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Supplemental Information

The Selectivity Filter Is Involved in the U-Type Inactivation Process of Kv2.1 and Kv3.1 Channels

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Supplemental Information

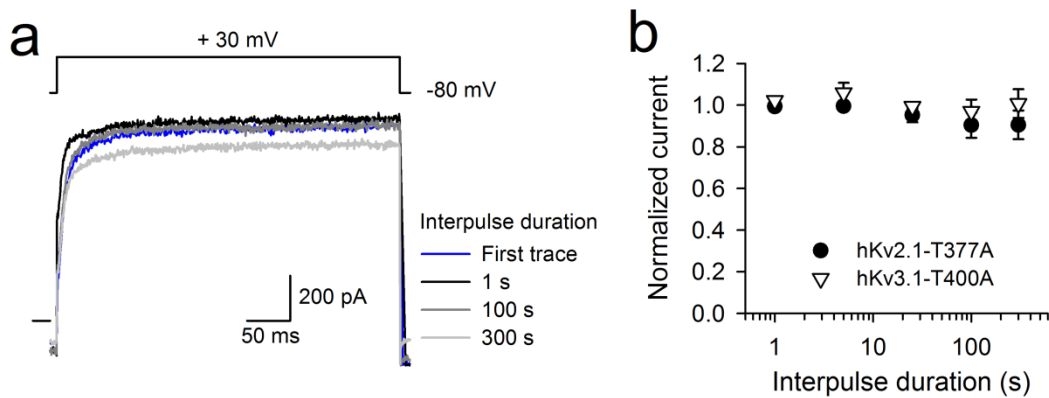


Figure S1: Prolonged membrane hyperpolarization does not reveal closed-state inactivation in hKv2.1-T377A and hKv3.1-T400A. **a.** Representative current recording of hKv2.1-T377A obtained by applying the pulse protocol shown on top. Inter-pulse duration was increased from 1 s (black trace) to 100 s (dark gray trace) and 300 s (light gray trace) at -80 mV. The current elicited by the first pulse is depicted in blue. Aside of a very small decrease in current amplitude after 300 s at -80 mV, prolonged hyperpolarizations did not decrease hKv2.1-T377A currents. **b.** Normalized current amplitude at +30 mV for Kv2.1-T377A and Kv3.1-T400A as a function of inter-pulse duration. There is no significant decrease in current amplitude with prolonged hyperpolarization at -80 mV. Data is shown as mean values \pm SEM from 4 and 6 individual cells for hKv2.-T377A and hKv3.1-T400A, respectively.

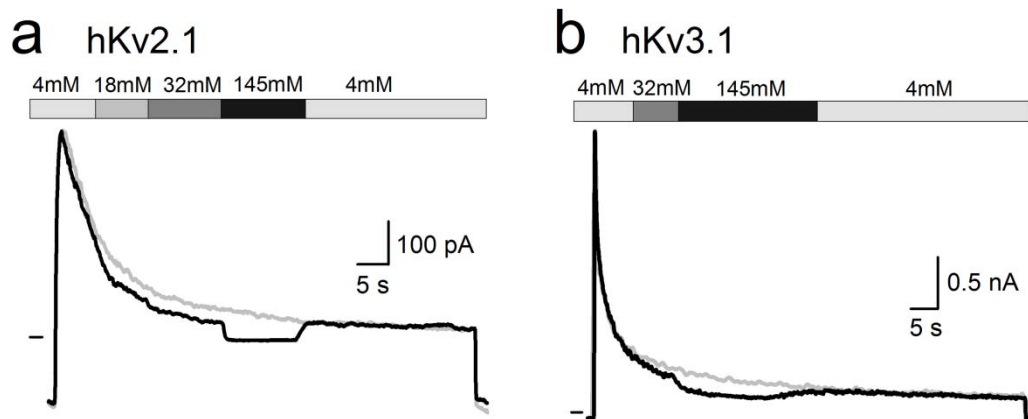


Figure S2: Increased external K⁺ reduces current amplitude in hKv2.1 and hKv3.1.

a. Representative hKv2.1 current recording showing the effect of increasing the external K⁺ concentration on current amplitude. During a prolonged depolarization to +20 mV, starting from -80 mV holding potential, the external K⁺ concentration was changed, using a pressurized perfusion system, from 4 mM to 145 mM as indicated on top. The current amplitude decreased with increasing K⁺ concentrations, as is predicted from a reduced effective driving force ($V_{\text{effective}} = V_{\text{applied}} - V_{\text{reversal}}$). The gray trace is the current elicited by the same pulse protocol in 4 mM [K⁺]. **b.** A representative hKv3.1 recording during prolonged depolarization to +40 mV and increasing the extracellular K⁺ concentration. Gray recording is again the reference current obtained in 4 mM [K⁺].

