hz, 2H), 7.07 (dd, *J* = 8.9, 2.1 Hz, 2H), 2.46 (s, 3H). HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₀H₁₇NO₃SCI: 386.0612, found: 386.0598.

Isolated product

The title compound was obtained as a white powder by flash column chromatography (P.E.:EtOAc – 6:1) (168 mg 87%). ¹H NMR (400 MHz, CDCl₃) δ = 8.16 (d, *J* = 7.3 Hz, 2H), 7.97 (d, *J* = 8.4 Hz, 2H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.48–7.38 (m, 4H), 7.29 (d, *J* = 6.8 Hz, 2H), 7.09 (d, *J* = 8.9 Hz, 2H), 2.51 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 171.8, 147.8, 146.1, 135.2, 133.2, 133.0, 132.7, 130.1, 129.9, 129.8, 128.4, 128.3, 124.4, 21.9. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇NO₃SCI: 386.0612, found: 386.0605. IR (neat, cm⁻¹): 3062, 1650, 1483, 1248.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 9.2 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.53–7.42 (m, 6H), 7.07–6.97 (m, 2H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇BrNO₃S: 430.0107, found: 430.0096.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.2, 2H), 7.95 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.52–7.43 (m, 5H), 7.30 (t, *J* = 2.1 Hz, 1H), 7.27 (t, *J* = 8.2 Hz, 1H), 7.08 (ddd, *J* = 8.3, 2.3, 0.9 Hz, 1H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇BrNO₃S: 430.0107, found: 430.0096.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

While the reaction proceeded to full conversion in 30 min, some hydrolysis of the final product was obtained as well.

t = 30 min		%
	Conversion	>95
Of which	Starting material	0
	Product	96.5
	Hydrolysis of product	3.5

¹H NMR (400 MHz, CD₃CN) δ = 8.07 (d, *J* = 7.2 Hz, 2H), 8.01 (d, *J* = 8.4 Hz, 2H), 7.66-7.58 (m, 2H), 7.58-53 (m, 1H), 7.53-7.44 (m, 4H), 7.42 (td, *J* = 7.9, 1.6 Hz, 1H), 7.25 (td, *J* = 8.0, 1.4 Hz, 1H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇BrNO₃S: 430.0107, found: 430.0093.

Isolated product

The title compound was obtained as a white powder from flash column chromatography (CH₂Cl₂:P.E. - 5:1) (171 mg, 79%). ¹H NMR (400 MHz, CDCl₃) δ = 8.13 (d, *J* = 7.1 Hz, 2H), 8.03 (d, *J* = 8.4 Hz, 2H), 7.67 (dd, *J* = 8.2, 1.4 Hz, 1H), 7.57 - 7.49 (m, 2H), 7.40 (dd, *J* = 16.3, 8.2 Hz, 4H), 7.33 (d, *J* = 8.2 Hz, 1H), 7.15 (td, *J* = 7.9, 1.4 Hz, 1H), 2.48 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 171.3, 147.1, 146.1, 135.3, 134.0, 133.8, 132.7, 130.0, 129.9, 128.8, 128.6, 128.4, 128.3, 125.3, 116.6, 21.9. HRMS (ESI) *m/z* [*M*+H]⁺ calcd for C₂₀H₁₇BrNO₃S: 430.0107, found: 430.0099. IR (neat): 3078, 1642, 1468, 1271 cm⁻¹.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 7.3 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 2H), 7.68 (d, *J* = 8.9 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.50–7.43 (m, 4H), 6.87 (d, *J* = 8.9 Hz, 2H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₇INO₃S: 477.9968, found: 477.9957.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.07 (d, *J* = 7.9 Hz, 2H), 7.94 (d, *J* = 8.0 Hz, 2H), 7.58 (t, *J* = 7.3 Hz, 1H), 7.49-7,44 (m, 4H), 7.38 (d, *J* = 8.3 Hz, 2H), 7.02 (d, *J* = 8.4 Hz, 2H), 2.46 (s, 3H), 1.26 (s, 9H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₄H₂₆NO₃S: 408.1628, found: 408.1618.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08-8.01 (m, 4H), 7.57 (t, *J* = 7.4 Hz, 1H), 7.54–7.51 (m, 1H), 7.51–7.40 (m, 5H), 7.23–7.15 (m, 2H), 2.46 (s, 3H), 1.34 (s, 9H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₄H₂₆NO₃S: 408.1628, found: 408.1617.

Isolated product

The title compound was obtained as a yellow oil from flash column chromatography on silica. (P.E.:CH₂Cl₂ - 15:85) (189 mg, 0.46 mmol, 92%). ¹H NMR (400 MHz, CDCl₃) δ = 8.13–8.05 (m, 4H), 7.58–7.53 (m, 1H), 7.53–7.47 (m, 1H), 7.43–7.36 (m, 5H), 7.20–7.12 (m, 2H), 2.47 (s, 3H), 1.37 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ = 171.6, 149.3, 145.6, 141.1, 135.5, 135.3, 132.5, 130.2, 129.8, 128.2, 128.13, 128.09, 127.4, 126.3, 120.8, 34.8, 30.5, 21.9. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₄H₂₆NO₃S: 408.1628, found: 408.1616. IR (neat, cm⁻¹): 3068, 2918, 1642, 1597, 1270



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 8.0 Hz, 2H), 7.97-7.93 (m, 4H), 7.59 (t, *J* = 7.3 Hz, 1H), 7.50-7.42 (m, 4H), 7.19 (d, *J* = 8.3 Hz, 2H), 3.84 (s, 3H), 2.46 (s, 3H). HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₂H₂₀NO₅S: 410.1057, found: 410.1043.



3t

Hydrolysis occurred during the monitoring of the reaction. Full conversion of the starting material towards the product was achieved within 5 min. At this time, hydrolysis made up 18% of the final mixture composition. The peak relations in ¹H NMR were confirmed by COSY.

t = 5 min		%
	Conversion	>95
Of which	Starting material	0
	Product	82
	Hydrolysis of product	18

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 7.3 Hz, 2H), 7.93 (d, *J* = 8.3 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.49-7.42 (m, 4H), 7.30 (d, *J* = 8.3 Hz, 2H), 7.03 (t, *J* = 8.9 Hz, 2H), 4.50 (s, 1.5H)*, 4.39 (s, 0.5H)*, 2.46 (s, 3H). *(belong to benzyl CH₂). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₂₀NO₄S: 382.1108, found: 382.1093.



3u 4-hydroxy phenylboronic acid product

While the reaction proceeded to full conversion in 30 min, some hydrolysis of the final product was obtained as well. Moreover, LC-MS analysis also showed a product corresponding whereby loss of the boronic acid occurred.

t = 30 min		%
	Conversion	>95
Of which	Starting material	0
	Product	92
	Hydrolysis of product	8

Reaction mixture

¹H NMR (400 MHz, CD₃CN) δ = 8.13-8.06 (m, 2H), 7.94-7.89 (m, 2H), 7.67 (d, *J* = 7.6 Hz, 1H), 7.62–7.54 (m, 1H), 7.51–7.42 (m, 4H), 7.38-7,31 (m, 1H), 6.92 (d, *J* = 7.8 Hz, 1H), 6.83 (d, *J* = 8.3 Hz, 1H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₀H₁₉BNO₅S: 396.1072, found: 396.1058.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.08 (d, *J* = 7.5 Hz, 2H), 7.97 (d, *J* = 7.9 Hz, 2H), 7.68 (d, *J* = 8.2 Hz, 2H), 7.59 (t, *J* = 7.3 Hz, 1H), 7.54–7.42 (m, 4H), 7.29 (d, *J* = 8.2 Hz, 2H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₇F₃NO₃S: 420.0876, found: 420.0864.



