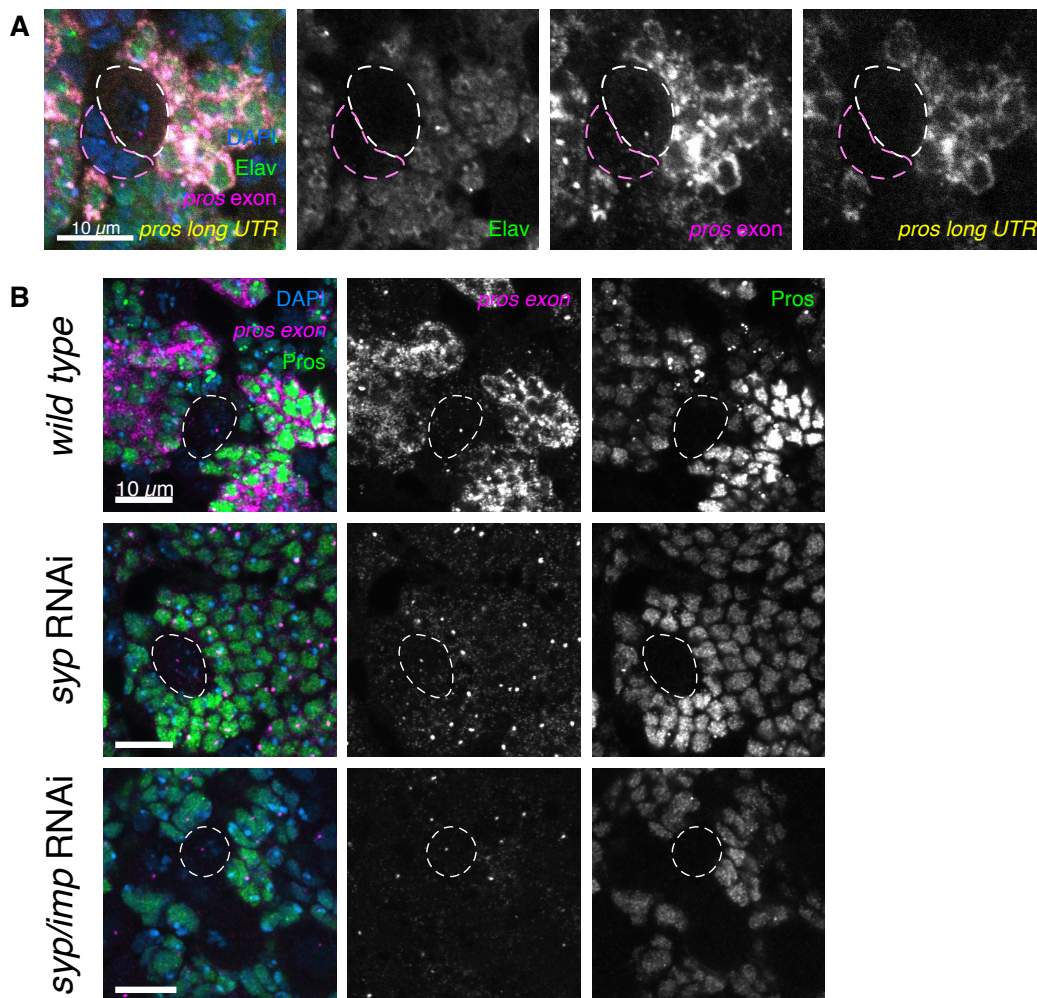