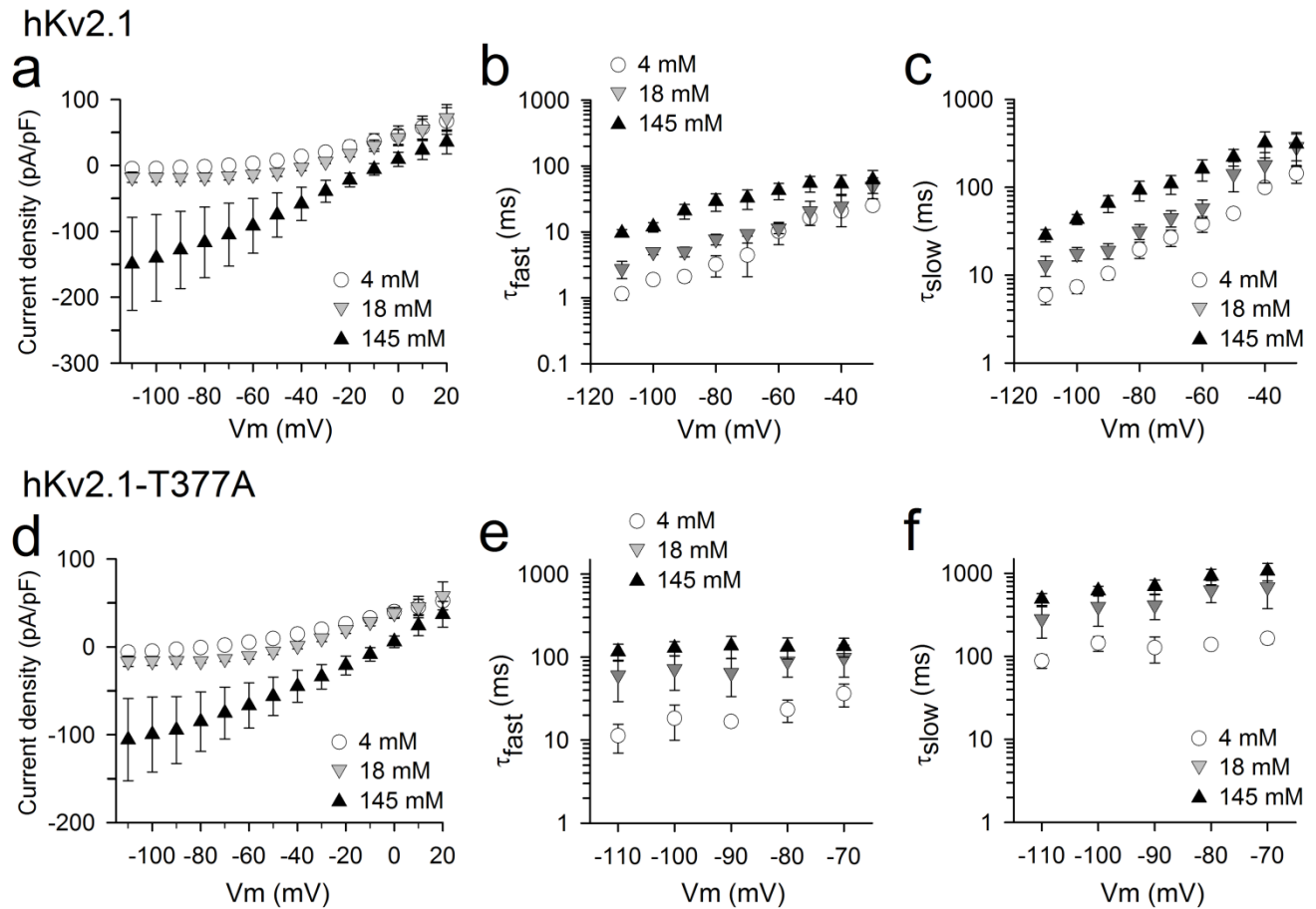


Figure S3: Increased external K⁺ concentration slowed down deactivation in both hKv2.1 and hKv2.1-T377A. **a.** Current-voltage (IV) relationship of hKv2.1 obtained in different external K⁺ concentrations (4 mM, 18 mM and 145 mM). As expected for K⁺ selective channels, the reversal potential shifts from -80 mV to -45 and 7 mV at 4 mM, 18 mM and 145 mM external K⁺ respectively. **b and c.** Time constants of deactivation at 4 mM (white circles), 18 mM (gray inverted triangles) and 145 mM (black triangles) external K⁺ for hKv2.1. Values were obtained by fitting the current decay with a double exponential function yielding a fast (panel b) and slow (panel c) component. Time constants are represented as mean \pm SEM from 3 to 6 independent recordings. Both the fast and the slow component are slowed down by increased extracellular K⁺. **d.** IV relationship of hKv2.1-T377A obtained in 4 mM, 18 mM and 145 mM external K⁺ concentration. **e and f.** Time constants of deactivation at 4 mM (white circles), 18 mM (gray inverted triangles) and 145 mM (black triangles) external K⁺ for hKv2.1-T377A. Note that the T-to-A mutant behaved as WT and the deactivation kinetics gradually slowed down with increasing extracellular K⁺ concentrations.