The reaction suffered from incomplete conversion of the starting material, as well as hydrolysis of the product. Conversion increased over time, but the amount of desired product decreased over time, due to hydrolysis of said product. At t=30 min, an optimum was reached.

t = 30 min		%
	Conversion	89
Of which	Starting material	11
	Product	85
	Hydrolysis of product	4

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 9.96 (s, 1H), 8.10 (d, *J* = 8.3, 2H), 7.98 (d, *J* = 8.5 Hz, 2H), 7.89 (d, *J* = 8.7 Hz, 2H), 7.60 (t, *J* = 7.4 Hz, 1H), 7.53-7.46 (m, 4H), 7.31 (d, *J* = 8.6 Hz, 2H), 2.48 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₈NO₄S: 380.0951, found: 380.0940.



3х

The reaction was monitored continuously via ¹H NMR. After 30 min, an optimum in conversion was reached, while hydrolysis of the product was not too significant. After this time, conversion did not improve significantly, while hydrolysis of the product increased.

t = 30 min		%
	Conversion	89
Of which	Starting material	11
	Product	84
	Hydrolysis of product	5

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.05 (d, *J* = 8.4 Hz, 2H), 7.95 (d, *J* = 8.4 Hz, 2H), 7.89 (d, *J* = 8.8 Hz, 2H), 7.80 (d, *J* = 8.9 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.50–7.41 (m, 4H), 7.27 (d, *J* = 8.8 Hz, 2H), 7.00 (d, *J* = 8.9 Hz, 2H), 4.73–4.64 (m, 1H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₉H₂₈NO₆S₂: 550.1353, found: 550.1335.



Reaction mixture

The reaction was monitored continuously. After 30 min, an optimum in conversion was reached, while hydrolysis of the product was not too significant. After this time, conversion did not improve significantly, while hydrolysis of the product increased.

T = 70 min		%
	Conversion	87
Of which	Starting material	13
	Product	84
	Hydrolysis of product	3

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.09 (d, *J* = 7.4 Hz, 2H), 7.97 (d, *J* = 8.3 Hz, 2H), 7.74 (d, *J* = 8.7 Hz, 2H), 7.61 (t, *J* = 7.4 Hz, 1H), 7.54–7.44 (m, 4H), 7.29 (t, *J* = 7.7 Hz, 3H), 2.48 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₇N₂O₃S: 377.0954, found: 377.0935.

Isolated product

The title compound was obtained as a white powder from flash column chromatography (CH₂Cl₂: P.E. – 5:1) (60 mg, 33%). ¹H NMR (400 MHz, CDCl₃) δ = 8.12 (d, *J* = 7.1 Hz, 2H), 7.97 (d, *J* = 8.4 Hz, 2H), 7.63 (d, *J* = 8.8 Hz, 2H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.47-7.39 (m, 4H), 7.28 (d, *J* = 8.8 Hz, 2H), 2.50 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 171.6, 152.6, 146.5, 134.9, 134.0, 133.05, 132.96, 130.3, 129.8, 128.4, 128.2, 124.1, 117.9, 111.5, 21.9. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₇N₂O₃S: 377.0954, found: 377.0951. IR (neat): 3069, 2225, 1649, 1595, 1490, 1266 cm⁻¹.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.12–8.06 (m, 2H), 7.97 (d, *J* = 8.4 Hz, 2H), 7.69 (d, *J* = 7.7 Hz, 1H), 7.61 (t, *J* = 7.4 Hz, 1H), 7.57–7.51 (m, 2H), 7.51–7.45 (m, 4H), 7.43 (ddd, *J* = 8.3, 2.3, 1.0 Hz, 1H), 2.49 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₇N₂O₃S: 377.0954, found: 377.0940.



3z

The reaction was monitored continuously. While conversion steadily grew, hydrolysis of the product eventually increased at a faster pace than conversion. When the reaction progress was compared between 3 and 21 h, conversion went up slightly, albeit at the cost of a higher proportion of hydrolysis. The results are presented after 21 h after the addition of DBU. After this time, conversion did not improve significantly, while hydrolysis of the product increased.

t = 21 h		%
	Conversion	88
Of which	Starting material	12
	Product	60
	Hydrolysis of product	28

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.09 (d, J = 8.2, 2H), 7.99-7.99 (m, 2H), 7.62 (t, J = 7.4 Hz, 1H), 7.55-7.48 (m, 4H), 7.21 (s, 2H), 7.11 (s, 1H), 2.49 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₂H₁₆F₆NO₃S: 488.0750, found: 488.0728.



3aa

Data was taken from a sample reacted with 10 equivalents of phenol, as the conversion for the 1:1 reaction was very slow. Over time, no desired SuFEx product was formed. Instead, the hydrolysis product was obtained as the sole product, without forming the desired product as an intermediate. The data obtained are therefore in concurrence with the hydrolysis product.

Reaction mixture

(from crude reaction mixture, also containing 10 equivalents of DBU and 10 equivalents of phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.02 (d, J = 8.3, 2H), 7.87 – 7.81 (m, 2H), 7.40 – 7.28 (m, 3H), 7.21 (d, J = 8.0 Hz, 1.25 Hz) 2H), 6.99 (s, 2H)* 2.38 (s, 3H), 2.25 (s, 3H)*, 1.44 (s, 18H)*. HR-MS (ESI) m/z [M-H]⁻ calc for C₁₂H₁₂NO₃S, 274.0543. found 274.0545.

* overlaps with excess 2,6-di-tert-butyl-4-methyl-phenol.



3ab

The reaction with 2,6-dichlorophenol was very slow and hydrolysis of the product occurred instantaneously. While conversion did increase over time, hydrolysis of the product occurred faster, hence the results after 22 h are presented.

T = 22h		%
	Conversion	45
Of which	Starting material	55
	Product	39
	Hydrolysis of product	6

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.10 (d, J = 8.0, 2H), 7.92 (d, J = 8.0 Hz, 2H), 7.57 (d, J = 8.3 Hz, 3H), 7.48 (d, J = 8.0 Hz, 2H), 7.57 (d, J = 8.3 Hz, 3H), 7.48 (d, J = 8.0 Hz, 2H), 7.57 (d, J = 8.3 Hz, 3H), 7.48 (d, J = 8.0 Hz, 2H), 7.57 (d, J = 8.3 Hz, 3H), 7.48 (d, J = = 8.1 Hz, 2H), 7.43 (t, J = 7.7 Hz, 2H), 7.30 (dd, J = 8.6, 7.6 Hz, 1H), 2.49 (s, 3H). HRMS (ESI) m/z [M+H]⁺ calcd for C₂₀H₁₆Cl₂NO₃S: 420.0222, found: 420.0207.



3ac

Reaction mixture

Using non-anhydrous conditions, the conversion was determined using ¹H-NMR.

The reaction was monitored continuously. The reaction mixture analysis after 70 min is presented, because after this time the yield of product did not significantly increase. However, hydrolysis did increase significantly.

T = 70 min		%
	Conversion	56
Of which	Starting material	44
	Product	35
	Hydrolysis of product	21

Isolated product

The product was successfully isolated after performing the reaction in anhydrous conditions. This somewhat suppressed the formation of the hydrolysis product, but it was still present.

