

**Stem Cell Reports, Volume 14**

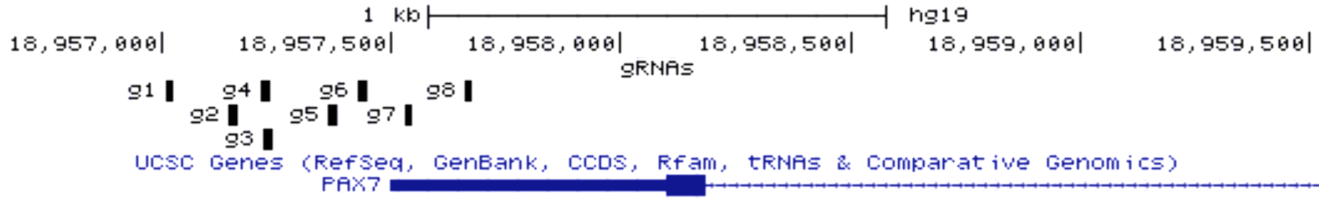
**Supplemental Information**

**Myogenic Progenitor Cell Lineage Specification by CRISPR/Cas9-  
Based Transcriptional Activators**

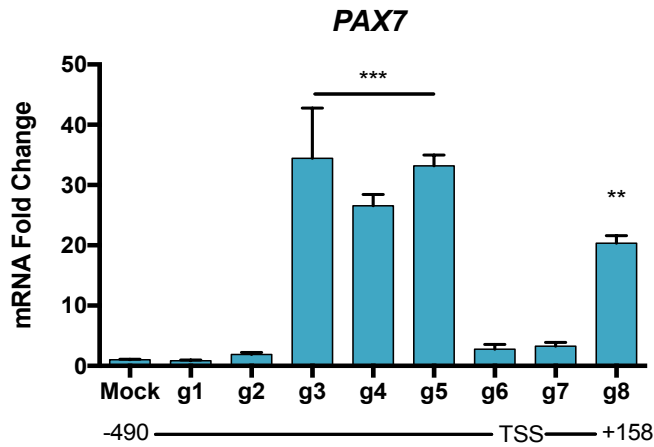
**Jennifer B. Kwon, Ashish Vankara, Adarsh R. ETTYREDDY, Joel D. Bohning, and Charles A. Gersbach**

