

Supplementary Material

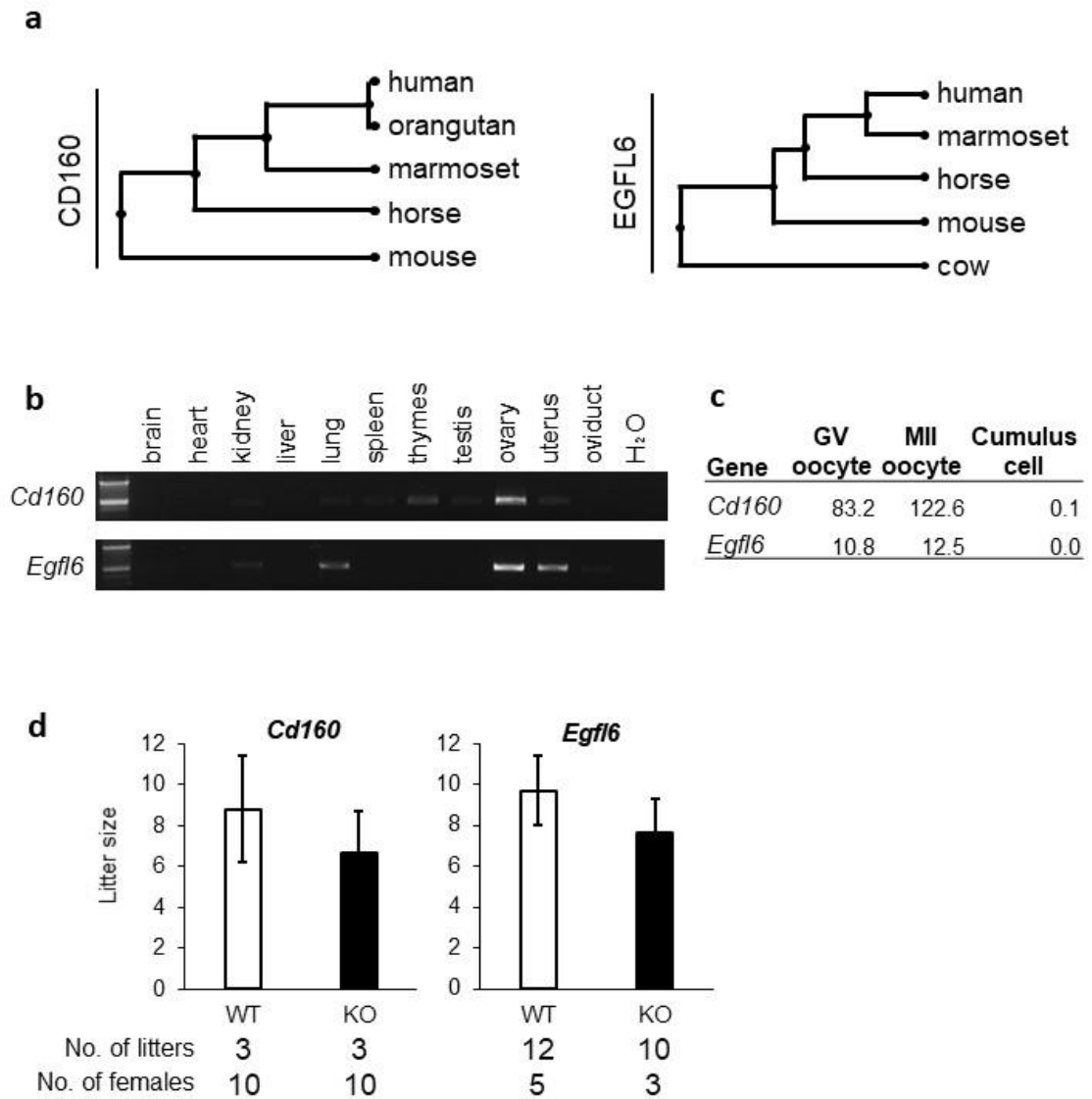


Figure S1. Conservation of *Cd160* and *Egfl6* between species, tissue expression analysis, and KO mice fertility. (a) Phylogenetic trees. (b) RT-PCR analysis using multiple tissues. (c) Expression levels in germinal vesicle (GV) oocytes, metaphase II (MII) oocytes and cumulus cells. (d) Average litter size of control and KO females.

Table S1. Primers used for RT-PCR analysis.