The title compound was obtained as a white powder from flash column chromatography (CH₂Cl₂:P.E. - 5:1), providing the product (145 mg, 71% yield). ¹H NMR (400 MHz, CDCl₃) δ = 8.07 (d, *J* = 7.5 Hz, 2H), 7.99 (d, *J* = 8.4 Hz, 2H), 7.73–7.70 (m, 1H), 7.68 (d, *J* = 8.4 Hz, 1H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.45 (dd, *J* = 8.4, 2.0 Hz, 1H), 7.44-7.39 (m, 4H), 2.49 (s, 3H), 2.43 (s, 3H). ¹³C NMR (100 MHz, C₆D₆) δ = 171.4, 145.9, 143.3, 139.4, 138.1, 135.8, 134.5, 133.8, 132.7, 130.2, 130.1, 128.5, 128.4, 126.5, 125.8, 21.3, 20.1. HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₉N₂O₅S: 411.1009, found: 411.1002. IR (neat): 3068, 1642, 1530, 1266 cm⁻¹.



3ad *p*-nitrophenol product

Using non-anhydrous conditions, the conversion was determined using ¹H-NMR.

The reaction mixture analysis after 45 min is presented, because after this time the yield of product did not significantly increase. However, hydrolysis did increase significantly and was already the predominant product.

T = 45 min		%
	Conversion	77
Of which	Starting material	23
	Product	24
	Hydrolysis of product	53

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.21 (d, *J* = 9.2 Hz, 2H), 8.10 (d, *J* = 7.1 Hz, 2H), 8.03-7.99 (m, 2H), 7.64–7.59 (m, 1H), 7.52–7.46 (m, 4H), 7.40–7.35 (m, 2H), 2.49 (s, 3H). HRMS (ESI) *m*/*z* [*M*-H]⁻ calcd. for C₂₀H₁₆N₂O₅S: 395.0696, found 395.0696.



The conversion with pentachlorophenol was very low and product was very easily hydrolyzed.

With mass analysis, the only product found were starting material and the hydrolyzed product, while the desired product was not found. Therefore, no assigned ¹H NMR data are presented. HRMS (ESI) m/z [M+H]⁺ calcd. for C₂₀H₁₂Cl₅NO₃SNa: 543.8873, found: 543.8876.

Natural phenol substrates



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.07 (d, *J* = 7.3 Hz, 2H), 7.91 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.51–7.40 (m, 4H), 7.23 (d, *J* = 8.2 Hz, 1H), 6.80 (s, 1H), 6.76 (dd, *J* = 8.2, 1.8 Hz, 1H), 5.96 (ddt, *J* = 17.0, 10.3, 6.7 Hz, 1H), 5.13 – 5.02 (m, 2H), 3.46 (s, 3H), 3.30 (d, *J* = 6.0 Hz, 2H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₄H₂₄NO₄S: 422.1421, found: 422.1409.



Reaction mixture

(from crude reaction mixture, also containing 2 equivalents of DBU and excess phenol)

1H NMR (400 MHz, CD₃CN) δ = 10.50 (br. s, 1H), 8.08 (d, *J* = 8.0 Hz, 2H), 7.90 (d, *J* = 7.9 Hz, 2H), 7.59 (t, *J* = 7.3 Hz, 1H), 7.53 (s, 1H), 7.50-7.45 (m, 3H), 7.42 (d, *J* = 8.0 Hz, 2H), 7.22 (d, *J* = 8.2 Hz, 1H), 3.47 (s, 3H), 2.45 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₂H₂₀NO₆S: 426.1006, found: 426.0991.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.09 (d, *J* = 8.2, , 2H), 7.94 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.51-7.43 (m, 4H), 7.18 (d, *J* = 8.6 Hz, 2H), 7.00 (d, *J* = 8.6 Hz, 2H), 2.83 – 2.76 (m, 2H), 2.74 – 2.70 (m, 2H), 2.47 (s, 3H), 2.05 (s, 3H). HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₄H₂₄NO₄S: 4422.1421, found: 422.1412.



The reaction proceeded quickly to full conversion, within 3 minutes. Some hydrolysis product was obtained at that time, which was confirmed by HPLC-MS analysis.

t = 3 min		%
	Conversion	>99
Of which	Starting material	0
	Product	93
	Hydrolysis of product	7

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.09 (d, *J* = 7.1 Hz, 2H), 7.94 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.51–7.43 (m, 4H), 7.21 (d, *J* = 8.5 Hz, 2H), 7.00 (d, *J* = 8.6 Hz, 2H), 3.64 (t, *J* = 6.8 Hz, 2H), 2.74 (t, *J* = 6.7 Hz, 2H), 2.47 (s, 3H). HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₂H₂₂NO₄S: 396.1264, found: 396.1252.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.10 (d, *J* = 7.2 Hz, 2H), 7.94 (d, *J* = 8.4 Hz, 2H), 7.60 (t, *J* = 7.4 Hz, 1H), 7.50-7.44 (m, 4H), 6.74 (d, *J* = 8.5 Hz, 1H), 6.62 (d, *J* = 2.3 Hz, 1H), 6.53 (dd, *J* = 8.5, 2.4 Hz, 1H), 5.98 (s, 2H), 2.48 (s, 3H). HRMS (ESI) *m*/*z* [*M*+H]⁺ calcd. for C₂₁H₁₈NO₅S: 396.0900, found: 396.0885.



Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 9.79 (br. s, 1H), 8.08 (d, *J* = 8.0 Hz, 2H), 7.92 (d, *J* = 8.0 Hz, 2H), 7.59 (t, *J* = 7.3 Hz, 1H), 7.51–7.40 (m, 4H), 7.26 (d, *J* = 8.3 Hz, 1H), 7.17 (d, *J* = 15.8 Hz, 1H), 7.10 (s, 1H), 7.06 (d, *J* = 8.4 Hz, 1H), 6.45 (d, *J* = 15.9 Hz, 1H), 3.49 (s, 3H), 2.46 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₄H₂₂NO₆S: 452.1162, found: 452.1148.



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Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 8.07 (d, *J* = 7.1 Hz, 2H), 7.96 (d, *J* = 8.4 Hz, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.52–7.42 (m, 4H), 7.26 (d, *J* = 8.0 Hz, 1H), 7.13 (d, *J* = 7.9 Hz, 1H), 7.03 (s, 1H), 3.29 – 3.18 (m, 1H), 2.47 (s, 3H), 2.24 (s, 3H), 1.09 (d, *J* = 6.9 Hz, 3H), 1.00 (d, *J* = 6.9 Hz, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₄H₂₆NO₃S: 408.1628, found: 408.1616.



The reaction with salicyl aldehyde did not proceed to full completion. After 60 min of reaction time, conversion nearly halted and hydrolysis of the product occurred instead. When the reaction was left for longer, the conversion increased minutely, while hydrolysis of the product increased significantly.

t = 60 min		%
	Conversion	>95
Of which	Starting material	4
	Product	94
	Hydrolysis of product	2

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 10.32 (s, 1H), 8.08 (d, *J* = 7.1 Hz, 2H), 7.97 (d, *J* = 8.4 Hz, 2H), 7.90 (dd, *J* = 7.7, 1.7 Hz, 1H), 7.67–7.56 (m, 2H), 7.52–7.46 (m, 5H), 7.15 (d, *J* = 7.5 Hz, 1H), 2.49 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₁H₁₈NO₄S: 380.0951, found: 380.0937.



The reaction with vanillin did not proceed to full completion. Instead, hydrolysis of the product overtook the formation of desired product. After 4 h of reaction time, an optimum between conversion to the desired product and hydrolysis of said product was reached. When the reaction was left for longer, the conversion increased slightly, while hydrolysis of the product increased significantly.

t = 4h		%
	Conversion	93
Of which	Starting material	7
	Product	88
	Hydrolysis of product	5

Reaction mixture

(from crude reaction mixture, also containing DBU and excess phenol)

¹H NMR (400 MHz, CD₃CN) δ = 9.93 (s, 1H), 8.08 (d, *J* = 7.7 Hz, 2H), 7.95 (d, *J* = 8.2 Hz, 2H), 7.60 (t, *J* = 7.4 Hz, 1H), 7.56 (d, *J* = 8.3 Hz, 1H), 7.54–7.43 (m, 6H), 3.56 (s, 3H), 2.47 (s, 3H). HRMS (ESI) *m/z* [*M*+H]⁺ calcd. for C₂₂H₂₀NO₅S: 410.1057, found: 410.1043.

