





Supplemental Figure S1. A phylogenetic tree was constructed from the amino acid sequence of DXS from the following species : Adonis aestivalis (ABK35283.1); Amomum villosum (ACR02668.1); Andrographis paniculata (AAP14353.1); Arabidopsis thaliana (CLA1 At4g15560; DXS2 At3g21500; DXS3 AT5G11380); Artemisia annua (AAD56390.2); Antirrhinum majus (AAW28999.1); Brassica rapa (ABE60813.1); Capsicum annuum (CAA75778.1); Catharanthus roseus (DXS2.1 renamed DXS2A CAA09804.2, DXS2.2 renamed DXS2B ABI35993.1); Chlamydomonas reinhardtii (CAA07554.1); Chrysanthemum x morifolium (BAE79547.1) ; Croton stellatopilosus (BAF75640.1) ; Deinococcus radiodurans R1 (NP_295198.1) ; Elaeis guineensis (AAS99588.1); Glycine max (ACO72582.1); Ginkgo biloba (DXS1 AAS89341.1, DXS2 AAR95699.1); Gossypium barbadense (ABN13970.1); Hordeum vulgare (BAJ94232.1. BAJ98888.1); Hevea brasiliensis (DXS1 clade AAS94123.1, DXS2 clade ABF18929.1; BAF98288.1); Medicago truncatula (DXS1, CAD22530.1, DXS2, CAN89181.1); Mentha piperita (AAC33513) ; Morinda citrifolia (AAL32062.1) ; Narcissus pseudonarcissus (CAC08458.1); Narcissus tazetta (ADD82535.1); Nicotiana tabacum (CBA12009.1); Oryza sativa (DXS1 NP 001055524, DXS2 NP 001059086, DXS3 clade BAD67657.1); Ostreococcus lucimarinus (XP 001416190.1); Physcomitrella patens (XP 001771607, XP 001756357, XP 001760423, XP 001754004); Picea abies (ABS50519.1; ABS50518.1); Picea sitchensis (ACN39837.1); Picrorhiza kurrooa LAMIALES (ACB55417.1); Pinus densiflora (DXS1 ACC54557.1, DXS2 ACC54554.1); Pinus taeda (DXS1 ACJ67021.1, DXS2 ACJ67020.1); Populus trichocarpa (DXS1 XP 002312717.1, DXS2 XP 002303416.1, XP 002331678.1, DXS3 XP_002308644.1); Pueraria montana (AAQ84169.1); Ricinus communis (DXS1 XP_002516843.1, DXS2 XP 002533688.1, DXS3 XP 002514364.1; DXS4 XP 002532384.1); Salvia miltiorrhiza (DXS1 ACF21004.1, DXS2 ACQ66107.1); Selaginella moellendorffii (EFJ14322.1); Sorghum bicolor (XP 002441088); Solanum lvcopersicum (CAZ66648; AF143812 1); Solanum tuberosum (DXS1 ADK73609.1); Stevia rebaudiana (ACI43010); Taxus x media (AAS89342.1); Vitis vinifera (DXS1, XP 002277919; DXS2, XP 002266925; DXS3, XP 002271585; DXS4, XP 002282428; DXS5, XP 002270336; DXS6, XP 002271782); Zea mays (DXS1 ABP88134.1, DXS2 : ABP88135.1, DXS3 ADN 22972). Note that C. roseus DXS2.1 and DXS2.2 have been renamed DXS2A and DXS2B in this tree.

