

Article

Supplementary information

A systematic review: Is *Aedes albopictus* an efficient bridge vector for zoonotic arboviruses?

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Table S1. List of the 16 articles found by searching Google Scholar to characterize the types of natural breeding sites exploited by *Ae. albopictus*.

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Table S2. Typology and number of reported natural containers exploited by *Ae. albopictus* from articles found in PubMed.

Type of natural container	Number of reported breeding sites	References
Bamboo stumps	127	
	1	Delatte, H.; Dehecq, J.S.; Thiria, J.; Domerg, C.; Paupy, C.; Fontenille, D. Geographic Distribution and Developmental Sites of <i>Aedes albopictus</i> (Diptera: Culicidae) During a Chikungunya Epidemic Event. <i>Vector Control Southeast Asia</i> 2008 , <i>8</i> , 25–34.
	46	Dev, V. Dengue vectors in urban and suburban Assam, India: entomological observations. <i>WHO South-East Asia J. Public Heal.</i> 2014 , <i>3</i> , 5838.
	17	Edillo, F.E.; Roble, N.D.; Otero, N.D. The key breeding sites by pupal survey for dengue mosquito vectors, <i>Aedes aegypti</i> (Linnaeus) and <i>Aedes albopictus</i> (Skuse), in Guba, Cebu City, Philippines. <i>Southeast Asian J. Trop. Med. Public Health</i> 2012 , <i>43</i> , 1365–1374.
	12	Gilotra, S.K.; Rozeboom, L.E.; Bhattacharya, N.C. Observations on possible competitive displacement between populations of <i>Aedes aegypti</i> Linnaeus and <i>Aedes albopictus</i> Skuse in Calcutta. <i>Bull. World Health Organ.</i> 1967 , <i>37</i> , 437–446.
	21	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972 , <i>46</i> , 477–483.
	30	Sota, T.; Mogi, M.; Hayamizu, E. Seasonal distribution and habitat selection by <i>Aedes albopictus</i> and <i>Aedes riversi</i> (Diptera: Culicidae) in Northern Kyushu, Japan. <i>J. Med. Entomol.</i> 1992 , <i>29</i> , 296–304.
Bromeliads	291	
	3	Forattini, O.P.; Brito, M. de B. Brief Communication an Unusual Ground Larval Habitat of <i>Aedes albopictus</i> . <i>Rev. Inst. Med. Trop. Sao Paulo</i> 1998 , <i>40</i> , 121–122.
	275	Marques, G.R.; Forattini, O.P. <i>Aedes albopictus</i> in soil bromeliads in Ilhabela, coastal area of Southeastern Brazil. <i>Rev Saude Publica</i> 2005 , <i>39</i> , 548–552.
	7	Mocellin, M.G.; Simões, T.C.; do Nascimento, T.F.S.; Teixeira, M.L.F.; Lounibos, L.P.; de Oliveira, R.L. Bromeliad-inhabiting mosquitoes in an urban botanical garden of dengue endemic Rio de Janeiro - Are bromeliads productive habitats for the invasive vectors <i>Aedes aegypti</i> and <i>Aedes albopictus</i> ? <i>Mem. Inst. Oswaldo Cruz</i> 2009 , <i>104</i> , 1171–1176.
	5	Oliveira, V.C. de; Almeida Neto, L.C. de Ocorrência de <i>Aedes aegypti</i> e <i>Aedes albopictus</i> em bromélias cultivadas no Jardim Botânico Municipal de Bauru, São Paulo, Brasil. <i>Cad. Saude Publica</i> 2017 , <i>33</i> , e00071016.
1	O'Meara, G.F.; Cutwa, M.M.; Evans, L.F. Bromeliad-inhabiting mosquitoes in south Florida: native and exotic plants differ in species composition. <i>J. vector Ecol.</i> 2003 , <i>28</i> , 37–46.	
Cacao shells	3	
	3	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005 , <i>42</i> , 726–731.
Coconut shells	838	
	8	Bagny, L.; Delatte, H.; Elissa, N.; Quilici, S.; Fontenille, D.; Adhami, J.; Murati, N.; Adhami, J.; Reiter, P.; Beltrame, A.; et al. <i>Aedes</i> (Diptera: Culicidae) vectors of arboviruses in Mayotte (Indian Ocean): distribution area and larval habitats. <i>J. Med. Entomol.</i> 2009 , <i>46</i> , 198–207.
	283	Banerjee, S.; Aditya, G.; Saha, G.K. Household disposables as breeding habitats of dengue vectors: Linking wastes and public health. <i>Waste Manag.</i> 2013 , <i>33</i> , 233–239.

Table S2. Continued

Type of natural container	Number of reported breeding sites	References
Coconut shells	838	
	7	Edillo, F.E.; Roble, N.D.; Otero, N.D. The key breeding sites by pupal survey for dengue mosquito vectors, <i>Aedes aegypti</i> (Linnaeus) and <i>Aedes albopictus</i> (Skuse), in Guba, Cebu City, Philippines. <i>Southeast Asian J. Trop. Med. Public Health</i> 2012 , <i>43</i> , 1365–1374.
	144	Rao, B.B.; George, B. Breeding patterns of <i>Aedes stegomyia albopictus</i> in periurban areas of Calicut, Kerala, India. <i>Southeast Asian J. Trop. Med. Public Health</i> 2010 , <i>41</i> , 536–540.
	36	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972 , <i>46</i> , 477–483.
	20	Shriram, A.N.; Sivan, A.; Sugunan, A.P. Spatial distribution of <i>Aedes aegypti</i> and <i>Aedes albopictus</i> in relation to geo-ecological features in South Andaman, Andaman and Nicobar Islands, India. <i>Bull. Entomol. Res.</i> 2018 , <i>108</i> , 166–174.
	1	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: Culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005 , <i>42</i> , 726–731.
Dead cow horns	316	Thavara, U.; Tawatsin, a; Chansang, C.; Kong-ngamsuk, W.; Paosriwong, S.; Boon-Long, J.; Rongsriyam, Y.; Komalamisra, N. Larval occurrence, oviposition behavior and biting activity of potential mosquito vectors of dengue on Samui Island, Thailand. <i>J. Vector Ecol.</i> 2001 , <i>26</i> , 172–180.
	22	Vijayakumar, K.; Sudheesh Kumar, T.K.; Nujum, Z.T.; Umarul, F.; Kuriakose, A. A study on container breeding mosquitoes with special reference to <i>Aedes</i> (<i>Stegomyia</i>) <i>aegypti</i> and <i>Aedes albopictus</i> in Thiruvananthapuram district, India. <i>J. Vector Borne Dis.</i> 2014 , <i>51</i> , 27–32.
	3	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: Culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005 , <i>42</i> , 726–731.
Dead leaves	3	
	5	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: Culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005 , <i>42</i> , 726–731.
Ground cavity	1	Bagny, L.; Delatte, H.; Elissa, N.; Quilici, S.; Fontenille, D.; Adhami, J.; Murati, N.; Adhami, J.; Reiter, P.; Beltrame, A.; et al. <i>Aedes</i> (Diptera: Culicidae) vectors of arboviruses in Mayotte (Indian Ocean): distribution area and larval habitats. <i>J. Med. Entomol.</i> 2009 , <i>46</i> , 198–207.
	1	Forattini, O.P.; Brito, M. de B. Brief Communication an Unusual Gground Larval Habitat of <i>Aedes albopictus</i> . <i>Rev. Inst. Med. Trop. Sao Paulo</i> 1998 , <i>40</i> , 121–122.
Hollow log	1	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: Culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005 , <i>42</i> , 726–731.
	1	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972 , <i>46</i> , 477–483.
Leaf axils	16	
	1	Banerjee, S.; Aditya, G.; Saha, G.K. Household disposables as breeding habitats of dengue vectors: Linking wastes and public health. <i>Waste Manag.</i> 2013 , <i>33</i> , 233–239.
	4	Delatte, H.; Dehecq, J.S.; Thiria, J.; Domerg, C.; Paupy, C.; Fontenille, D. Geographic Distribution and Developmental Sites of <i>Aedes albopictus</i> (Diptera: Culicidae) During a Chikungunya Epidemic Event. <i>Vector Control Southeast Asia</i> 2008 , <i>8</i> , 25–34.