# Figure S1

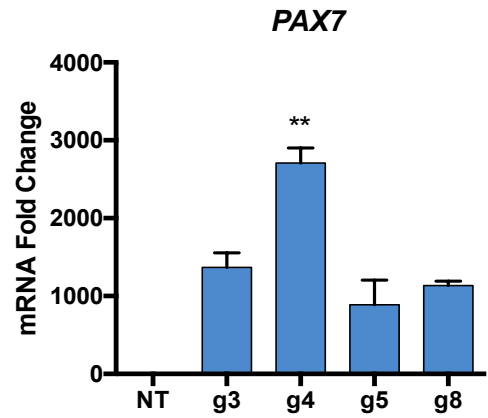
**A**



**B**

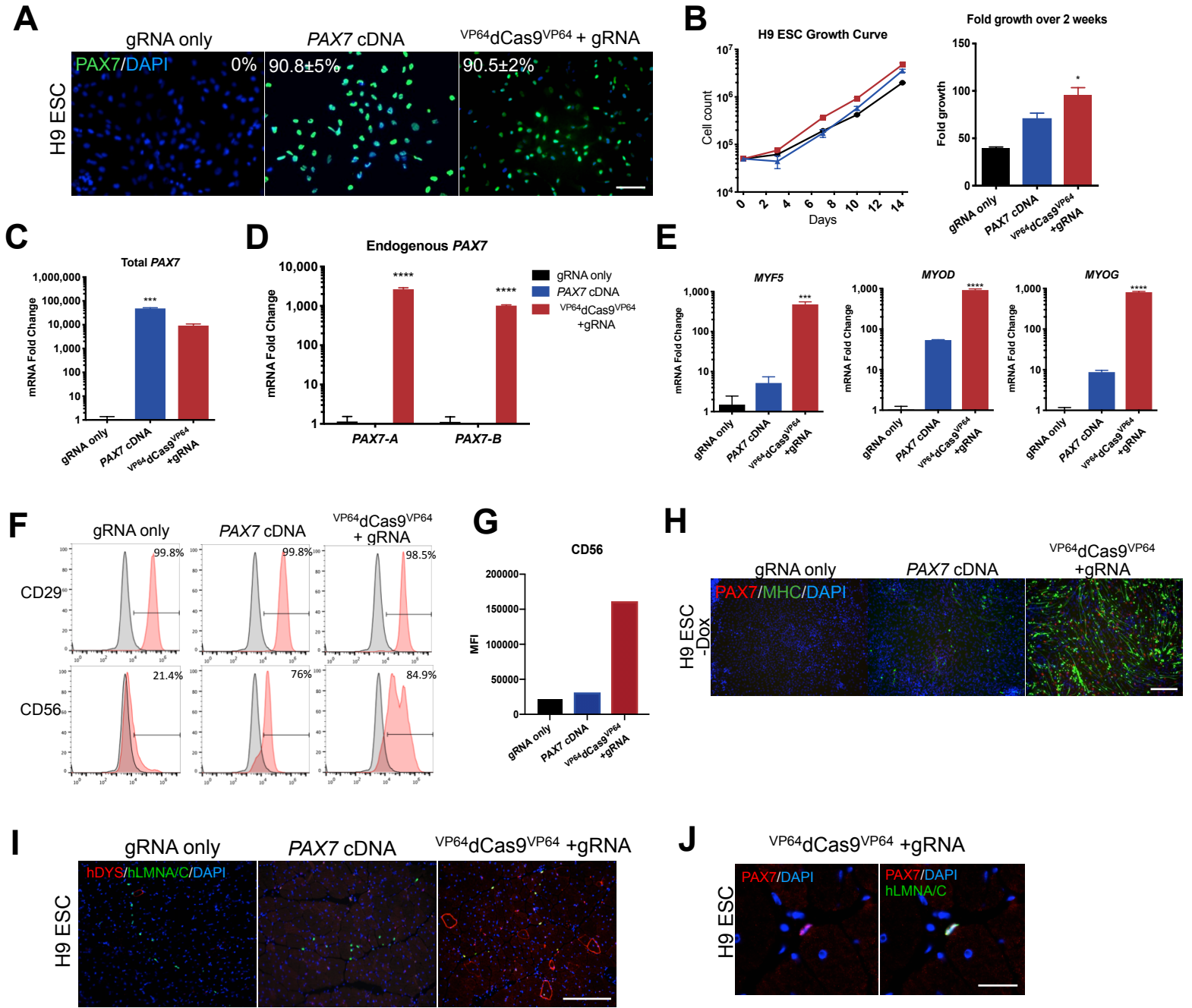


**C**



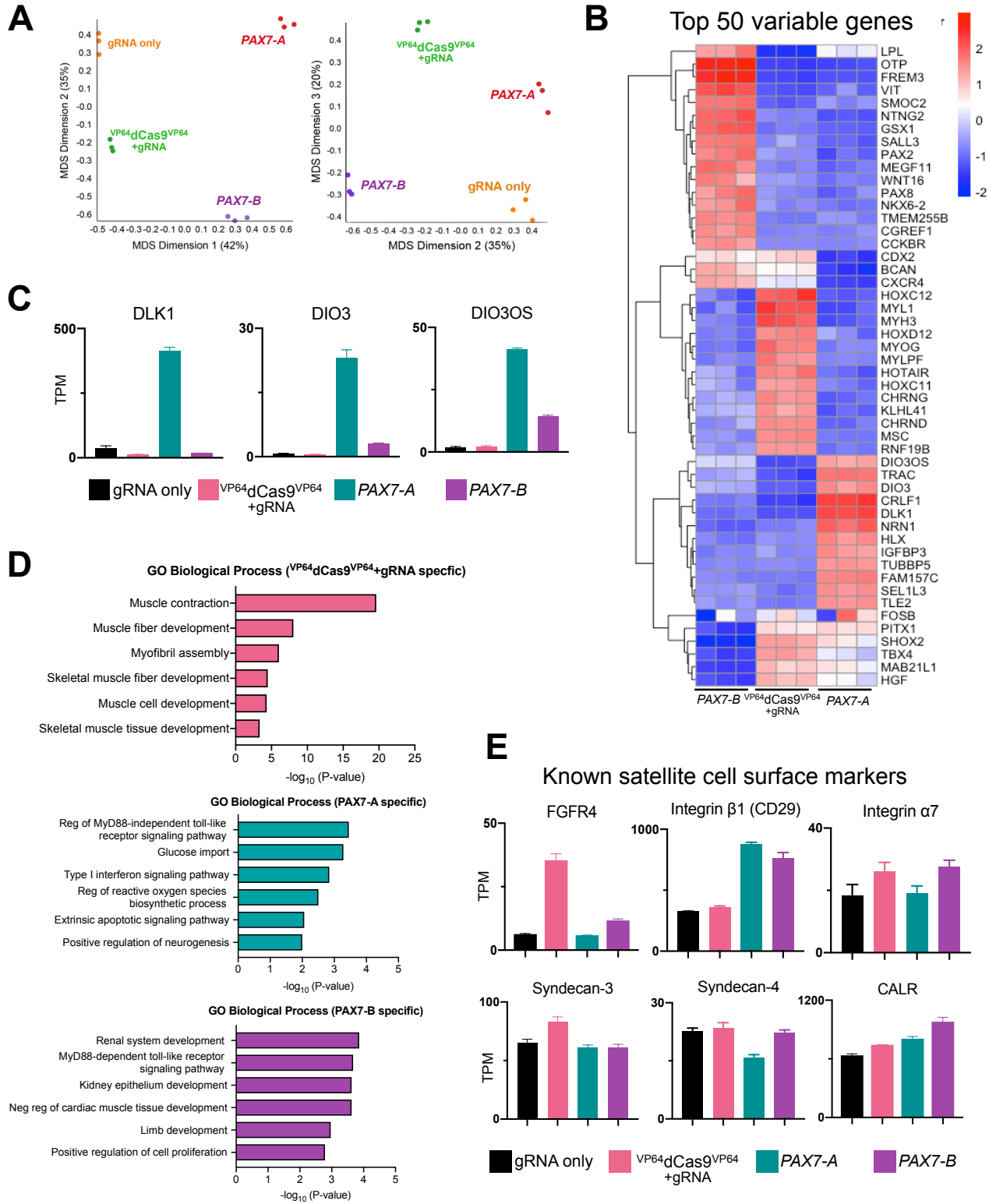
**Figure S1. Screening gRNAs for *PAX7* activation with  $^{VP64}dCas9^{VP64}$ , related to Figure 1.** (A) gRNA target sites relative to genome browser position of the human *PAX7* gene. (B) Cells expressing  $^{VP64}dCas9^{VP64}$  were treated for two days with CHIRON99021 and lipofected with *PAX7*-targeting gRNAs. Cells were harvested for qRT-PCR analysis after 6 days. gRNA 3, 4, 5 and 8 significantly upregulated *PAX7* compared to mock transfection, but were not significantly different from each other. (C) Lentiviral transduction of gRNAs in paraxial mesoderm cells expressing  $^{VP64}dCas9^{VP64}$  and gRNAs for 1 week. gRNA 4 significantly outperformed the other gRNAs. P values were determined by one-way ANOVA followed by Tukey's post hoc test;  $p < 0.05$  (mean  $\pm$  SEM,  $n = 3$  independent replicates).

# Figure S2



**Figure S2. Characterization and transplantation of myogenic progenitors derived from H9 ESCs via  $VP64dCas9^{VP64}$ -mediated activation of endogenous *PAX7* or exogenous *PAX7* cDNA expression, related to Figure 2 and 3.