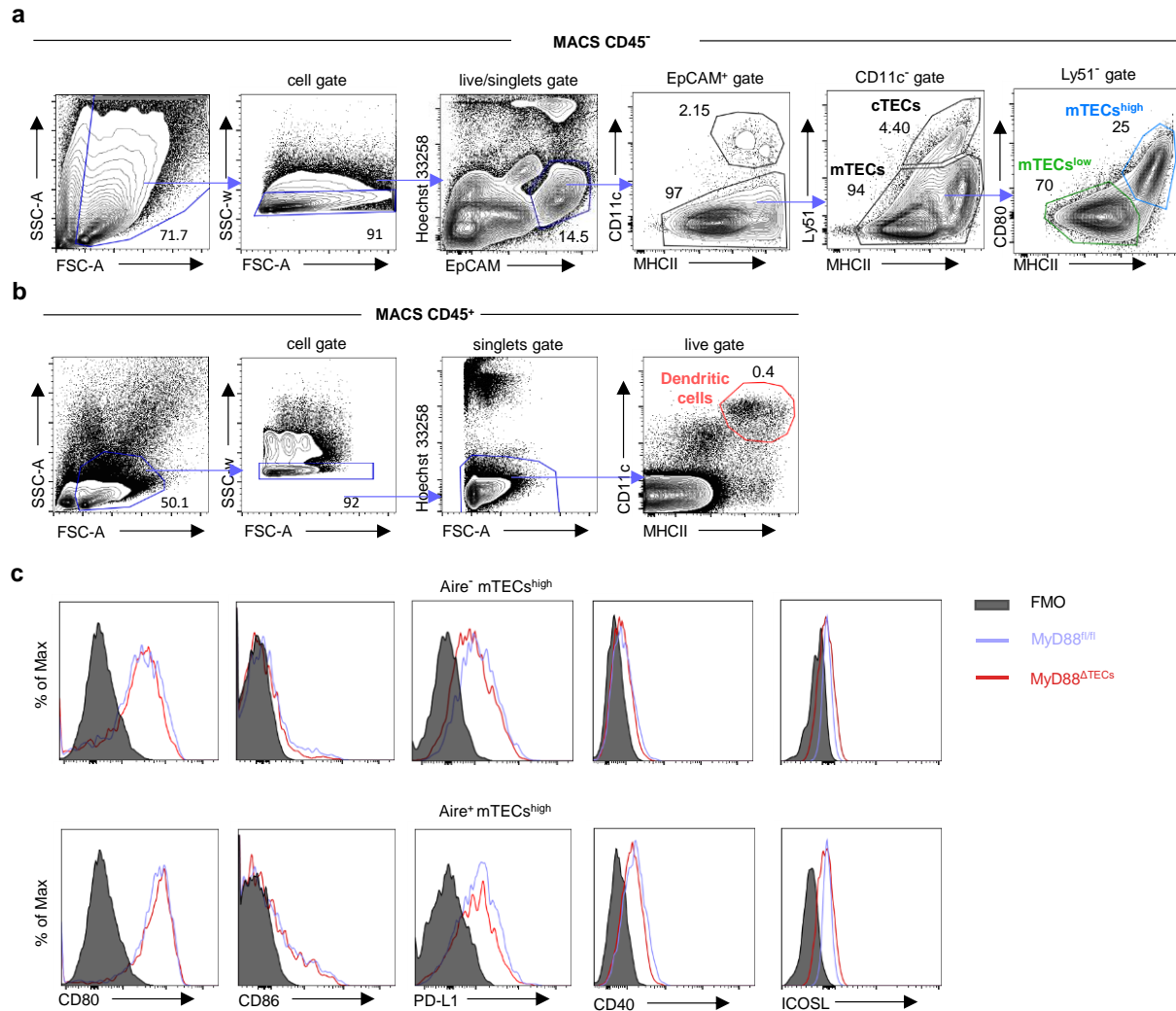


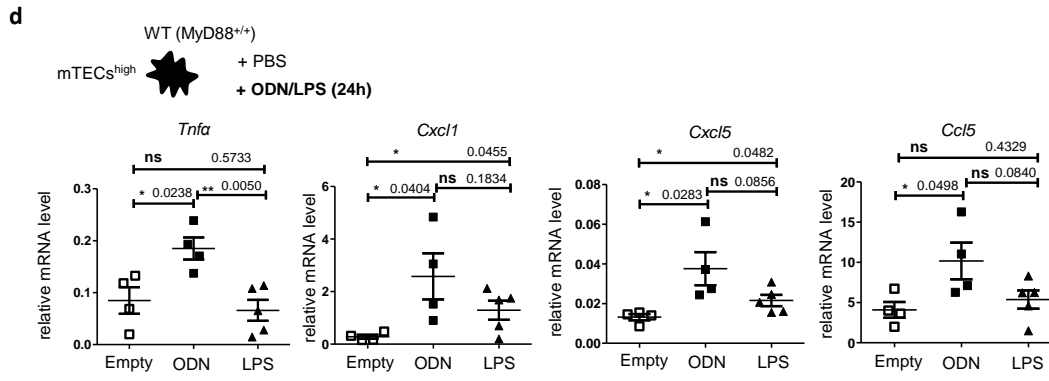
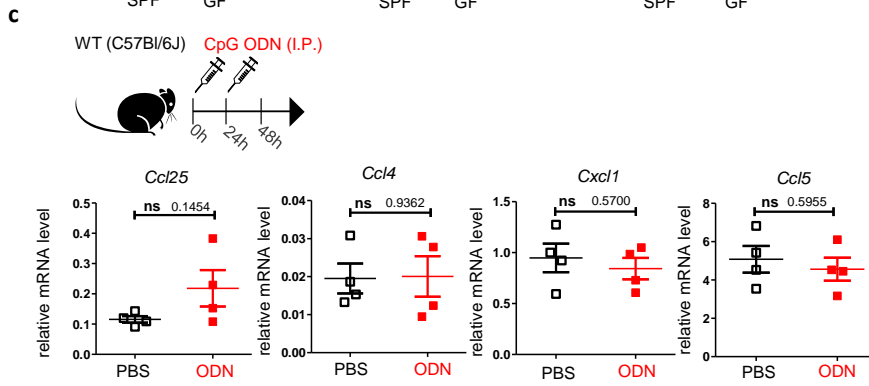
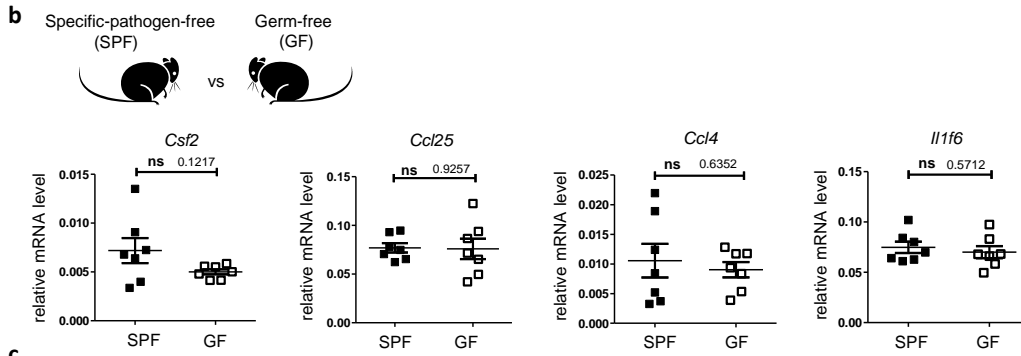
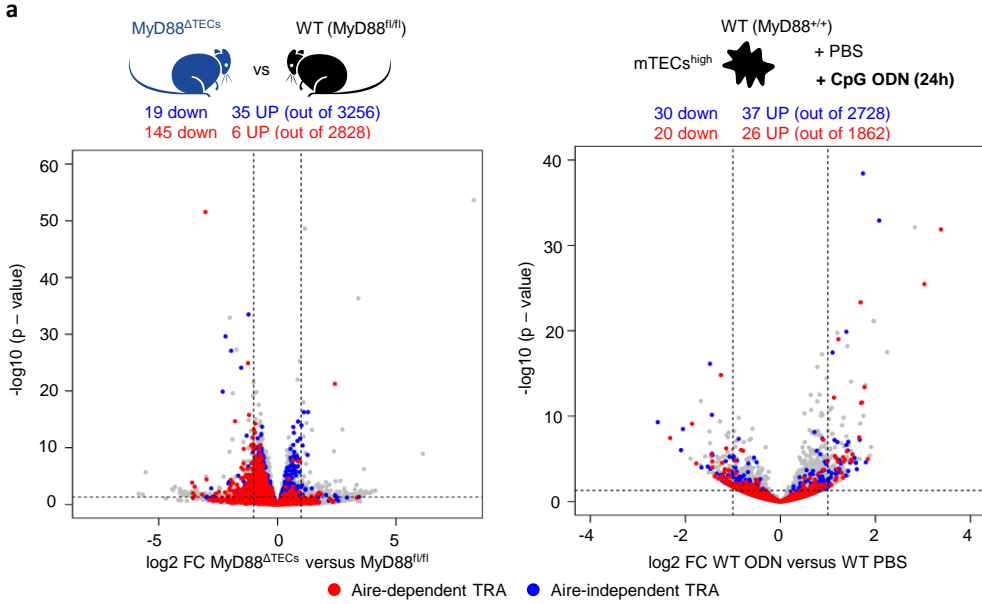
Supplementary information

Toll-like receptor signaling in thymic epithelium controls monocyte-derived dendritic cell recruitment and Treg generation

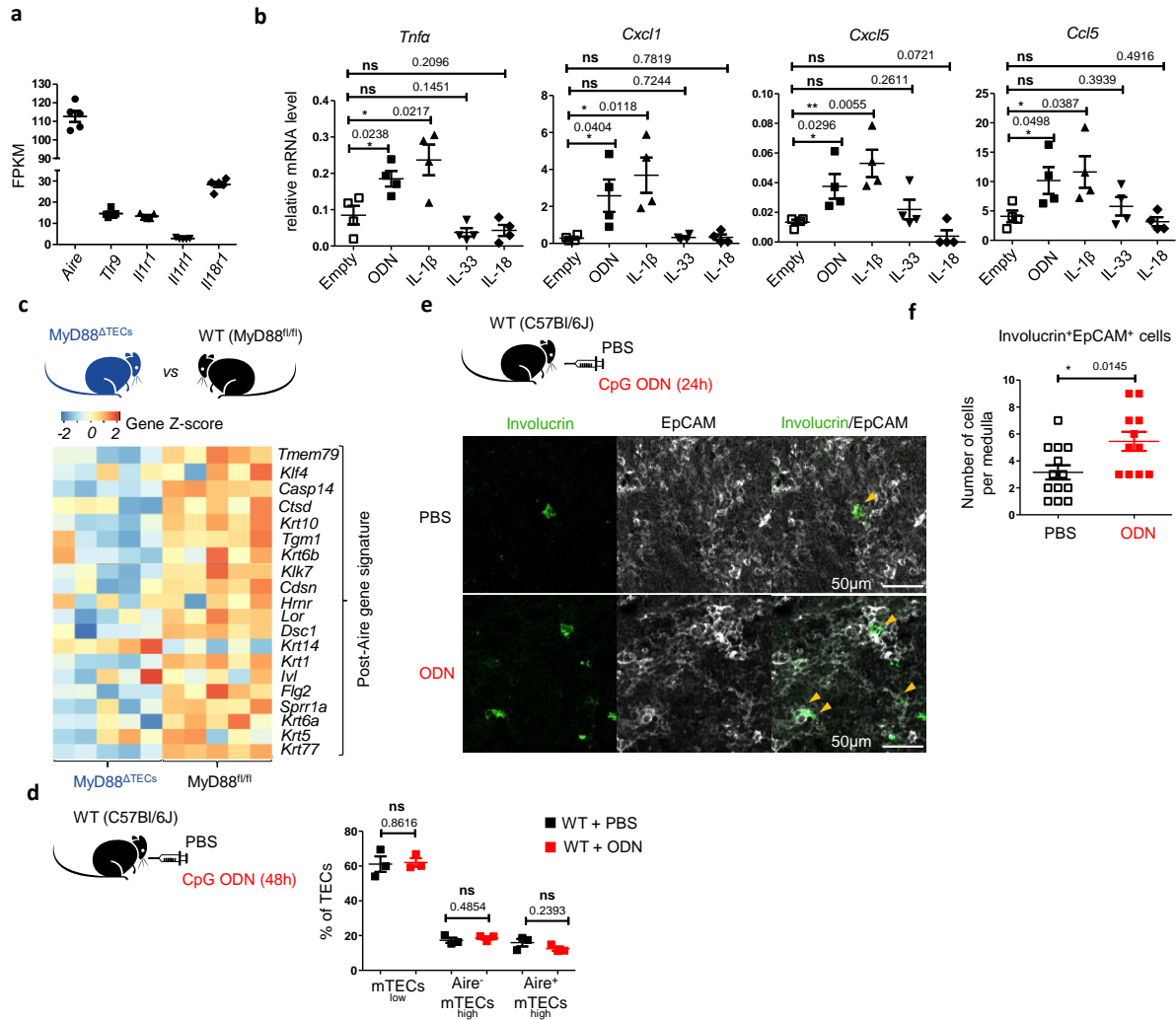
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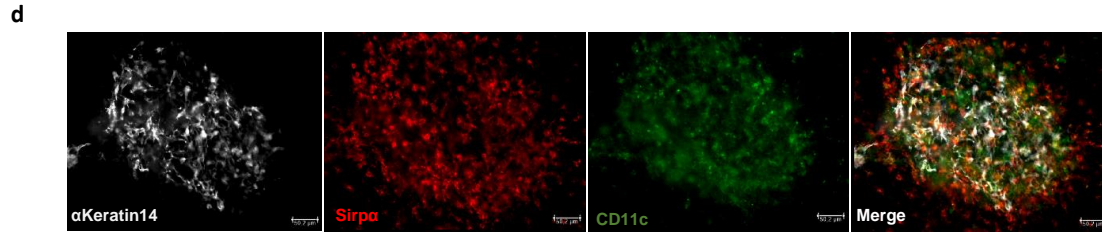
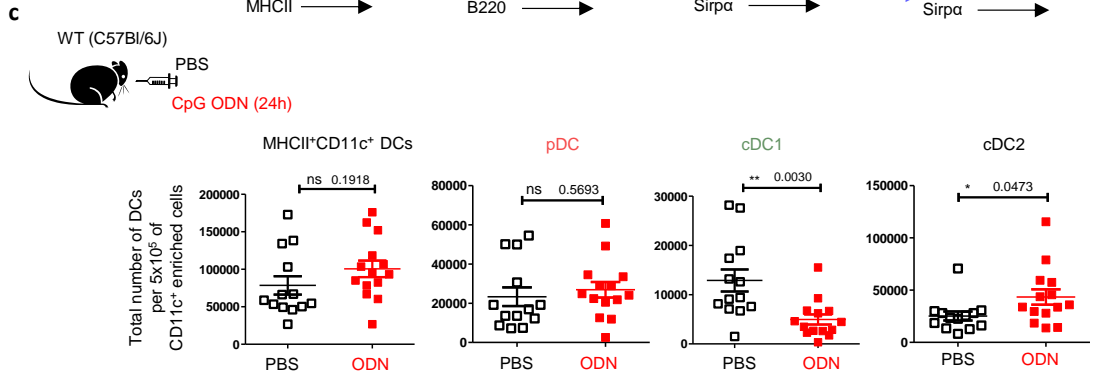
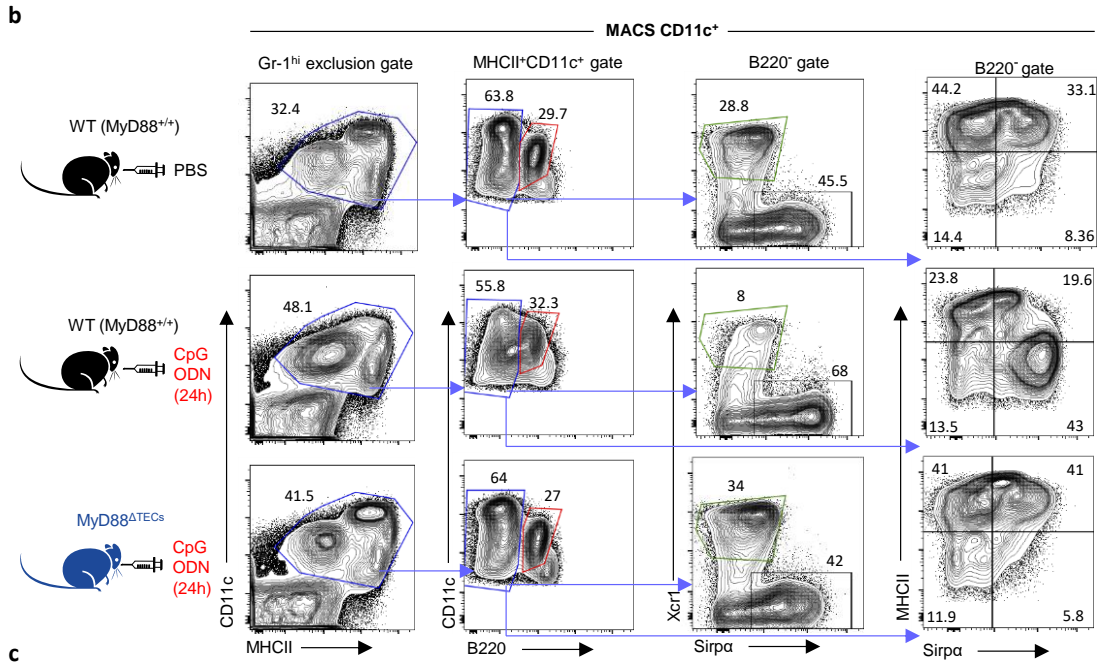
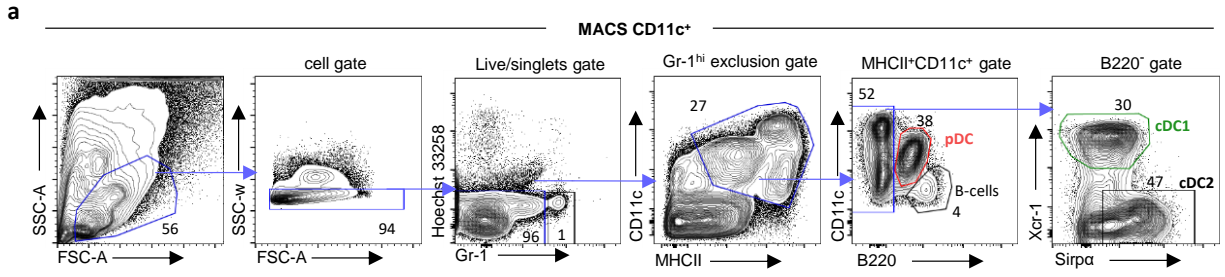
Supplementary Fig. 1, Related to Fig. 1 | Gating strategies and MyD88 independent expression of co-stimulatory molecules by mTECs. a, b Complete gating