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Data was taken from a sample reacted with 10 equivalents of salicylic acid, as the conversion for the 1:1 reaction was very slow. Over time, no desired SuFEx product was formed. Instead, the hydrolysis product was obtained as the sole product, without forming the desired product as an intermediate. The data obtained are therefore in concurrence with the hydrolysis product.

Reaction mixture

(from crude reaction mixture, also containing 10 equivalents of DBU and 10 equivalents of salicylic acid). ¹H NMR (400 MHz, CD₃CN) δ = 7.97 (d, *J* = 7.1 Hz, 2H), 7.80 (s, 2H), 7.43 – 7.35 (m, 3H), 7.32 (t, *J* = 7.3 Hz, 3H), 2.35 (s, 3H).

7 Reaction kinetics comparison between a Si-free SuFEx and traditional SuFEx reaction



Scheme S3. The SuFEx reaction of which the kinetics were studied.

The reaction rates of traditional SuFEx reactions were compared to that of the corresponding Si-free SuFEx reactions. As a representative substrate from both groups, TBDMS-protected 2-^{*t*}Bu-phenol and unprotected 2-^{*t*}Bu-phenol were compared. Kinetics were determined under pseudo-first-order conditions, whereby the phenol and DBU were added in 10-fold excess. Reactions were monitored using ¹H NMR spectroscopy. Mesitylene was added as an internal reference. Both reactions reached completion within 2 min at room temperature. To successfully monitor the kinetics, measurements were performed at -30 °C.

Equipment and procedures

The NMR machine was cooled to -30 °C. The NMR sample was pre-cooled using a cryostat set at -30 °C, which was also used to cool DBU. For t=0, a measurement was taken without the addition of DBU. Then, DBU was added, the NMR tube was shaken quickly and placed back in the NMR machine. Once the sample was in place, a measurement was taken (number of scans, NS = 8) immediately. Subsequent measurements were taken using a 'multizg' command (dummy scans, DS = 7, NS = 8, measuring time 1 min) until the reaction had reached completion.

For 2-^tBu-phenol

In an HPLC vial, sulfonimidoyl fluoride **1** (8.0 mg, 0.029 mmol) and 2-^tBu-phenol (43.3 mg, 0.29 mmol, 10 equiv) were dissolved in CD₃CN (0.55 mL, 0.05M). Mesitylene (4.1 μ L, 0.029 mmol) was added as an internal reference and the mixture was shaken well. The solution was transferred to an NMR tube. The solution was cooled to -30 °C. DBU (43 μ L, 0.29 mmol, 10 equiv) was added to start the reaction and ¹H NMR measurements were commenced immediately.

NMR data were collated and globally processed using MestReNova software. Mesitylene was integrated between δ = 2.260 and 2.160 and set to 1.0000 (Me = s, 9H). Starting material was integrated between δ = 8.190 and 8.085 (signal = dd, 4H). Product was integrated between δ = 8.085 and 8.000 (signal = dd, 4H). The integral-data was used in the determination of the conversion and rate constant.

For TBDMS-2-^{*t*}Bu-phenol

In an HPLC vial, sulfonimidoyl fluoride **1** (8.0 mg, 0.029 mmol) and TBDMS-2-^tBu-phenol (76.4 mg, 0.29 mmol, 10 equiv) were dissolved in CD₃CN (0.55 mL, 0.05M). Mesitylene (4.1 μ L, 0.029 mmol) was added as an internal reference and the mixture was shaken well. The solution was transferred to an NMR tube. The solution was cooled to -30 °C, whereby the mixture become opaque and rather solid. DBU (43 μ L, 0.29 mmol, 10 equiv) was added, whereby the sample became liquid again. ¹H NMR measurements were commenced immediately.

NMR data were collated and globally processed using MestReNova software.

Mesitylene was integrated between δ = 6.780 and 6.620 and set to 1.0000 (Ar-H = s, 3H). Starting material was integrated between δ = 8.320 and 8.100 (signal = dd, 4H). Product was integrated between δ = 8.100 and 7.990 (signal = dd, 4H). This integral-data was used in the determination of the conversion and rate constant.

Rate constant determination

Equation 1 describes the second-order rate for the studied reaction (mol⁻¹·s⁻¹), where [1] is the concentration of sulfonimodoyl fluoride and [2] the concentration of phenol, and k_2 is the second–order rate constant (M⁻¹s⁻¹). Since, the amount of phenol derivative (10 equivalents) is in large excess compared to the amount of sulfonimidoyl fluoride, it is assumed to be essentially constant throughout the entire kinetics experiment (equation 2). Thus, the SuFEx reaction can be reduced to pseudo–first order kinetics, with rate constant, k'. Therefore, from plots of ln [$(I_{\infty} - I_t)/(I_{\infty} - I_0)$] versus time, the resulting pseudo first–order rate constant (k') can be obtained directly from the slope, as shown in equation 3.

$$v = k_2[\mathbf{1}][\mathbf{2}] = \frac{d[\mathbf{1}]}{dt}$$
 (1)

$$v = k'[1]$$
 (since [2] >> [1]) (2)

$$\ln\left(\frac{I_{\infty}-I_{t}}{I_{\infty}-I_{0}}\right) = k't \tag{3}$$

So:
$$k' = k_2[2]$$
,
 $k_2 = \frac{k'}{[2]}$

7.1 Si-free SuFEx reaction rate determination





Figure S6. The conversion and Ln-plot of the Si-free SuFEx reaction between 1 and 2-'Bu-phenol, at -30 °C.

From the conversion and the In-plot (Figure S6), we can determine the k' as 0.001576 s⁻¹. With [**2**] being 0.524 M/L, k₂ becomes 0.00300 M⁻¹s⁻¹.

From the duplicate experiment, we obtained a $k^2 = 0.001607 \text{ s}^{-1}$. Using the same concentration for [**2**], k₂ becomes 0.00306 M⁻¹s⁻¹.

Therefore, an average rate of 3.03 x 10⁻³ M⁻¹s⁻¹ was found for the Si-free SuFEx reaction at -30 °C.



7.2 Traditional SuFEx reaction rate determination



Figure S7. The conversion and Ln-plot of the Si-free SuFEx reaction between 1 and TBDMS-2-tBu-phenol, at -30 °C.

The conversion plot for the traditional SuFEx reaction (Figure S7) shows the conversion as an S-plot, meaning the reaction goes through an activation phase before the reaction truly takes off. We hypothesize that this is due to desilylation of the substrate, prior to the occurrence of the reaction. Once the rate-determining desilylation has taken place, the SuFEx reaction occurs rapidly, thereby freeing F⁻ anion, driving the reaction forward. At -30 °C, desilylation by F⁻ is also very rapid, which was demonstrated by a 1:1 reaction between TBDMS-*p*-cresol and tetramethyl ammonium fluoride (TMAF, (CH₃)₄NF). Full desilylation occurred within 2 min at this temperature. In our reactions, DBU can play a double role, as it has been reported to catalyze the desilylation, ^{10,11} while as a base, of course, it also increases the nucleophilicity of the phenol.

