

Neural activity mapping of bumble bee (*Bombus ignitus*) brains during foraging flight using immediate early genes

Shiori Iino¹, Yurika Shiota¹, Masakazu Nishimura², Shinichi Asada^{3*}, Masato Ono^{2*} and Takeo Kubo^{1*}

¹ Department of Biological Sciences, Graduate School of Science, The University of Tokyo, Bunkyo-Ku, Tokyo, 113-0033, Japan.

² Laboratory of Entomology, Graduate School of Agriculture, Tamagawa University, Machida-Shi, Tokyo, 194-8610, Japan.

³ Bioresource Sciences Major, Graduate School of Agriculture, Tamagawa University, Machida-Shi, Tokyo, 194-8610, Japan.

Correspondence and requests for materials should be addressed to T.K. (email: stkubo@bs.s.u-tokyo.ac.jp)

*: Corresponding authors

Supplementary table S1

Mean \pm SEM	MB		Other brain region	
	Nurse bee	Forager	Nurse bee	Forager
<i>BiHR38</i>	1 \pm 0.22	1.86 \pm 0.13	0.13 \pm 0.02	0.31 \pm 0.02
<i>BiEgr1</i>	1 \pm 0.14	3.70 \pm 0.31	0.48 \pm 0.07	1.09 \pm 0.07
<i>BiEcR</i>	1 \pm 0.10	1.44 \pm 0.11	1.39 \pm 0.10	1.67 \pm 0.09

Supplementary Table S1. The gene expression of *B. ignitus* in the greenhouse sampling normalized using *Actin*

Relative gene expression in each brain tissue of workers normalized to *BiActin* in the *B. ignitus* greenhouse sampling compared with the MBs of nurse bees.

Supplementary table S2

Mean \pm SEM	MB				Other brain region			
	8:30		8:37	9:00	8:30		8:37	9:00
	Nurse bee	Forager			Nurse bee	Forager		
<i>BiHR38</i>	1 \pm 0.16	2.38 \pm 0.40	2.00 \pm 0.31	2.47 \pm 0.80	0.16 \pm 0.01	0.22 \pm 0.05	0.19 \pm 0.02	0.30 \pm 0.03
<i>preBiHR38</i>	1 \pm 0.14	1.82 \pm 0.28	2.03 \pm 0.54	6.44 \pm 1.54	0.20 \pm 0.04	0.49 \pm 0.19	0.65 \pm 0.34	1.76 \pm 0.67
<i>BiEgr1</i>	1 \pm 0.15	2.30 \pm 0.48	1.94 \pm 0.44	9.76 \pm 3.38	0.65 \pm 0.06	0.70 \pm 0.10	0.53 \pm 0.05	1.10 \pm 0.12
<i>BiEcR</i>	1 \pm 0.09	1.03 \pm 0.10	1.14 \pm 0.10	1.29 \pm 0.23	1.55 \pm 0.14	1.83 \pm 0.12	1.41 \pm 0.13	1.72 \pm 0.08

Supplementary Table S2. The gene expression of *B. ignitus* in the laboratory flight-cage sampling normalized using *Actin*

Relative gene expression in each brain tissue of workers normalized to *BiActin* in the *B. ignitus* laboratory flight-cage sampling compared with the MBs of nurse bees collected at 8:30.