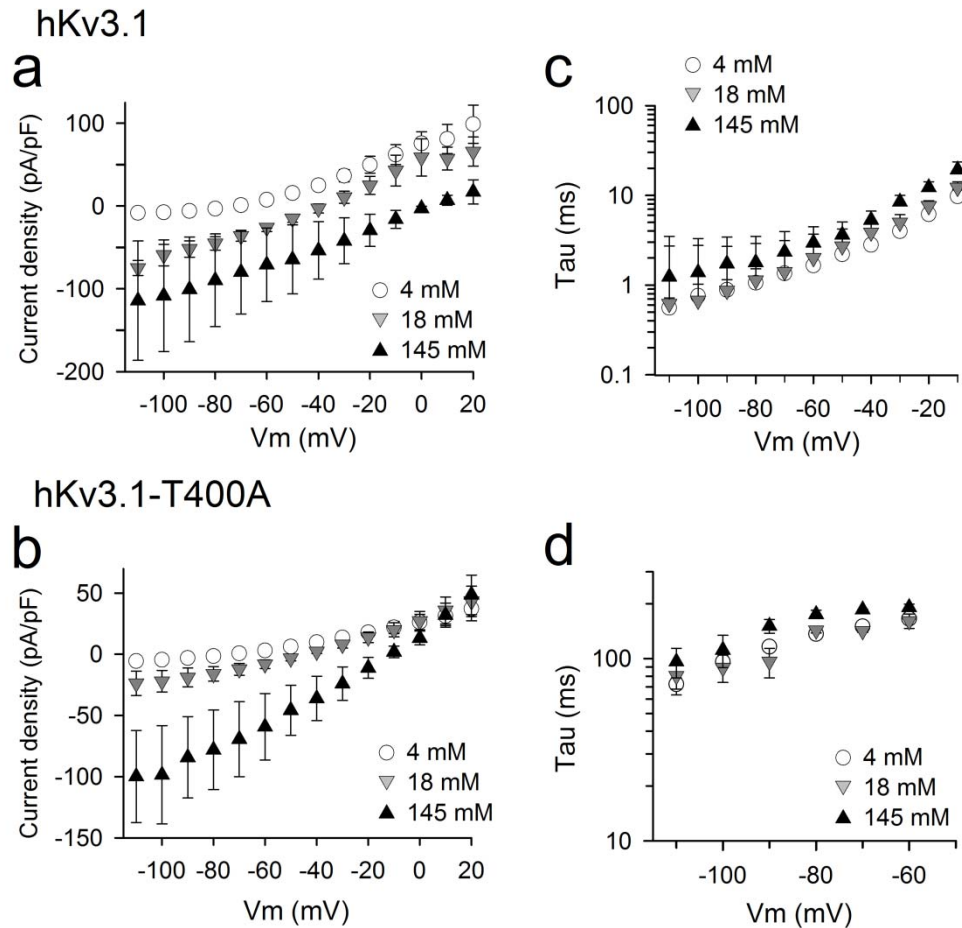


Figure S4: Increased external K^+ concentration did not affect deactivation in both hKv3.1 and hKv3.1-T400A. **a and b.** IV relationships of hKv3.1 and hKv3.1-T400A at different external K^+ concentrations: 4 mM (white circles), 18 mM (gray inverted triangles) and 145 mM (black triangles). As expected for K^+ selective channels, the reversal potential shifts from -80 mV to -45 and 7 mV at 4 mM, 18 mM and 145 mM external K^+ , respectively. **c and d.** Time constants of deactivation in 4 mM (white circles), 18 mM (gray inverted triangles) and 145 mM (black triangles) external K^+ for WT hKv3.1 (panel c) and hKv3.1-T400A (panel d). Current decay was fitted with a single exponential function and time constants are depicted as mean \pm SEM from 4 to 5 independent cells. Elevated external $[K^+]$ appeared to have no significant effect on deactivation kinetics in both the WT and the T-to-A mutant channels.