

Supplementary Information for:

**Inner ear sensory system changes as extinct crocodylomorphs transitioned from land to water**

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## 1. Specimens Examined

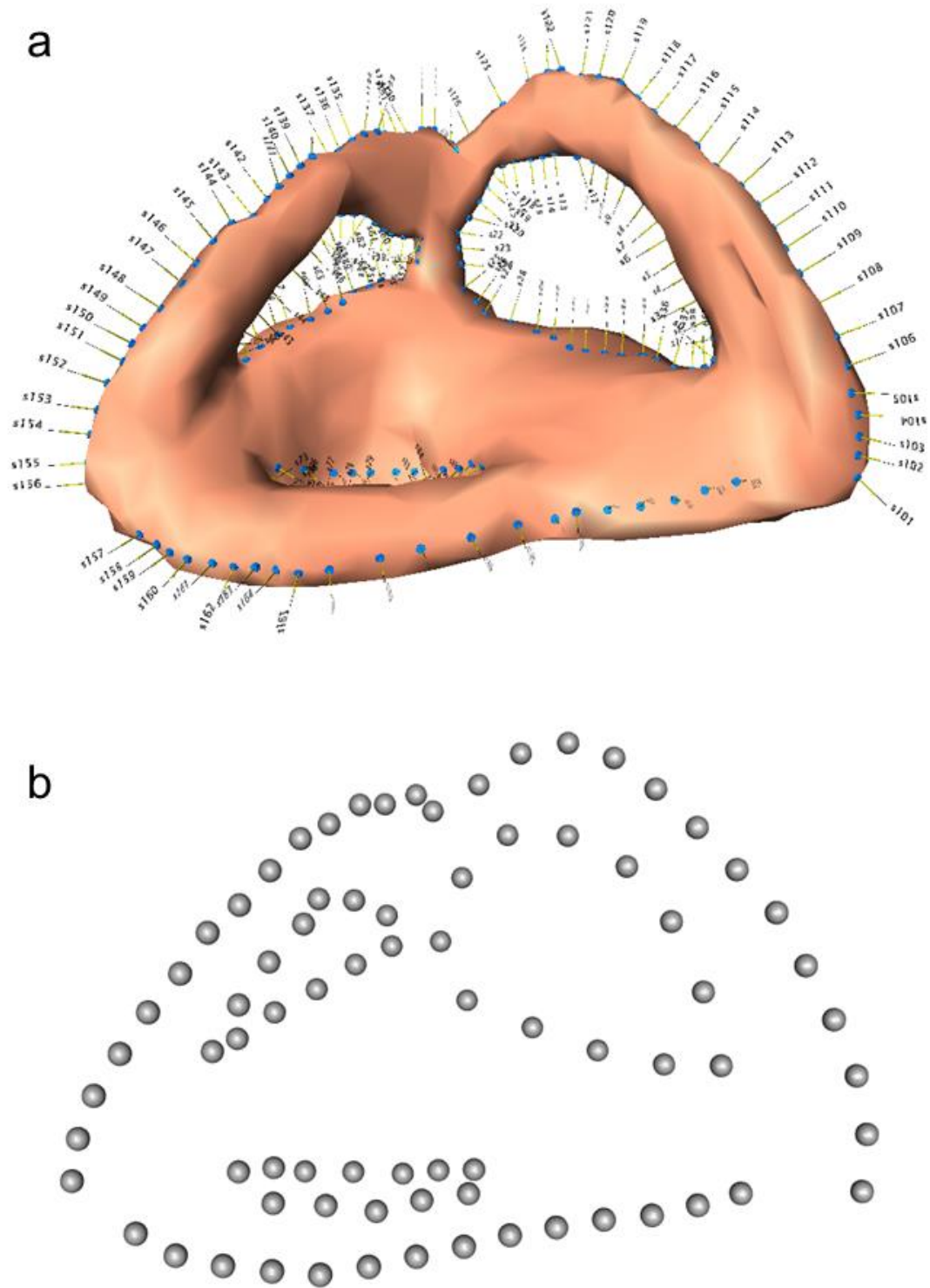
**Table S1.** Detailed information on the specimens we scanned; their museum specimen numbers, ages, and CT scanning parameters. The abbreviations are the notations used in the Figures in the main text and the Supplementary Tables.

Taxa	ID	Abbr.	voxel size (mm)	facility
<i>Junggarsuchus sloani</i>	IVPP V14010	JuSl	0.13	Institute of Vertebrate Paleontology and Paleoanthropology (IVPP) of the Chinese Academy of Sciences
Undescribed sphenosuchian	NCSM 21722	UnSp	0.039327	Nikon XTH 225 ST $\mu$ CT scanner, Shared Materials Instrumentation Facility, Duke University, Durham, NC
<i>Eopneumatosuchus colberti</i>	MNA V2460	EoCo	0.092	General Electric eXplore Locus in vivo Small Animal $\mu$ CT scanner, Ohio University $\mu$ CT Scanning Facility
<i>Protosuchus haughtoni</i>	BP/1/4770	PrHa	0.0538	Nikon Metrology XTH 225/320 LC dual source industrial CT system, University of the Witwatersrand, Johannesburg
' <i>Steneosaurus</i> ' cf. <i>gracilirostris</i>	NHMUK PV OR 33095	StGr	0.089	Nikon XT H 225S CTsystem, Natural History Museum, London
<i>Steneosaurus bollensis</i>	BSPG 1984 I258	StBo1	0.043	Nanotom Scan, Zoologische Staatssammlung München
<i>Steneosaurus bollensis</i>	MCZ 1063	StBo2	0.404297	micro CT scanner at the American Museum of Natural History
<i>Steneosaurus pictaviensis</i>	LPP.M.35	StPi	0.140036	RX-solutions EasyTom XL Duo, Plateforme de Microtomographie, University of Poitiers
<i>Pelagosaurus typus</i>	NHMUK PV OR 32599	PtTy	0.098627983	Nikon XT H 225S CTsystem, Natural History Museum, London
<i>Metriorhynchus superciliosus</i>	MNHN.F RJN 256	MeSu1	0.10358673X 0.265	Muséum national d'Histoire naturelle, Paris, France
<i>Metriorhynchus superciliosus</i>	NHMUK PV R11999	MeSu2	0.12	$\mu$ -VIS X-Ray Imaging Centre, University of Southampton
<i>Metriorhynchus superciliosus</i>	AMNH 997	MeSu3	0.14177665	micro CT scanner at the American Museum of Natural History
<i>Cricosaurus araucanensis</i>	MLP 72-IV-7-1	CrAu1	0.448X0.448	SNSB X-ray facility
<i>Cricosaurus araucanensis</i>	MLP 76-XI-19-1	CrAu2	0.39	SNSB X-ray facility
<i>Cricosaurus schroederi</i>	MMGLV #	CrSc	0.5	Leibniz Institute for Zoo and Wildlife Research, Berlin
' <i>Metriorhynchus</i> ' <i>brachyrhynchus</i>	NHMUK PV OR 32618	MeBr	0.0546	$\mu$ -VIS X-Ray Imaging Centre, University of Southampton
<i>Torvoneustes coryphaeus</i>	MJML K1863	ToCo	0.2267	$\mu$ -VIS X-Ray Imaging Centre, University of Southampton
<i>Shamosuchus djadochtaensis</i>	IGM 100-1195	ShDj	0.011260684	micro CT scanner at the American Museum of Natural History
<i>Alligator mississippiensis</i>	USNM 211232	AlMi4	0.625	Ohio Health O'Bleness Hospital
<i>Alligator mississippiensis</i>	USNM 211233	AlMi5	0.625	Ohio Health O'Bleness Hospital
<i>Alligator mississippiensis</i>	OUVG 9761	AlMi6	0.5X1	Ohio Health O'Bleness Hospital
<i>Caiman crocodilus</i>	FMNH 73711	CaCr	0.065X0.142	University of Texas, Austin
<i>Crocodylus acutus</i>	FMNH 59071	CrAc	0.625	Ohio Health O'Bleness Hospital
<i>Crocodylus johnstoni</i>	TMM M-6807	CrJo	0.223	University of Texas High-Resolution X-ray CT Facility
<i>Crocodylus moreletii</i>	TMM M-4980	CrMo	0.1904X0.5	High-Resolution X-ray CT Facility, University of Texas
<i>Crocodylus rhombifer</i>	MNB AB50.0171	CrRh	0.1748X0.5	High-Resolution X-ray CT Facility, University of Texas
<i>Mecistops cataphractus</i>	TMM M-3529	CrCa	0.165X0.5	High-Resolution X-ray CT Facility, University of Texas
<i>Gavialis gangeticus</i>	UF-herp-118998	GaGa2	0.14654672	Florida Museum of Natural History Herpetology
<i>Gavialis gangeticus</i>	TMM M-5490	GaGa1	0.228	High-Resolution X-ray CT Facility, University of Texas
<i>Osteolaemus tetraspis</i>	FMNH 98936	OsTe	0.0546875X0.1108	High-Resolution X-ray CT Facility, University of Texas
<i>Tomistoma schlegelii</i>	TMM M-6342	ToSc1	0.165X0.46	High-Resolution X-ray CT Facility, University of Texas
<i>Tomistoma schlegelii</i>	USNM 211322	ToSc2	0.625	Ohio Health O'Bleness Hospital