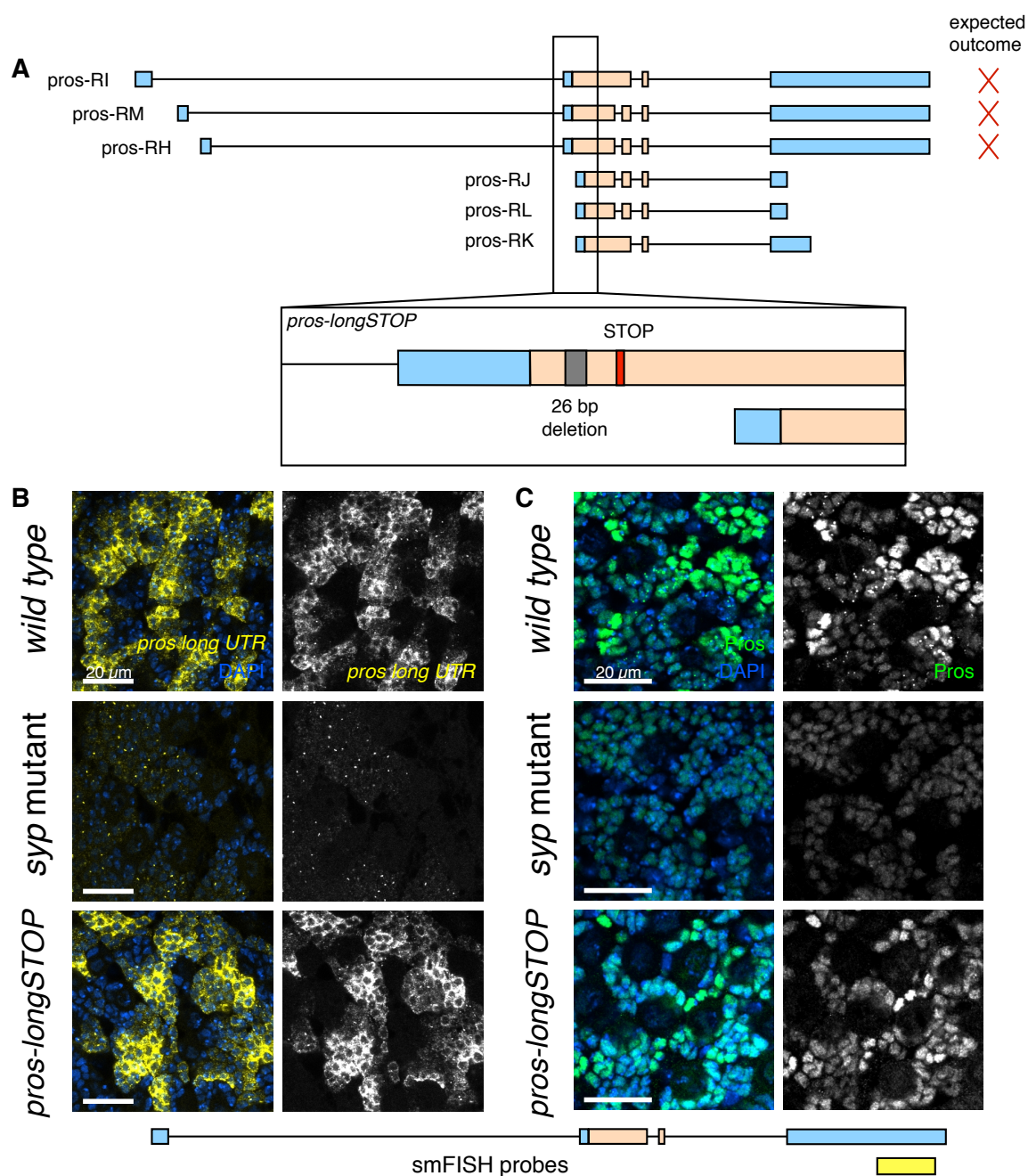


