In the format provided by the authors and unedited.

Glucose-responsive insulin patch for the regulation of blood glucose in mice and minipigs

Jicheng Yu^{1,2}, Jinqiang Wang^{1,3,4}, Yuqi Zhang^{1,2}, Guojun Chen^{1,3,4}, Weiwei Mao², Yanqi Ye^{1,2}, Anna R. Kahkoska⁵, John B. Buse⁵, Robert Langer^(0,6,7,8,9,10) and Zhen Gu^(0,1,3,4,11,12*)

¹Joint Department of Biomedical Engineering, University of North Carolina at Chapel Hill and North Carolina State University, Raleigh, NC, USA. ²Zenomics Inc., Los Angeles, CA, USA. ³Department of Bioengineering, University of California, Los Angeles, Los Angeles, CA, USA. ⁴California NanoSystems Institute, University of California, Los Angeles, Los Angeles, CA, USA. ⁵Department of Medicine, University of North Carolina School of Medicine, Chapel Hill, NC, USA. ⁶Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge, MA, USA. ⁷David H. Koch Institute for Integrative Cancer Research, Massachusetts Institute of Technology, Cambridge, MA, USA. ⁸Department of Anesthesiology, Boston Children's Hospital, Boston, MA, USA. ⁹Division of Health Science and Technology, Cambridge, MA, USA. ¹¹Jonsson Comprehensive Cancer Center, University of California, Los Angeles, Los Angeles, Los Angeles, CA, USA. ¹²Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, CA, USA. ¹²Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, Los Angeles, CA, USA. ¹²Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, Los Angeles, CA, USA. ¹²Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, Los Angeles, Los Angeles, Los Angeles, CA, USA. ¹²Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, Los Angeles, CA, USA. ¹²Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, CA, USA. ¹⁴Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, CA, USA. ¹⁵Center for Minimally Invasive Therapeutics, University of California, Los Angeles, Los Angeles, CA, USA. ¹⁵Center for Minimally Invasive Therapeutics, University of California, Los Angeles, CA, USA. ¹⁵Center for Minimally Invasive Therapeutics, Figure S1. Mass spectrum analysis of the native insulin and insulin extracted from GR-MN.

Figure S2. Photographs of GR-MN with a 1:4 ratio of DMAEA to 3APBA pre-incubated and post-incubated in PBS with varying glucose concentrations for 1 h.

Figure S3. *In vitro* accumulated insulin release from the polymeric matrix with a ratio of DMAEA to 3APBA at a 1:1 ratio or a 1:20 ratio across different glucose concentrations at 37 °C.

Figure S4. Insulin release from the polymeric matrix with a 1:4 ratio of DMAEA to 3APBA under different pH values at 37 °C.

Figure S5. *In vitro* accumulated insulin release from the polymeric matrix with a ratio of DMAEA to 3APBA at a 1:3 ratio or a 1:5 ratio across different glucose concentrations at 37 °C.

Figure S6. Insulin release of the polymeric matrix with a 1:4 ratio of DMAEA to 3APBA as a function of glucose concentrations.

Figure S7. (a) PGLs in STZ-induced diabetic mice after treatment with GR-MN with a ratio of DMAEA to 3APBA at 1:3, 1:4 or 1:5. (b) *In vivo* intraperitoneal glucose tolerance test in diabetic mice at 4 h post-administration of GR-MN with a ratio of DMAEA to 3APBA at 1:3, 1:4 or 1:5 in comparison with the healthy control mice.

Figure S8. (a) PGLs in the STZ-induced diabetic mice after treatment with GR-MN with different insulin doses. (b) *In vivo* intraperitoneal glucose tolerance test in diabetic mice at 4 h post-administration of GR-MN with different insulin doses in comparison with healthy control mice.

Figure S9. Hematoxylin and eosin (H&E)-stained section of mouse skin pre- and post-treatment of GR-MN.

Figure S10. The daily glucose pattern of the healthy Göttingen minipig recorded by calibrated CGMS.