**Table S2.** Information of the habitat (ecomorphology) and age of each species used in our sample.

Taxa	habitat	Age	Reason	Reference
<i>Junggarsuchus sloani</i>	terrestrial	M. Jurassic	Inferred, from deposit, and limb-dimensions and cranial shape	1
Undescribed sphenosuchian	terrestrial	Carnian	Inferred, from deposit, and limb-dimensions and cranial shape	NCSM 21722
<i>Eopneumatosuchus colberti</i>	semiaquatic	Sinemurian*	Inferred, from deposit and cranial shape	2
<i>Protosuchus haughtoni</i>	terrestrial	Hettangian	Inferred, from post-cranial skeleton	3
' <i>Steneosaurus</i> ' <i>gracilirostris</i>	semiaquatic	Toarcian	Inferred, from a marine deposit, but has extant-like limbs, and has osteoderms	4
<i>Steneosaurus bollensis</i>	semiaquatic	Toarcian	Inferred, from a marine deposit, but has extant-like limbs, and has osteoderms	4
<i>Steneosaurus pictaviensis</i>	semiaquatic	Toarcian	Inferred, from a marine deposit and is a teleosaurid	5
<i>Pelagosaurus typus</i>	semiaquatic	Toarcian	Inferred, from a marine deposit, but has extant-like limbs, and has osteoderms	4
<i>Metriorhynchus superciliosus</i>	pelagic	Callovian	Inferred, marine deposit, has flippers and a tail-fin	5
<i>Cricosaurus araucanensis</i>	pelagic	Tithonian	Inferred, marine deposit, has flippers and a tail-fin	6, 7
<i>Cricosaurus schroederi</i>	pelagic	Valanginian	Inferred, marine deposit and is a metriorhynchid	8
' <i>Metriorhynchus</i> ' <i>brachyrhynchus</i>	pelagic	Callovian	Inferred, marine deposit and has a tail-fin	5
<i>Torvoneustes coryphaeus</i>	pelagic	Kimmeridgian	Inferred, marine deposit and is a metriorhynchid	9
<i>Shamosuchus djadochtaensis</i>	semiaquatic	L. Cretaceous	Inferred, based on post-cranial anatomy	10
<i>Alligator mississippiensis</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Caiman crocodilus</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Crocodylus acutus</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Crocodylus johnstoni</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Crocodylus moreletii</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Crocodylus rhombifer</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Mecistops cataphractus</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Gavialis gangeticus</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Osteolaemus tetraspis</i>	semiaquatic	Recent	extant, observed	e.g. 11
<i>Tomistoma schlegelii</i>	semiaquatic	Recent	extant, observed	e.g. 11

\*The age of the Kayenta Formation may be Pliensbachian-Toarcian, as has been presented in a conference abstract (Marsh et al., 2014, Society of Vertebrate Paleontology Annual Meeting).



**Fig. S1.** Landmark placing on the right labyrinth of *Metriorhynchus superciliosus* (NHMUK PV R11999). a, landmark placement in the IDAV Landmark software; b, evenly spaced semilandmarks on the three semicircular canals, 11 on the internal surface and 12 on the external surface (see methods section for details).

### 3. Principal component analysis

**Table S3.** Results of the Principal Component Analysis. PCA coordinates for the first 31 PC axes.

	PC1	PC2	PC3	PC4	PC5	PC6	PC7	PC8
<b>AlMi4</b>	-0.025	-0.024	0.035	-0.038	0.013	-0.002	0.009	0.000
<b>AlMi5</b>	-0.022	-0.050	0.025	-0.044	0.000	-0.007	0.010	-0.003
<b>CaCr</b>	-0.051	-0.043	-0.011	-0.037	-0.025	-0.001	0.010	0.028
<b>CrAc</b>	-0.016	-0.056	0.015	0.025	0.022	-0.030	-0.020	-0.006
<b>CrMo</b>	-0.061	-0.021	-0.019	0.013	0.011	0.012	-0.014	-0.004
<b>CrJo</b>	-0.059	-0.003	-0.004	-0.059	0.012	0.000	0.012	-0.028
<b>CrCa</b>	-0.014	-0.085	0.022	0.047	0.011	-0.023	0.005	-0.004
<b>OsTe</b>	-0.030	-0.002	0.049	-0.003	-0.035	-0.044	-0.012	0.034
<b>GaGa1</b>	-0.028	-0.056	0.054	-0.006	0.004	-0.008	0.005	0.004
<b>ToSc1</b>	-0.051	-0.069	0.013	0.027	0.013	0.014	-0.007	0.014
<b>ToSc2</b>	-0.022	-0.071	0.035	0.025	0.009	0.027	-0.017	0.021
<b>GaGa2</b>	-0.032	-0.010	0.058	-0.023	0.037	0.028	-0.019	0.005
<b>AlMi6</b>	-0.038	0.013	0.028	-0.045	0.001	-0.011	0.042	-0.030
<b>CrRh</b>	-0.033	-0.053	0.002	0.039	0.001	0.036	0.001	-0.005
<b>PeTy</b>	0.014	0.084	0.028	0.006	-0.043	0.010	-0.016	-0.002
<b>JuSI</b>	-0.115	0.019	-0.064	0.031	-0.033	-0.056	0.012	0.002
<b>MeSu2</b>	0.057	0.015	-0.016	-0.041	0.038	0.003	0.048	0.032
<b>StBo1</b>	0.011	0.056	0.058	0.023	-0.011	0.013	-0.017	0.013
<b>CrSc</b>	0.104	-0.092	-0.062	-0.005	-0.077	0.060	-0.011	-0.047
<b>EoCo</b>	-0.048	0.066	-0.030	0.009	0.014	0.046	-0.020	-0.021
<b>UnSp</b>	-0.087	0.039	-0.065	-0.008	-0.081	-0.015	0.016	0.012
<b>MeSu1</b>	0.038	0.064	-0.037	-0.004	0.001	0.031	0.031	0.033
<b>StBo2</b>	0.014	0.097	0.059	0.018	-0.012	-0.024	0.006	-0.056
<b>StGr</b>	0.059	0.066	0.029	0.051	-0.018	0.001	0.005	0.001
<b>MeBr</b>	0.072	0.063	-0.009	0.000	0.040	-0.006	0.049	0.005
<b>MeSu3</b>	0.028	0.039	-0.022	0.005	-0.004	0.041	0.029	0.031
<b>CrAu1</b>	0.136	0.008	0.021	-0.074	-0.026	-0.023	-0.073	0.037
<b>PrHa</b>	-0.071	0.101	-0.096	-0.005	0.060	-0.002	-0.071	-0.005
<b>ToCo</b>	0.115	-0.065	-0.110	0.057	0.025	-0.042	0.005	0.017
<b>CrAu2</b>	0.087	-0.028	-0.032	-0.047	0.019	-0.044	-0.003	-0.052
<b>StPi</b>	0.073	0.031	0.068	0.069	0.005	-0.011	0.002	-0.013
<b>ShDj</b>	-0.003	-0.035	-0.025	-0.007	0.027	0.025	0.002	-0.016

	PC9	PC10	PC11	PC12	PC13	PC14	PC15	PC16
<b>AlMi4</b>	0.028	-0.006	-0.019	0.014	-0.035	-0.013	-0.002	0.039
<b>AlMi5</b>	0.016	-0.002	0.026	-0.018	-0.009	-0.018	0.008	0.008
<b>CaCr</b>	0.008	-0.005	-0.012	-0.020	0.014	0.003	-0.024	0.008
<b>CrAc</b>	0.042	-0.010	0.014	-0.023	0.040	0.005	0.015	0.002
<b>CrMo</b>	-0.023	-0.018	-0.003	-0.014	0.011	0.015	0.006	0.005
<b>CrJo</b>	-0.030	-0.022	-0.008	0.022	-0.004	0.008	0.026	-0.013
<b>CrCa</b>	-0.032	-0.010	0.024	0.005	0.012	-0.010	0.023	0.001
<b>OsTe</b>	0.012	-0.040	0.003	-0.001	-0.022	0.034	-0.015	-0.012
<b>GaGa1</b>	0.014	0.018	0.010	0.004	-0.002	0.007	-0.001	0.006
<b>ToSc1</b>	-0.013	0.017	0.005	0.006	0.010	-0.017	-0.003	-0.005
<b>ToSc2</b>	0.032	0.013	0.006	0.024	-0.020	-0.036	-0.005	-0.026
<b>GaGa2</b>	0.008	0.028	-0.012	-0.011	0.014	0.026	-0.015	-0.013
<b>AlMi6</b>	-0.002	-0.034	-0.017	-0.022	0.009	-0.024	-0.022	-0.017
<b>CrRh</b>	-0.025	-0.015	-0.004	0.017	-0.006	0.018	0.004	0.024
<b>PeTy</b>	-0.017	-0.014	0.002	-0.011	0.007	-0.022	-0.013	0.015
<b>JuSl</b>	0.005	0.045	-0.053	-0.014	-0.010	-0.003	0.027	-0.004
<b>MeSu2</b>	-0.023	0.009	0.024	-0.006	-0.015	0.008	0.026	0.009
<b>StBo1</b>	-0.016	0.021	0.007	-0.017	-0.030	0.015	0.001	-0.004
<b>CrSc</b>	0.036	-0.009	-0.008	-0.010	-0.006	0.013	0.018	-0.003
<b>EoCo</b>	-0.007	0.001	0.004	-0.006	-0.030	-0.008	-0.005	-0.015
<b>UnSp</b>	-0.007	0.005	0.041	0.044	0.013	0.003	-0.012	-0.005
<b>MeSu1</b>	0.028	0.012	-0.013	0.012	0.026	-0.010	-0.006	0.017
<b>StBo2</b>	0.006	0.012	0.013	-0.009	-0.002	0.012	-0.003	0.008
<b>StGr</b>	-0.022	-0.011	0.002	-0.022	0.005	-0.032	0.012	-0.002
<b>MeBr</b>	0.033	-0.024	-0.013	0.022	0.002	0.006	0.016	-0.021
<b>MeSu3</b>	-0.001	0.020	0.018	-0.025	0.011	0.017	0.000	-0.007
<b>CrAu1</b>	-0.025	0.002	-0.021	0.011	0.012	-0.008	0.017	-0.005
<b>PrHa</b>	0.027	-0.018	0.015	0.004	0.003	-0.002	0.003	0.008
<b>ToCo</b>	-0.010	-0.016	-0.002	-0.015	-0.025	0.001	-0.028	-0.001
<b>CrAu2</b>	-0.005	0.045	0.023	0.009	0.000	-0.001	-0.020	0.000
<b>StPi</b>	0.011	0.001	-0.020	0.040	0.012	0.011	-0.006	0.005
<b>ShDj</b>	-0.048	0.004	-0.031	0.011	0.016	0.002	-0.023	0.000