Table S2. Continued

Type of natural container	Number of reported breeding sites	References
Leaf axils	16	
	6	Dev, V. Dengue vectors in urban and suburban Assam, India: entomological observations. <i>WHO South-East Asia J. Public Heal.</i> 2014 , <i>3</i> , 5838.
	1	Edillo, F.E.; Roble, N.D.; Otero, N.D. The key breeding sites by pupal survey for dengue mosquito vectors, <i>Aedes aegypti</i> (Linnaeus) and <i>Aedes albopictus</i> (Skuse), in Guba, Cebu City, Philippines. <i>Southeast Asian J. Trop. Med. Public Health</i> 2012 , <i>43</i> , 1365–1374.
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	3	Shriram, A.N.; Sivan, A.; Sugunan, A.P. Spatial distribution of <i>Aedes aegypti</i> and <i>Aedes albopictus</i> in relation to geo-ecological features in South Andaman, Andaman and Nicobar Islands, India. <i>Bull. Entomol. Res.</i> 2018 , <i>108</i> , 166–174.
Palm bracts	7	
	7	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972 , <i>46</i> , 477–483.
Palm leaves	55	
	55	Delatte, H.; Dehecq, J.S.; Thiria, J.; Domerg, C.; Paupy, C.; Fontenille, D. Geographic Distribution and Developmental Sites of <i>Aedes albopictus</i> (Diptera: Culicidae) During a Chikungunya Epidemic Event. <i>Vector Control Southeast Asia</i> 2008 , <i>8</i> , 25–34.
Puddle	1	
	1	Marquetti, M.; Bisset, J.; Leyva, M.; Garcia, A.; Rodriguez, M. Comportamiento estacional y temporal de <i>Aedes aegypti</i> y <i>Aedes albopictus</i> en La Habana, Cuba. 2008 , <i>60</i> , 62–67.
Rock holes	49	
	1	Bagny, L.; Delatte, H.; Elissa, N.; Quilici, S.; Fontenille, D.; Adhami, J.; Murati, N.; Adhami, J.; Reiter, P.; Beltrame, A.; et al. <i>Aedes</i> (Diptera: Culicidae) vectors of arboviruses in Mayotte (Indian Ocean): distribution area and larval habitats. <i>J. Med. Entomol.</i> 2009 , <i>46</i> , 198–207.
	33	Delatte, H.; Dehecq, J.S.; Thiria, J.; Domerg, C.; Paupy, C.; Fontenille, D. Geographic Distribution and Developmental Sites of <i>Aedes albopictus</i> (Diptera: Culicidae) During a Chikungunya Epidemic Event. <i>Vector Control Southeast Asia</i> 2008 , <i>8</i> , 25–34.
	8	O'Meara, G.F.; Evans Jr., L.F.; Womack, M.L. Colonization of rock holes by <i>Aedes albopictus</i> in the southeastern United States. <i>J Am Mosq Control Assoc</i> 1997 , <i>13</i> , 270–274.
	3	Pena, C.J.; Gonzalez, G.; Chadee, D.D. Seasonal prevalence and container preferences of <i>Aedes albopictus</i> in Santo Domingo City, Dominican Republic. <i>J. Vector Ecol.</i> 2003 , <i>28</i> , 208–212.
	1	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972 , <i>46</i> , 477–483.
	1	Shriram, A.N.; Sivan, A.; Sugunan, A.P. Spatial distribution of <i>Aedes aegypti</i> and <i>Aedes albopictus</i> in relation to geo-ecological features in South Andaman, Andaman and Nicobar Islands, India. <i>Bull. Entomol. Res.</i> 2018 , <i>108</i> , 166–174.
	2	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005 , <i>42</i> , 726–731.

Table S2. Continued

Type of natural container	Number of reported breeding sites	References
Snail shells	10	
	9	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972, 46, 477–483.
	1	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005, 42, 726–731.
Tree holes	125	
	1	Bagny, L.; Delatte, H.; Elissa, N.; Quilici, S.; Fontenille, D.; Adhami, J.; Murati, N.; Adhami, J.; Reiter, P.; Beltrame, A.; et al. <i>Aedes</i> (Diptera: Culicidae) vectors of arboviruses in Mayotte (Indian Ocean): distribution area and larval habitats. <i>J. Med. Entomol.</i> 2009, 46, 198–207.
	32	Gomes, A.; Forattini, O.; Kakitani, I.; Marques, G.; Marques, C.; Marucci, D.; Brito, M. Microhabitats de <i>Aedes albopictus</i> (Skuse) na região do Vale do Paraíba, Estado de São Paulo, Brasil. <i>Rev. Saúde Pública S. Paulo</i> 1992, 26, 108–118.
	1	Delatte, H.; Dehecq, J.S.; Thiria, J.; Domerg, C.; Paupy, C.; Fontenille, D. Geographic Distribution and Developmental Sites of <i>Aedes albopictus</i> (Diptera: Culicidae) During a Chikungunya Epidemic Event. <i>Vector Control Southeast Asia</i> 2008, 8, 25–34.
	12	Müller, G.C.; Kravchenko, V.D.; Junnila, A.; Schlein, Y. Tree-hole breeding mosquitoes in Israel. <i>J. Vector Ecol.</i> 2012, 37, 102–109.
	5	O'Meara, G.F.; Evans Jr., L.F.; Womack, M.L. Colonization of rock holes by <i>Aedes albopictus</i> in the southeastern United States. <i>J Am Mosq Control Assoc</i> 1997, 13, 270–274.
	13	Rozeboom, L.E.; Bridges, J.R. Relative population densities of <i>Aedes albopictus</i> and <i>A. guamensis</i> on Guam. <i>Bull. World Health Organ.</i> 1972, 46, 477–483.
	26	Shriram, A.N.; Sivan, A.; Sugunan, A.P. Spatial distribution of <i>Aedes aegypti</i> and <i>Aedes albopictus</i> in relation to geo-ecological features in South Andaman, Andaman and Nicobar Islands, India. <i>Bull. Entomol. Res.</i> 2018, 108, 166–174.
	2	Simard, F.; Nchoutpouen, E.; Toto, J.C.; Fontenille, D. Geographic distribution and breeding site preference of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: culicidae) in Cameroon, Central Africa. <i>J Med Entomol</i> 2005, 42, 726–731.
	8	Sivan, A.; Shriram, A.N.; Sugunan, A.P.; Anwesh, M.; Muruganandam, N.; Kartik, C.; Vijayachari, P. Natural transmission of dengue virus serotype 3 by <i>Aedes albopictus</i> (Skuse) during an outbreak in Havelock Island: Entomological characteristics. <i>Acta Trop.</i> 2016, 156, 122–129.
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Table S3: List of references used to analyse the host feeding preferences of *Aedes albopictus*.