Supplementary table S3

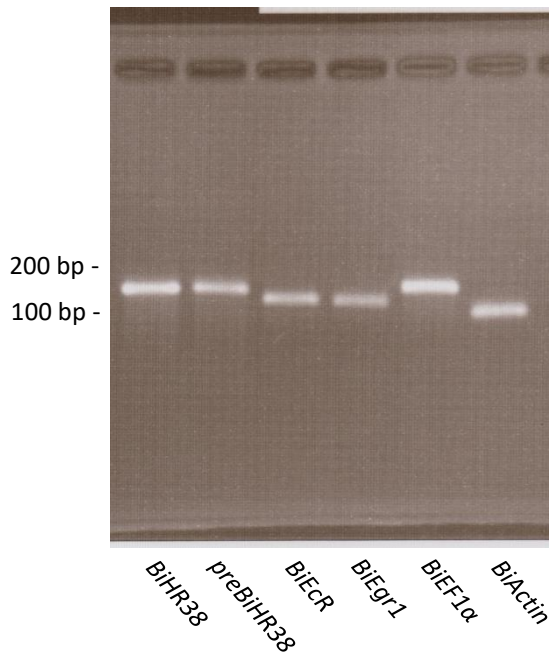
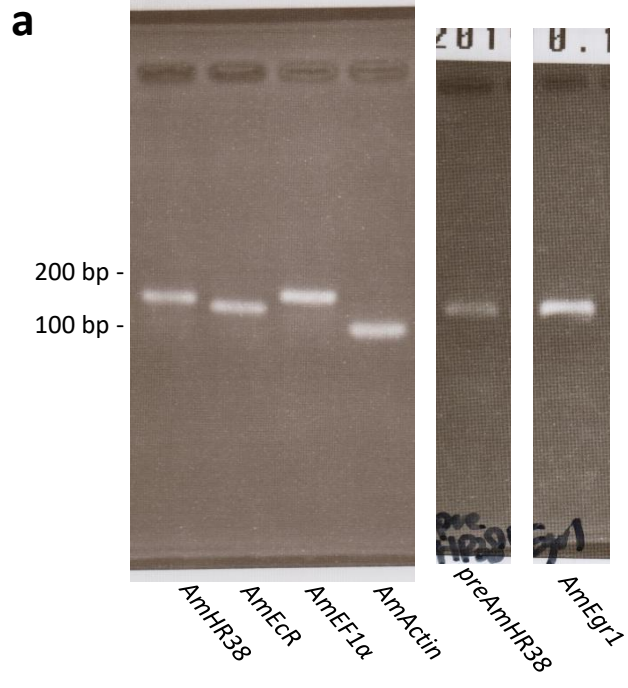
Mean ± SEM	MB									
	6:30		8:30	9:00	9:30		12:30		22:30	
	Nurse bee	Forager	Forager	Forager	Nurse bee	Forager	Nurse bee	Forager	Nurse bee	Forager
<i>AmHR38</i>	1±0.06	2.31±0.48	3.15±1.11	4.71±2.55	0.91±0.10	2.74±0.49	2.28±0.93	11.79 ±2.01	2.75±0.33	2.93±0.41
<i>preAmHR 38</i>	1±0.10	1.93±0.43	3.14±0.60	15.17 ±9.01	1.17±0.21	6.26±1.71	1.32±0.15	7.88±1.74	0.76±0.17	1.08±0.21
<i>AmEgr1</i>	1±0.29	0.86±0.21	1.54±0.36	3.46±2.12	0.76±0.06	1.79±0.31	0.96±0.09	6.18±1.53	0.76±0.14	0.90±0.37
<i>AmEcR</i>	1±0.30	0.75±0.11	1.00±0.27	1.84±0.81	0.65±0.05	0.94±0.10	0.99±0.13	2.82±0.51	0.59±0.08	0.58±0.05