	PC17	PC18	PC19	PC20	PC21	PC22	PC23	PC24
<b>AlMi4</b>	-0.013	0.000	-0.005	-0.001	-0.005	0.018	0.002	0.002
<b>AlMi5</b>	-0.012	0.000	0.018	0.009	-0.003	-0.006	-0.015	-0.015
<b>CaCr</b>	-0.011	-0.008	0.008	0.001	0.033	-0.004	0.002	0.011
<b>CrAc</b>	-0.007	0.007	0.006	-0.007	-0.002	0.007	-0.009	0.013
<b>CrMo</b>	-0.011	0.010	0.004	-0.004	0.004	0.001	0.009	-0.019
<b>CrJo</b>	-0.004	-0.002	0.016	0.011	0.001	-0.011	0.011	0.017
<b>CrCa</b>	0.012	-0.011	-0.003	-0.002	0.003	0.015	0.011	0.000
<b>OsTe</b>	0.027	0.014	-0.008	-0.008	-0.002	-0.001	-0.005	0.001
<b>GaGa1</b>	0.012	-0.003	0.002	0.004	-0.002	-0.015	-0.002	-0.009
<b>ToSc1</b>	-0.007	0.001	-0.023	-0.023	0.005	-0.008	0.000	0.002
<b>ToSc2</b>	-0.001	0.006	0.004	0.004	-0.008	-0.009	0.005	0.007
<b>GaGa2</b>	0.005	-0.029	-0.014	0.022	-0.003	0.005	0.003	-0.006
<b>AlMi6</b>	0.008	0.003	-0.004	-0.003	-0.014	0.012	0.010	-0.004
<b>CrRh</b>	0.014	0.004	0.011	0.001	-0.003	-0.013	0.000	-0.007
<b>PeTy</b>	0.006	0.002	-0.009	0.013	-0.002	-0.014	-0.006	0.010
<b>JuSl</b>	0.007	0.002	0.000	0.004	-0.004	-0.001	-0.004	-0.001
<b>MeSu2</b>	0.011	-0.007	-0.020	-0.001	0.000	0.003	-0.009	0.010
<b>StBo1</b>	-0.026	0.021	-0.006	0.000	0.006	0.006	0.016	0.002
<b>CrSc</b>	0.002	-0.001	-0.015	0.002	0.000	0.002	0.001	0.001
<b>EoCo</b>	0.017	-0.015	0.019	-0.018	0.015	0.013	-0.013	-0.001
<b>UnSp</b>	-0.015	-0.009	-0.005	0.005	-0.004	0.010	-0.003	-0.005
<b>MeSu1</b>	0.021	0.000	0.007	-0.012	-0.002	-0.003	0.017	-0.003
<b>StBo2</b>	-0.012	-0.021	0.001	-0.021	-0.010	-0.015	0.003	0.003
<b>StGr</b>	0.006	0.002	-0.006	0.016	0.011	0.001	-0.003	-0.007
<b>MeBr</b>	-0.017	0.004	-0.009	-0.003	0.013	-0.010	-0.005	-0.012
<b>MeSu3</b>	-0.004	0.014	0.022	0.002	-0.017	0.006	-0.002	0.006
<b>CrAu1</b>	-0.004	-0.009	0.009	-0.011	-0.006	0.002	0.000	-0.006
<b>PrHa</b>	0.002	0.006	-0.011	0.009	-0.003	-0.002	0.003	0.001
<b>ToCo</b>	-0.011	-0.018	0.007	0.005	-0.008	-0.005	0.005	0.002
<b>CrAu2</b>	0.017	0.023	0.001	0.003	0.013	0.001	0.005	-0.003
<b>StPi</b>	0.000	0.001	0.013	0.011	0.005	0.014	-0.007	0.006
<b>ShDj</b>	-0.011	0.014	-0.009	-0.005	-0.010	0.000	-0.019	0.003

	PC25	PC26	PC27	PC28	PC29	PC30	PC31
<b>AlMi4</b>	-0.001	-0.002	-0.014	0.005	0.000	0.002	0.001
<b>AlMi5</b>	0.001	-0.003	0.010	-0.007	0.001	-0.010	0.008
<b>CaCr</b>	0.010	-0.001	0.002	0.001	-0.004	-0.003	-0.006
<b>CrAc</b>	-0.010	-0.004	0.000	0.001	0.010	0.008	-0.001
<b>CrMo</b>	-0.015	-0.005	-0.005	-0.008	-0.015	0.005	-0.002
<b>CrJo</b>	-0.004	-0.004	-0.002	0.005	0.000	0.002	0.008
<b>CrCa</b>	0.018	0.006	-0.007	-0.010	0.002	-0.004	0.001
<b>OsTe</b>	0.002	-0.008	-0.004	-0.001	-0.001	-0.004	0.005
<b>GaGa1</b>	0.010	0.015	0.003	0.007	-0.006	0.018	0.003
<b>ToSc1</b>	-0.010	0.007	-0.001	0.011	-0.003	-0.010	0.008
<b>ToSc2</b>	0.000	-0.010	-0.001	-0.007	-0.004	0.001	-0.009
<b>GaGa2</b>	-0.005	-0.005	-0.005	-0.001	0.005	-0.003	0.001
<b>AlMi6</b>	-0.002	0.009	0.011	0.004	0.000	-0.001	-0.005
<b>CrRh</b>	-0.004	0.000	0.004	0.006	0.013	-0.006	-0.011
<b>PeTy</b>	-0.010	0.015	-0.009	-0.012	0.002	0.001	0.001
<b>JuSl</b>	0.000	0.002	0.001	-0.002	0.001	-0.002	-0.002
<b>MeSu2</b>	-0.007	-0.003	0.010	-0.003	-0.006	0.002	-0.006
<b>StBo1</b>	0.002	0.004	0.012	-0.003	0.009	0.004	0.003
<b>CrSc</b>	0.003	0.001	0.003	0.000	-0.003	-0.001	0.002
<b>EoCo</b>	-0.004	0.004	-0.002	-0.001	0.003	0.005	0.002
<b>UnSp</b>	-0.003	-0.002	0.001	0.002	0.003	0.004	-0.001
<b>MeSu1</b>	0.000	-0.009	0.005	-0.006	0.004	0.002	0.008
<b>StBo2</b>	0.006	-0.009	-0.001	-0.001	-0.004	-0.002	-0.004
<b>StGr</b>	0.004	-0.017	-0.003	0.014	-0.001	0.003	0.001
<b>MeBr</b>	0.003	0.008	-0.009	-0.003	0.007	-0.001	-0.003
<b>MeSu3</b>	0.006	0.007	-0.013	0.007	-0.006	-0.006	-0.001
<b>CrAu1</b>	0.000	0.003	0.001	0.002	0.001	0.000	-0.004
<b>PrHa</b>	0.011	0.003	0.006	0.003	-0.004	-0.004	-0.001
<b>ToCo</b>	-0.005	0.002	0.001	0.001	0.001	0.002	0.003
<b>CrAu2</b>	-0.005	-0.002	-0.006	0.000	0.002	-0.003	-0.002
<b>StPi</b>	-0.005	0.006	0.011	0.000	-0.011	-0.005	0.002
<b>ShDj</b>	0.014	-0.009	0.001	-0.006	0.002	0.005	0.003

**Table S4.** Results of the Principal Component Analysis. Variance (%) and Cumulative Proportion (%) for the first 31 principal component axes.

	Variance (%)	Cumulative Proportion (%)
PC1	22.269	22.269
PC2	18.443	40.713
PC3	12.386	53.099
PC4	7.336	60.435
PC5	5.641	66.076
PC6	4.605	70.681
PC7	4.183	74.864
PC8	3.487	78.351
PC9	3.100	81.450
PC10	2.368	83.819
PC11	2.124	85.943
PC12	1.951	87.894
PC13	1.702	89.596
PC14	1.525	91.121
PC15	1.454	92.575
PC16	1.006	93.581
PC17	0.918	94.498
PC18	0.769	95.267
PC19	0.748	96.015
PC20	0.597	96.612
PC21	0.540	97.152
PC22	0.483	97.635
PC23	0.409	98.044
PC24	0.374	98.418
PC25	0.325	98.743
PC26	0.309	99.053
PC27	0.262	99.315
PC28	0.197	99.511
PC29	0.196	99.707
PC30	0.167	99.874
PC31	0.126	100.000

#### 4. Morphospace clustering analysis

**Table S5.** Results of the PERMANOVA, to test if the three habitat groups are statistically significantly different from each other.

pairs	F Model	R2	p value	p adjusted
<b>semiaquatic vs. terrestrial</b>	4.875	0.223	0.002	0.003
<b>semiaquatic vs. pelagic</b>	5.302	0.218	0.002	0.003
<b>terrestrial vs. pelagic</b>	4.543	0.431	0.012	0.012

## 5. Canonical variate analysis

**Table S6.** Results for the canonical variate analysis (CVA). Showing the classification of the three habitat groups (pelagic, semiaquatic, terrestrial) with an overall classification accuracy of 100%.

	pelagic	semiaquatic	terrestrial
pelagic	100	0	0
semiaquatic	0	100	0
terrestrial	0	0	100

overall classification accuracy: 100 %

	CV 1	CV 2
AlMi4	-1.033	0.846
AlMi5	-0.485	1.594
CaCr	-0.128	-0.364
CrAc	-2.027	0.536
CrMo	-3.402	0.029
CrJo	-2.502	-0.035
CrCa	-3.322	1.787
OsTe	-3.384	-0.204
GaGa1	-2.031	1.215
ToSc1	-2.747	0.397
ToSc2	-1.644	0.679
GaGa2	-2.243	2.108
AlMi6	-2.406	2.322
CrRh	-3.277	1.411
PeTy	-1.567	0.197
JuSI	-1.680	-7.193
MeSu2	5.803	1.926
StBo1	-1.803	1.280
CrSc	5.133	1.732
EoCo	-1.782	-1.412
UnSp	0.071	-7.272
MeSu1	5.550	-1.380
StBo2	-2.583	0.189
StGr	-0.300	2.021
MeBr	4.587	0.826
MeSu3	4.047	0.695
CrAu1	4.832	0.948
PrHa	-0.600	-6.291
ToCo	6.925	-0.480
CrAu2	5.415	-0.885
StPi	-1.109	1.514
ShDj	-0.307	1.266

## 6. Phylogenies used for phylogenetic comparative methods

The main phylogenetic framework used herein is based on the results from Young *et al.* (12). This dataset (the ‘H+Y dataset’) is the latest in the ongoing Crocodylomorph SuperMatrix Project, a continuously updated dataset investigating the evolutionary relationships of crocodylomorph archosaurs. It was first presented in Ristevski *et al.* (13), and has been subsequently updated (e.g. 14–17). It is one of three datasets that form the core of the Crocodylomorph SuperMatrix Project, the other two also being presented in a modified form in Ósi *et al.* (14).