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Table S4: List of references that reported infection, infections rate, dissemination rate, dissemination efficiency, transmissions rate, transmission efficiency or transovarial transmission in *Ae. albopictus* for the indicated arboviruses.

Virus Name	References
Arumowot	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.
BeAn 100049 (Urucuri)	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.
Bujaru	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.
Bussuquara	Tesh, R.B.; Shroyer, D.A. The mechanism of arbovirus transovarial transmission in mosquitoes: San Angelo virus in <i>Aedes albopictus</i> . <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 1394–1404.
Cache Valley	Armstrong, P.M.; Anderson, J.F.; Farajollahi, A.; Healy, S.P.; Unlu, I.; Crepeau, T.N.; Gaugler, R.; Fonseca, D.M.; Andreadis, T.G. Isolations of Cache Valley virus from <i>Aedes albopictus</i> (Diptera: Culicidae) in New Jersey and evaluation of its role as a regional arbovirus vector. <i>J. Med. Entomol.</i> 2013 , <i>50</i> , 1310–1314.
Chandipura	Ramachandra, R.; Sing, K.; Dhanda, V.; Bhattacharya, N. Experimental transmission of Chandipura Virus by mosquitoes. <i>Ind. Jour. Med. Res.</i> 1967 , <i>55</i> , 1306–1310.
Chilibre	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.
Eastern equine encephalomyelitis	Mitchell, C.J.; McLean, R.G.; Nasci, R.S.; Crans, W.J.; Smith, G.C.; Caccamise, D.F. Susceptibility parameters of <i>Aedes albopictus</i> to per oral infection with eastern equine encephalitis virus. <i>J Med Entomol</i> 1993 , <i>30</i> , 233–235. Moncayo, a C.; Edman, J.D.; Turell, M.J. Effect of eastern equine encephalomyelitis virus on the survival of <i>Aedes albopictus</i> , <i>Anopheles quadrimaculatus</i> , and <i>Coquilletidia perturbans</i> (Diptera: Culicidae). <i>J. Med. Entomol.</i> 2000 , <i>37</i> , 701–706. Scott, T.W.; Lorenz, L.H.; Weaver, S.C. Susceptibility of <i>Aedes albopictus</i> to infection with eastern equine encephalomyelitis virus. <i>J. Am. Mosq. Control Assoc.</i> 1990 , <i>6</i> , 274–278. Turell, M.; Beaman, J.; Neely, G. Experimental transmission of Eastern equine encephalitis virus by strains of <i>Aedes albopictus</i> and <i>A. taeniorhynchus</i> (Diptera: Culicidae). <i>J. Med. Entomol.</i> 1994 , <i>31</i> , 287–290.
Getah	Takashima, I.; Hashimoto, N. Getah virus in several species of mosquitoes. <i>Trans. R. Soc. Trop. Med. Hyg.</i> 1985 , <i>79</i> , 546–550.
Icoaraci	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.
Ilheus	Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666.
Itaporanga	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.
Jamestown Canyon	Grimstad, P.R. Recently Introduced <i>Aedes albopictus</i> in the United States: Potential vector of La Crosse Virus (Bunyaviridae: California Serogroup)1. 1989 , <i>5</i> , 422–427.
Japanese encephalitis	Rosen, L.; Tesh, R.B.; Lien, J.; John, H. Transovarial Transmission of Japanese Encephalitis Virus by Mosquitoes. <i>Am. Assoc. Adv. Sci.</i> 1978 , <i>199</i> , 909–911. Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666. Weng, M.H.; Lien, J.C.; Wang, Y.M.; Wu, H.L.; Chin, C. Susceptibility of three laboratory strains of <i>Aedes albopictus</i> (Diptera: Culicidae) to Japanese encephalitis virus from Taiwan. <i>J Med Entomol</i> 1997 , <i>34</i> , 745–747.
Karimabad	Wispelaere, M.; Desprès, P.; Choumet, V. European <i>Aedes albopictus</i> and <i>Culex pipiens</i> Are Competent Vectors for Japanese Encephalitis Virus. <i>PLoS Negl. Trop. Dis.</i> 2017 , <i>11</i> , e0005294.
Keystone	Tesh, R.B. Multiplication Phlebotomus phlebotomus fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4. Grimstad, P.R. Recently Introduced <i>Aedes albopictus</i> in the United States: Potential vector of La Crosse Virus (Bunyaviridae: California Serogroup)1. 1989 , <i>5</i> , 422–427.