## SUPPLEMENTARY INFORMATION



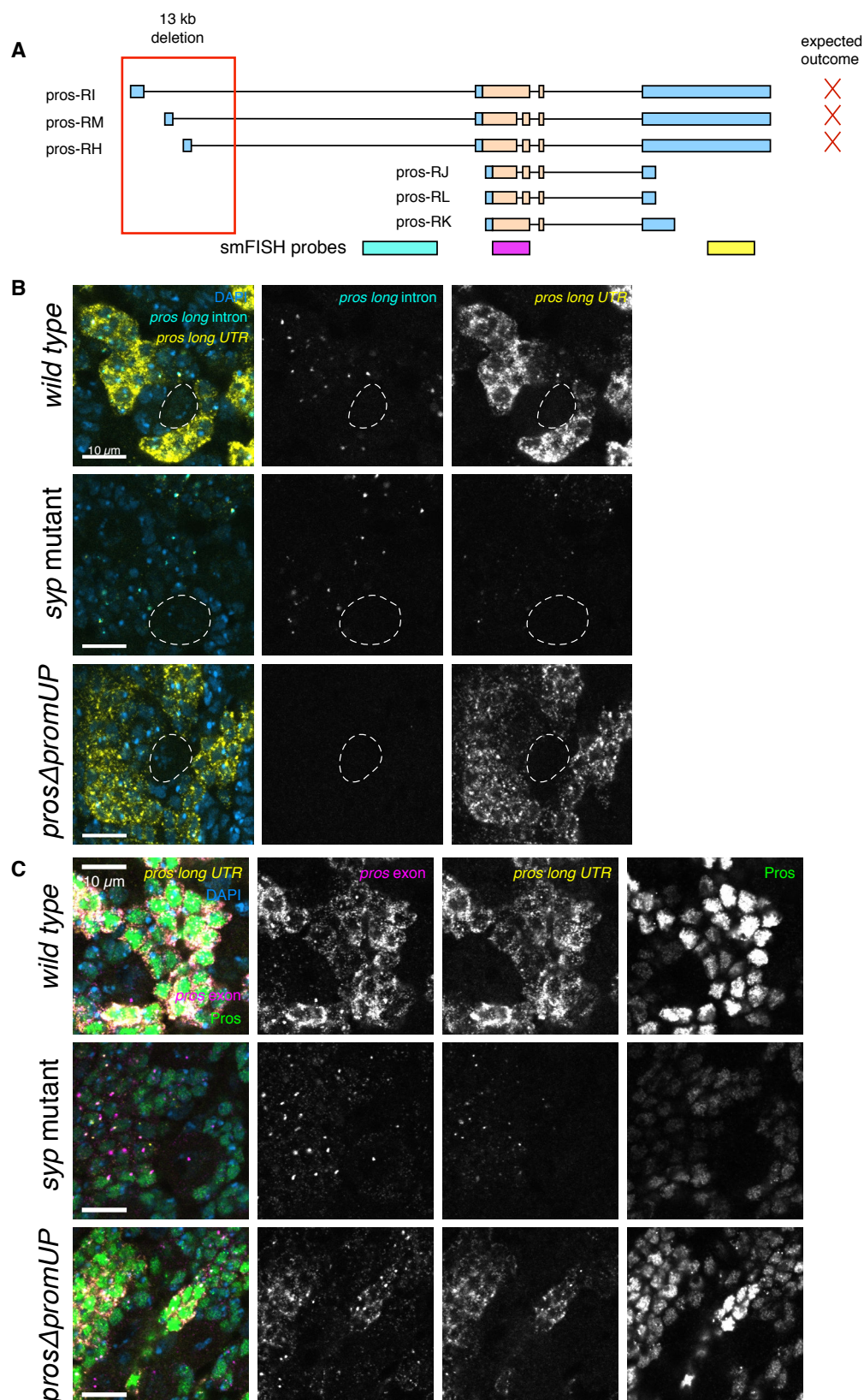
### Figure S1: Syp stabilises *pros* directly, not via Imp

**A** Staining with Elav IF (marking post-mitotic neurons), and *pros* exon and *pros long UTR* smFISH. *pros<sup>long</sup>* is only expressed in Elav+ cells, and is not expressed in the NBs (white outline) or GMCs (pink outline) **B** Brains stained with Pros protein and *pros* exon smFISH. *pros* is lost in the *syp* RNAi knockdown, and in the *syp/imp* double knockdown brains. This shows that *pros* is regulated directly by Syp, not via its downregulation of Imp. RNAi constructs are driven by *insc-GAL4* (Betschinger *et al.*, 2006).