From Figure S7, it is clear that the Ln plot is non-linear. However, if we – for the assumption of getting some semiquantitative information – assume that we can still plot it as linear, then we can determine the k' = 0.000551 s⁻¹. With [TBDMS-2] being 0.524 M/L, k₂ = 0.001052 M⁻¹s⁻¹. From the duplo experiment, we obtained a k' = 0.000508 s⁻¹. Using the same concentration for [TBDMS-2], k₂ = 0.00097 M⁻¹s⁻¹.

Therefore, an average rate of $1.01 \times 10^{-3} \text{ M}^{-1} \text{s}^{-1}$ was found for the traditional SuFEx reaction at -30 °C. We do not hold this number to provide any other quantitative information that that it supports that the Si-protected phenol in the traditional SuFEx reactions reacts appreciably slower than phenolates in the Si-free SuFEx reaction.
8 Mechanistic studies

Combining previous reports,¹² three likely mechanisms were proposed (Figure S8). If one enantiomer is used as the substrate for the reaction, a racemized product will be obtained through the S_N1 reaction pathway. Conversely, a stereospecific product will be obtained through the S_N2 or addition-elimination pathway.



Figure S8. Possible mechanisms of SuFEx reaction

8.1 Separation of sulfonimidoyl fluoride enantiomers

The two enantiomers of the sulfonimidoyl fluoride were isolated by HPLC.

HPLC resolution conditions of the sulfonimidoyl fluoride: Chiralpak IA column, hexane/propan-2-ol 90:10, flow rate 0.5 mL/min, oven temperature 40 °C, detector wavelength 240 nm; retention time **isomer-1** $t_1 = 16.8$ min, **isomer-2** $t_2 = 18.8$ min. The purity of each isolated enantiomer exceeded 99% (Figure S9).



a)

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Figure S9. Chromatograms of a) the racemate b) isomer-1 and c) isomer-2.

8.2 Stereoselectivity investigation of Si-Free SuFEx reactions

Two reaction substrates with widely different reaction rates were selected as the research targets. When *p*-cresol **2b**-*p* was used as a substrate, the reaction was completed in 5 min. However, when 4-methyl-2-nitrophenol **2ac** was used as a substrate, the reaction took more than 18 h to complete (Scheme S4).

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Scheme S4. Model reactions in mechanism researches

To a solution of one enantiomer of **1** (1 mL, 3.61 mM, 1 equiv) in CH₃CN, a solution of *p*-cresol or 4-methyl-2nitrophenol (0.5 mL, 14.4 mM, 1 equiv) in CH₃CN and DBU (0.25 mL, 14.4 mM, 1 equiv) were added. After 1.5 h, 0.5 mL reaction mixture was taken out and quenched with 0.5 mL of a saturated aqueous solution of ammonium chloride. The aqueous phase was extracted with ethyl acetate (2 x 0.2 mL). All the organic phases were combined and concentrated. The residue oil was dissolved with a mixed solution of *n*-hexane and isopropanol (90:10) as the sample for HPLC analysis.

% ee was calculated by

$$ee = \frac{majority - minority}{majority + minority} \times 100\%$$
 (4)

where majority and minority refer to the integral area of the respective peaks of the majority and minority products.

8.2.1 Reactions with *p*-cresol **2b**-*p*

HPLC resolution conditions of the compound 3b-p:

Chiralpak IA column, hexane/propan-2-ol 98:2, flow rate 0.5 mL/min, oven temperature 40 °C, detector wavelength 240 nm; retention time $t_1 = 39.0$ min, $t_2 = 50.0$ min (Figure 10).



When **isomer-1** ($t_R = 16.8$ min for sulfonimidoyl fluoride) was used as a starting material, the % *ee* of the product was 73% (Figure S11).

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Figure S11. a) HPLC chromatogram and b) the result of the reaction with isomer-1.

When **isomer-2** ($t_R = 18.8$ min for sulfonimidoyl fluoride) was used as a starting material, the % *ee* of the product was 70% (Figure S12).

a)



b)

Peak	RetTime	Sig	Туре	Area	Height	Area
#	[min]			[mAU*s]	[mAU]	%
1	39.369	1	BB	3.47654e4	453.98547	85.2143
2	50.643	1	BB	6032.22852	66.83584	14.7857

Figure S12. a) HPLC chromatogram and b) the result of the reaction with isomer-2.

The above experiments showed that when the reactant is a single enantiomer, the reaction has enantioselectivity (although % *ee* of final products was only around 70%). If the reaction was via the S_N1 route, the final product should be a racemic mixture. However, no matter which configuration of sulfonimidoyl fluorides was used as the reaction substrate, more than 85% of the products were in a single configuration.

Based on these data, concerted S_N2 or addition-elimination mechanisms thus look more reasonable.

8.2.2 Reactions with 4-methyl-2-nitrophenol 2ac

HPLC resolution conditions of the sulfonimidoyl fluoride: Chiralpak IA column, MeO'Bu/methanol 400:1, flow rate 1.0 mL/min, oven temperature 40 °C, detector wavelength 240 nm; retention time of sulfonimidoyl fluoride **isomer-1** $t_1 = 4.7 \text{ min}$, **isomer-2** $t_2 = 5.3 \text{ min}$ (Figure S13).



Figure S13. Chromatogram of the sulfonimidoyl fluoride racemate 3ac.

HPLC resolution conditions of the product: Chiralpak IA column, MeO'Bu/methanol 400:1, flow rate 1.0 mL/min, oven temperature 40 °C, detector wavelength 240 nm; retention time $t_1 = 7.9$ min, $t_2 = 9.5$ min (Figure S14).

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Figure S14. Chromatogram of the racemate of compound 3ac.

The HPLC result of reaction with isomer-1 (retention time of the sulfonimidoyl fluoride was 4.7 min) (Figure S15).



Figure S15. a) HPLC chromatogram and b) the result of the reaction with isomer-1.

The HPLC result of the reaction with isomer-2 (retention time of the sulfonimidoyl fluoride is 5.3 min) (Figure S16).

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Figure S16. a) HPLC chromatogram and b) the result of the reaction with isomer-2.

First, the chromatogram of this reaction appeared to be more complicated, mainly because of the lower reaction conversion and the presence of unreacted 4-methyl-2-nitrophenol and the sulfonimidoyl fluoride. The above experiments showed that no matter which enantiomer was used as a reaction substrate, the enantioselectivity was not observed (% ee was less than 5%).

The above experimental results were summarized as follows (Table S4)

$R^{1} \longrightarrow OR^{3} +$ 2b-p $R^{1} = Me, R^{2} = H, R^{3} = H$ Min 2ac $R^{1} = Me, R^{2} = NO_{2}, R^{3} = H$ e	nantiomer-1/enantiomer-2	(1 equiv) $(3.6 mM),$ Me 3
Enantiomer ^[a]	phenol	% ee ^[b]
opontiomor 1	2b- <i>p</i>	73%
enantionier-i	2ac	< 2%
enantiomer-2	2b- <i>p</i>	71%
	2ac	< 2%

Table S4. Enantioselective silicon-free SuFEx reactions

Reaction conditions: **enantiomer-1/enantiomer-2** (1 mL, 3.6 mM in CH₃CN), phenolic derivative (1 equiv), DBU (1 equiv), rt.

[a] **enantiomer-1** and **enantiomer-2** refer to two enantiomers with retention times of 16.8 and 18.8 min with chiral HPLC, respectively.

[b] % ee determined using chiral HPLC and calculated by

% ee = $\frac{majority - minority}{majority + minority} \times 100\%$,

[c] 10 equiv **2b-***p* used.

Kinetics of the racemization

Surprisingly, however, in the reaction with 4-methyl-2-nitrophenol, unreacted sulfonimidoyl fluoride also appeared to be racemized. If the sulfonimidoyl fluoride suffered racemization, this could also cause the non-stereospecificity of the product formation (Scheme S5).