Enzyme	Primer	Sequence (5' – 3')	Amplification	
	DXS2forA	GGAGATAGTGTATTCTGTGGCCA	3' end with T7	
DV62.2	DXS2-rev1	CTATCTCCTCCCTTAATTCGTCAGC	5' end with M13 reverse	
DA52.2	DXS2-full-for	CGACGAGTACTCTAGCATTCCTATCCTC	Eull longth ODE	
	DXS2-full-rev CCTAATTCCACAATTGTATTCATCACCAAGTA		Full-length OKF	
	CMS1	GGNCAGCCWATTGCHTTRTA	Laternal aDNA for survey	
	CMS3	TCATCRGGTGTHGTNACCTT	Internal CDNA fragment	
CMS	CMS7	AGCGGAGGATGTTGAGAAGGT	3' end with T7	
CMS	CMS5	TCAGGCATGCGGGAGAAAGTA	5' end with M13 reverse	
	CMS11	AGAAGATGTCAATTCTTCGGC	Full-length ORF	
	CMS12	GAATTACTATGCAGAGATGGG		
	CMKfor	CATGAYYTRGCITCHYTITTTCAT	Internal cDNA fragment	
	CMKrev	GTTGCWGCATTRCTRCTYCCWCC		
CMK	3CMKfor1	GGTTCATCTCGATAAAAAGGTGCC	3' end with T7	
UNIK	5CMKrev1	GTTATCAGTGCCAGTCTTCTTCCTAT	5' end with M13 reverse	
	CMK-full-for	GGATAGACAAAAAGGCATATCTTGAATG	· Full-length ORF	
	CMK-full-rev	TGGTTTTGGCATTTATAACCATTTTC		
	LytCR8	GAGCGTGCWGTYCARATTG	Internal cDNA fragment	
	LytCR10	GAGCGTGCWGTYCARATTG		
IIDD	LytCR12	GATCCAGATGAGGATCTCAT	3' end with T7	
HDK	LytCR13	GATTGTGAATAATTTCATTCGTA	5' end with M13 reverse	
	HDR1	ACAATGGCAATCTCTCTCCAA	Eull lon oth ODE	
	HDR2	AATCTATGCCAGTTGAAGGGC	run-iengin OKr	

Supplemental Table S1. Details of primer sequences and cloning procedure used to amplify the *DX2.2, CMS, CMK* and *HDR* cDNAs.

Enzyme	Primer sequence (5'-3')	Restriction site added	Plasmids cloning sites	Complemented <i>E. coli</i> strain	
DXS2.2	pQE-DXS2-Hindfor GCAAGCTTGCTGCTGCTCATGGGCATGCT	HindIII	nOF 30	EcAB4-2 (MG1655 <i>dxs::CAT</i> <i>MVA</i> +)	
	pQE-DXS2-Hindrev GCAAGCTTCTAAAACTGGAGAACTTGAAGAG TATTCTT	HindIII	HindIII		
	pQE-CMSBam TCGGATCCGAGAAAAGTGTCTCGGTAATTCT	BamHI pOF 30		EcAB4-7	
CMS	pQE-CMSHind TCAAGCTTAGCAGAGATGGGATTCAAAATTC T	HindIII	BamHI-HindIII	(MG1655 <i>ygbP::CAT</i> <i>MVA</i> +)	
СМК	pQE-CMKBam ACGGATCCTGGAATAAGCTAGCTGATGAAGT A	BamHI	pQE-30	EcAB4-9 (MG1655 <i>ychB::CAT</i> <i>MVA</i> +)	
	pQE-CMKKpn TCGGTACCTTCAGAAGACTGGGAGAAGTCT	KpnI	BamH1-Kpn1		
HDR	рQE-HDR-Bam2 GCGGATCCGAATTCGATGCTAAGAAATTCAG GCAC	BamHI	pQE-30 BamHI-HindIII	MG1655 ara⇔ispH	
	pQE-HDR-Bam3 GCGGATCCGAACAGAGCACAGAGCTGATGAA C	BamHI			
	pQE-HDR-Hind GCAAGCTTCTATGCCAGTTGAAGGGCTTCTTCAC G	HindIII			

Supplemental Table S2. Details of primer sequences and cloning procedures used to express proteins for functional validation.