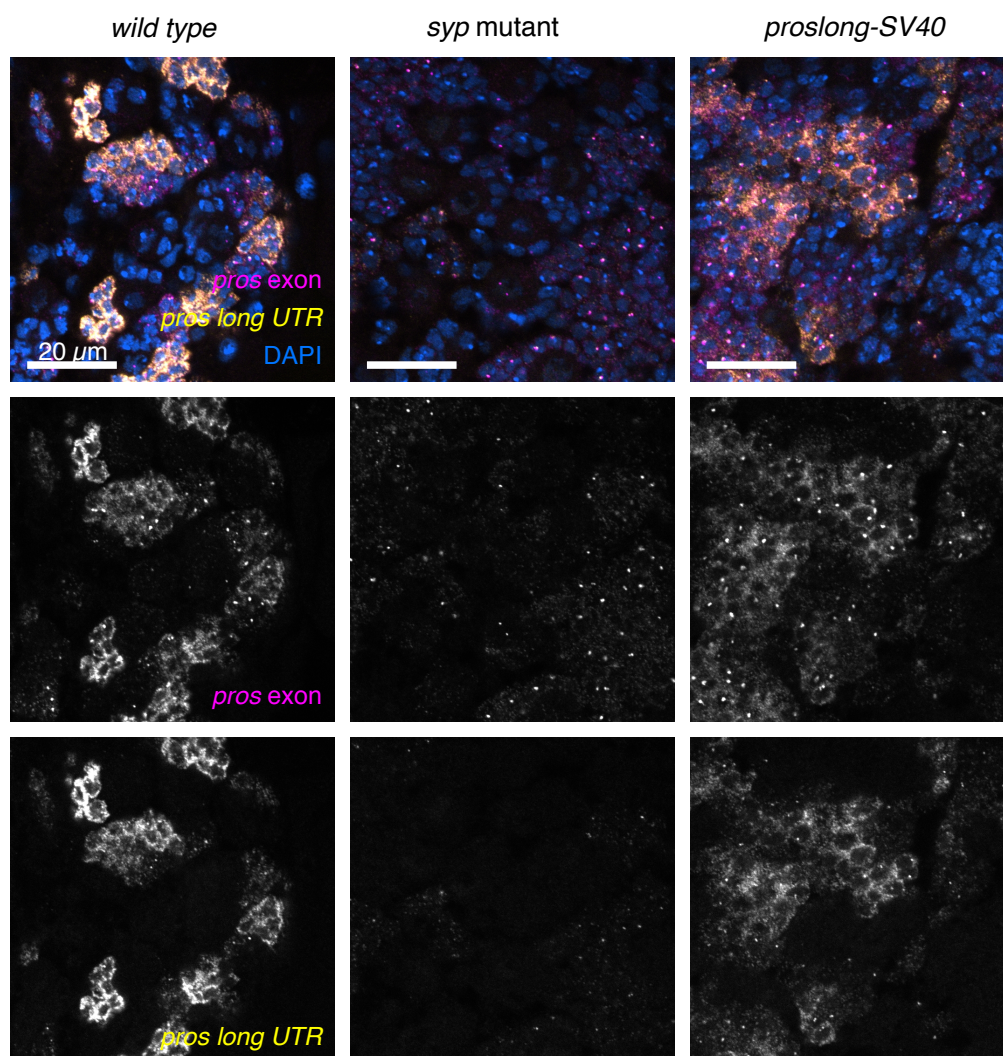
### Figure S2: Introducing an NMD mutation does not remove *pros<sup>long</sup>*

**A** We used CRISPR/Cas9 to make a double stranded DNA break in the upstream coding region unique to the *pros<sup>long</sup>* isoforms. Repair by non-homologous end joining resulted in a 26 bp deletion, leading to a frame shift which introduces a premature stop codon 80 bp downstream (*pros-longSTOP*). This should produce a truncated polypeptide and induce NMD of the *pros<sup>long</sup>* mRNA isoforms. The *pros-longSTOP* mutant was stained with **B** *pros long* UTR smFISH and **C** Pros protein. Neither *pros<sup>long</sup>* or Pros protein were decreased compared to *wild type*.



**Figure S3: The deletion of the upstream *pros* promoters uncovers a new isoform**

**A** We deleted the three upstream transcription start sites, which are annotated to produce *pros<sup>long</sup>*. (*pros $\Delta$ prosUP*). **B** Staining the with *pros-long* intron and *pros long* UTR smFISH showed that transcription from the upstream promoters was abolished, but low levels of *pros long* UTR signal remained. **C** Staining with Pros protein additionally showed that the neuronal upregulation of Pros protein was not disrupted.



**Figure S4: Insertion of an SV40 transcriptional terminator reduces, but does not abolish, *pros<sup>long</sup>* expression**

Staining for *pros* exon and *pros long* UTR smFISH shows a reduction in *pros<sup>long</sup>* expression in the *proslong-SV40* line compared to *wild type*. However the *pros<sup>long</sup>* level is not reduced as much as in the *syp* mutant.

**Table S1 - Differential expression analysis of *wild type* vs *syp* mutant third instar larval brains**

[Click here to Download Table S1](#)

**Table S2: Primers for generating Northern blot probes**

The above primers are used to generate radioactive Northern blot probes. ORF - Open Reading Frame of *pros* transcript; UTR Long are used to generate probes that are specific to the 15 kb *pros* 3' UTR (probe LONG); UTR Short are used to generate probe that is against region that is common to all *pros* 3' UTRs (probe ALL).

Name	Sequence (5' - 3')
<i>ORF F1</i>	AGAAGCGCAAGCTCTACCAG
<i>ORF R1</i>	GTCTTGGGTTTTAGGGGCGA
<i>ORF F2</i>	AAGAAACCCGGCATGGACTT
<i>ORF R2</i>	CGTCACCATCTCCGGTCAAA
<i>UTR Long1 F</i>	GACGATGGTGAACGCGAAAG
<i>UTR Long1 R</i>	TGTGGCTGTGTTCTTGTGGT
<i>UTR Long2 F</i>	ATTTCCCAATCGGCGTCCTT
<i>UTR Long2 R</i>	TTGCCTGTGCGATTGCTCTGT
<i>UTR Short1 F</i>	TTGGATGGGAACACCGCTAC
<i>UTR Short1 R</i>	GTGCTCCAAAATCGGGCTTG
<i>UTR Short2 F</i>	CGCAGGCCAAAGCTAAAAGG
<i>UTR Short2 R</i>	ACCAACGGCGAGTACAGTTT
<i>actin F</i>	GGTCGCGATTTAACCGACTACCTGAT
<i>actin R</i>	AGCAGATGTGGATCTCGAAGCAAGAG