strategies of TECs and thymic classical DCs related to Fig. 1a. Thymic cell fraction were MACS depleted according to the expression of CD45. CD45⁻ fraction was sequentially gated according to FSC-A and SSC-A, singlets, live EpCAM⁺ and CD11c⁻ cells. The EpCAM⁺CD11c⁻ fraction representing TECs was then gated as cTECs (Ly51⁺) and mTECs (Ly51⁻). mTECs consist of two major populations: mTECs^{low} (MHCII^{low}CD80^{low}) and mTECs^{high} (MHCII^{high}CD80^{high}). CD45⁺ fraction was gated according to the FSC-A, SSC-A, SSC-W and Hoechst 33258 to live, single cells. Thymic conventional DCs were gated as MHCII⁺CD11c^{high}. **c** Representative flow cytometry histograms of co-stimulatory molecules (CD80, CD86, PD-L1, CD40 and ICOSL) expression on Aire⁻ mTECs^{high} and Aire⁺ mTECs^{high} of MyD88^{fl/fl} and MyD88^{ΔTECs} mice, *n*=2 independent experiments.



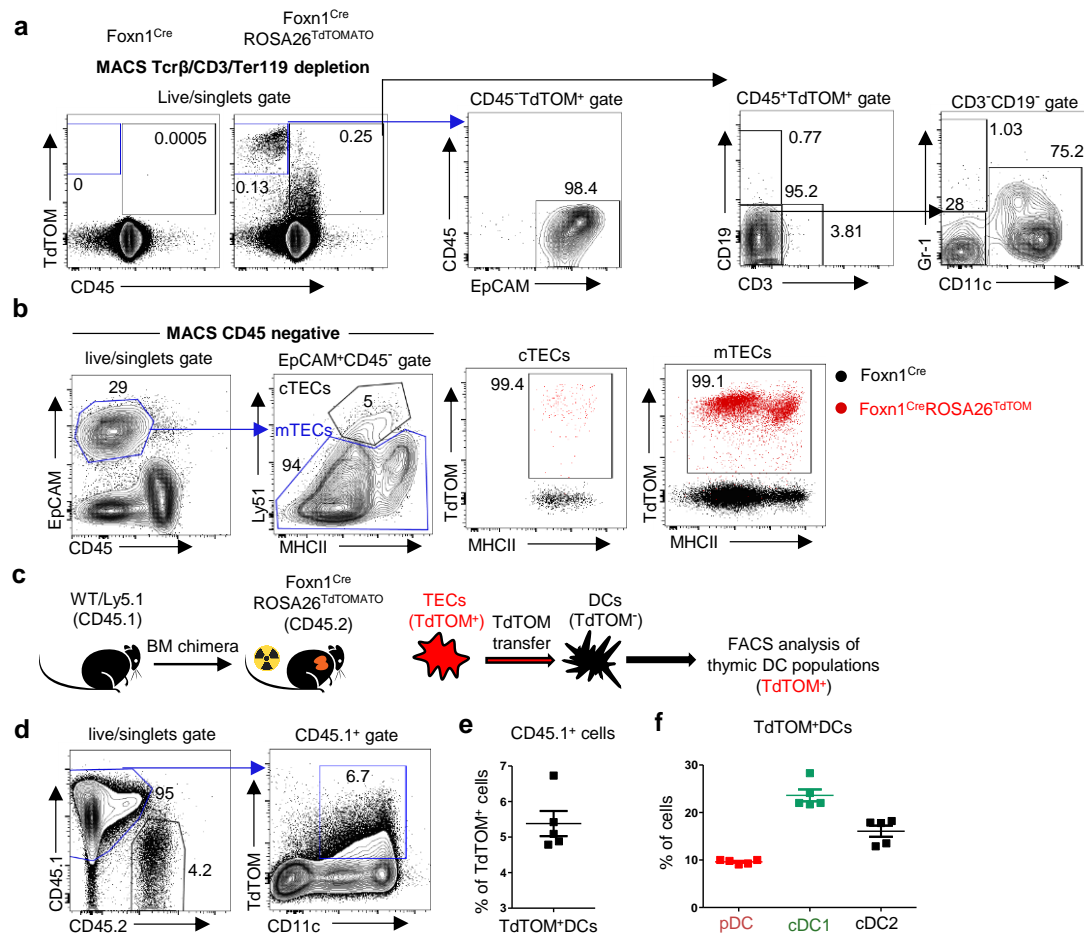
Supplementary Fig. 2, Related to Fig. 2 | TLR/MyD88 signaling in mTECs^{high} drives the expression of cytokines and chemokine independently on presence of microbiota. **a** Volcano plot analysis of RNA-sequencing data from Fig. 2b (left plot) and 2e (right plot) showing comparison of Aire-dependent (red) and Aire-independent (blue) TRA genes, non TRA genes are in grey. Statistical analysis was performed by Wald test, Fold-change cutoff of $\log_2 = \pm 1,0$ and p-value: 0,05 are marked by dashed lines ($n=5$ samples for left plot and $n=4$ samples for right plot). **b** qRT-PCR analysis of mRNA expression (normalized to Cacs3) of genes selected based on the Fig. 2b from mTEC^{high} isolated from Specific-pathogen-free (SPF) or Germ-free (GF) WT (C57Bl/6J) mice (mean \pm SEM, $n=7$ mice). **c** qRT-PCR analysis of mRNA expression (normalized to Cacs3) of genes selected based on the Fig. 2b and e from intraperitoneally CpG ODN stimulated WT (C57Bl/6J) mice (mean \pm SEM, $n=4$ mice). **d** qRT-PCR analysis of mRNA expression (normalized to Cacs3) of genes selected based on the Fig. 2e from *in vitro* CpG ODN or LPS stimulated mTECs^{high} (mean \pm SEM, $n=4$ for Empty and ODN and $n=5$ for LPS stimulated samples). Statistical analysis in b, c and d was performed by unpaired, two-tailed Student's t-test, $p \leq 0.05 = *$, $p \leq 0.01 = **$, ns = not significant.