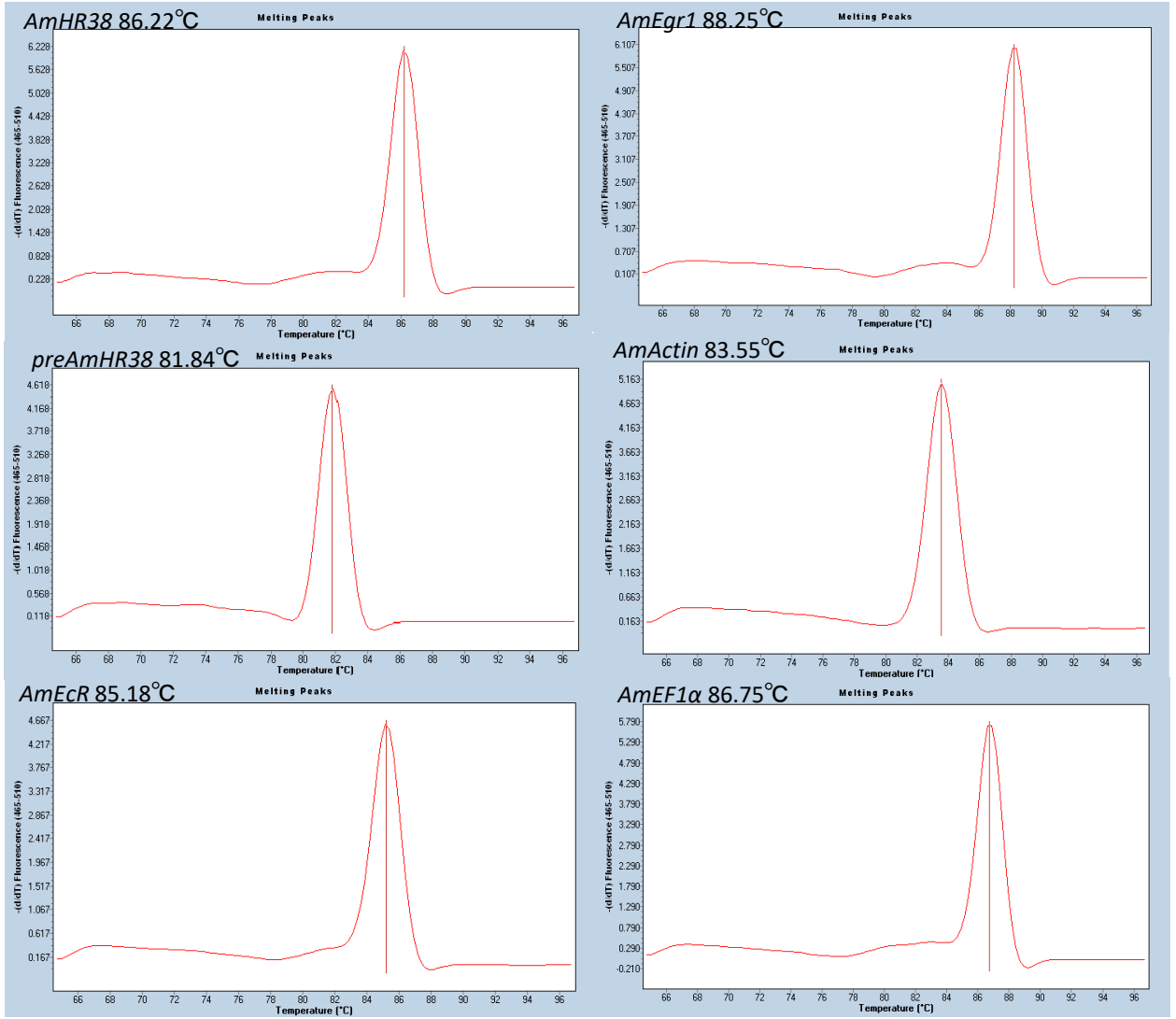
Mean ± SEM	Other brain region									
	6:30		8:30	9:00	9:30		12:30		22:30	
	Nurse bee	Forager	Forager	Forager	Nurse bee	Forager	Nurse bee	Forager	Nurse bee	Forager
<i>AmHR38</i>	0.17±0.06	0.36±0.13	0.54±0.12	0.53±0.15	0.24±0.04	0.57±0.08	0.43±0.10	1.39±0.33	0.45±0.05	0.23±0.00
<i>preAmHR 38</i>	0.09±0.01	0.18±0.03	1.07±0.05	1.14±0.35	0.14±0.03	1.34±0.25	0.18±0.05	1.34±0.35	0.13±0.03	0.07±0.01
<i>AmEgr1</i>	0.27±0.04	0.42±0.10	0.72±0.12	0.70±0.08	0.26±0.02	0.66±0.05	0.46±0.04	1.10±0.15	0.55±0.06	0.31±0.03
<i>AmEcR</i>	0.55±0.06	0.96±0.18	0.83±0.09	0.88±0.10	0.65±0.13	0.66±0.02	1.17±0.08	1.09±0.23	1.43±0.42	0.75±0.10