Enzyme	Primer	Séquence (5'-3')
DVC2 1	qDXS-up	TTGATGTAGGTATTGCCGAGCA
DA52.1	qDXS-down	ACAATGAGTGGGTCCATCTGC
DVG22	qDXS2-up	GCTACAGTTGTTGATGCCCGA
DA52.2	qDXS2-down	TTCAAAGCCAAGAACTGTGCC
DVD	qDXR-up	GCAGCAAATGAGAAGGCAGTT
DXR	qDXR-down	GGGCTCAAACCAAGACTGTTC
CMS	qCMS-up	GCAACTTTGTCTCCTCAGCATT
CMS	qCMS-down	GCAACTTTGTCTCCTCAGCATT
CMR	qCMK-up	AAAGATTGAAGCAGCGTGTGC
CMK	qCMK-down	AGAGGCTTCAGACAAGGAAACG
MECS	qMECS-up	TAGAGGATGCGAGGCTCACT
MECS	qMECS-down	TCATAACCTGCCTCGTCCATC
IIDC	qHDS-up	GTCCCTTACTGAACCTCCAGAG
HDS	qHDS-down	AATCACCTGTCCTGCGTTGG
IIDD	qHDR-up	GATTGAGAGGACGATGATGCGT
НИК	qHDR-down	AGTTCCACCCTCCAACGACT
RPS9	qRPS9-for	TTACAAGTCCCTTCGGTGGT
AJ749993	aRPS9-rev	TTACAAGTCCCTTCGGTGGT

Supplemental Table S3. The sequence of primers used in the qRT-PCR analysis.

Enzyme	Primer sequence (5'-3')	Restriction site added	Plasmids cloning sites	Fusion protein
DXS2.1	DXS-GFP-S ATAGATCTTATGGCGGTTTCCGGGGCT	BglII	pSCA-cassette-GFPi	DXS-GFP DXS-YFP
	DXS-GFP-AS cgactagtttgaagtttaagggttct	SpeI	Bg/III-SpeI	
DXS2.2	DXS2-GFP-S AGGCTAGCATGGCGGTGGCATCGAAT	NheI	pSCA-cassette-GFPi	DXS2-GFP DXS2-YFP
	DXS2-GFP-AS cggctagcaaactggagaacttgaagag	NheI	Spel	
DVD	DXR-GFP-S ggagatcttatggctttgaattcgctgtcc	BglII	pSCA-cassette-GFPi	DXR-GFP DXR-YFP
DAK	DXR-GFP-AS ggactagtggcagggctcaaaccaagact	SpeI	BglII-SpeI	
CMS	CMS-GFP-S CGAGATCTTATGTCAATTCTTCGGCTC	<i>Bgl</i> II	pSCA-cassette-GFPi	CMS-GFP
CMS	CMS-GFP-AS CGACTAGTAGCAGAGATGGGATTCAA	SpeI	BglII-SpeI	
	CMK-GFP-S GCACTAGTATGGCTTCCTCTCAGTCTCTATG TGGT	SpeI	pSCA-cassette-GFPi	CMK-GFP
CIVIK	CMK-GFP-AS GCACTAGTCTATTCAGAAGACTGGGAGAAGT CTGT	SpeI	Spel	
MECS	MECS-GFP-S TCAGATCTTATGGCTATGGCGACTTCT	BglII	pSCA-cassette-GFPi	MECS-GFP
	MECS-GFP-AS gcactagtcttcctcataagaagtac	SpeI	SpeI	
HDS	HDS-GFP-S ggagatctgatggcgaccggaacagttc	BglII	pSCA-cassette-GFPi	HDS-GFP HDS-YFP
	HDS-GFP-AS GGACTAGTGTCTTCTGCAGGAGGATCAACC	SpeI	BglII-SpeI	
	HDR-GFP-S GCAGATCTTATGGCAATCTCTCTCCAA	BglII	pSCA-cassette-GFPi	HDR-GFP
HDR	HDR-GFP-AS TAACTAGTTGCCAGTTGAAGGGCTTC	SpeI	BglII-SpeI	

Supplemental Table S4. Details of primer sequences and cloning procedure used to express full-length MEP pathway enzyme fused to FP.