Table S4. Continued

Virus Name	References
Keystone	Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666.
kokobera	Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666.
Kunjin	Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666. Cully Jr., J.F.; Streit, T.G.; Heard, P.B. Transmission of La Crosse virus by four strains of <i>Aedes albopictus</i> to and from the eastern chipmunk (<i>Tamias striatus</i>). <i>J Am Mosq Control Assoc</i> 1992 , <i>8</i> , 237–240. Grimstad, P.R. Recently Introduced <i>Aedes albopictus</i> in the United States: Potential vector of La Crosse Virus (Bunyaviridae: California Serogroup)1. 1989 , <i>5</i> , 422–427.
La Crosse	Hughes, M.T.; Gonzalez, J.A.; Reagan, K.L.; Carol, D.; Beaty, B.J.; Beaty, B.J. Comparative Potential of <i>Aedes triseriatus</i> , <i>Aedes albopictus</i> , and <i>Aedes aegypti</i> (Diptera: Culicidae) to Transovarially Transmit La Crosse Virus Comparative Potential of <i>Aedes triseriatus</i> , <i>Aedes albopictus</i> , and <i>Aedes aegypti</i> (Diptera: Culicidae). <i>Entomol. Soc. Am.</i> 2006 , <i>43</i> , 757–761. Lambert, A.J.; Blair, C.D.; D’Anton, M.; Ewing, W.; Harborth, M.; Seiferth, R.; Xiang, J.; Lanciotti, R.S. La Crosse Virus in <i>Aedes albopictus</i> Mosquitoes, Texas, USA, 2009. <i>Emerg. Infect. Dis.</i> 2010 , <i>16</i> , 856–858. Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666.
Mayaro	Smith, G.C.; Francly, D.B. Laboratory studies of a Brazilian strain of <i>Aedes albopictus</i> as a potential vector of Mayaro and Oropouche viruses. <i>J Am Mosq Control Assoc</i> 1991 , <i>7</i> , 89–93.
Oropouche	Smith, G.C.; Francly, D.B. Laboratory studies of a Brazilian strain of <i>Aedes albopictus</i> as a potential vector of Mayaro and Oropouche viruses. <i>J Am Mosq Control Assoc</i> 1991 , <i>7</i> , 89–93.
Orungo	Tomori, O.; Aitken, T.H. Orungo virus: transmission studies with <i>Aedes albopictus</i> and <i>Aedes aegypti</i> (Diptera: Culicidae). <i>J Med Entomol</i> 1978 , <i>14</i> , 523–526.
Pacui	Tesh, R.B. Multiplication <i>Phlebotomus phlebotomus</i> fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4. Francly, D.; Krabatsos, N.; Wesson, C.; Moore, J.; Lazuiick, J.; Niebylski, T.; Tsai, T.; Craig, J. A new Arbovirus from <i>Aedes albopictus</i> , an Asian mosquito Established in the United States. <i>Science (80-)</i> . 1990 , <i>250</i> , 1738–1740.
Potosi	Heard, P.B.; Niebylski, M.L.; Francly, D.B. Transmission of a Newly Recognized Virus (Bunyaviridae, Bunyavirus) Isolated from <i>Aedes albopictus</i> (Diptera: Culicidae) in Potosi, Missouri. <i>J. Med. Entomol.</i> 1991 , <i>28</i> , 601–605. Mitchell, C.J.; Smith, G.C.; Miller, B.R. Vector Competence of <i>Aedes Albopictus</i> for a Newly Recognized Bunyavirus From Mosquitoes Collected in Potosi, Missouri1. <i>J. Am. Mosq. Control Assoc.</i> 1990 , <i>6</i> , 523–527.
Rift Valley	Brustolin, M.; Talavera, S.; Nunez, A.; Santamaría, C.; Rivas, R.; Pujol, N.; Valle, M.; Verdun, M.; Brun, A.; Pages, N.; et al. Rift Valley fever virus and European mosquitoes: vector competence of <i>Culex pipiens</i> and <i>Stegomyia albopicta</i> (= <i>Aedes albopictus</i>). <i>Med. Vet. Entomol.</i> 2017 , <i>31</i> , 365–372. Turell, M.J.; Bailey, C.L.; Beaman, J.R. Vector competence of a Houston, Texas strain of <i>Aedes albopictus</i> for Rift Valley fever virus. <i>J. Am. Mosq. Control Assoc.</i> 1988 , <i>4</i> , 94–96. Mitchell, C.J.; Gubler, D.J. Vector competence of geographic strains of <i>Aedes albopictus</i> and <i>Aedes polynesiensis</i> and certain other <i>Aedes</i> (<i>Stegomyia</i>) mosquitoes for Ross River virus. <i>J Am Mosq Control Assoc</i> 1987 , <i>3</i> , 142–147.
Ross River	Mitchell, C.J.; Miller, B.R.; Gubler, D.J. Vector competence of <i>Aedes albopictus</i> from Houston, Texas, for dengue serotypes 1 to 4, yellow fever and Ross River viruses. <i>J. Am. Mosq. Control Assoc.</i> 1987 , <i>3</i> , 460–465.
Salehabad	Tesh, R.B. Multiplication <i>Phlebotomus phlebotomus</i> fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4. Shroyer, D.A. Transovarial Maintenance of San Angelo Virus in Sequential Generations of <i>Aedes albopictus</i> . <i>Am J Trop Med Hyg.</i> 1986 , <i>35</i> , 408–417.
San Angelo	Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666. Tesh, R.B. Multiplication <i>Phlebotomus phlebotomus</i> fever group arboviruses in mosquitoes after intrathoracic inoculation. <i>J. Med. Entomol.</i> 1975 , <i>12</i> , 1–4.

Table S4. Continued

Virus Name	References
Tensaw Virus	Mitchell, C.J. Vector competence of North and South American strains of <i>Aedes albopictus</i> for certain arboviruses: a review. <i>J. Am. Mosq. Control Assoc.</i> 1991 , <i>7</i> , 446–451.
Trivittatus Virus	Grimstad, P.R. Recently Introduced <i>Aedes albopictus</i> in the United States: Potential vector of La Crosse Virus (Bunyaviridae: California Serogroup)1. 1989 , <i>5</i> , 422–427.
Uganda S	Tesh, R.B. Experimental studies on the transovarial transmission of Kunjin and San Angelo viruses in mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1980 , <i>29</i> , 657–666.
Usutu virus	Puggioli, A.; Bonilauri, P.; Calzolari, M.; Lelli, D.; Carrieri, M.; Urbanelli, S.; Pudar, D.; Bellini, R. Does <i>Aedes albopictus</i> (Diptera: Culicidae) play any role in Usutu virus transmission in Northern Italy? Experimental oral infection and field evidences. <i>Acta Trop.</i> 2017 , <i>172</i> , 192–196.
Venezuelan equine encephalitis	Beaman, J.R.; Turell, M.J. Transmission of Venezuelan Equine Encephalomyelitis Virus by Strains of <i>Aedes albopictus</i> (Diptera: Culicidae) Collected from North and South America. <i>J. Med. Entomol.</i> 1991 , <i>28</i> , 161–164.
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West Nile	Baqar, S.; Hayes, C.G.; Murphy, J.R.; Watts, D.M. Vertical transmission of West Nile virus by <i>Culex</i> and <i>Aedes</i> species mosquitoes. <i>Am. J. Trop. Med. Hyg.</i> 1993 , <i>48</i> , 757–762.
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Yellow fever	Amraoui, F.; Ayed, W. Ben; Madec, Y.; Faraj, C.; Himmi, O.; Btissam, A.; Sarih, M.; Failloux, A.B. Potential of <i>aedes albopictus</i> to cause the emergence of arboviruses in Morocco. <i>PLoS Negl. Trop. Dis.</i> 2019 , <i>13</i> , e0006997.
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Table S4. Continued

Virus Name	References
Yellow fever	Lourenço de Oliveira, R.; Vazelle, M.; Maria, A.N.A.; Filippis, B.D.E.; Failloux, A. Large genetic differentiation and low variation in vector competence for dengue and yellow fever viruses of <i>Aedes albopictus</i> from Brazil, the United States, and the Cayman Islands. <i>Am. J. Trop. Med. Hyg.</i> 2003, 69, 105–114.
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Table S5: Natural detection or isolation of arboviruses in *Ae. albopictus* from field-collected mosquitoes.