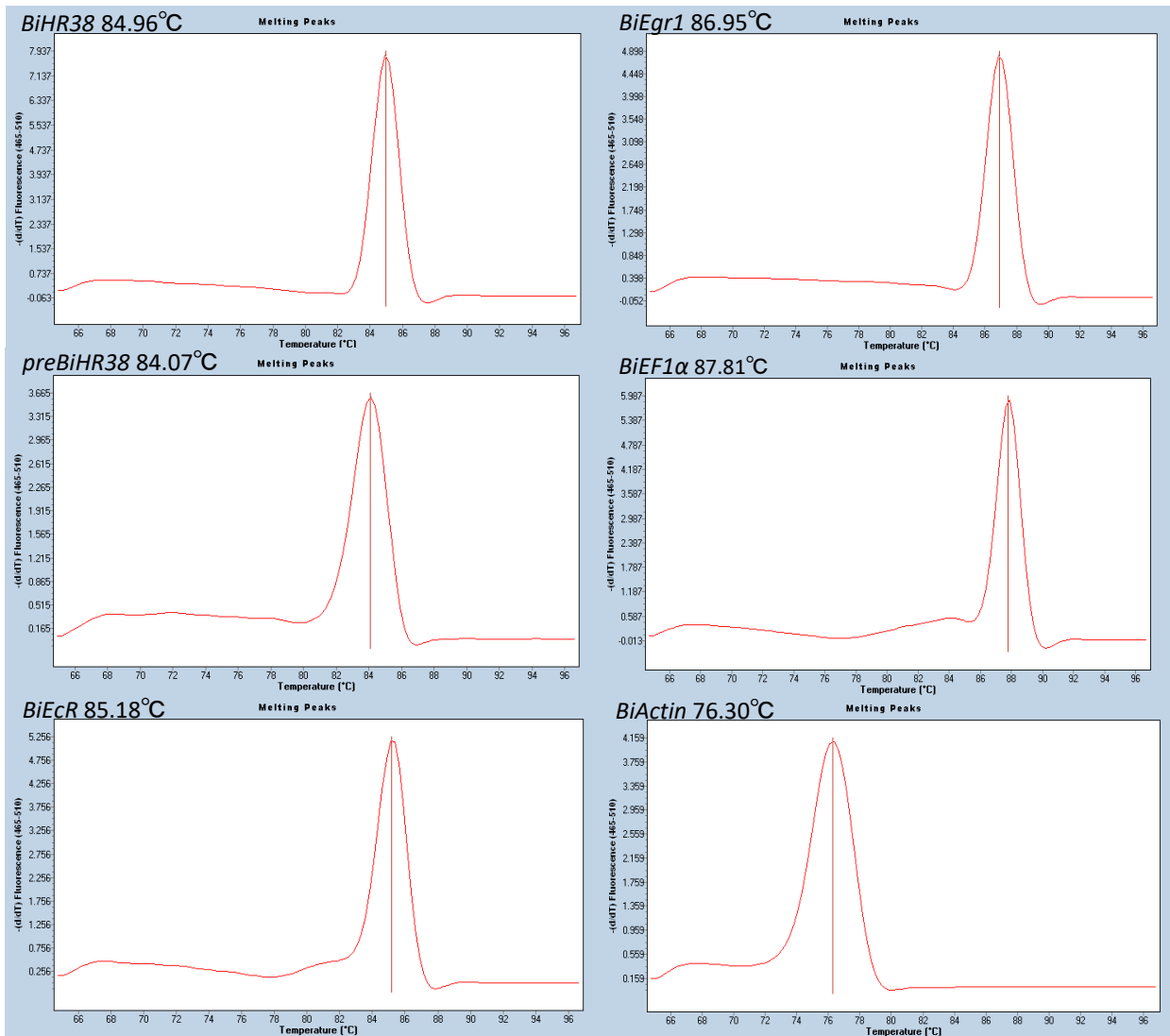
Supplementary Table S3. The gene expression of *A. mellifera* normalized using *Actin*