** (A) Representative immunostaining of PAX7 at 5 days post-sort. Scale bar = 100  $\mu$ m. (B) Growth curve of purified myogenic progenitors during post-sort expansion phase was monitored over 2 weeks. (C) Relative amount of total *PAX7* mRNA was determined by qRT-PCR using primers complementary to sequences present in the gene body. (D) Endogenous *PAX7* mRNA was detected using primers complementary to sequencing in the 3' UTR of either *PAX7-A* or *PAX7-B* isoforms. (E) The mRNA expression levels of myogenic markers *MYF5*, *MYOD*, and *MYOG* during the expansion phase. (F) Representative FACS analysis of CD29 and CD56 surface marker expression during the expansion phase. (G) Mean fluorescence intensity (MFI) of CD56 staining intensity across treatments. (H) Representative immunostaining of PAX7 and MHC in differentiated H9 ESCs after 4 passages in the presence of dox. Scale bar = 200  $\mu$ m. (I) Detection of human-derived fibers in  $VP64dCas9^{VP64}$ -treated cells 1 month after intramuscular injection of  $5 \times 10^5$  differentiated ESCs into NSG mice pre-injured with  $BaCl_2$ . Sections are stained with human-specific dystrophin and lamin A/C antibodies to mark donor-derived fibers and nuclei. Scale bar = 100  $\mu$ m. (J) Identification of donor-derived satellite cells expressing PAX7 and human-specific lamin A/C. All P values were determined by one-way ANOVA followed by Tukey's post hoc test (mean  $\pm$  SEM, n = 3 independent replicates). Scale bar = 25  $\mu$ m.