Virus Name	Detection	Isolation	References
CVV	-	+	Armstrong, P.M.; Anderson, J.F.; Farajollahi, A.; Healy, S.P.; Unlu, I.; Crepeau, T.N.; Gaugler, R.; Fonseca, D.M.; Andreadis, T.G. Isolations of Cache Valley virus from <i>Aedes albopictus</i> (Diptera: Culicidae) in New Jersey and evaluation of its role as a regional arbovirus vector. <i>J. Med. Entomol.</i> 2013 , <i>50</i> , 1310–1314.
	-	+	Mitchell, C.J.; Haramis, L.D.; Karabatsos, N.; Smith, G.C.; Starwalt, V.J. Isolation of La Crosse, Cache Valley, and Potosi Viruses from <i>Aedes</i> Mosquitoes (Diptera: Culicidae) Collected at Used-Tire Sites in Illinois during 1994–1995. <i>J. Med. Entomol.</i> 1998 , <i>35</i> , 573–577.
EEEV	-	+	Mitchell, C.J.; Niebylski, M.L.; Smith, G.C.; Karabatsos, N.; Martin, D.; Mutebi, J.P.; Craig, G.B.J.; Mahler, M.J. Isolation of eastern equine encephalitis virus from <i>Aedes albopictus</i> in Florida. <i>Science</i> 1992 , <i>257</i> , 526–527.
KEYV	-	+	Mitchell, C.J.; Niebylski, M.L.; Smith, G.C.; Karabatsos, N.; Martin, D.; Mutebi, J.P.; Craig, G.B.J.; Mahler, M.J. Isolation of eastern equine encephalitis virus from <i>Aedes albopictus</i> in Florida. <i>Science</i> 1992 , <i>257</i> , 526–527.
	+	-	Westby, K.M.; Fritzen, C.; Paulsen, D.; Poindexter, S.; Moncayo, A.C. La Crosse Encephalitis Virus Infection in Field-Collected <i>Aedes albopictus</i> , <i>Aedes japonicus</i> , and <i>Aedes triseriatus</i> in Tennessee. <i>J. Am. Mosq. Control Assoc.</i> 2015 , <i>31</i> , 233–241.
LACV	-	+	Gerhardt, R.R.; Gottfried, K.L.; Apperson, C.S.; Davis, B.S.; Erwin, P.C.; Smith, A.B.; Panella, N.A.; Powell, E.E.; Nasci, R.S. First isolation of La Crosse virus from naturally infected <i>Aedes albopictus</i> . <i>Emerg. Infect. Dis.</i> 2001 , <i>7</i> , 807–811.
	-	+	Lambert, A.J.; Blair, C.D.; D’Anton, M.; Ewing, W.; Harborth, M.; Seiferth, R.; Xiang, J.; Lanciotti, R.S. La Crosse Virus in <i>Aedes albopictus</i> Mosquitoes, Texas, USA, 2009. <i>Emerg. Infect. Dis.</i> 2010 , <i>16</i> , 856–858.
	-	+	Mitchell, C.J.; Haramis, L.D.; Karabatsos, N.; Smith, G.C.; Starwalt, V.J. Isolation of La Crosse, Cache Valley, and Potosi Viruses from <i>Aedes</i> Mosquitoes (Diptera: Culicidae) Collected at Used-Tire Sites in Illinois during 1994–1995. <i>J. Med. Entomol.</i> 1998 , <i>35</i> , 573–577.
POTV	-	+	Francy, D.; Krabatsos, N.; Wesson, C.; Moore, J.; Lazwick, J.; Niebylski, T.; Tsai, T.; Craig, J. A new Arbovirus from <i>Aedes albopictus</i> , an Asian mosquito Established in the United States. <i>Science (80-.)</i> . 1990 , <i>250</i> , 1738–1740.
	-	+	Harrison, B.A.; Mitchell, C.J.; Apperson, C.S.; Smith, G.C.; Karabatsos, N.; Engber, B.R.; Newton, N.H. Isolation of potosi virus from <i>Aedes albopictus</i> in North Carolina. <i>J Am Mosq Control Assoc</i> 1995 , <i>11</i> , 225–229.
	-	+	Mitchell, C.J.; Haramis, L.D.; Karabatsos, N.; Smith, G.C.; Starwalt, V.J. Isolation of La Crosse, Cache Valley, and Potosi Viruses from <i>Aedes</i> Mosquitoes (Diptera: Culicidae) Collected at Used-Tire Sites in Illinois during 1994–1995. <i>J. Med. Entomol.</i> 1998 , <i>35</i> , 573–577.
TENV	-	+	Mitchell, C.J.; Niebylski, M.L.; Smith, G.C.; Karabatsos, N.; Martin, D.; Mutebi, J.P.; Craig, G.B.J.; Mahler, M.J. Isolation of eastern equine encephalitis virus from <i>Aedes albopictus</i> in Florida. <i>Science</i> 1992 , <i>257</i> , 526–527.
USUV	+	-	Puggioli, A.; Bonilauri, P.; Calzolari, M.; Lelli, D.; Carrieri, M.; Urbanelli, S.; Pudar, D.; Bellini, R. Does <i>Aedes albopictus</i> (Diptera: Culicidae) play any role in Usutu virus transmission in Northern Italy? Experimental oral infection and field evidences. <i>Acta Trop.</i> 2017 , <i>172</i> , 192–196.
	+	-	Cupp, E.W.; Hassan, H.K.; Yue, X.; Oldland, W.K.; Lilley, B.M.; Unnasch, T.R. West Nile Virus Infection in Mosquitoes in the Mid-South USA, 2002–2005. <i>J. Med. Entomol.</i> 2007 , <i>44</i> , 117–125.
WNV	+	-	Farajollahi, A.; Kesavaraju, B.; Price, D.C.; Williams, G.M.; Healy, S.P.; Gaugler, R.; Nelder, M.P. Field efficacy of BG-Sentinel and industry-standard traps for <i>Aedes albopictus</i> (Diptera: Culicidae) and West Nile virus surveillance. <i>J. Med. Entomol.</i> 2009 , <i>46</i> , 919–25.
	+	-	Holick, J.; Kyle, A.; Ferraro, W.; Delaney, R.; Marta, I. Discovery of <i>Aedes albopictus</i> infected with west nile virus in Southeastern Pennsylvania. <i>J. Am. Mosq. Control Assoc. INC.</i> 2002 , <i>18</i> , 131.