Scheme S5. Racemization of the sulfonimidoyl fluoride and SuFEx reactions.

In order to better evaluate the effect of the racemization reaction in SuFEx reaction, its kinetics were studied. Importantly, in the absence of the DBU, the enantiomers of the sulfonimidoyl fluoride were very stable, both in protic solvents such as isopropanol, as well as aprotic solvents, such as acetonitrile.

The kinetics of DBU-promoted sulfonimidoyl fluoride racemization were studied (Scheme S6).



Scheme S6. DBU-promoted sulfonimidoyl fluoride racemization

8.2.3 Modeling the racemization¹³

We assume that this racemization is second order related with the enantiomer and DBU, hence, its reaction rate can be given by

$$r = k \times c[\text{DBU}] \times c[\text{isomer}]$$
(5)

where c[DBU] is the DBU concentration, and c[isomer] is the concentration of the **S** or **R** enantiomer, k is the second order rate constant.

If DBU is used in large excess in this racemization, then the reaction rate can be simplified to

$$r = k' \times c[\text{isomer}] \tag{6}$$

where k' is the pseudo-first order rate constant.

The concentration of the two enantiomers in solution can be described as

$$\frac{dc_S}{dt} = k'(c_S - c_R) \tag{7}$$

$$\frac{dc_R}{dt} = k'(c_S - c_R) \tag{8}$$

where c_S and c_R are the concentrations of the S and R enantiomer, respectively.

$$c_S + c_R = c_0 \tag{9}$$

As the sum of the concentrations of two enantiomers is a constant, i.e. the initial reactant concentration c_0 . The enantiomeric excess is defined by

$$ee = \frac{c_S - c_R}{c_S + c_R} \tag{10}$$

Eq 3 minus eq 4 gives

$$\frac{d(ee)}{dt} = -2k'(ee) \tag{11}$$

Integrating eq 10 with $ee(0) = ee_0$

$$ln(ee) = -2k't + \ln(ee_0)$$
(12)

8.2.4 Kinetic experiments

To the solution of **isomer-1** in CH₃CN (>99% ee,1 mM, 3 mL, 1 equiv) was added DBU (9 μ L, 9.1 mg, 20 equiv) and the resulting solution was stirred at room temperature (20 ± 0.5 °C). To monitor reaction progress, aliquots were removed at 1, 2, 3, 5, 8, 12, 15, 21, 26 min via syringe and analyzed by HPLC employing chiral column (Chiralpak IA column, hexane/propan-2-ol 85:15, flow rate 0.6 mL/min, oven temperature 40 °C, detector wavelength 240 nm; retention time **isomer-1** t₁ = 12.3 min, **isomer-2** t₂ = 13.6 min). This reaction was performed twice. A plot of ln [% ee] versus time was linear to ~3 half-lives with a pseudo first-order rate constant of k'= (1.1 ± 0.1)× 10⁻³ s⁻¹ obtained by a least–squares fit (R² > 0.95) (Figure S17). The second–order rate constant (k₂ = 0.056 ± 0.004 M⁻¹s⁻¹) is derived from here using the concentration of DBU (20 mM).

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Figure S17. Pseudo first-order plots for the duplicate experiments for the racemization of isomer-1 in CH₃CN at 20 °C.

The same racemization experiment was performed at -30 °C twice again (Figure S18).

a)



b)



Figure S18. Pseudo first-order plots for the racemization of isomer-1 in CH₃CN at -30 °C in duplicate.

A plot of ln [% ee] versus time was linear to ~3 half-lives with a pseudo first-order rate constant of k'= (3.82 ± 0.02) × 10^{-5} s⁻¹ obtained by a least–squares fit (R² > 0.95) (Figure S18). The second–order rate constant (k₂ = (1.9 ± 0.1) × 10^{-3} M⁻¹s⁻¹) is derived from here using the concentration of DBU (20 mM).

8.3 Stereospecific Si-free SuFEx reactions

The rate of this racemization reaction is around 0.002 $M^{-1}s^{-1}$ at -30 °C, which is comparable to the rate of Sifree SuFEx reactions, but only slightly slower. If racemization was the main reason of low enantioselectivity, then the S_N2 reaction (either concerted or addition-elimination) and racemization are a pair of competitive reactions. Therefore, it follows that when the concentration of the nucleophile used is increased, the racemization reaction can be prevented. Consistent to the experimental results, 95% *ee* was achieved when 10 equiv *p*-cresol was used (Table S5 and Figure S19-20).

In addition, if DBU promoted the racemization of **1**, stereospecific results should be obtained in asymmetric SuFEx without DBU as base. When sodium phenolate **6** was used, >99% *ee* was found (Table S5, and Figure S21-22).



Table S5. Enantioselective silicon-free SuFEx reactions

Enantiomer ^[a]	phenol	% ee ^[b]
enantiomer-1	10 equiv 2b- <i>p</i>	95
enantionier-1	6	> 99%
	7	> 99%
	10 equiv 2b- <i>p</i>	95 ^[c]
enantiomer-2	6	> 99%
	7	> 99%

Reaction conditions: **enantiomer-1/enantiomer-2** (1 mL, 3.6 mM in CH₃CN), phenolic derivative (1 equiv), DBU (1 equiv), rt.

[a] **enantiomer-1** and **enantiomer-2** refer to two enantiomers with retention times of 16.8 and 18.8 min with chiral HPLC, respectively.

[b] % ee determined using chiral HPLC and calculated by

% ee = $\frac{majority - minority}{majority + minority} \times 100\%$,



Figure S19. a) HPLC chromatogram and b) the result of the reaction with 10 equiv p-cresol and isomer-1

a)



Figure S20. a) HPLC chromatogram and b) the result of the reaction with 10 equiv p-cresol and isomer-2.







As expected therefore, >99% *ee* was also achieved when sodium 4-methyl-2-nitrophenolate **7** was adopted (Table S5), even though 4-methyl-2-nitrophenol **2ac** was a really poor nucleophile.

Sodium 4-methyl-2-nitrophenolate **7** was prepared according to the following procedure. A flame-dried Schlenk flask, under Ar flow, was charged with NaH (60% suspension in mineral oil, 50 mg, 1.25 mmol). To remove the mineral oil, the solid was washed with anhydrous Et_2O (3x 2 mL), and the ether was removed via syringe. 4-methyl-2-nitrophenol **2ac** (210 mg, 1.37 mmol, 1.1 equiv) was dissolved in 1.2 mL anhydrous MeCN and was added dropwise to the washed NaH. The resulting orange suspension, with a maximum concentration of 1M phenolate was kept under Ar flow. The exact concentration of phenolate was not determined.

Unlike the previous reaction with phenolate **6**, these reactions were much slower. After 19 hours, there was still some fluoride **1** remaining. However, compound **1** was not racemized, while only stereospecific products were obtained (Figure S23-24).

a)							
	Peak	RetTime	Sig	Туре	Area	Height	Area
	#	[min]			[mAU*s]	[mAU]	%
	1	3.109	1	MF	427.45465	35.44067	7.8458
	2	3.449	1	FM	1013.53381	109.61823	18.6032
	3	9.719	1	MM	4007.17725	135.96382	73.5509
b)							
	Peak	RetTime	Sig	Туре	Area	Height	Area
	#	[min]			[mAU*s]	[mAU]	%
	1	3.120	1	BV	483.68060	27.24945	9.2345
	2	3.838	1	VB	1131.13550	126.20741	21.5959
	3	7.403	1	BB	3622.91235	176.37514	69.1695

Figure S23. Results of reactions of a) isomer-1 and b) isomer-2 with phenolate 7.