Figure S11. (a) PGLs in STZ-induced diabetic mice during a 48-hour consecutive administration of GR-MN. (b) PGLs in three STZ-induced diabetic minipigs during a 48-hour consecutive administration of GR-MN.

Figure S12. Porcine C-peptide levels in diabetic minipigs after an intravenous glucose challenge at 4 h post-administration of GR-MN.

Figure S13. The *in vivo* glucose-responsive insulin release promoted by intravenous glucose challenge at 4 h post-administration of GR-MN for multiple rounds in three individual experiments.

Figure S14. (a) Representative SEM image of the MN array after removal from the skin. (b) H&E-stained section indicates that insignificant fragments were found in the skin tissue after the removal of GR-MN.

Figure S15. H&E-stained section of minipig skin pre- and post-treatment of GR-MN.

Table S1. Leachable monomers from the GR-MN patch with 20×20 array of microneedles.

Table S2. Pharmacokinetic parameters of insulin after administration of GR-MN, CR-MN, and subcutaneous injection on diabetic mice.

Table S3. The plasma glucose levels of diabetic pigs.

Table S4. Gradient HPLC method for NVP, 3APBA and EGDMA.

Table S5. Gradient HPLC method for DMAEA.



Supplementary Figure 1. Mass spectrum analysis of the native insulin and insulin extracted from GR-MN.



Supplementary Figure 2. Photographs of GR-MN with a 1:4 ratio of DMAEA to 3APBA preincubated and post-incubated in PBS with varying glucose concentrations for 1 h. Scale bar: 100 μ m.



Supplementary Figure 3. *In vitro* accumulated insulin release from the polymeric matrix with a ratio of DMAEA to 3APBA at a 1:1 ratio (left) or a 1:20 ratio (right) across different glucose concentrations at 37 °C. Data are presented as mean \pm s.d. (*n*=3).



Supplementary Figure 4. Insulin release from the polymeric matrix with a 1:4 ratio of DMAEA to 3APBA under different pH values at 37 °C. Data are presented as mean \pm s.d. (*n*=3).



Supplementary Figure 5. *In vitro* accumulated insulin release from the polymeric matrix with a ratio of DMAEA to 3APBA at a 1:3 ratio (left) or a 1:5 ratio (right) across different glucose concentrations at 37 °C. Data are presented as mean \pm s.d. (*n*=3).



Supplementary Figure 6. Insulin release of the polymeric matrix with a 1:4 ratio of DMAEA to 3APBA as a function of glucose concentrations. Data are presented as mean \pm s.d. (*n*=3).



Supplementary Figure 7. (a) PGLs in STZ-induced diabetic mice after treatment with GR-MN with a ratio of DMAEA to 3APBA at 1:3, 1:4 or 1:5. Insulin dose: 0.5 mg. Data are presented as mean \pm s.d. (*n*=5). (b) *In vivo* intraperitoneal glucose tolerance test in diabetic mice at 4 h post-administration of GR-MN with a ratio of DMAEA to 3APBA at 1:3, 1:4 or 1:5 in comparison with the healthy control mice. Glucose dose: 1.5 g/kg. Data are presented as mean \pm s.d. (*n*=5).



Supplementary Figure 8. (a) PGLs in the STZ-induced diabetic mice after treatment with GR-MN with different insulin doses. Data are presented as mean \pm s.d. (*n*=5). (b) *In vivo* intraperitoneal glucose tolerance test in diabetic mice at 4 h post-administration of GR-MN with different insulin doses in comparison with healthy control mice. Glucose dose: 1.5 g/kg. Data are presented as mean \pm s.d. (*n*=5).



Supplementary Figure 9. Hematoxylin and eosin (H&E)-stained sections of mouse skin preand post-treatment of GR-MN. Scale bar: 400 µm.



Supplementary Figure 10. The daily glucose pattern of the healthy Göttingen minipig recorded by calibrated CGMS.