# Figure S3



**Figure S3. RNA-seq analysis, related to Figure 6.** (A) Multidimensional scaling (MDS) of the top 500 differentially expressed genes. (B) Heatmap showing differential expression of top 50 variable genes between the 3 PAX7-expressing groups. The color bar indicates z-score. (C) Expression profile from selected genes overexpressed in response to cDNA encoding *PAX7-A* from RNA-seq (mean  $\pm$  SEM, n = 3 independent replicates). (D) GO biological process terms for genes specifically enriched in cells treated with <sup>VP64</sup>dCas9<sup>VP64</sup>+gRNA, *PAX7-A* cDNA, or *PAX7-B* cDNA, corresponding to Venn diagram in Figure 4C. (E) Additional expression profiles of known satellite cell surface markers.

Table S1. gRNAs used in this study

gRNA #	Protospacer Sequence (5' - 3')	Position Relative to TSS
1	GGCCGGGGACTCGGCGGATC	-490
2	TCCCCGGCTCGACCTCGTTT	-351
3	CCAGGGCGCAAGGGAGCGG	-278
4	TCCTCCGCTCCCTTGCGCCC	-282
5	GGGGCGCGAGTGATCAGCT	-137
6	CGGGTTTCAGGGCTGGACGG	-70
7	TGGTCCGGAGAAAGAAGGCG	+30
8	AGCGCCAGAGCGCGAGAGCG	+158

# Table S2. qRT-PCR and ChIP-qPCR primers and conditions

Target	Forward Primer (5' - 3')	Reverse Primer (5' - 3')	Cycling Conditions
<i>GAPDH</i>	GAAGGTGAAGGTCGGAGTC	GAAGATGGTGATGGGATTTTC	95°C 5s 58°C 20s x40
<i>PAX7</i>	CAGCAAGCCCAGACAGGTGG	GCACGCGGCTAATCGAACTC	95°C 5s 58°C 20s x40
<i>MYF5</i>	AATTTGGGGACGAGTTTGTG	CATGGTGGTGGACTTCCTCT	95°C 5s 58°C 20s x40
<i>MYOD</i>	AGACTGCCAGCACTTTGCTA	GTAGCTCCATATCCTGGCGG	95°C 5s 58°C 20s x40
<i>MYOG</i>	GGTGCCCAGCGAATGC	TGATGCTGTCCACGATGGA	95°C 5s 58°C 20s x40
Endogenous <i>PAX7-A</i>	AGCTACAAGGTGGTGTCAGGGT	GAGCCATAGTACGGAAGCAGAG	95°C 5s 58°C 20s x40
Endogenous <i>PAX7-B</i>	TCTGGCCAAAAATGTGAGCCT	GGGTCAGTTAGGGTTGGGC	95°C 5s 58°C 20s x40
<i>T</i>	TGCTTCCCTGAGACCCAGTT	GATCACTTCTTTCTTTGCATCAAG	95°C 5s 58°C 20s x40
<i>TBX6</i>	CAACCCCGCATACACCTAGT	CGTCTCGCTCCCTCTTACAG	95°C 5s 58°C 20s x40
<i>MSGN1</i>	AACCTGCGCGAGACTTTCC	ACAGCTGGACAGGGAGAAGA	95°C 5s 58°C 20s x40
<i>PAX3</i>	CTCACCTCAGGTAATGGGACT	CGTGGTGGTAGGTTCCAGAC	95°C 5s 58°C 20s x40
PAX7 ChIP 1 -731bp	CGGGGCTCTGACATTACACA	GCCAGAGTCCGCCCTATTTTC	95°C 5s 60°C 20s x40
PAX7 ChIP 2 -289bp	TATTGGTCCTCCGCTCCCTT	GTGAGCGCGATCTGATAGGT	95°C 5s 60°C 20s x40
PAX7 ChIP 3 +562bp	TTGCCGACTTTGGATTGTC	TCCAAAGGGAATCCCGTGC	95°C 5s 60°C 20s x40
PAX7 ChIP 4 +926	CGCAGGGCTGAAATTCTGGT	AGAGCCGAGAAACTGTCAGG	95°C 5s 60°C 20s x40