Relative gene expression in each brain tissue of workers normalized with *AmActin* in the *A. mellifera* sampling compared with the MBs of nurse bees collected at 6:30.

Supplementary Figure S1



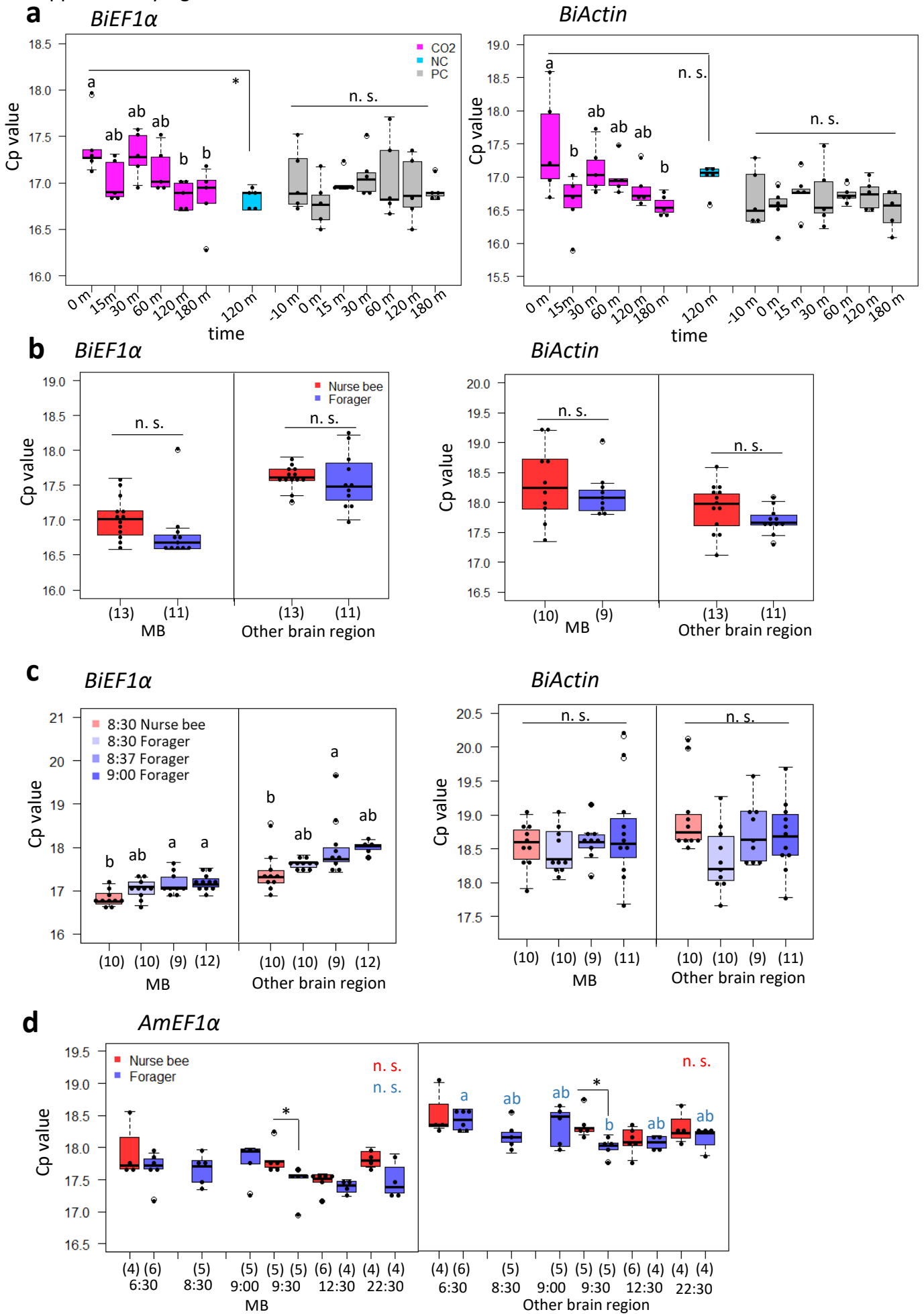
b

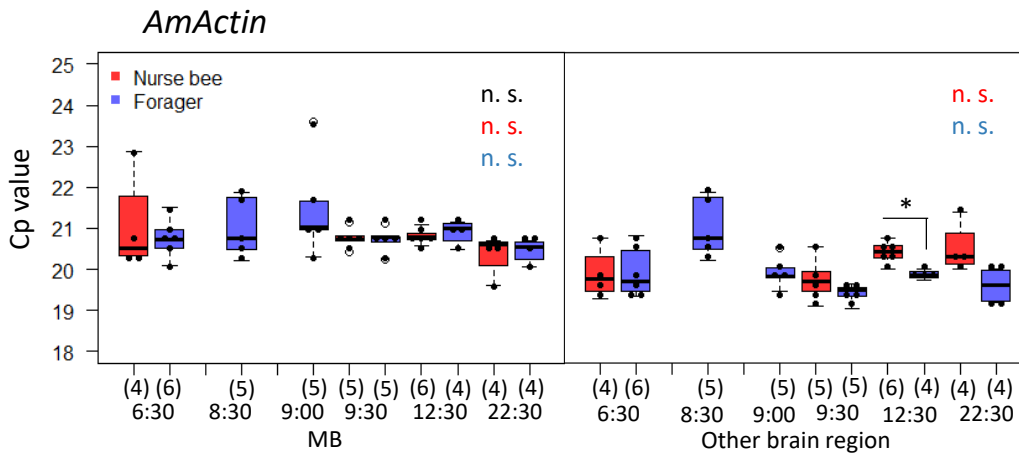


Supplementary Figure S1. Validation selectivity of gene-specific primers

(a) Agarose gel electrophoresis of each transcript of RT-PCR using gene-specific primers. The upper panel shows RT-PCR transcript using *A. mellifera* cDNA, and the lower panel shows that using *B. ignitus* cDNA. Full-length gel images were cropped from different gels and grouped with white spaces. (b) Melting peaks of qRT-PCR.