However, there are some taxa we use in our datasets that are known to have differing positions phylogenetically. The first of which is *Eopneumatosuchus colberti*. It has been recovered as the basal-most thalattosuchian in the ‘H+Y dataset’. However, in Young *et al.* (12), the position of *Eopneumatosuchus* can change depending on use of implied weighting or using Bayesian methods. In those analyses, *Eopneumatosuchus* is recovered as the sister taxon to Shartegosuchoidea + Mesoeucrocodylia. To test whether this second possible position of *Eopneumatosuchus* alters our result, we ran a series of sensitivity analyses with *Eopneumatosuchus* placed as the sister taxon to Thalattosuchia + (*Shamosuchus* + Crocodylia).

The second species we ran sensitivity analyses on was *Pelagosaurus typus*. While the more recent and large-sampled thalattosuchians phylogenetic datasets recover *Pelagosaurus* as the basal-most member of Metriorhynchoidea (papers using the ‘H+Y’ dataset, 18), some earlier datasets and pre-phylogenetic opinions considered *Pelagosaurus* to be a teleosauroid (e.g. 4, 19; although see Buffetaut (20) for a metriorhynchid-like opinion). We tested this alternate position of *Pelagosaurus* to see if it impacted upon our results.

There are also some competing hypotheses for the relationships of extant crocodylians in our dataset. In particular, there is no consensus on whether *Osteolaemus tetraspis* is more closely related to *Mecistops* or *Crocodylus*, in either molecular or morphological phylogenies (21). Additionally, the position of *Gavialis gangeticus* differs between molecular and morphological datasets. Molecular datasets recover *Gavialis* as the sister taxon to *Tomistoma* (being within Crocodylidae) – a position the Young *et al.* (12) dataset also recovers (e.g. 21, 22). However, most morphology-only datasets recover *Gavialis* as the basal-most living species, being the sister taxon to Alligatoridae + Crocodylidae (for further analyses and discussion see 23). However, as these both concern small nuances of the relationships of extant crocodylians—all of which have semiaquatic-type ears—these rearrangements would not affect our results.

## 7. Pagel's Lambda analysis

**Table S7.** Results for the phylogenetic influence tested with the Pagel's lambda tested for three different phylogenies: our primary phylogeny (first listed) and two alternative trees, for sensitivity analysis (see Section 6 above). 0 → no correlation between species; 1 → correlation between species equal to the Brownian expectation/the structure of the phylogeny alone can explain changes in traits.

### Primary phylogeny

	lambda	logL	logL0	p value
PC1	0.99996	28.705	16.435	0.00000073
PC2	0.69646	31.134	22.893	0.00004913
PC3	0.99993	31.597	18.921	0.00000048

### *Eopneumatosuchus colberti* placed as the sister taxon to Thalattosuchia + (*Shamosuchus* + Crocodylia)

	lambda	logL	logL0	p value
PC1	0.77400	27.769	16.506	2E-06
PC2	0.65831	30.953	22.949	6E-05
PC3	0.99993	32.243	19.051	3E-07

### *Pelagosaurus typus* positioned as a teleosauroid

	lambda	logL	logL0	p value
PC1	0.77400	27.769	16.506	2E-06
PC2	0.65831	30.953	22.949	6E-05
PC3	0.99993	32.243	19.051	3E-07



## 8. pGLS regressions

**Table S8.** Results for Ordinary least squares (OLS) and phylogenetic generalised least squares (pGLS), using our primary phylogeny. First correlating the raw PC scores and centroid size (labyrinth size); second, correlating the raw PC scores and habitat; third correlating the residuals from the PC scores and centroid size with habitat.

### Primary phylogeny

Model	Type	AICc	intercept	p value	slope	p value	R2
PC1~centroid size	OLS	-61.319	-4.706	0.111	4.662	0.112	0.071
	pGLS	-78.643	-5.078	0.034	4.993	0.035	0.111
PC2~centroid size	OLS	-64.049	-0.531	0.845	0.527	0.845	-0.044
	pGLS	-76.99	8.307	0.002	-8.208	0.002	0.418
PC3~centroid size	OLS	-88.324	7.205	0.0002	-7.149	0.0002	0.451
	pGLS	-81.948	5.787	0.011	-5.751	0.011	0.315
PC1~pelagic	OLS	-78.162	-0.028	0.012	0.116	2.65e-05	0.559
	pGLS	-70.159	-0.035	0.370	0.083	0.042	0.385
PC2~pelagic	OLS	-64.329	0.0033	0.81	-0.016	0.59	0.013
	pGLS	-60.347	0.002	0.974	-0.060	0.222	-0.165
PC3~pelagic	OLS	-76.934	0.0035	0.734	-0.046	0.056	0.156
	pGLS	-68.628	-0.031	0.445	-0.050	0.226	-0.193
PC1.residuals~pelagic	OLS	-72.660	-0.019	0.110	0.089	0.00143	0.377
	pGLS	-59.892	0.018	0.703	0.089	0.0005	-0.061
PC2.residuals~pelagic	OLS	-64.505	0.004	0.769	-0.019	0.522	0.019
	pGLS	-49.136	-0.038	0.522	0.007	0.801	-0.861
PC3.residuals~pelagic	OLS	-88.418	0.001	0.895	-0.005	0.773	0.004
	pGLS	-68.295	0.003	0.940	0.008	0.847	-1.304

**Table S9.** Results for phylogenetic generalised least squares (pGLS), using the two alternative phylogenies. First correlating the raw PC scores and centroid size (labyrinth size); second, correlating the raw PC scores and habitat; third correlating the residuals from the PC scores and centroid size with habitat.

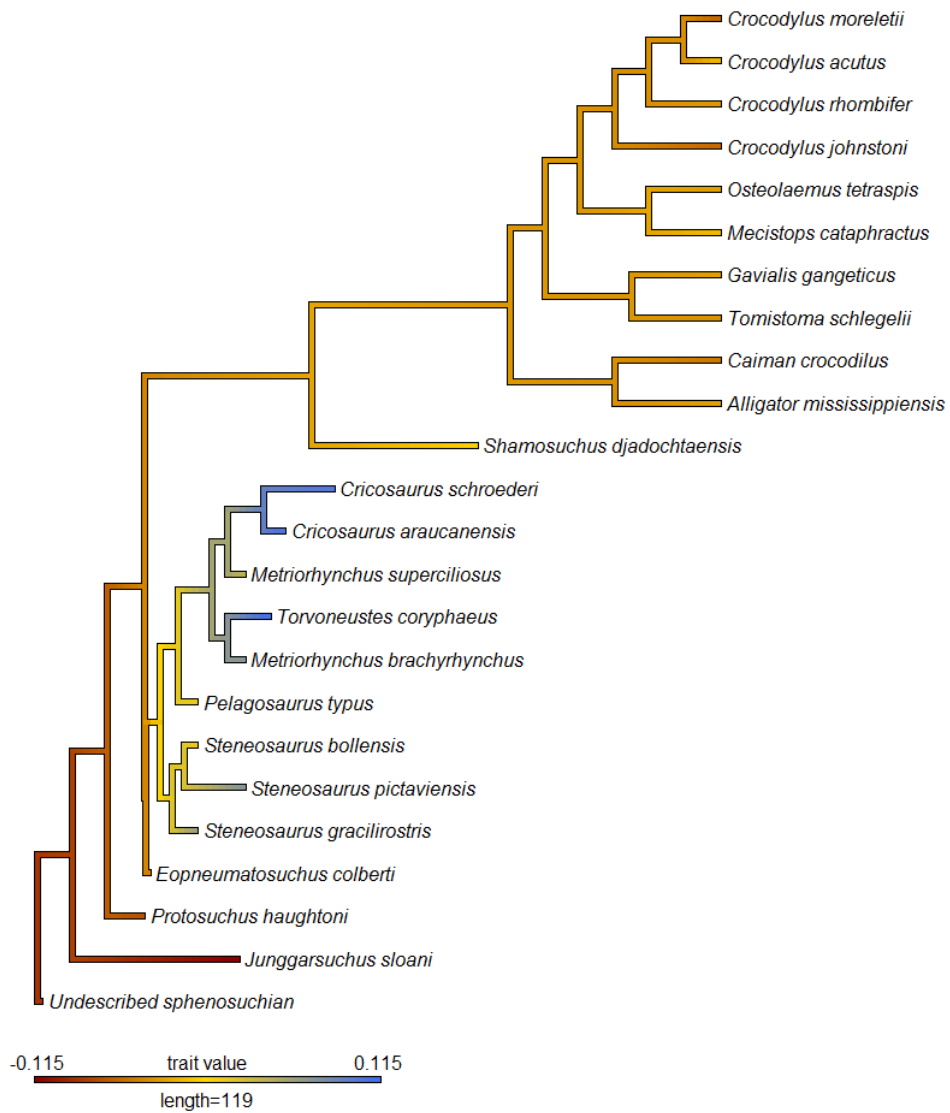
***Eopneumatosuchus colberti* placed as the sister taxon to Thalattosuchia + (*Shamosuchus* + *Crocodylia*)**