CCV, Cache Valley virus; EEEV, Eastern Equine Encephalomyelitis virus; KEYV, Keystone virus; LACV, La Crosse virus; POTV, Potosi virus; TENV, Tensaw virus; USUV, Usutu virus; WNV, West Nile virus.

Table S6. Geographic distribution, vertebrate hosts and potential vectors of arboviruses isolated or tested for vector competence in *Ae. albopictus*. Information on hosts and virus isolation from field-collected mosquitoes are from the arbocat database (<https://wwwn.cdc.gov/arbocat>). Arbovirus isolated (+) or not (-) from *Ae. albopictus*.

Virus	Virus Family	Virus Genus	Geographic Distribution	Natural Host Family	Arthropod Isolation
Cache Valley(+)	Peribunyaviridae	Orthobunyavirus	Canada USA Jamaica	Atelidae	<i>Aedes sollicitans</i>
				Canidae	<i>Aedes taeniorhynchus</i>
				Cervidae	<i>Anopheles crucians</i>
				Equidae	<i>Anopheles grabhamii</i>
				Humans	<i>Anopheles quadrimaculatus</i>
				Procyonidae	<i>Culiseta inornata</i>
				Other unidentified animals	<i>Psorophora confinnis</i>
				Eastern equine encephalomyelitis(+)	Togaviridae
Canidae	<i>Aedes fulvus pallens</i>				
Cricetidae	<i>Aedes mitchellae</i>				
Didelphidae	<i>Aedes sollicitans</i>				
Echimyidae	<i>Aedes sticticus</i>				
Equidae	<i>Aedes taeniorhynchus</i>				
Humans	<i>Aedes vexans</i>				
Muridae	<i>Anopheles crucians</i>				
Phasianidae	<i>Coquillettidia perturbans</i>				
Other unidentified animals	<i>Culex nigripalpus</i>				
	<i>Culex quinquefasciatus</i>				
	<i>Culex restuans</i>				
	<i>Culex salinarius</i>				
	<i>Culex taeniopus</i>				
	<i>Culiseta melanura</i>				
	<i>Dermanyssus gallinae</i>				
	<i>Eomenocanthus stramineus</i>				
	<i>Eusimulium johannseni</i>				
	<i>Simulium meridionale</i>				
Keystone(+)	Peribunyaviridae	Orthobunyavirus	USA	Cervidae	<i>Aedes atlanticus</i>
				Cricetidae	<i>Aedes aurifer</i>
				Other unidentified animals	<i>Aedes infirmatus</i>
					<i>Aedes taeniorhynchus</i>
					<i>Anopheles crucians</i>
	<i>Culex nigripalpus</i>				
La Crosse(+)	Peribunyaviridae	Orthobunyavirus	USA	Humans Sciuridae	Unknown
Potosi(+)	Peribunyaviridae	Orthobunyavirus	USA	Unknown	Unknown
Tensaw(+)	Peribunyaviridae	Orthobunyavirus	USA	Bovidae	<i>Aedes atlanticus</i>
				Bovidae	<i>Aedes atlanticus</i>
				Canidae	<i>Aedes infirmatus</i>
				Cricetidae	<i>Aedes mitchellae</i>
				Humans	<i>Aedes taeniorhynchus</i>
				Leporidae	<i>Anopheles crucians</i>
				Phasianidae	<i>Anopheles punctipennis</i>
				Procyonidae	<i>Anopheles quadrimaculatus</i>
				Other unidentified animals	<i>Culex nigripalpus</i>
					<i>Culex salinarius</i>
					<i>Mansonia perturbans</i>
					<i>Psorophora confinnis</i>

Table S6. Continued

Virus	Virus Family	Virus Genus	Geographic Distribution	Natural Host Family	Arthropod Isolation
West Nile(+)	Flaviviridae	Flavivirus	Mozambique	Camelidae	<i>Aedes cantans</i>
			Cent. Af. Rep.	Equidae	<i>Anopheles coustani</i>
			Congo	Humans	<i>Anopheles subpictus</i>
			Egypt	Muridae	<i>Anopheles maculipennis</i>
			Uganda	Other	<i>Argas hermanni</i>
			Nigeria	unidentified	<i>Coquillettidia metallica</i>
			Turkey	animals	<i>Culex antennatus</i>
			Cyprus		<i>Culex modestus</i>
			Israel		<i>Culex quinquefasciatus</i>
			France		<i>Culex univittatus</i>
			Portugal		<i>Culex vishmii</i>
			Eastern Europe		<i>Culex weschei</i>
			Albania		<i>Hyalomma plumbeum</i>
			India		
			Malaysia		
Philippines					
Thailand					
Arumowot(-)	Phenuiviridae	Phlebovirus	Cent. Af. Rep.	Muridae	<i>Culex antennatus</i>
			Ethiopia	Turdidae	<i>Culex rubinotus</i>
			Kenya	Other	<i>Mansonia uniformis</i>
			Nigeria	unidentified	
			Rhodesia	animals	
South Africa					
Sudan					
Bujaru(-)	Phenuiviridae	Phlebovirus	Brazil	Echimyidae	
Bussuquara(-)	Flaviviridae	Flavivirus	Brazil	Atelidae	<i>Culex (Mel) sp.</i>
			Colombia	Echimyidae	<i>Culex (Mel) taeniopus</i>
			Panama	Humans	<i>Culex crybda</i>
				Other	<i>Culex sp.</i>
				unidentified	<i>Culex vomerifer</i>
					<i>Mansonia titillans</i>
		<i>Trichoprosopon sp.</i>			
Chandipura(-)	Rhabdoviridae	Vesiculovirus	India	Bovidae	<i>Phlebotomus sp.</i>
			Nigeria	Camelidae	
				Cercopithecoid	
				Equidae	
				Erinaceidae	
				Humans	
Chilibre(-)	Phenuiviridae	Phlebovirus	Panama	Unknown	<i>Lutzomyia ssp.</i>
					<i>Lutzomyia trapidoi</i>
					<i>Lutzomyia ylephilatrix</i>
Getah(-)	Togaviridae	Alphavirus	Australia	Humans	<i>Aedes vexans nip.</i>
			Cambodia		<i>Anopheles amictus</i>
			Japan		<i>Culex (Culex) gelidus</i>
			Malaysia		<i>Culex tritaeniorhynchus</i>
			Philippines		<i>Culex vishmii</i>