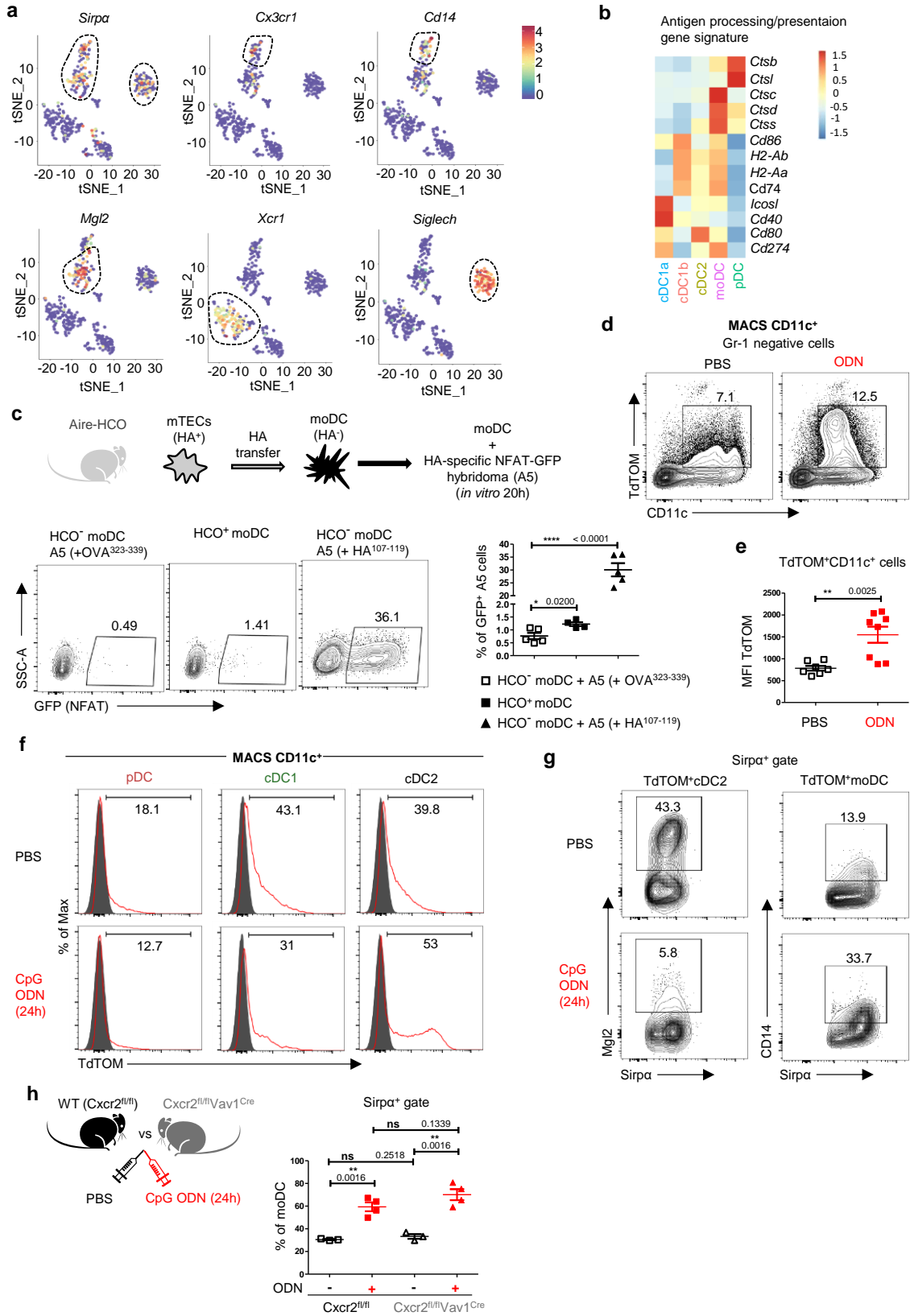
Supplementary Fig. 3, Related to Fig. 2 | TLR/MyD88 signaling affects the development of post-Aire mTECs. **a** Expression of indicated genes in mTECs^{high} depicted as FPKM (Fragments normalized per kilobase of feature length per million mapped fragments) (mean \pm SEM, $n=5$ samples). **b** qRT-PCR analysis of mRNA expression (normalized to *Cac3*) of genes selected based on the Fig. 2e from *in vitro* CpG ODN, IL-1 β , IL-33 and IL-18 stimulated mTECs^{high} (mean \pm SEM, $n=4$ samples). **c** Comparison of the expression of signature genes associated with post-Aire mTECs between MyD88 Δ TECs and MyD88^{fl/fl} mice. Data are derived from RNAseq analysis shown in Fig. 2b. **d** The frequencies of mTECs (gated as in Fig. 1e) isolated from CpG ODN or PBS intrathymically stimulated WT (C57Bl/6J) mice and analyzed by flow cytometry (mean \pm SEM, $n=3$ mice). **e** Microscopic examinations of thymic sections isolated from CpG ODN or PBS intrathymically stimulated WT (C57Bl/6J) mice. Cryosections were stained with Involucrin (green) and EpCAM (white). Scale bar represents 50 μ m. **f** Enumeration of Involucrin⁺EpCAM⁺ cells in the medullar region of the cryosections shown in e (mean \pm SEM, $n=11$ for ODN and $n=13$ for PBS stimulated medullary regions from 3 independent experiments). Statistical analysis in b, d and f was performed by unpaired, two-tailed Student's t-test, $p \leq 0.05 = *$, $p \leq 0.01 = **$, ns = not significant.