Figure S24. HPLC chromatograms of a) racemate of **3ac**, b) racemate of fluoride **1**, c) **2ac**, the reaction of d) **isomer-1** and e) **isomer-1** with phenolate **7**. Enantioselectivity was monitored using an HPLC Chiralpak IA column, Conditions were: *n*-hexane/methyl *tert*-butyl ether = 90:10, flow rate 2.0 mL/min, oven temperature 40 °C, detector wavelength 240 nm.

9 Computational studies

All calculations were performed with Gaussian 16 at the wb97xd/6–311+G(d,p) level of theory, using the SMD solvent model to represent CH₃CN as implemented in there.¹⁴ Images of the 3D structures of molecules were generated using CYLview.¹⁵ Frequency calculations were conducted for all stationary points to confirm them as either a minimum or a TS. Even if DBU was brought close to S atom, and with that an energy increase of 19 kcal/mol was observed, then the F still does not want to leave, nor is there an eventual lowering of the energy. So, a transition state closely related to **COMP1** was not obtained (Scheme S7).



Scheme S7. Potential energy surface for phenolate in SuFEx. B.O. is short for Wiberg bond order.

REACT

Zero-point correction = 0.298941 (Hartree/Particle) Thermal correction to Energy = 0.320260 Thermal correction to Enthalpy = 0.321204 Thermal correction to Gibbs Free Energy = 0.246386 Sum of electronic and zero-point Energies = -1511.191721 Sum of electronic and thermal Energies = -1511.170402 Sum of electronic and thermal Enthalpies= -1511.169458 Sum of electronic and thermal Free Energies = -1511.244275

Center	Co	ordinates (Angstro	oms)
Number	х	Y	Z
	-1 /87603	-1 5/1/76	-0 820876
N2	0.021638	-1.341470	-0.629070
C3	0.021030	-1.331314	0.009923
C4	2 160907	1.077410	0.3147
05	2.109097	-1.518818	1 600180
C6	2 072640	-1.038007	1.009109
C7	1 22882	-0.830072	1.400094
C8	4.55005	-0.658358	0.113136
C9	4.51142	-0.0000000	-1 02578
C10	2 7/6103	-0.094101	-0.011/02
010	2.740103	1 697509	-0.911402
E12	-1.935465	-1.007.000	-2.189518
C13	-1.912313	-2.932322	-0.070102
C14	-2.546905	-0.433021	1 445529
C15	-2.024711	-0.400079	1.440020
C16	-3.449022	0.4333000	2.003032
C17	-4.17502	1.300744	0.046257
C18	-4.000011	0.479209	-0.040237
H19	-3.204117	0.470290	-0.700425
H20	2.010907	-1.173130	2.400240
H21	4.900090	-0.79940	2.200103
H22	0.979009 1 55919	-0.494931	2.004657
H23	2 1170/1	-0.555140	-2.004037
H24	2.117941	-0.910040	-1.792200
H25	-2.044409	-1.199493	2.003097
H26	-3.323722	0.419002	3.104090
H27	4.010021	2.072137	0.620149
H28	-4.001711	2.105059	-0.020148
029	-3.101300	0.490099	-1.701149
C30	-0.790232	1.007007	-2.002099
C31	0.107072	2.003302	-1.113030
C32	1.47904	2.200109	-1.443917
C33	2.401103	2.505041	-0.400723
C3/	2.100121	2.01000	0.0/0102
C35	0.003049	2.301341	1.241124
НЗЕ	1 75/567	2.030013	0.200U2 I
1100	1.704007	2.20/000	-2.49010

H37	3.48228	2.651181	-0.792266
H38	2.902152	2.665523	1.629368
H39	0.528241	2.299541	2.293416
H40	-1.209436	1.934445	0.599575

TS1

Zero-point correction = 0.299211 (Hartree/Particle) Thermal correction to Energy = 0.320292Thermal correction to Enthalpy = 0.321236Thermal correction to Gibbs Free Energy = 0.247211Sum of electronic and zero-point Energies = -1511.187360Sum of electronic and thermal Energies = -1511.16628Sum of electronic and thermal Enthalpies = -1511.165336Sum of electronic and thermal Free Energies = -1511.239361Imaginary frequency = -111.1 cm⁻¹

Center	Coc	ordinates (Angstrom	s)
Number	Х	Y	Z
S1	-1.32895	-0.777262	-1.1735
N2	0.18706	-0.859747	-0.89436
C3	0.803153	-1.344107	0.225035
C4	2.299023	-1.235444	0.152051
O5	0.259426	-1.832991	1.20792
C6	3.047743	-1.660861	1.249342
C7	4.43313	-1.566437	1.231866
C8	5.080936	-1.04752	0.114423
C9	4.338565	-0.6244	-0.98398
C10	2.952535	-0.717025	-0.96605
011	-1.69671	-0.517509	-2.54425
F12	-1.7289	-2.44129	-1.05504
C13	-2.64867	-0.35627	-0.01996
C14	-2.5935	-0.754587	1.305119
C15	-3.66674	-0.432058	2.129956
C16	-4.76563	0.251547	1.623174
C17	-4.80038	0.621311	0.281275
C18	-3.73446	0.322057	-0.55566
H19	2.531537	-2.060452	2.113832
H20	5.008295	-1.895804	2.08978
H21	6.162553	-0.971853	0.100316
H22	4.840003	-0.218256	-1.85509
H23	2.368949	-0.379053	-1.81272
H24	-1.73189	-1.292477	1.677973
H25	-3.64078	-0.726523	3.172512
H26	-5.59884	0.494677	2.27227
H27	-5.65724	1.151385	-0.11737
H28	-3.73805	0.618404	-1.59557
O29	-1.07374	1.53153	-0.94482

C30	-0.00535	2.000343	-0.36178
C31	0.904381	2.841043	-1.05012
C32	2.051415	3.326326	-0.43763
C33	2.356037	2.993063	0.883003
C34	1.473669	2.171703	1.582356
C35	0.317356	1.691014	0.982716
H36	0.684423	3.093105	-2.08349
H37	2.724751	3.966799	-1.00045
H38	3.258477	3.366061	1.354368
H39	1.690741	1.900047	2.611555
H40	-0.36006	1.053553	1.542192

INT1

Zero-point correction = 0.300126 (Hartree /Particle) Thermal correction to Energy = 0.321717 Thermal correction to Enthalpy = 0.322661 Thermal correction to Gibbs Free Energy = 0.247354 Sum of electronic and zero-point Energies= -1511.193957 Sum of electronic and thermal Energies = -1511.172365 Sum of electronic and thermal Enthalpies = -1511.171421 Sum of electronic and thermal Free Energies = -1511.246728

Center	Coordinates (Angstroms)				
Number	Х	Y	Z		
 S1	1 250062	0 131004	0 951593		
N2	-0.19112	-0.540119	0.95423		
C3	-0.7213	-1.284186	-0.03215		
C4	-2.21568	-1.423205	0.101888		
O5	-0.13474	-1.836103	-0.96779		
C6	-2.85376	-2.503684	-0.504545		
C7	-4.23502	-2.637344	-0.424047		
C8	-4.98879	-1.679281	0.247613		
C9	-4.35642	-0.590481	0.841767		
C10	-2.97435	-0.465829	0.775656		
O11	1.442445	1.072248	2.047286		
F12	1.869392	-1.357059	2.017405		
C13	2.802018	-0.25205	0.074016		
C14	2.84364	-1.139426	-0.987109		
C15	4.066121	-1.358243	-1.614014		
C16	5.210496	-0.69283	-1.186154		
C17	5.137394	0.198732	-0.121514		
C18	3.92494	0.426912	0.521188		
H19	-2.25849	-3.235477	-1.038042		
H20	-4.72414	-3.48622	-0.888656		
H21	-6.06718	-1.778697	0.304392		
H22	-4.94193	0.164242	1.355041		
H23	-2.47439	0.385239	1.22247		