Table S6. Continued

Virus	Virus Family	Virus Genus	Geographic Distribution	Natural Host Family	Arthropod Isolation
Icoaraci(-)	Phenuiviridae	Phlebovirus	Brazil	Bovidae Cricetidae Echimyidae	Unknown
Ilheus(-)	Flaviviridae	Flavivirus	Argentina Brazil Colombia Guatemala Honduras Panama Trinidad	Bovidae Equidae Suidae Humans Other unidentified	<i>Aedes</i> spp. <i>Coquillettidia</i> spp. <i>Culex</i> spp. <i>Haemagogus</i> spp. <i>Psorophora</i> spp. <i>Sabethes</i> spp. <i>Trichopropion</i> spp. <i>Wyeomia</i> spp.
Itaporanga(-)	Phenuiviridae	Phlebovirus	Brazil French Guiana Trinidad	Didelphidae Humans Muridae Other unidentified	<i>Coquillettidia venezuelensis</i> <i>Culex caudelli</i> <i>Culex eastor</i>
Jamestown canyon(-)	Peribunyaviridae	Orthobunyavirus	USA Canada	Cervidae Humans	<i>Aedes cantator</i> <i>Aedes vexans</i> <i>Aedes abserratus</i> <i>Aedes communis</i> <i>Aedes</i> spp. <i>Aedes stimulans</i> <i>Aedes triseriatus</i> <i>Chrysops cincticornis</i> <i>Culiseta inornata</i>
Japanese Encephalitis(-)	Flaviviridae	Flavivirus	Easter Asia	Ardeidae Equidae Hipposiderida Humans Mnipteridae Suidae	<i>Anopheles barbirostris</i> <i>Anopheles hyrcanus</i> <i>Culex annulus</i> <i>Culex fuscocephala</i> <i>Culex gelidus</i> <i>Culex pipiens</i> <i>Culex tritaeniorhynchus</i> <i>Culex vishnui</i>
Karimabad(-)	Phenuiviridae	Phlebovirus	Bangladesh Eastern Europe Egypt Iran Pakistan Sudan	Bovidae Columbidae Humans Muridae Phasianidae	<i>Phlebotomus</i> spp
Keystone(-)	Peribunyaviridae	Orthobunyavirus	USA	Unknown	<i>Chrysops obsoletus</i>
Kokobera(-)	Flaviviridae	Flavivirus	Australia New Guinea	Bovidae Equidae Macropodidae Humans	<i>Aedes</i> spp. <i>Aedes vigilax</i> <i>Culex annulirostris</i>
Kujin(-)	Flaviviridae	Flavivirus	Australia Borneo Indonesia Malaysia New Guinea Sarawak	Bovidae Equidae Humans Oriolidae Phasianidae	<i>Culex annulirostris</i> <i>Culex pseudovishnui</i> <i>Culex squamosus</i>

Table S6. Continued

Virus	Virus Family	Virus Genus	Geographic Distribution	Natural Host Family	Arthropod Isolation
Mayaro(-)	Togaviridae	Alphavirus	USA	Callitrichidae	<i>Culex spp.</i>
			Bolivia	Cebidae	<i>Gigantolaelaps sp.</i>
			Brazil	Columbidae	<i>Haemagogus spp.</i>
			Colombia	Didelphidae	<i>Mansonia venezuelensis</i>
			Guyana	Humans	<i>Psorophora spp.</i>
			Surinam	Pitheciidae	<i>Sabethini spp.</i>
			Trinidad	Telidae	
			Venezuela	Tropiduridae Other unidentified	
Oropuche(-)	Peribunyaviridae	Orthobunyavirus	Brazil	Atelidae	<i>Aedes serratus</i>
			Colombia	Bradypodidae	<i>Culex quinquefasciatus</i>
			Trinidad	Cebidae	<i>Culicoides paranensis</i>
				Humans	<i>Mansonia venezuelensis</i>
				Other unidentified	
Orungo(-)	Reoviridae	Orbivirus	Cent. Af. Rep.	Bovidae	<i>Aedes aegypti</i>
			Central African	Camelidae	<i>Aedes dentatus</i>
			Ivory Coast	Erinaceidae	<i>Anopheles funestus</i>
			Nigeria	Humans	<i>Anopheles gambiae</i>
			Senegal	Soricidae	<i>Culex perfuscus</i>
			Sierra Leone		<i>Culicoides sp.</i>
			Uganda		
Pacui(-)	Phenuiviridae	Phlebovirus	Brazil	Cricetidae	<i>Lutzomyia flaviscutellata</i>
			Trinidad	Didelphidae	<i>Lutzomyia infraspinosa</i>
				Echimyidae	
				Other unidentified	
Rift Valley Fever(-)	Phenuiviridae	Phlebovirus	Egypt	Bovidae	<i>Aedes (Neomelanicionion)</i>
			Kenya	Hipposiderid	<i>Aedes (Ochlerotatus) caballus</i>
			Nigeria	Humans	<i>Culex (Culex) theileri</i>
			South Africa	Pteropodidae	<i>Culicoides spp.</i>
			Sudan	Other	<i>Eretmapodites chrysogaster</i>
			Uganda		
Ross River(-)	Togaviridae	Alphavirus	American Samoa	Canidae	<i>Aedes normanensis</i>
			Australia	Equidae	<i>Aedes vigilax</i>
			New Guinea	Humans	<i>Aedes vigilax</i>
				Macropodida	<i>Anopheles amictus</i>
				Monarchidae	<i>Culex annulirostris</i>
				Petrociidae	<i>Culex spp.</i>
				Suidae	<i>Mansonia uniformis</i>
Saint Louis encephalitis(-)	Phenuiviridae	Flavivirus	Argentina	Anatidae	<i>Aedes dorsalis</i>
			Brazil	Apodidae	<i>Anopheles crucians</i>
			Canada	Canidae	<i>Culex caudelli</i>
			Haiti	Cardinalidae	<i>Culex coronator</i>
			Jamaica	Columbidae	<i>Culex nigripalpus</i>
			Mexico	Turdidae	<i>Culex peus</i>
			Surinam	Corvidae	<i>Culex pipiens</i>
			Trinidad	Cricetidae	<i>Culex quinquefasciatus</i>
			USA	Felidae	<i>Culex spissipes</i>
			Venezuela	Humans	<i>Culex taeniopus</i>
				Mimidae	<i>Culex tarsalis</i>
				Passeridae	<i>Culex virgultus</i>
				Pipridae	<i>Deinocerites pseudes</i>