Model	Type	AICc	intercept	p value	slope	p value	R2
PC1~centroid size	pGLS	-76.721	-4.918	0.069	4.838	0.069	0.532
PC2~centroid size	pGLS	-77.183	7.885	0.005	-7.790	0.005	0.422
PC3~centroid size	pGLS	-83.430	5.577	0.019	-5.544	0.019	0.356
PC1~pelagic	pGLS	-70.159	-0.035	0.370	0.083	0.042	0.385
PC2~pelagic	pGLS	-60.347	0.002	0.974	-0.060	0.222	-0.165
PC3~pelagic	pGLS	-68.628	-0.031	0.445	-0.049	0.226	-0.192
PC1.residuals~pelagic	pGLS	-55.657	0.009	0.859	0.092	0.001	-0.266
PC2.residuals~pelagic	pGLS	-47.624	-0.032	0.594	0.013	0.666	-0.983
PC3.residuals~pelagic	pGLS	-70.355	0.003	0.939	0.011	0.544	-1.114

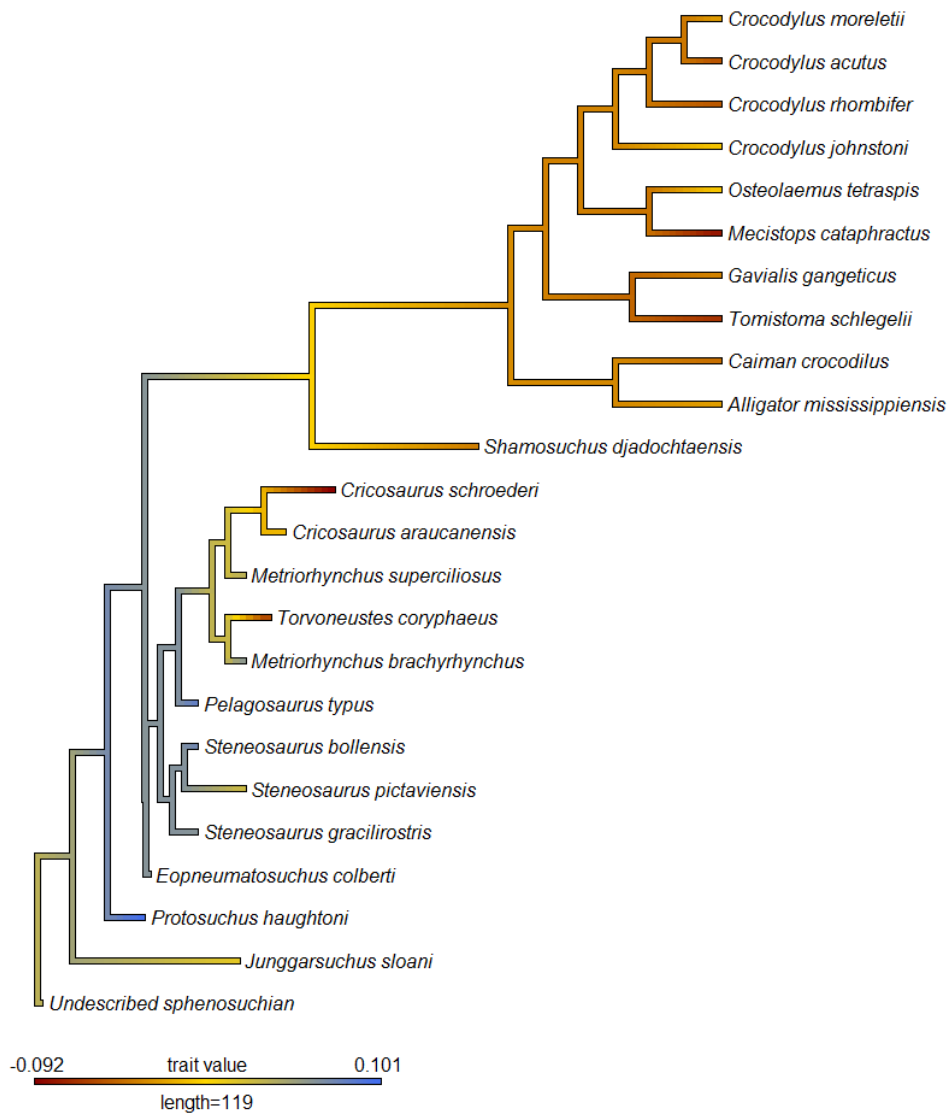
***Pelagosaurus typus* positioned as a teleosauroid**

Model	Type	AICc	intercept	p value	slope	p value	R2
PC1~centroid size	pGLS	-80.371	-5.328	0.023	5.230	0.024	0.598
PC2~centroid size	pGLS	-77.218	8.343	0.002	-8.242	0.002	0.423
PC3~centroid size	pGLS	-82.687	5.655	0.012	-5.627	0.012	0.336
PC1~pelagic	pGLS	-71.177	-0.042	0.279	0.078	0.041	0.410
PC2~pelagic	pGLS	-60.562	0.0002	0.997	-0.059	0.214	-0.154
PC3~pelagic	pGLS	-69.841	-0.038	0.345	-0.052	0.174	-0.134
PC1.residuals~pelagic	pGLS	-55.436	0.008	0.887	0.089	0.001	-0.278
PC2.residuals~pelagic	pGLS	-48.265	-0.039	0.536	0.007	0.809	-0.930
PC3.residuals~pelagic	pGLS	-70.095	0.004	0.916	0.005	0.782	-1.137

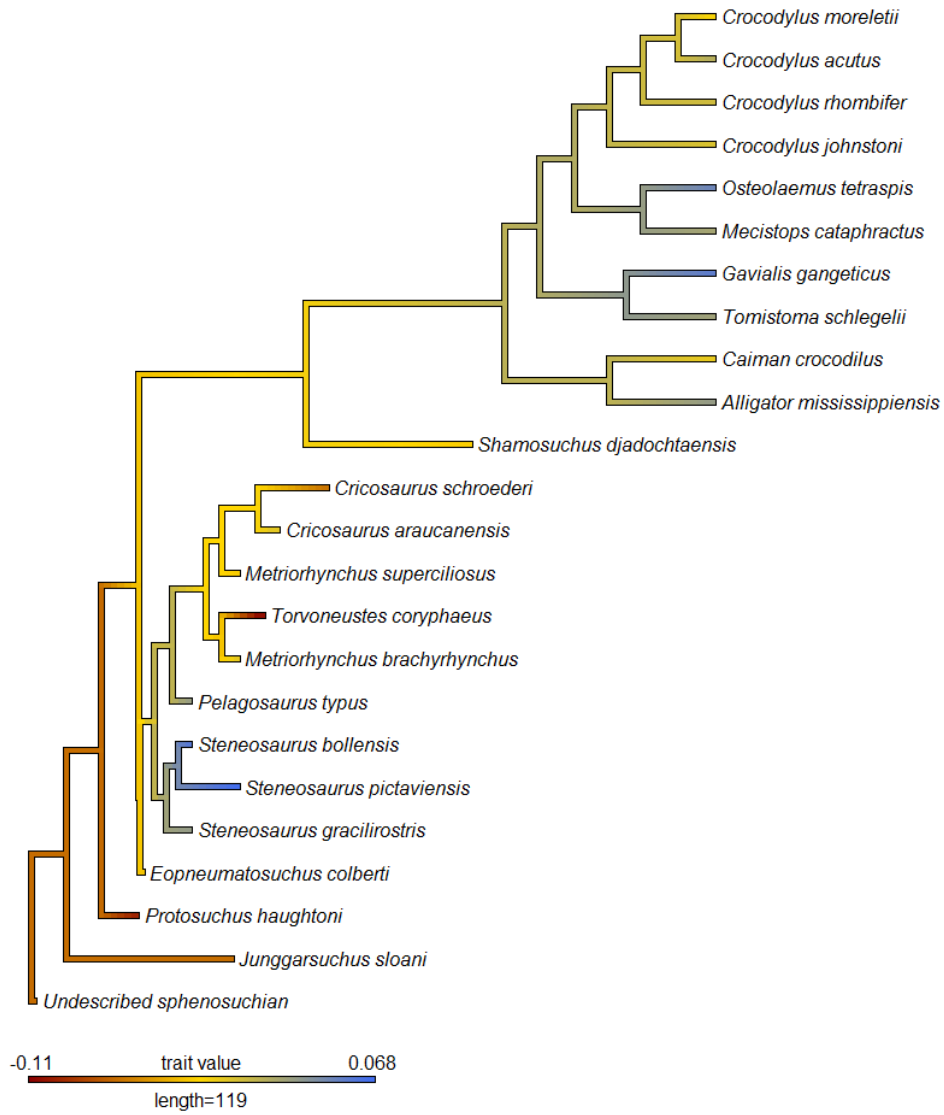
## 9. Labyrinth character optimization on phylogeny