Supplementary Figure S2

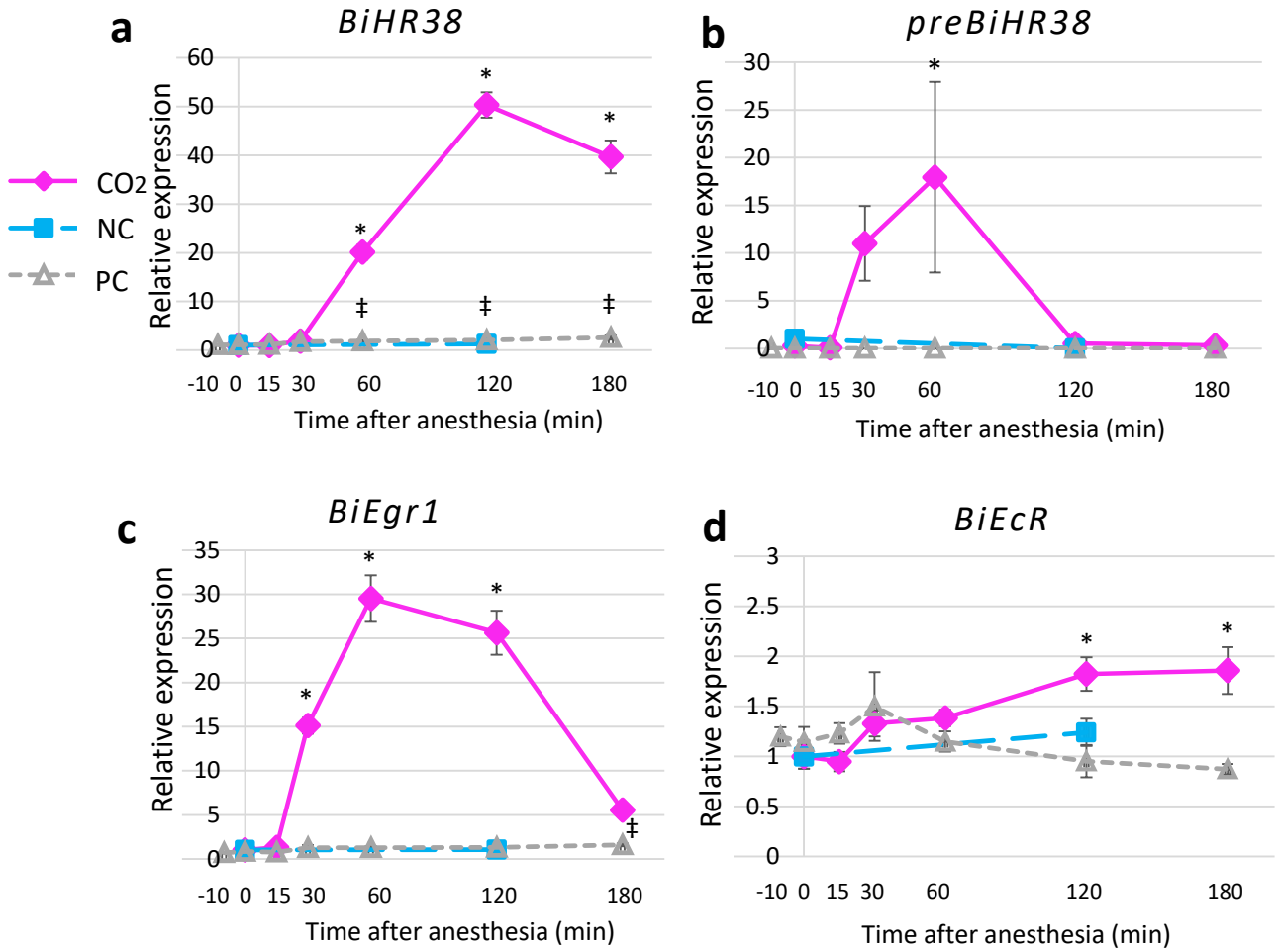




Supplementary Figure S2. The expression levels of *EF1a* and *Actin*

Time-course of the expression after awakening from CO₂ anesthesia (a). Magenta indicates the group anesthetized with CO₂ (“CO₂”), light blue indicates the negative control (“NC”, continuously anesthetized with CO₂ for 120 min), and gray indicates the positive control (“PC”, exposed to just an air flow). Significant differences are indicated using different letters among experimental groups ($p < 0.05$, Tukey-Kramer test) or asterisks between “CO₂” 0 min and “NC” 120 min ($p < 0.05$, Student’s t test or Welch’s t test after the F test). $n = 5$ for each sample. (b) The greenhouse experiment for *B. ignitus*. Significant differences are indicated with asterisks for each tissue ($p < 0.05$, Student’s t test or Welch’s t test after the F test). (c) The flight-cage experiment for *B. ignitus*. Significant differences are indicated using different letters for each brain tissue ($p < 0.05$, Tukey-Kramer test). (d) The experiment for *A. mellifera*. Significant differences are indicated using different letters for nurse bees (red) and foragers (blue) during the time-course ($p < 0.05$, Tukey-Kramer test), and asterisks (black) ($p < 0.05$, Student’s t test or Welch’s t test after the F test at each time-point) for each tissue. The sample size is indicated by the number in parentheses below the horizontal axis. n. s., not significant.

Supplementary Figure S3

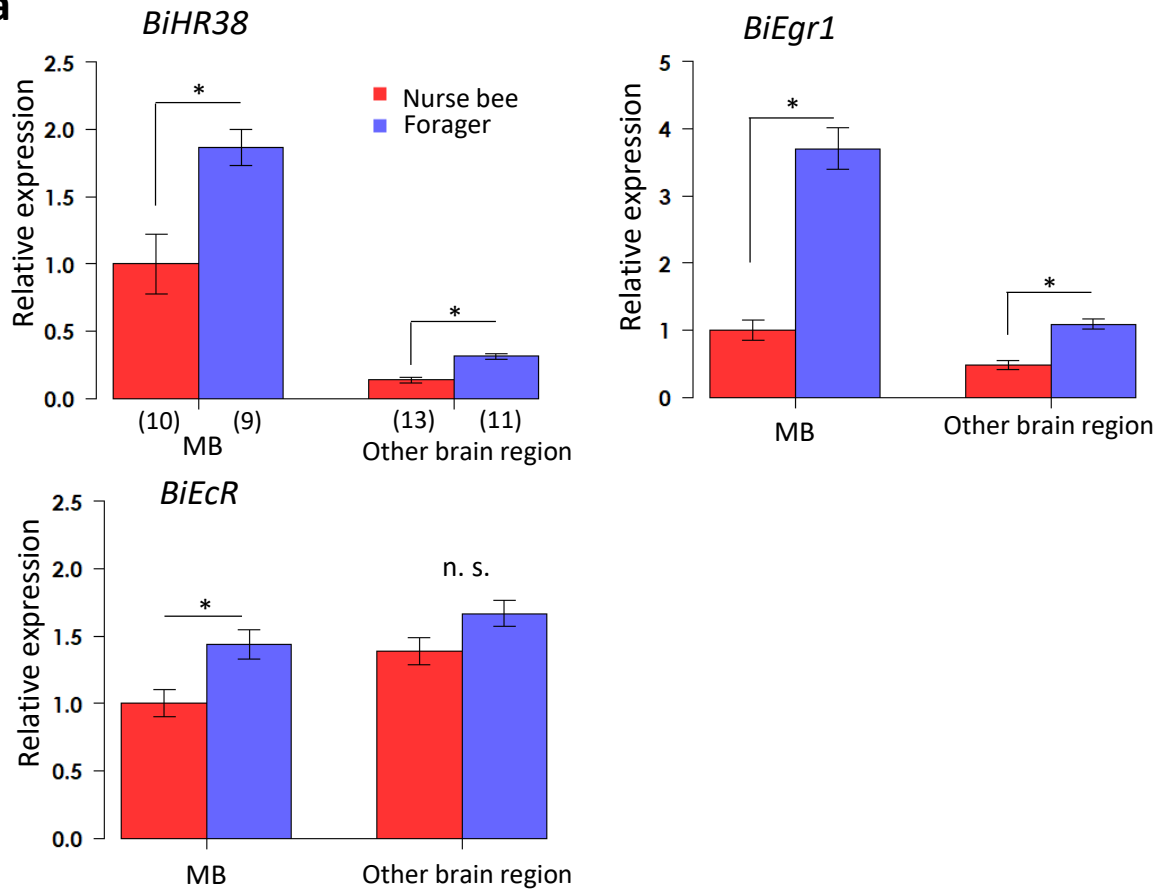


Supplementary Figure S3. Analysis of *BiHR38*, *preBiHR38*, *BiEgr1*, and *BiEcR* expression levels after seizure induction