**Table S3: Primers used for RT-qPCR for Syncrip and IgG immunoprecipitation experiments**

*pros* and housekeeping gene *rp49* was used to assess the efficiency and specificity of Syncrip binding to *pros*.

Name	Sequence (5' - 3')
<i>rp49 F</i>	GCTAAGCTGTCGCACAAA
<i>rp49 R</i>	TCCGGTGGGCAGCATGTG
<i>pros F</i>	TATGCACGACAAGCTGTCACC
<i>pros R</i>	CGACCACGAAGCGGAAATTC

**Table S4 - Sequence used to generate Stellaris® DNA probe sets**

[Click here to Download Table S4](#)

**Table S5: gRNA constructs**

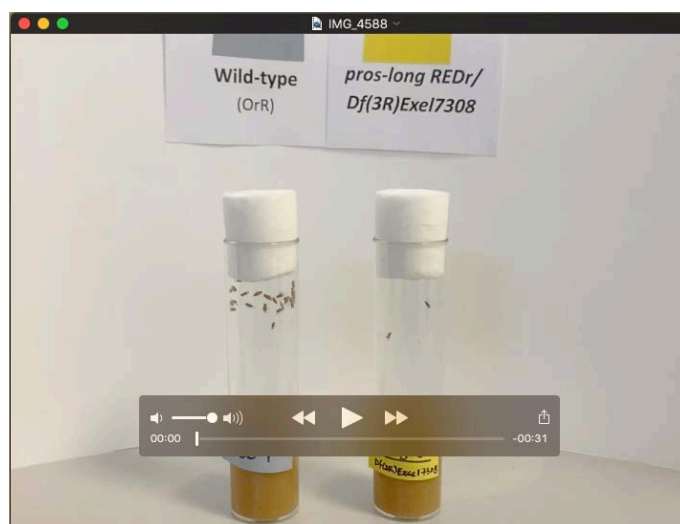
Guide RNA constructs used to produce the *pros<sup>long</sup>* deletion lines.

<b><i>pros</i> deletion lines</b>	<b>gRNA</b>	<b>Use</b>
<i>pros-longSTOP</i>	GTGTGACCGTTGCTGCTCGG_CGG	NMD mutant
<i>prosΔpromUP</i> (upstream)	TTCCTACTAACTCATGCACA_TGG	Upstream promoter deletion, upstream cut
<i>prosΔpromUP</i> (downstream)	GTTGGGCTACTAGAACTACA_AGG	Upstream promoter deletion, downstream cut
<i>prosΔUTR</i> (upstream)	TTTAGTGAGATGTGTGAAGG_TGG	Long UTR deletion, upstream cut. SV40 insertion site.
<i>prosΔUTR</i> (downstream)	ACTAAAAATGTGTAATGGAA_TGG	Long UTR deletion, downstream cut



### Movie 1 - *proslong-REDr* flies exhibit defects in their activity

1-3 day old mixed sex flies were transferred to empty vials and allowed to acclimatise for two hours without disturbance. The vial on the left contains *wild type* flies and the vial on the right contains *proslong-REDr* homozygous flies. After disturbance, by tapping the vial to bring all flies to the bottom, the *wild type* flies immediately crawl up the sides of the vial. However the activity of the *proslong-REDr* flies is impaired and the flies remain at the bottom of the vial.



### Movie 2 - hemizygous *proslong-REDr* flies phenocopy the homozygous *proslong-REDr* adult activity defect

1-3 day old mixed sex flies were transferred to empty vials and allowed to acclimatise for two hours without disturbance. The vial on the left contains *wild type* flies and the vial on the right contains *proslong-REDr/Df(3R)Exel7308* flies. The deficiency (*Df(3R)Exel7308*) includes the entire *pros* gene region. After disturbance, by tapping the vial to bring all flies to the bottom, the *wild type* flies immediately crawl up the sides of the vial. The activity of the hemizygous *proslong-REDr* flies is impaired, similarly to the *proslong-REDr* flies in Movie S1. This result suggests that the phenotype of *proslong-REDr* is not due to a CRISPR off-target effect.