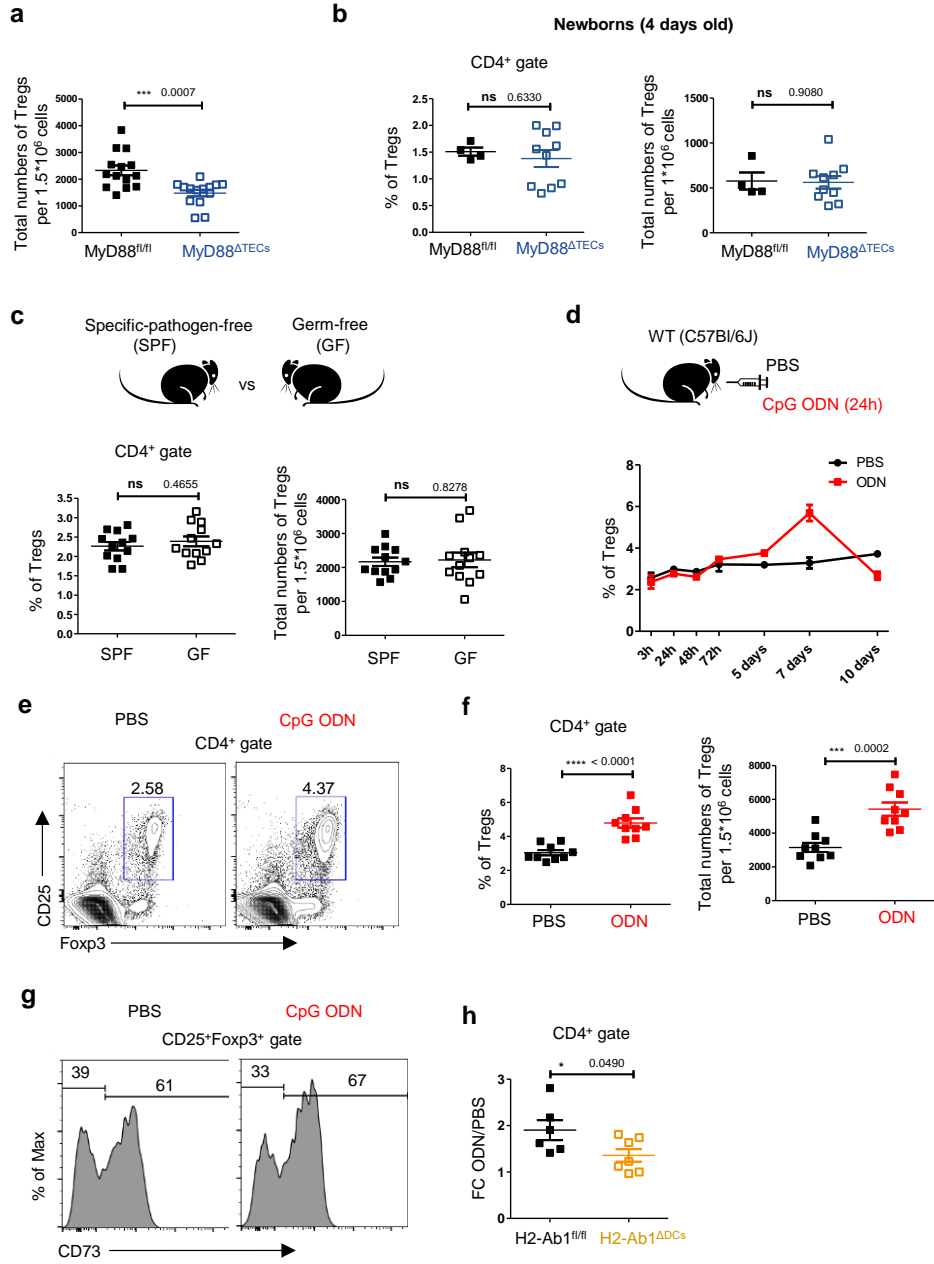
Supplementary Fig. 4, Related to Fig. 3 | Representative gating strategies and localization of thymic DCs. **a** Complete gating strategy of thymic DCs subpopulations. Thymic cells were MACS enriched for CD11c⁺ cells and sequentially gated according to FSC-A and SSC-A, singlets and live cells. Total thymic DCs were gated as Gr-1⁻CD11c⁺MHCII⁺. Thymic DCs were then divided into three major populations; pDC = CD11c^{low}MHCII^{low}B220⁺, cDC1 = CD11c⁺MHCII⁺B220⁻Xcr1⁺Sirpα⁻ and cDC2 = CD11c⁺MHCII⁺B220⁻Xcr1⁻Sirpα⁺. **b** Representative flow cytometry plots showing frequency of thymic DC subpopulations in CpG ODN or PBS intrathymically stimulated MyD88^{fl/fl} or MyD88^{ΔTECs} mice. **c** Quantification of the total number of thymic DCs, isolated as CD11c⁺ MACS enriched population from CpG ODN or PBS intrathymically stimulated WT (C57Bl/6J) mice (mean ± SEM, *n*=13 for PBS and *n*=14 for ODN stimulated mice). Statistical analysis was performed by unpaired, two-tailed Student's t-test, *p* ≤ 0.05 = *, *p* ≤ 0.01 = **, ns = not significant. **d** Microscopic analysis of the localization of Sirpα⁺mDCs in WT (C57Bl/6J) thymic medullary region determined by Keratin-14 staining. Scale bar represents 50.2 μm (*n*=3 independent experiments).