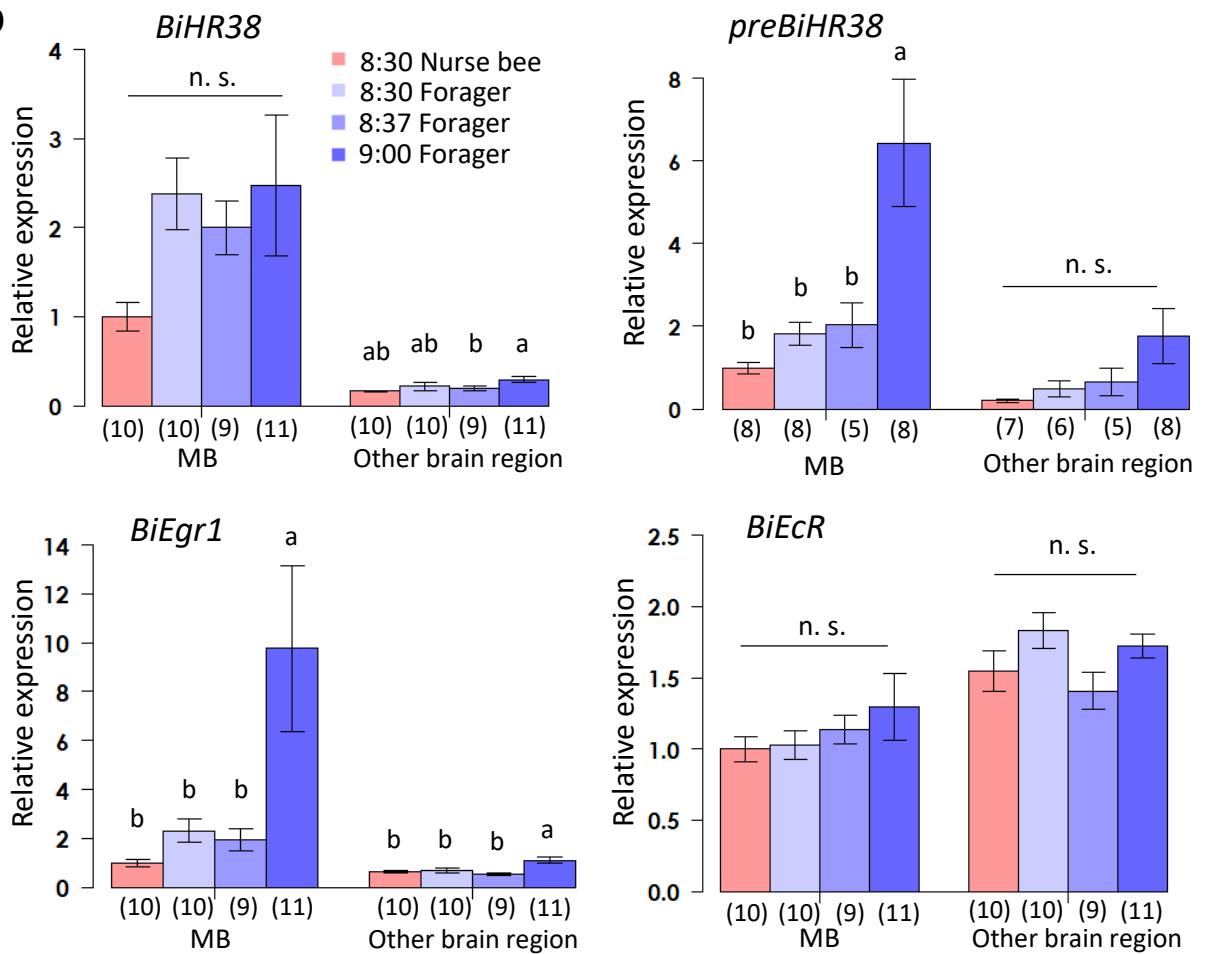
Time-course of the expression of *BiHR38* (a), *preBiHR38* (b), *BiEgr1*(c), and *BiEcR* (d) after awakening from CO₂ anesthesia. The expression level of each gene was analyzed by qRT-PCR and normalized using *BiActin*. Magenta lines indicate the group anesthetized with CO₂ (“CO₂”), light blue dashed lines indicate the negative control (“NC”, continuously anesthetized with CO₂ for 120 min), and gray dotted lines indicate the positive control (“PC”, exposed to an air flow). All data indicate means \pm SEM. Significant differences are indicated using Dunnett’s test after ANOVA (*:p<0.05 for CO₂ group, ‡: p<0.05 for PC group). Student’s t test and Welch’s t test revealed no significant difference between the NC group and CO₂ 0-min group. n=5 for each sample.