Gene	Upstream	Downstream	Band size (bp)
<i>Oosp1</i>	GTCTGCAATGTGCCGACCAC	CTCAGACCTGCTCGTTTCAG	464
<i>Oosp2</i>	AGTCTGCCGGATGGGGTCT	TCCGAGAGGCAGAACAACACTGCAA	374
<i>Oosp3</i>	ATGAAGGCCTTCGTTGCTTC	AGTACCAGGAACCAGCATTG	470
<i>Cd160</i>	GCTGGCAATTGTGAACTTCC	AACTGAGAGTGCCGTTGATA	428
<i>Egfl6</i>	GGTGCAAGTTCGGTGAGTGT	CGGGTATTGAGGCAATTGGC	517
<i>Actb</i>	CATCCGTAAAGACCTCTATGCCAAC	ATGGAGCCACCGATCCACA	171

Table S2. sgRNAs used for generating knockout mice. sgRNAs used for generating *Oosp* family knockout mice targeted the gene's upstream (U) and downstream (D) regions. EP: electroporation

Gene	sgRNA (5'→3')		PAM	Mutation
<i>Oosp1</i>	-		-	COMP tm1b
<i>Oosp2</i>	GTGCGTGTATTTGAGTTACA		CGG	-11/-11
<i>Oosp3</i>	GCTGCCCAGTAACTTCCATA		AGG	-26/-26
<i>Oosp</i> family	U	TATGCCACTATCTACTGAGG	AGG	-64159/
	D	ATGCTGCTGTGGCAAGTCAA	CGG	-64159
<i>Cd160</i>	AGGGCACAGCAGCTTTGGCC		AGG	-20/-20
<i>Egfl6</i>	CAGCCTGGGGTCTGTCAGTA		TGG	+1/+1

Table S3. Efficiency of generating knockout mice. EP: electroporation.

Gene	method	No. of transferred egg	GMO / pups genotyped
<i>Oosp1</i>	COMP tm1b	*ES	*ES 24/28
<i>Oosp2</i>	Plasmid injection	87	2/19 (11%)
<i>Oosp3</i>	Plasmid injection	102	3/26 (12%)
<i>Oosp</i> family	EP	93	7/33 (21%)
<i>Cd160</i>	Plasmid injection	72	1/3 (33%)
<i>Egfl6</i>	Plasmid injection	75	1/6 (17%)

Table S4. Primers used for genotyping analysis and conditions of PCR.

Gene	Allele	Forward primer	Reverse primer	Annealing	Elongation	Band Size (bp)
<i>Oosp1</i>	WT	GTGCCGACCACTGGTTC CATC	GCACTGTTACAGCACAG CCTCTC	65°C,30s	72°C,30s	532
	KO	CACACCTCCCCCTGAAC CTGAAAC	GCACTGTTACAGCACAG CCTCTC	65°C,30s	72°C,30s	331
<i>Oosp2</i>	WT	GTCCATCCTTTTCGTCAC AGCTCC	CATGCATATGGATCAGT TAAAGGACC	60°C,30s	72°C,30s	406
	KO	GTCCATCCTTTTCGTCAC AGCTCC	CATGCATATGGATCAGT TAAAGGACC	60°C,30s	72°C,30s	395
<i>Oosp3</i>	WT	AACTTCTGAGTGTTTTG	ATTATCCTATACATGGC AAG	60°C,30s	72°C,30s	577
	KO	AACTTCTGAGTGTTTTG	ATTATCCTATACATGGC AAG	60°C,30s	72°C,30s	551
<i>Oosp family</i>	WT	TGCCTACCTCCTGTCCT AACAACTG	CTTGCAGTGATTCTGTT GTCTCTG	60°C,30s	72°C,30s	746
	KO	TGCCTACCTCCTGTCCT AACAACTG	CACTGATAGTTATTTTC ATAGGTATTACAAAAGG	60°C,30s	72°C,30s	366
<i>Cd160</i>	WT	AACACGGGCTATTCACC AAAGGAGC	GACTTTGTGAAATGCCT TGGCTC	60°C,30s	72°C,30s	653
	KO	AACACGGGCTATTCACC AAAGGAGC	GACTTTGTGAAATGCCT TGGCTC	60°C,30s	72°C,30s	633
<i>Egfl6</i>	WT	CTAACTCAGAATTCT TTATCCACAGTCCAT	GTGAAGTGCAACCACT CTGGACACTGCAC	60°C,30s	72°C,30s	304
	KO	CTAACTCAGAATTCT TTATCCACAGTCCAT	GTGAAGTGCAACCACT CTGGACACTGCAC	60°C,30s	72°C,30s	305