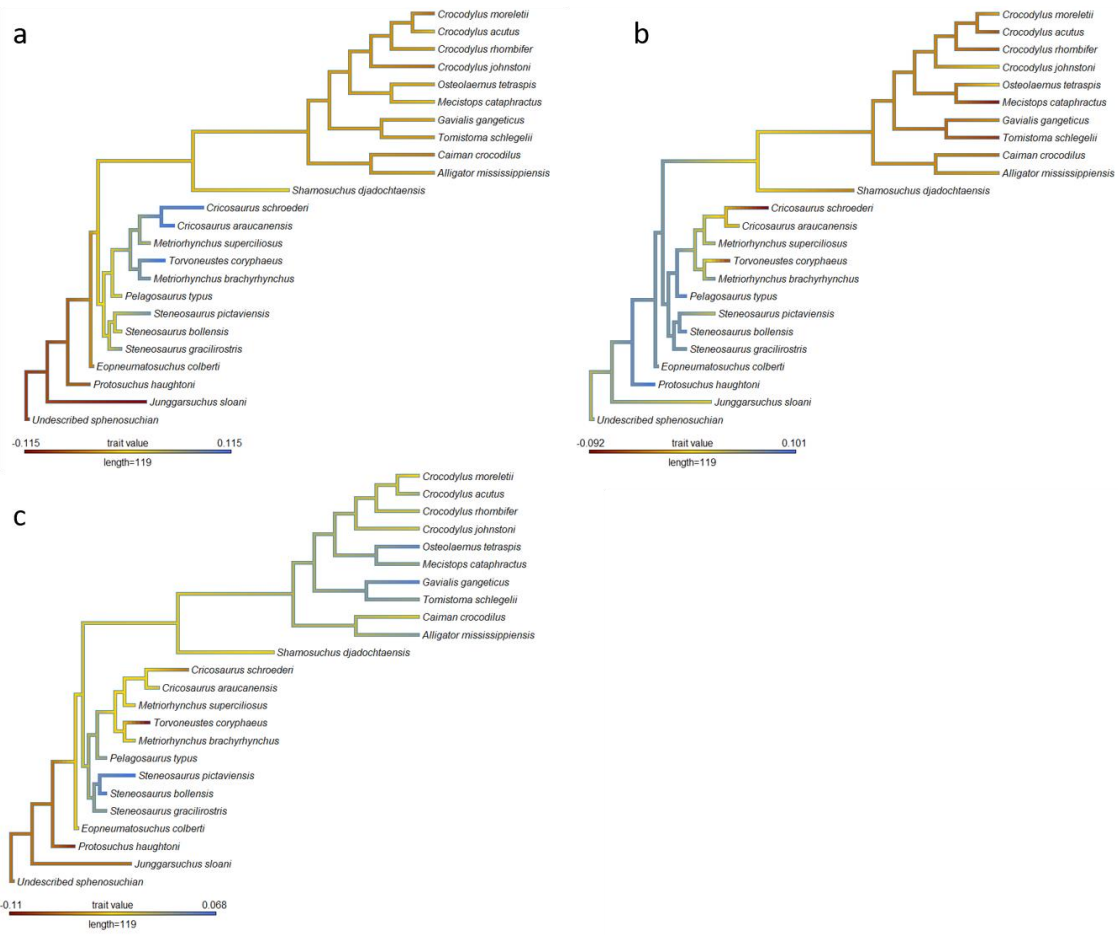
**Fig. S2.** PC1 scores optimized on our primary phylogeny to predict ancestral states for the major clades and assess evolutionary trends.



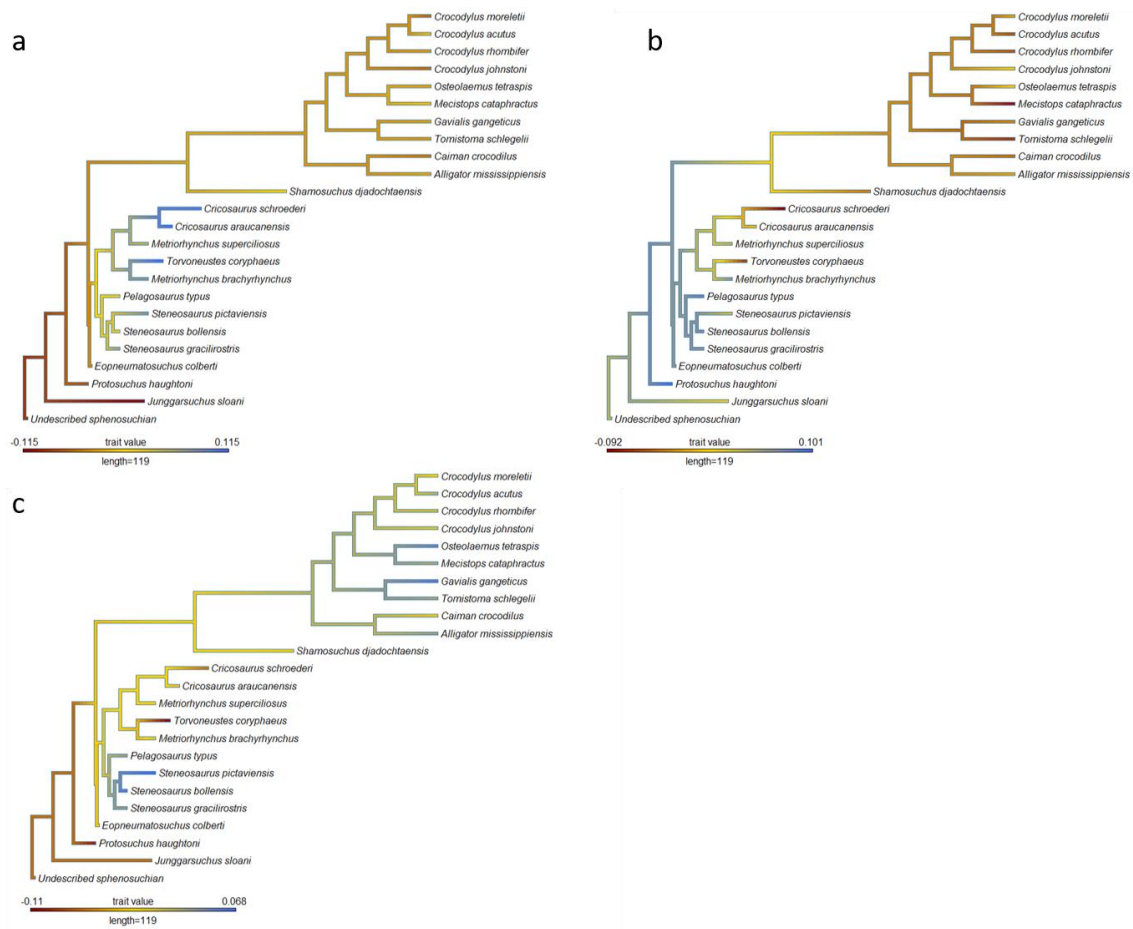
**Fig. S3.** PC2 scores optimized on our primary phylogeny to predict ancestral states for the major clades and assess evolutionary trends.



**Fig. S4.** PC3 scores optimized on our primary phylogeny to predict ancestral states for the major clades and assess evolutionary trends.



**Fig. S5.** PC scores of the first three principal components, optimized on the phylogeny with *Eopneumatosuchus colberti* placed as the sister taxon to Thalattosuchia + (*Shamosuchus* + *Crocodylia*), to predict ancestral states for the major clades and assess evolutionary trends. a, PC1; b, PC2; c, PC3.



**Fig. S6.** PC3 scores optimized on the phylogeny with *Pelagosaurus typus* positioned as a teleosauroid, to predict ancestral states for the major clades and assess evolutionary trends. a, PC1; b, PC2; c, PC3.

## 10. Evolutionary model fitting

**Table S10.** Five standard models fitted on the first three principal component axes showing the AIC scores. The models are: BM (Brownian motion), OU (Ornstein-Uhlenbeck), EB (Early-burst), trend (Brownian motion with trend) and lambda (Pagel).

### Primary phylogeny

	BM	OU	EB	trend	lambda
PC1	-74.485	-71.856	-79.996	-63.232	-71.856
PC2	-69.521	-68.680	-69.084	-64.574	-67.822
PC3	-74.591	-73.483	-78.519	-64.725	-71.962

### Primary phylogeny (fossil specimens only)

	BM	OU	EB	trend	lambda
PC1	-36.607	-33.298	-33.297	-39.191	-33.305
PC2	-35.977	-34.494	-32.667	-38.418	-33.666
PC3	-42.606	-40.841	-39.524	-36.884	-39.297

*Eopneumatosuchus colberti* placed as the sister taxon to Thalattosuchia + (*Shamosuchus* + Crocodylia)

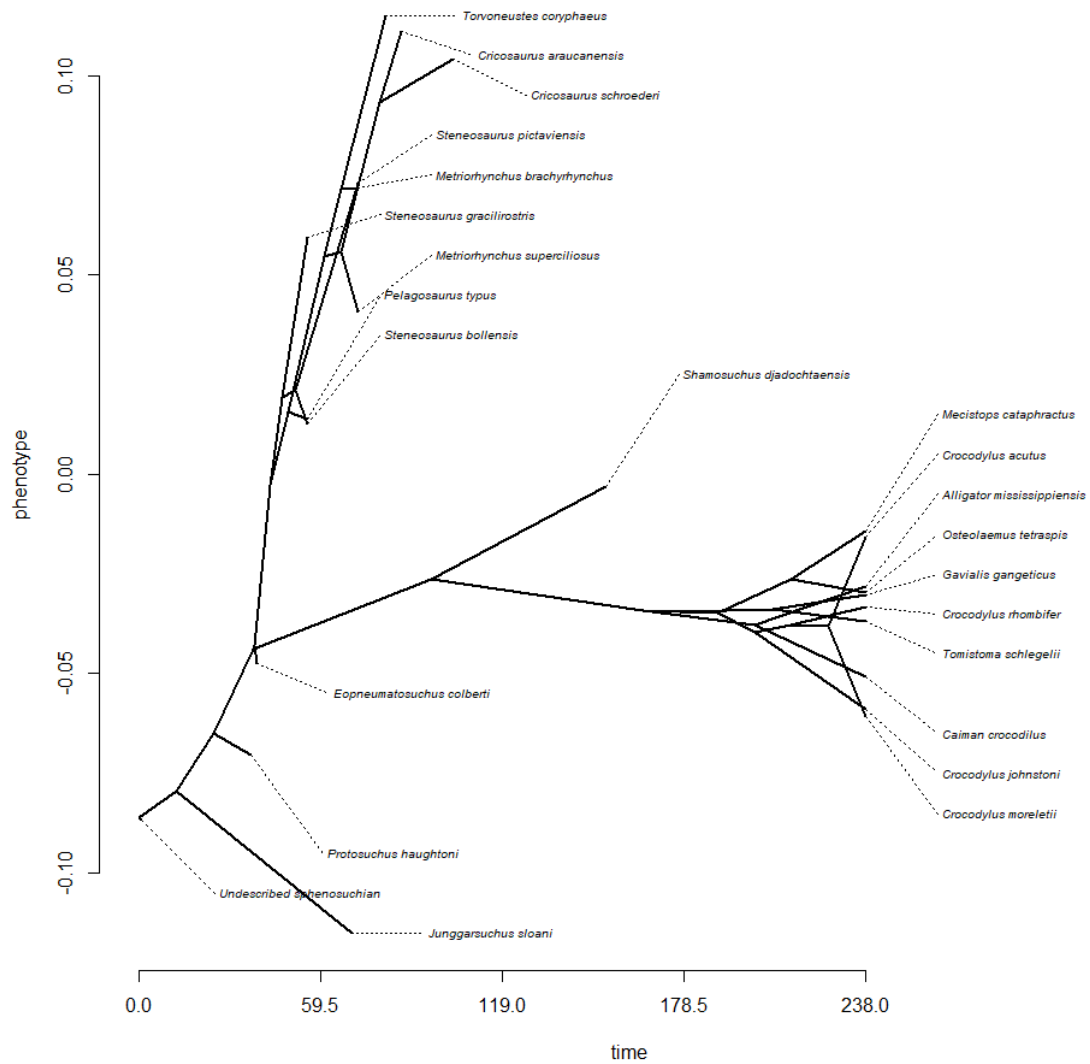
	BM	OU	EB	trend	lambda
PC1	-72.911	-70.283	-78.911	-61.215	-70.283
PC2	-72.719	-71.107	-71.698	-67.207	-70.091
PC3	-78.059	-76.365	-80.978	-67.787	-75.429