Enzyme	Primer sequence (5'-3')	Restriction site added	Plasmids cloning sites	Fusion protein
DXS2.1	DXS-GFP-S ATAGATCTTATGGCGGTTTCCGGGGGCT	BglII	nSCA_cassette_GEPi	tpDXS-GFP
	peptideL-DXSrev gcactagttgctctaagttctgcagcgagct g	SpeI	BglII-SpeI	
DXS2.2	DXS2-GFP-S Aggctagcatggcggtggcatcgaat	NheI	nSCA_cassette_GEPi	tpDXS2-GFP
	peptideL-DXS2rev gcgctagcgatggtatccaaaataggagtaa aaggctt	NheI	SpeI	
DXR	DXR-GFP-S GGAGATCTTATGGCTTTGAATTCGCTGTCC	BglII	pSCA-cassette-GFPi	tpDXR-GFP tpDXR-YFP
	peptideL-DXRrev GCACTAGTATTCTCAGCAACTATATCTAGTG TCTG	SpeI	pSCA-cassette-YFPi Bg/II-SpeI	
HDS	HDS-GFP-S ggagatctgatggcgaccggaacagttc	BglII	pSCA-cassette-GFPi	tpHDS-GFP tpHDS-YFP
	peptideL-HDSrev GCACTAGTGTCACTACCAAGGGCTACATTCC C	SpeI	pSCA-cassette-YFPi BglII-SpeI	

Supplemental Table S5. Details of primer sequences and cloning procedure used to fuse the first 100-residues of DXS2.1, DXS2.2, DXR and HDS to GFP.

Enzyme	Primer sequence (5'-3')	Restriction site added	Plasmids <i>cloning sites</i>	Fusion protein
DXR	DXR-GFP-S GGAGATCTTATGGCTTTGAATTCGCTGTCC	BglII	nSCA-cassette-VFPi	tpDXR(1-83)- YFP
	peptideL-DXRrev2 gcactagtaactattgaaatgggcttctgac cttc	SpeI	BglII-SpeI	
	DXR-C-for2 GCACTAGTGGCTCTACAGGCTCAGTAGGAAC TCAG	SpeI	pSCA-cassette-YFPi- tpDXR(1-83)	Tp(1-83)-YFP- DXR(84-474)
	DXR-GFP-AS ggactagtggcagggctcaaaccaagact	SpeI	NheI	
HDS	HDS-GFP-S ggagatctgatggcgaccggaacagttc	BglII	nSCA-cassette-VEPi	tpHDS(1-100)- YFP
	peptideL-HDSrev gcactagtgtcactaccaagggctacattcc c	SpeI	BglII-SpeI	
	HDS-C-for2 GCACTAGTATGGTTGGGAATGTAGCCCTTGG TAGTGAC	SpeI pSCA-cassette-YFPi- tpHDS(1-100)		tp(1-100)-YFP-
	HDS-GFP-AS GGACTAGTGTCTTCTGCAGGAGGATCAACC	SpeI	N heI	нрэ(91-740)

Supplemental Table S6. Details of primer sequences and cloning procedures used to create internal YFP fusion for DXR and HDS.

Enzyme	Primer sequence (5'-3')	Restriction site added	Plasmids cloning sites	Fusion protein
CAB1	CAB1-for gcactagtatggccgcctcaacaatggc	SpeI	nSCA_cassette_CEPi	
	CAB1-rev gcactagtctttccgggaacaaagttggtgg c	SpeI	SpeI SpeI	
TIC40	TIC40-for gcactagtatggagaaccttaccctagtttc ttgct	SpeI	pSCA-cassette-CFPi	TIC-CFP
	TIC40-rev gcactagtacccgtcattcctgggaagagc	SpeI	Spel	
CCD4	CCD4-for GCACTAGTATGGACTCTGTTTCTTCTTC CTTCC	SpeI pSCA-cassette-CFPi		CCD4 CEP
	CCD4-rev GCACTAGTAAGCTTATTAAGGTCACTTTCCT	SpeI	SpeI	CCD4-CFP

Supplemental Table S7. Details of primer sequences and cloning procedures used to create plastid subcompartment CFP-markers of. The CAB1 (chlorophyll a/b binding protein 1, At1g29930), TIC40 (translocon at the inner envelope of chloroplasts, At5g16620), CCD4 (carotenoid cleavage dioxygenase 4, At4g19170) proteins of *A. thaliana* were used as a marker of thylakoid, inner membrane of plastid and plastoglobuli, respectively. The coding sequence of these proteins was amplified using retro-transcribed RNA extracted from *A. thaliana* as a matrix.



© 2020 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).