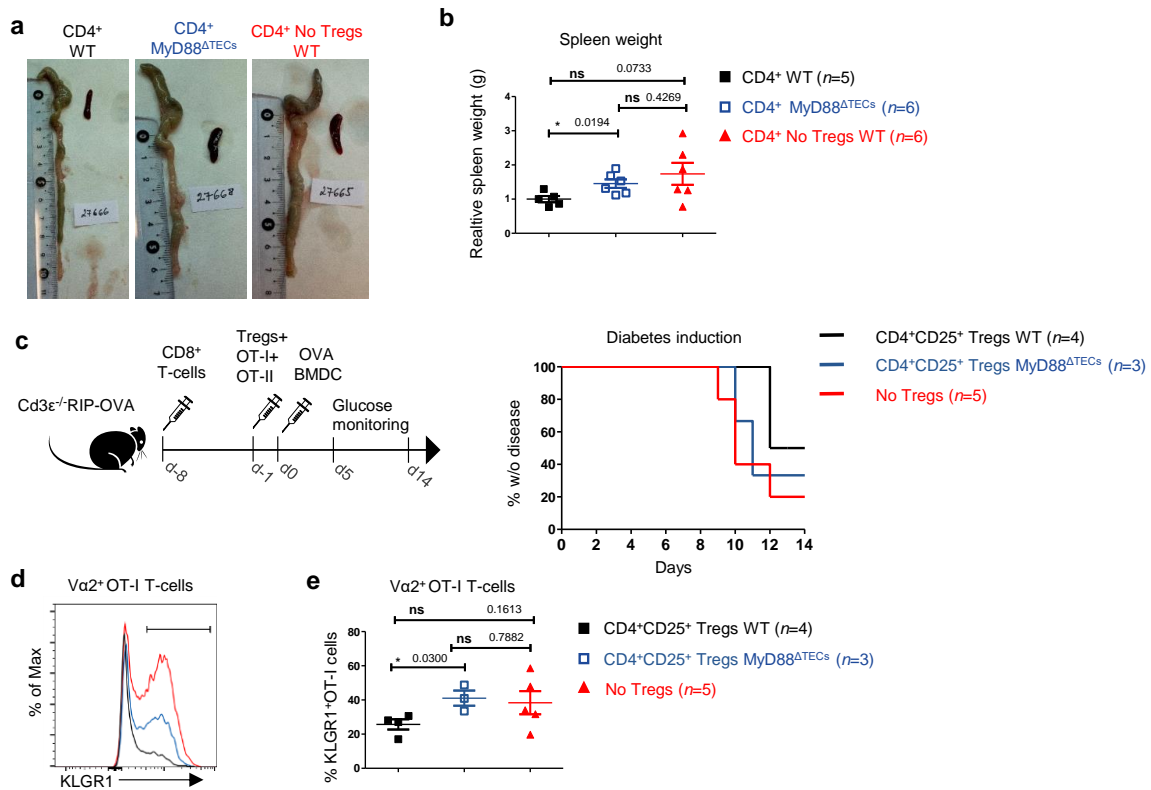
Supplementary Fig. 5, Related to Fig. 4 | Foxn1^{Cre}ROSA26^{TdTOMATO} as a model of thymic cooperative antigen transfer. **a** Representative flow cytometry plots showing frequency of TdTOM⁺ cells among MACS TCR β , CD3 and Ter119 depleted thymic cells from WT (Foxn1^{Cre}) and Foxn1^{Cre}ROSA26^{TdTOMATO} mice ($n=3$ independent experiments). CD45⁺TdTOM⁺ cells were further divided by CD19, CD3, Gr-1 and CD11c protein staining. **b** Flow cytometry analysis of TdTOM expression in CD45⁻ depleted and EpCAM⁺ thymic epithelial populations. The level of TdTOM is projected as an overlay of gated subset (cTECs or mTECs, right panels) from Foxn1^{Cre} (black dots) and Foxn1^{Cre}ROSA26^{TdTOMATO} (red dots) mice ($n=3$ independent experiments). The number indicates the percentage of TdTOM⁺ cells from gated subset. **c** Design of experiment. **d** Representative flow cytometry plots showing the frequency of TdTOM⁺CD11c⁺ cells in the thymic MACS-enriched CD11c donor cells from WT (CD45.1⁺) to Foxn1^{Cre}ROSA26^{TdTOMATO} BM chimeras. **e** Quantification of the frequencies of CD45.1⁺TdTOM⁺ DCs (mean \pm SEM, $n=5$ mice). **f** Quantification of frequencies of CD45.1⁺TdTOM⁺ DCs among DC subsets (gated as in Supplementary Fig. 3a) (mean \pm SEM, $n=5$ mice).