Supplementary Figure S4

a



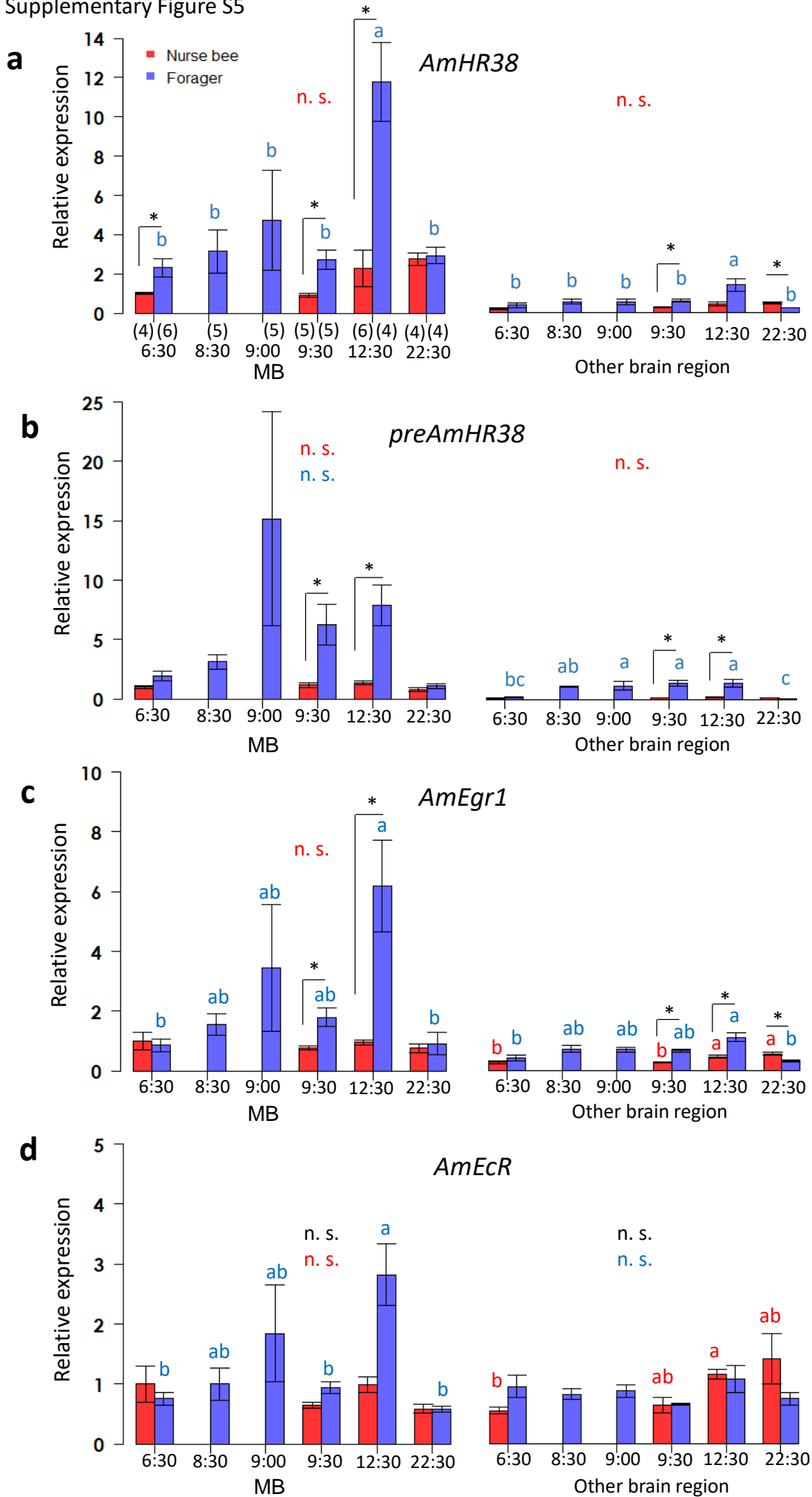
b



Supplementary Figure S4. Analysis of *BiHR38*, *preBiHR38*, *BiEgr1*, and *BiEcR* expression levels during foraging flight.

Expression analysis for the greenhouse experiment (a) and the laboratory experiment (b). The expression level of each gene was analyzed by qRT-PCR and normalized with that of *BiActin*. Each bar represents the mean \pm SEM. Significant differences are indicated by asterisks ($p < 0.05$, Student's t test or Welch's t test after the F test) on the error bars in (a), or different letters ($p < 0.05$, Tukey-Kramer test in (b), respectively). The sample size is shown below the horizontal axis in parentheses. n.s., not significant.

Supplementary Figure S5



Supplementary Figure S5 Analysis of *AmHR38*, *AmpreHR38*, *AmEgr1*, and *AmEcR* expression levels during foraging flight

Expression levels of *AmHR38* (a), *preAmHR38*(b), *AmEgr1*(c) and *AmEcR* (d) were analyzed by qRT-PCR and normalized with that of *AmActin*. Each bar represents the mean \pm SEM. Significant differences are indicated using different letters ($p < 0.05$, Tukey-Kramer test for each bee type during the time-course, in each brain tissue, respectively) or asterisks ($p < 0.05$, Student's t test or Welch's t test after the F test) on the error bars. The sample size is shown below the horizontal axis in parentheses. Some bars are too small to see in the *preAmHR38* graph. n.s., not significant.