*Pelagosaurus typus* positioned as a teleosauroid

	BM	OU	EB	trend	lambda
PC1	-76.211	-73.582	-81.262	-65.216	-73.582
PC2	-69.629	-68.789	-69.289	-64.601	-67.825
PC3	-75.373	-74.759	-79.190	-65.683	-72.745

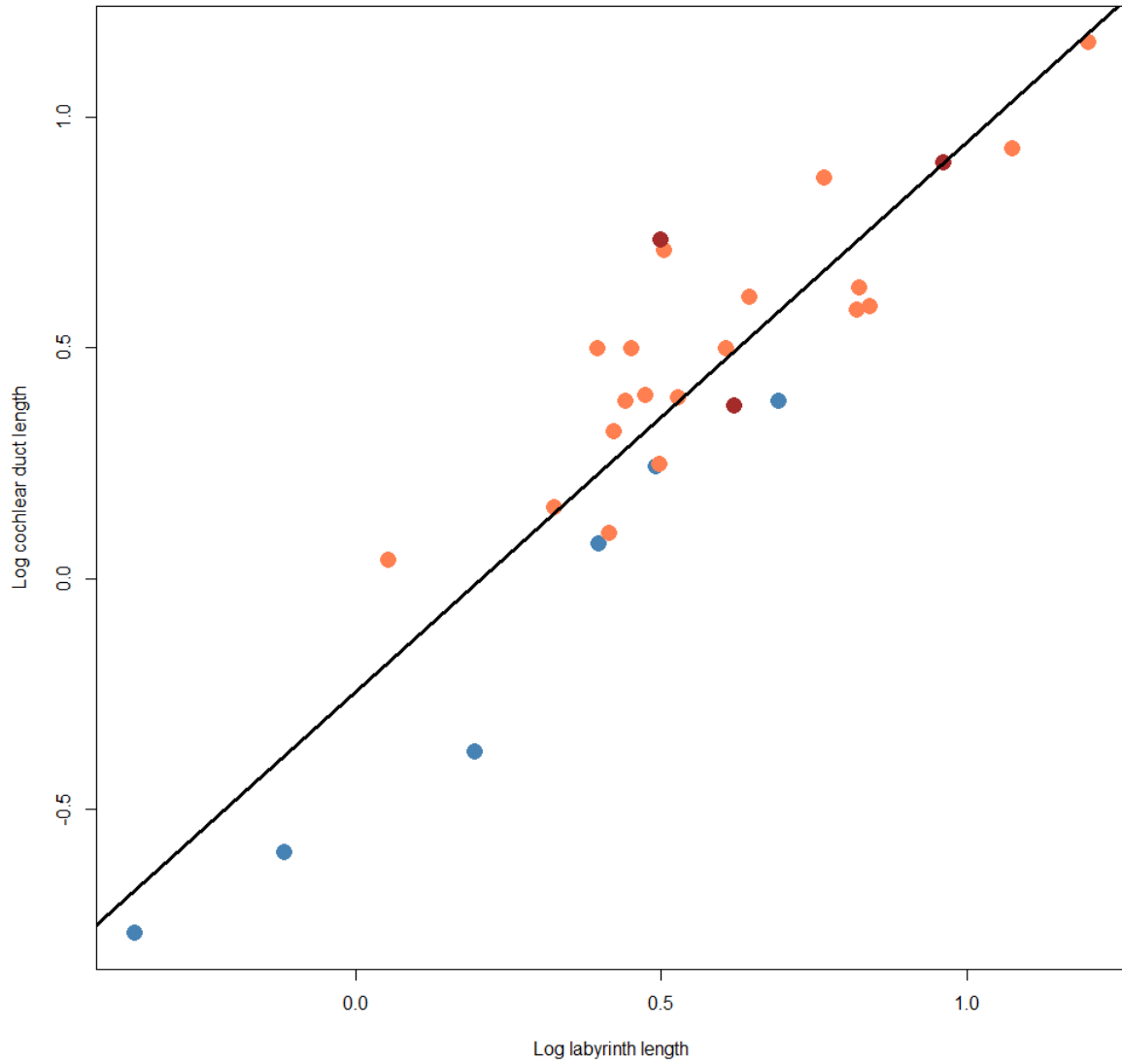


## 11. Evolutionary rates



**Fig. S7.** The primary phylogeny is plotted such that the x-axis is scaled to time and the y-axis to PC1 score.

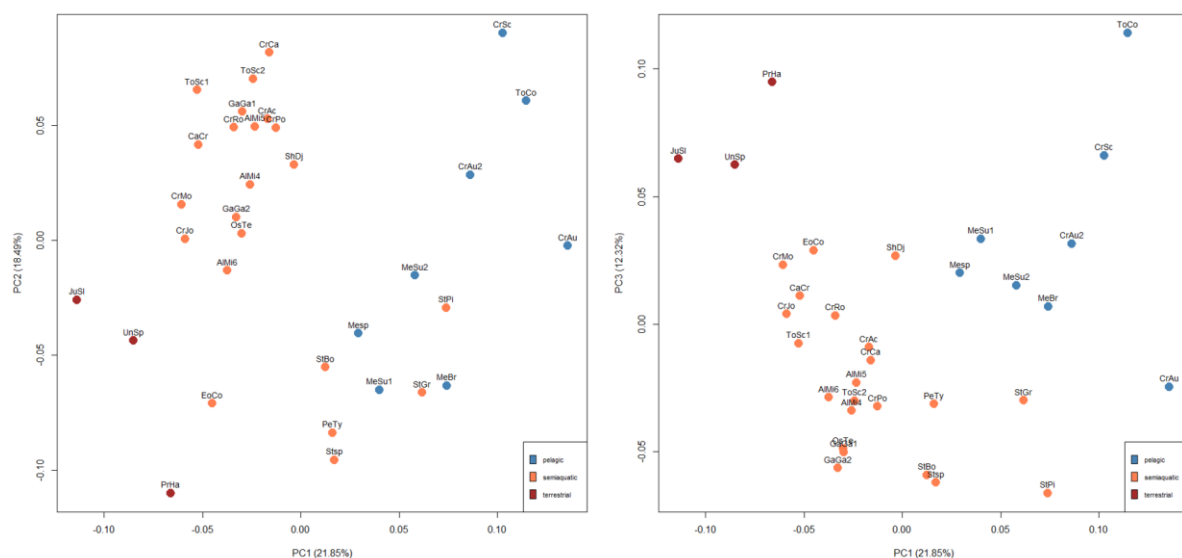
## 12. Cochlear measurements



**Fig. S8.** Logarithmic plot of labyrinth length and cochlear duct length. Blue, pelagic; orange, semiaquatic; red, terrestrial.

### 13. Inclusion of *Crocodylus porosus* juvenile

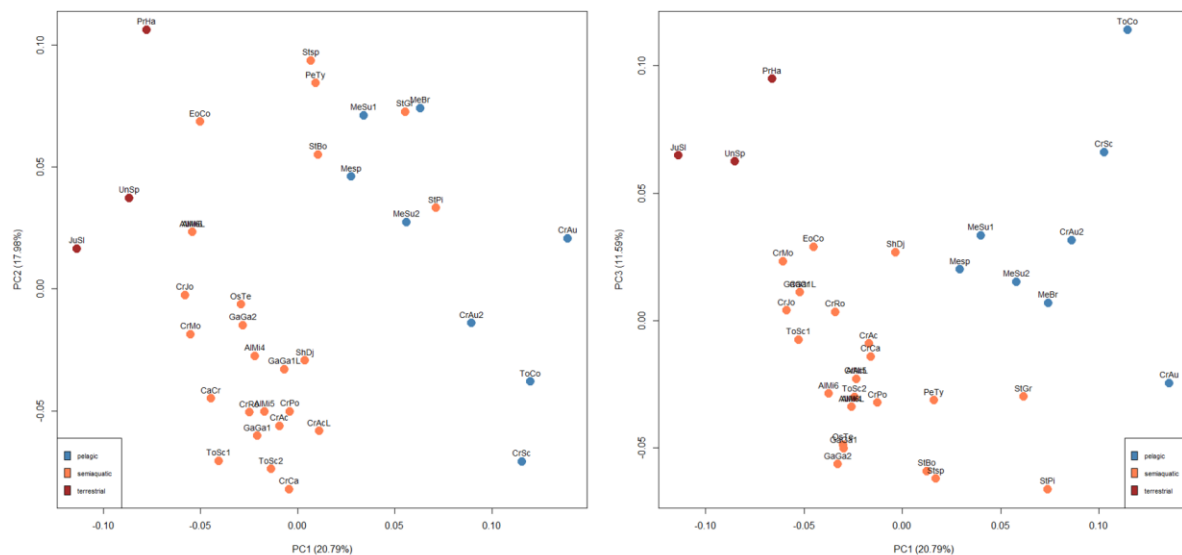
The extant crocodylian *Crocodylus porosus* often ventures further offshore than other extant species. We do not include *C. porosus* in our dataset as the scan we acquired is of a juvenile, and we are limiting our analysis to adults. However, to demonstrate that *C. porosus* has an inner ear that is similar in shape to other extant crocodylians, and distinct in shape from pelagic extinct thalattosuchians, we provide an auxiliary analysis in which this juvenile specimen is included in our PCA morphospace below (analysis conducted following the same protocol as our primary PCA analysis, described above and in the main text). This demonstrates that the exclusion of *C. porosus* should not greatly affect our results, as its ear is exceedingly similar to the many other extant crocodylians in our dataset. We also acknowledge that extant species other than *C. porosus* are frequently encountered in brackish or saline environments, including one of the species in our analysis, *C. acutus*, and that swimming behavior in extant crocodylians appears to be the same whether an individual is in fresh or salt water. Hence, we wouldn't expect substantial differences in inner ear morphology between coastal and fluviolacustrine extant forms.