Supplementary Fig. 6, Related to Fig. 5 | TLR/MyD88 signaling increases cooperative antigen transfer between TECs and CD14⁺ moDC subpopulation. **a** Heat-map projection of selected signature genes (based on the Fig. 5a) between individual clusters (dashed circles) onto the two-dimensional tSNE plot. **b** Heat-map analysis of the expression of antigen processing/presentation signature genes in each subset defined in Fig. 5a. **c** Indirect presentation of Influenza Haemagglutinin (HA) by CD14⁺moDC (sorted as CD11c⁺MHCII⁺B220⁻Xcr1⁻Cx3cr1⁺CD14⁺) using Aire-HCO mice model and A5 hybridoma T-cell line. Representative flow cytometry plots showing frequency of GFP⁺ A5 cells co-cultivates with moDCs sorted from Aire-HCO⁻ (pulsed with irrelevant OVA peptide, left plot), Aire-HCO⁺ mice (middle plot) and Aire-HCO⁻ (pulsed with HA peptide, right plot). The graph shows the quantification of frequency of GFP⁺ A5 cells (mean ± SEM, *n*=4 for HCO⁺moDC and *n*=5 for HCO⁻moDC + A5 independent experiments). **d** Representative flow cytometry plots comparing the frequency of Gr-1⁻TdTOM⁺CD11c⁺ cells between CpG ODN and PBS intrathymically stimulated Foxn1^{Cre}ROSA26^{TdTOMATO} mice. **e** Quantification of TdTOMATO Mean fluorescent intensity (MFI) in mice described in d (mean ± SEM, *n*=7 for PBS and *n*=8 for ODN mice). **f** Representative flow cytometry plots showing frequency of TdTOM⁺ cells among pDCs, cDC1 and cDC2 (gated as in Supplementary Fig. 4a) in mice described in Fig. 5d. Grey histograms represent control Foxn1^{Cre} mice. **g** Representative flow cytometry plots showing frequency of Mgl2⁺cDC2 or CD14⁺moDC cells among TdTOM⁺Sirpα⁺ DCs in CpG ODN or PBS intrathymically stimulated Foxn1^{Cre}ROSA26^{TdTOMATO} mice. **h** Quantification of frequencies of TdTOM⁺CD14⁺ moDC from CpG ODN or PBS intrathymically stimulated Cxcr2^{fl/fl} or Cxcr2^{fl/fl}Vav1Cre mice (mean ± SEM, *n*=3 for ODN⁻ and *n*=4 for ODN⁺ mice). Statistical analysis in c, e and h was performed by unpaired, two-tailed Student's t-test, *p* ≤ 0.05 = *, *p* ≤ 0.01 = **, *p* < 0.0001 = ****, ns = not significant.



Supplementary Fig. 7, Related to Fig. 6 | Development of CD25⁺Foxp3⁺ thymic Tregs is impaired in the absence of TLR/MyD88 signaling in TECs. **a** Quantification of the total number of CD4⁺CD25⁺Foxp3⁺ thymic Tregs in MyD88^{fl/fl} and MyD88^{ΔTECs} (mean ± SEM, *n*=14 mice). **b** Quantification of frequencies (left graph) and total numbers (right plot) of CD4⁺CD25⁺Foxp3⁺ thymic Tregs isolated from the newborn (4 days old) MyD88^{fl/fl} and MyD88^{ΔTECs} mice (mean ± SEM, *n*=4 for MyD88^{fl/fl} and *n*=10 for MyD88^{ΔTECs} mice). **c** Quantification of frequencies (left graph) and total numbers (right plot) of CD4⁺CD25⁺Foxp3⁺ thymic Tregs in Specific-pathogen-free (SPF) and Germ-free (GF) WT (C57Bl/6J) mice (mean ± SEM, *n*=12 mice). **d** Quantification of the frequencies of CD4⁺CD25⁺Foxp3⁺ thymic Tregs from CpG ODN or PBS intrathymically stimulated WT (C57Bl/6J) mice over the indicated period of time, from 3 hours to 10 days (mean ± SEM, *n*=2 for 48h, 72h and 10days PBS and 10days ODN and *n*=3 for other displayed items). **e** Representative flow cytometry plots comparing frequencies of CD4⁺CD25⁺Foxp3⁺ thymic Tregs in CpG ODN or PBS intrathymically stimulated WT (C57Bl/6J) mice 7 days after stimulation. **f** Quantification of frequencies and the total numbers of thymic Tregs from mice described in e, (mean ± SEM, *n*=9 mice). **g** Representative flow cytometry histograms showing the expression of CD73 by CD4⁺CD25⁺Foxp3⁺ thymic Tregs in CpG ODN or PBS intrathymically stimulated WT (C57Bl/6J) mice. **h** Fold-change increase in the cellularity of thymic Tregs in H2-Ab1^{fl/fl} and H2-Ab1^{ΔDCs} mice stimulated with CpG ODN (mean ± SEM, *n*=6 for H2-Ab1^{fl/fl} and *n*=7 for H2-Ab1^{ΔDCs} mice). Data were calculated from Fig. 6h. Statistical analysis in a, b, c, f and h was performed by unpaired, two-tailed Student's t-test, *p* ≤ 0.05 = *, *p* ≤ 0.001 = ***, *p* < 0.0001 = ****, ns = not significant.



Supplementary Fig. 8, Related to Fig. 7 | Tregs cells from MyD88^{ΔTECs} mice have reduced suppressive capacity and failed to prevent the development of T-cell induced colitis or diabetes. **a** Representative pictures showing the size of colon and spleen from indicated mice (described in Figure 7a). **b** Relative quantification (normalized to average of control mice from each experiment) of spleen weight of T-cell induced colitis experimental mice (mean ± SEM, *n*=5-6 mice). Statistical analysis was performed by unpaired, two-tailed Student's t-test, *p* ≤ 0.05 = *, ns = not significant. **c** Design of experiment and disease progression curve showing the percentage of non-diabetic mice. Number of *n* for each cohort is indicated. **d** Representative flow cytometry histograms of KLGR1 expression in Va2⁺OT-I T-cells isolated at day 14 from spleen of mice from c. **e** Quantification of frequencies of KLGR1⁺OT-I T-cells from d (mean ± SEM, *n*=3-5 mice). Statistical analysis was performed by unpaired, two-tailed Student's t-test, *p* ≤ 0.05 = *, ns = not significant.