Supplementary Figure 11. (a) PGLs in STZ-induced diabetic mice during a 48-hour consecutive administration of GR-MN. Insulin dose: 0.5 mg. The blue arrows indicate the time points of MN administration. Data are presented as mean \pm s.d. (n=5). (b) PGLs in three STZ-induced diabetic minipigs during a 48-hour consecutive administration of GR-MN. Insulin dose: 7 mg. The blue arrows indicate the time points of MN administration, and the pink arrows indicate the time points of feeding.



Supplementary Figure 12. Porcine C-peptide levels in diabetic minipigs after an intravenous glucose challenge at 4 h post-administration of GR-MN. Data are presented as mean \pm s.d. (*n*=3).



Supplementary Figure 13. The *in vivo* glucose-responsive insulin release promoted by intravenous glucose challenge at 4 h post-administration of GR-MN for multiple rounds in three individual experiments. Glucose dose for each round: 0.7 g/kg. The red arrows indicate the time points of glucose administration.



Supplementary Figure 14. (a) Representative SEM image of the MN array after removal from the skin. Scale bar: 500 μ m. (b) H&E-stained section indicates that insignificant fragments were found in the skin tissue after the removal of GR-MN. Scale bar: 200 μ m.



Supplementary Figure 15. H&E-stained sections of minipig skin pre- and post-treatment of GR-MN. Scale bar: 400 µm.

Monomer	Leachable monomer amount (mg)	% per patch
NVP	0.082 ± 0.020	0.24 ± 0.05
DMAEA	<0.001	<0.003
3APBA	0.055 ± 0.010	0.16 ± 0.04
EGDMA	<0.020	<0.06

Supplementary Table 1. Leachable monomers from the GR-MN patch with 20×20 array of microneedles.

Supplementary Table 2. Pharmacokinetic parameters of insulin after administration of GR-MN (insulin dose: 0.5 mg), CR-MN (insulin dose: 0.5 mg), and subcutaneous injection (insulin dose: 0.05 mg) in diabetic mice. Data are presented as mean \pm s.d. (*n*=5).

Group	AUC _{0→12} (μIU h ⁻¹ mL ⁻¹)	RBA (%)
GR-MN	3051.6 ± 429.9	11.6 ± 1.9
CR-MN	2966.2 ± 489.0	11.9 ± 1.6
SC insulin	2554.4 ± 376.1	100

 $AUC_{0\rightarrow 12}$, area under the plasma insulin concentration over time; RBA, relative bioavailability compared with subcutaneous injection.

PGL (mg/dL)	GR-MN		CR-MN			
Time (h)	1	2	3	1	2	3
3	35	37	38	36	72	35
6	38	121	37	213	154	39
9	49	51	48	296	241	172
12	35	37	36	271	299	181
24	155	307	367	253	252	364

Supplementary Table 3. The plasma glucose levels (PGLs) of diabetic pigs.

Time (min)	Mobile phase A (%)	Mobile phase B (%)	Flow (mL/min)
0.00	95.0	5.0	0.3
4.50	70.0	30.0	0.3
5.50	70.0	30.0	0.3
8.00	50.0	50.0	0.3
8.10	5.0	95.0	0.3
9.00	5.0	95.0	0.3
9.10	95.0	5.0	0.3

Supplementary Table 4. Gradient HPLC method for NVP, 3APBA and EGDMA.

A: water with 0.1% formic acid (v/v)

B: acetonitrile with 0.1% formic acid (v/v)

Injection volume was set to 10 $\mu L.$

Time (min)	Mobile phase A (%)	Mobile phase B (%)	Mobile phase C (%)	Flow (mL/min)
0.00	0.0	20.0	80.0	1.0
2.00	0.0	20.0	80.0	1.0
12.00	25.0	20.0	55.0	1.0
15.00	25.0	20.0	55.0	1.0
15.10	0.0	20.0	80.0	1.0
22.00	0.0	20.0	80.0	1.0

Supplementary Table 5. Gradient HPLC method for DMAEA.

A: acetonitrile

B: 50 mM perchloric acid

C: water

Injection volume was set to 10 μ L.