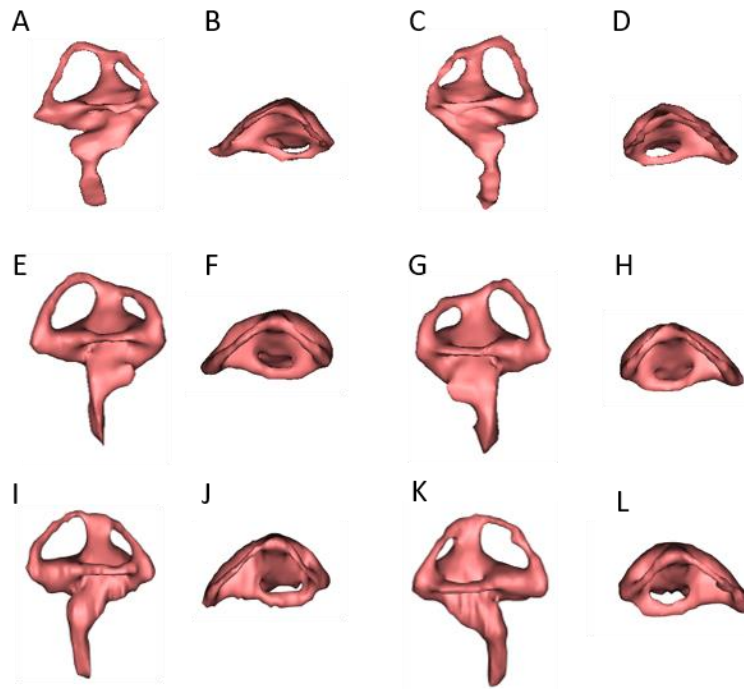
**Fig. S9.** PCA including the juvenile *Crocodylus porosus* (OUVC:10899; CrPo).

## 14. Left-Right Ear Asymmetry

To our knowledge, the only study testing whether left and right inner ear labyrinths of an archosaur are consistent in shape is a recent study of wild turkeys (24). The analyses in our paper use right labyrinths or mirrored left labyrinths. We here test whether mirrored left labyrinths resemble the shape of the right labyrinths in individuals of three key extant species, one from each major crocodylian subgroup. Importantly, we find that, for each individual, the mirrored left and actual right labyrinths fall out very close to each other in PCA morphospace (Fig. S10, analysis conducted following the same protocol as our primary PCA analysis, described above and in the main text), demonstrating that any asymmetry between left and right sides is minimal compared to the much greater amount of variation between species and between habitat groups. Thus, the inclusion of both right and mirrored left labyrinths in our dataset should not provide any serious bias. We visually show the actual left and right labyrinths of the three extant individuals in Fig. S11, which further demonstrates how similar in shape they are.



**Fig. S10.** PCA including the left mirrored labyrinths for *Alligator mississippiensis* (OUVC 9761; AIMi6L), *Crocodylus acutus* (FMNH 59071; CrAcL) and *Gavialis gangeticus* (TMM M-5490; GaGa1L).



**Fig. S11.** Bony labyrinths of *Alligator mississippiensis* (OUVC 9761), *Crocodylus acutus* (FMNH 59071) and *Gavialis gangeticus* (TMM M-5490). A, E, I left lateral view; B, F, J left dorsal view; C, G, K right lateral view; D, H, L right dorsal view.

## 15. Supplementary References

1. J. M. Clark, X. Xu, C. A. Forster, Y. Wang, A Middle Jurassic 'sphenosuchian' from China and the origin of the crocodylian skull. *Nature* **430**, 1021–1024 (2004).
2. A. W. Crompton, K. K. Smith, "A new genus and species of crocodylian from the Kayenta Formation (Late Triassic?) of Northern Arizona" in *Aspects of Vertebrate History*, L. L. Jacobs, Eds. (Museum of Northern Arizona Press, Flagstaff, 1980) pp. 193–217.
3. E. H. Colbert, C. C. Mook, The Ancestral crocodylian *Protosuchus*. *Bull. Am. Mus. Nat. Hist.* **97**, 143–182 (1951).
4. F. Westphal, Die Krokodilier des Deutschen und Englischen oberen Lias. *Palaeontographica A* **116**, 23–118 (1962).
5. C. W. Andrews, Descriptive Catalogue of the Marine Reptiles of the Oxford Clay: Part II (British Museum (Natural History), London, 1913).
6. Y. Herrera, M. Fernández, J. A. Varela, Morfología del miembro anterior de *Geosaurus araucanensis* Gasparini y Dellapé, 1976 (Crocodyliformes: Thalattosuchia). *Ameghiniana* **46**, 657–667 (2009).
7. Y. Herrera, M. S. Fernández, Z. Gasparini, Postcranial skeleton of *Cricosaurus araucanensis* (Crocodyliformes: Thalattosuchia): morphology and palaeobiological insights. *Alcheringa* **37**, 1–14 (2013).
8. H. Karl, E. Gröning, C. Brauckmann, N. Knötschke, Revision of the genus *Enaliosuchus* Koken, 1883 (Archosauromorpha: Metriorhynchidae) from the

- Early Cretaceous of northwestern Germany. *Studia Geológica Salmanticensia* **42**, 49–59 (2006).
9. M. T. Young, M.B. Andrade, S. Etches, B. L. Beatty, A new metriorhynchid crocodylomorph from the Lower Kimmeridge Clay Formation (Late Jurassic) of England, with implications for the evolution of dermatocranium ornamentation in Geosaurini. *Zool. J. Linn. Soc.* **169**, 820–848 (2013).
  10. D. Pol, A. H. Turner, M. A. Norell, Morphology of the Late Cretaceous Crocodylomorph *Shamosuchus djadochtaensis* and a Discussion of Neosuchian Phylogeny as Related to the Origin of Eusuchia. *Bull. Am. Mus. Nat. Hist.* **324**, 1–103 (2009).
  11. G. Grigg, D. Kirshner, *Biology and Evolution of Crocodylians* (CSIRO and Cornell University Press, 2015).
  12. M. T. Young, *et al.*, Convergent evolution and possible constraint in the posterodorsal retraction of the external nares in pelagic crocodylomorphs. *Zool. J. Linn. Soc.* (In press).
  13. J. Ristevski, M. T. Young, M. B. Andrade, A. K. Hastings, A new species of *Anteophthalmosuchus* (Crocodylomorpha, Goniopholididae) from the Lower Cretaceous of the Isle of Wight, United Kingdom, and a review of the genus. *Cretac. Res.* **84**, 340–383 (2018).
  14. A. Ósi, M. T. Young, A. Galácz, M. Rabi, A new large-bodied thalattosuchian crocodyliform from the Lower Jurassic (Toarcian) of Hungary, with further evidence of the mosaic acquisition of marine adaptations in Metriorhynchoidea. *PeerJ* **6**, e4668 (2018).

15. D. Foffa, M. M. Johnson, M. T. Young, L. Steel, S. L. Brusatte, A revision of the deep-water teleosauroid crocodylomorph *Teleosaurus megarhinus* Hulke, 1871 from the Kimmeridge Clay Formation (Late Jurassic) of England, UK. *PeerJ* **7**, e6646 (2019).
16. M. M. Johnson, M. T. Young, S. L. Brusatte, Re-description of two contemporaneous mesorostrine teleosauroids (Crocodylomorpha, Thalattosuchia) from the Bathonian of England, and insights into the early evolution of Machimosaurini. *Zoo. J. Linn. Soc.* zlz037 (2019).
17. S. Sachs, M. T. Young, P. Abel, H. Mallison, A new species of *Cricosaurus* (Thalattosuchia, Metriorhynchidae) from the Upper Jurassic of southern Germany. *Acta Palaeontologica Polonica* **64**, 345–356 (2019).
18. E. W. Wilber, What's in an Outgroup? The Impact of Outgroup Choice on the Phylogenetic Position of Thalattosuchia (Crocodylomorpha) and the Origin of Crocodyliformes. *Syst. Biol.* **64**, 624–637 (2015a).
19. E. W. Wilber, A new metriorhynchoid (Crocodylomorpha, Thalattosuchia) from the Middle Jurassic of Oregon and the evolutionary timing of marine adaptations in thalattosuchian crocodylomorphs. *J. Vert. Paleo.* 902846 (2015b).
20. E. Buffetaut, Position systématique et phylogénétique du genre *Pelagosaurus* Bronn, 1841 (Crocodylia, Mesosuchia), du Toarcien d'Europe. *Geobios* **13**, 783–786 (1980).
21. J. R. Oaks, A time-calibrated species tree of crocodylia reveals a recent radiation of the true crocodiles. *Evolution* **65**, 3285–3297 (2011).



22. J. Harshman, C. J. Huddleston, J. P. Bollback, T. J. Parsons, M. J. Braun True and false gharials: a nuclear gene phylogeny of crocodylia. *Syst Biol* **52**, 386–402 (2003).
23. M. S. Y. Lee, A. M. Yates Tip-dating and homoplasy: reconciling the shallow molecular divergences of modern gharials with their long fossil record. *Proc. R. Soc. B* **285**, 20181071 (2018).
24. D. G. Cerio, L. M. Witmer, Intraspecific variation and symmetry of the inner-ear labyrinth in a population of wild turkeys: implications for paleontological reconstructions. *PeerJ* **7**, e7355 (2019).