H24	1.938443	-1.640755	-1.305614
H25	4.119357	-2.05253	-2.444846
H26	6.158103	-0.867972	-1.682421
H27	6.024496	0.722591	0.215232
H28	3.856388	1.115319	1.353601
O29	0.937189	1.243626	-0.397048
C30	-0.28948	1.83855	-0.516962
C31	-0.75096	2.764794	0.417897
C32	-1.99674	3.354258	0.239587
C33	-2.77718	3.03991	-0.870131
C34	-2.30202	2.128651	-1.807222
C35	-1.0635	1.523322	-1.630318
H36	-0.13795	3.000712	1.278682
H37	-2.35857	4.067825	0.971871
H38	-3.74776	3.504073	-1.003154
H39	-2.90221	1.876607	-2.674589
H40	-0.68753	0.793732	-2.337978

TS2

Zero-point correction = 0.299896 (Hartree/Particle) Thermal correction to Energy = 0.32107Thermal correction to Enthalpy = 0.322014Thermal correction to Gibbs Free Energy = 0.24719Sum of electronic and zero-point Energies = -1511.193107Sum of electronic and thermal Energies = -1511.171934Sum of electronic and thermal Enthalpies = -1511.170989Sum of electronic and thermal Free Energies = -1511.245814Imaginary frequency = -72.0 cm⁻¹

Center	Coordinates (Angstroms)			
Number	Х	Y	Z	
S1	1.223303	0.258933	0.827675	
N2	-0.0949	-0.570583	0.912676	
C3	-0.67671	-1.268476	-0.098898	
C4	-2.15475	-1.450355	0.103468	
O5	-0.12717	-1.730923	-1.094802	
C6	-2.81905	-2.434204	-0.62786	
C7	-4.19359	-2.591076	-0.501648	
C8	-4.91446	-1.753461	0.344878	
C9	-4.25627	-0.763081	1.068269	
C10	-2.87975	-0.615264	0.953744	
O11	1.428638	1.100612	1.989199	
F12	2.111742	-1.504999	2.105418	
C13	2.79385	-0.189175	0.057877	
C14	2.838904	-1.010523	-1.055837	
C15	4.074614	-1.248418	-1.64862	
C16	5.227199	-0.659792	-1.139935	
C17	5.15038	0.173012	-0.027847	

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C18	3.926362	0.415812	0.582894
H19	-2.24938	-3.070234	-1.29507
H20	-4.704	-3.362906	-1.066571
H21	-5.98852	-1.870326	0.437811
H22	-4.81712	-0.102993	1.720315
H23	-2.36133	0.162096	1.501598
H24	1.929369	-1.457088	-1.436429
H25	4.130942	-1.895749	-2.516134
H26	6.18549	-0.84817	-1.610127
H27	6.04542	0.635537	0.371442
H28	3.853919	1.051632	1.455104
O29	0.884741	1.322601	-0.452403
C30	-0.39056	1.859798	-0.537647
C31	-0.85787	2.754721	0.418437
C32	-2.13828	3.27735	0.282139
C33	-2.93401	2.921695	-0.803363
C34	-2.44497	2.038684	-1.759914
C35	-1.17084	1.499152	-1.627384
H36	-0.22803	3.023288	1.257157
H37	-2.51355	3.969387	1.027457
H38	-3.93155	3.333666	-0.902994
H39	-3.05922	1.758379	-2.607907
H40	-0.77765	0.792912	-2.348445

PROD

Zero-point correction = 0.299943 (Hartree /Particle) Thermal correction to Energy = 0.322435 Thermal correction to Enthalpy = 0.323379 Thermal correction to Gibbs Free Energy = 0.242049 Sum of electronic and zero-point Energies = -1511.199643 Sum of electronic and thermal Energies = -1511.177150 Sum of electronic and thermal Enthalpies = -1511.176206 Sum of electronic and thermal Free Energies = -1511.257537

Center	(Coordinates (Angstroms)		
Number	Х	Y	Z	
S1	0.888703	0.833855	0.74649	
N2	-0.194945	-0.277544	0.704939	
C3	-0.5967	-0.859149	-0.47972	
C4	-1.574263	-1.975712	-0.286792	
O5	-0.224561	-0.521523	-1.594597	
C6	-2.087888	-2.610574	-1.418511	
C7	-2.998158	-3.649742	-1.283973	
C8	-3.3997	-4.062414	-0.016313	
C9	-2.888966	-3.43356	1.114969	
C10	-1.978758	-2.392489	0.982098	
O11	1.148759	1.249097	2.108888	
F12	2.576103	-3.719395	1.893197	
C13	2.418599	0.454131	-0.068791	

C14	2.569083	0.61215	-1.442126
C15	3.78561	0.263979	-2.012562
C16	4.813552	-0.241404	-1.222172
C17	4.639053	-0.397271	0.148729
C18	3.434004	-0.04439	0.740837
H19	-1.765721	-2.28078	-2.398798
H20	-3.394289	-4.139339	-2.166157
H21	-4.109899	-4.874861	0.089469
H22	-3.1989	-3.755486	2.102434
H23	-1.576439	-1.900535	1.8584
H24	1.756676	0.99273	-2.043545
H25	3.927871	0.385922	-3.079357
H26	5.75772	-0.51471	-1.678539
H27	5.440923	-0.788858	0.762159
H28	3.289361	-0.150283	1.808492
O29	0.460711	2.119357	-0.157053
C30	-0.864926	2.576077	-0.063226
C31	-1.260192	3.308246	1.04544
C32	-2.562499	3.793369	1.087372
C33	-3.436673	3.549346	0.032681
C34	-3.011989	2.817253	-1.071792
C35	-1.714901	2.319861	-1.1257
H36	-0.561265	3.495575	1.851063
H37	-2.890394	4.36748	1.945825
H38	-4.449666	3.932384	0.070776
H39	-3.69118	2.628314	-1.894672
H40	-1.359358	1.736429	-1.965318

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11 NMR spectra










































.0 8.5 8.0 7.5 7.0 6.5 6.0 4.0 1 3.5 3.0 2.5 2.0 0.0 5.5 5.0 1.5 1.0 0.5 4.5 f1 (ppm)



































^{0.0} 8.5 7.5 3.0 2.5 2.0 8.0 7.0 6.5 6.0 5.5 5.0 4.0 3.5 1.5 1.0 0.5 4.5 f1 (ppm)









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8.5 8.0 7.5 7.0 6.5 6.0 4.0 1 3.5 3.0 2.5 2.0 0.0 5.5 5.0 1.5 1.0 0.5 4.5 f1 (ppm)

SUPPORTING INFORMATION





5.5 f1 (ppm)

S106










^{0.0} 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 4.5 f1 (ppm)































SUPPORTING INFORMATION



7.5 3.0 2.5 0.0 10.5 10.0 9.5 9.0 8.5 8.0 7.0 6.5 6.0 5.0 4.5 4.0 3.5 2.0 1.5 1.0 0.5 5.5 f1 (ppm)











12 Author Contributions

All authors contributed to the writing of the paper and supporting information

Dong-Dong Liang: equal contribution - synthesis of starting materials, mechanistic studies, enantioselective SuFEx reactions.

Dieuwertje E. Streefkerk: equal contribution - competition reactions, substrate scope, kinetics.

Daan Jordaan and Jorden Wagemakers: supporting BSc students, who wrote BSc thesis on this topic - synthesis and isolation of starting materials and final SuFEx products.

Jacob Baggerman: supporting staff – instrumental for computational infrastructure.

Han Zuilhof: principal investigator - computational studies.