Table S6. Continued

Virus	Virus Family	Virus Genus	Geographic Distribution	Natural Host Family	Arthropod Isolation
Saint Louis encephalitis(-) (continued)				Thraupidae Other unidentified	<i>Dermacentor variabilis</i> <i>Dermanyssus americanus</i> <i>Dermanyssus gallinae</i> <i>Gigantolaelaps</i> sp. <i>Ornithonyssus sylviarum</i> <i>Psorophora ferox</i> <i>Sabethes belisarioi</i> <i>Sabethes chloropterus</i> <i>Sabethes</i> ssp. <i>Trichopropon</i> sp. <i>Wyeomia</i> sp.
Salehabad(-)	Phenuiviridae	Phlebovirus	Bangladesh Pakistan Iran	Bovidae Columbidae Humans Muridae Passeridae Phasianidae	<i>Phlebotomus</i> spp.
San Angelo(-)	Peribunyaviridae	Orthobunyavirus	USA	Canidae Cervidae Didelphidae Humans Phasianidae Procyonidae	<i>Anopheles pseudopunctipennis</i> <i>Psorophora signipennis</i>
Trivittatus(-)	Peribunyaviridae	Orthobunyavirus	USA	Cricetidae Cervidae	<i>Aedes atlanticus</i> <i>Aedes infirmatus</i> <i>Aedes taeniorhynchus</i> <i>Aedes triseriatus</i> <i>Aedes trivittatus</i> <i>Aedes vexas</i> <i>Culex pipiens</i> <i>Mansonia perturbans</i>
Uganda S(-)	Flaviviridae	Flavivirus	Cent. Af. Rep. Egypt Gambia India Indonesia Malaysia Mozambique Nigeria Senegal South Africa Tanganyika Uganda	Muscicapidae Humans Other unidentified	<i>Aedes ingrami</i> <i>Aedes longipalpis</i> <i>Aedes natronius</i>
Urucuri(-)	Phenuiviridae	Phlebovirus	Brazil	Dasyproctidae Cricetidae Echimyidae	Unknown
Usutu (+)	Flaviviridae	Flavivirus	South Africa Uganda Nigeria Cent. Af. Rep. Senegal Cameroun	Man Bovidae Muridae Bucerotidae Pycnonotidae Turdidae	<i>Culex neavei</i> <i>Coquillettidia aurites</i> <i>Culex pipens</i> <i>Culex perfuscus</i> <i>Culex perexiguus</i> <i>Mansonia africana</i> <i>Aedes caspius</i>

Table S7. List of references used to analyse the vector competence of several mosquito-virus pairs: *Aedes aegypti**CHIKV, *Aedes aegypti**DENV-1, *Aedes aegypti**DENV-2, *Aedes aegypti**ZIKV, *Aedes albopictus**CHIKV, *Aedes albopictus**DENV_1, *Aedes albopictus**DENV_2, *Aedes albopictus**ZIKV, *Culex pipiens**WNV, and *Haemagogus leucocelenus**YFV.

Mosquito-virus pairs

*Ae. aegypti**CHIKV

Paupy, C.; Ollomo, B.; Kamgang, B.; Moutailler, S.; Rousset, D.; Demanou, M.; Hervé, J.-P.; Leroy, E.; Simard, F. Comparative role of *Aedes albopictus* and *Aedes aegypti* in the emergence of Dengue and Chikungunya in central Africa. *Vector Borne Zoonotic Dis.* **2010**, *10*, 259–266.

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*Ae. aegypti**DENV-1

Calvez, E.; Guillaumot, L.; Girault, D.; Richard, V.; O'Connor, O.; Paoaafaite, T.; Teurlai, M.; Pocquet, N.; Cao-Lormeau, V.M.; Dupont-Rouzeyrol, M. Dengue-1 virus and vector competence of *Aedes aegypti* (Diptera: Culicidae) populations from New Caledonia. *Parasites and Vectors* **2017**, *10*, 1–8.

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*Ae. aegypti**DENV-2

Chepkorir, E.; Lutomiah, J.; Mutisya, J.; Mulwa, F.; Limbaso, K.; Orindi, B.; Ng'Ang'a, Z.; Sang, R. Vector competence of *Aedes aegypti* populations from Kilifi and Nairobi for dengue 2 virus and the influence of temperature. *Parasites and Vectors* **2014**, *7*, 435.

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*Ae. aegypti**ZIKV

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Table S7. Continued

Mosquito-virus pairs
<i>Ae. aegypti</i>*ZIKV (continued)
Heitmann, A.; Jansen, S.; Lühken, R.; Leggewie, M.; Badusche, M.; Pluskota, B.; Becker, N.; Vapalahti, O.; Schmidt-Chanasit, J.; Tannich, E. Experimental transmission of zika virus by mosquitoes from central Europe. <i>Eurosurveillance</i> 2017 , <i>22</i> , 14–17.
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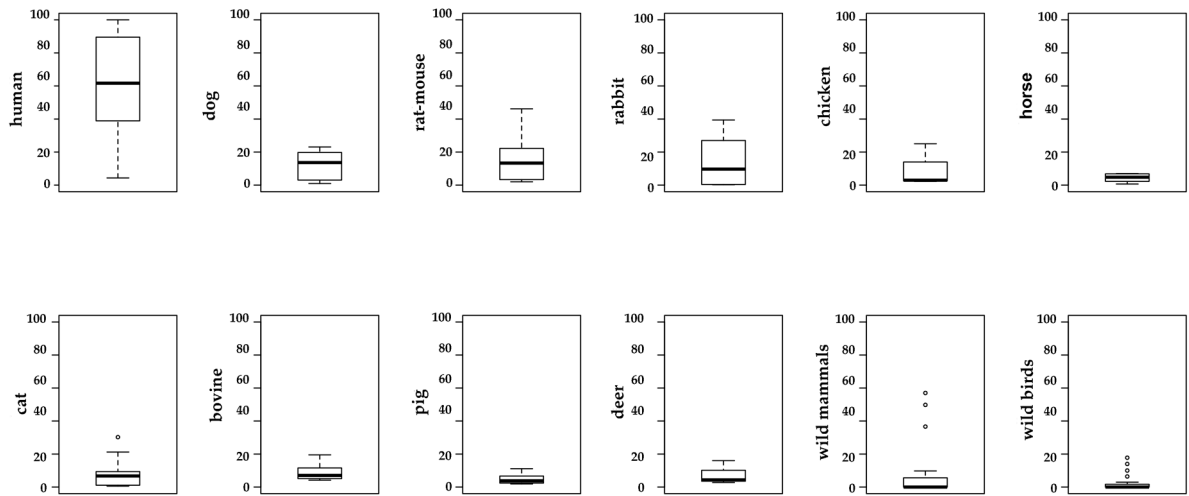
Table S7. Continued

Mosquito-virus pairs
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<p><i>Aedes albopictus</i>*DENV-2 Amraoui, F.; Aayed, W. Ben; Madec, Y.; Faraj, C.; Himmi, O.; Btissam, A.; Sarih, M.; Failloux, A.B. Potential of aedes albopictus to cause the emergence of arboviruses in Morocco. <i>PLoS Negl. Trop. Dis.</i> 2019, <i>13</i>, e0006997.</p> <p>Brustolin, M. et al. Experimental study of the susceptibility of a European <i>Aedes albopictus</i> strain to dengue virus under a simulated Mediterranean temperature regime. <i>Med. Vet. Entomol.</i> 2018, <i>32</i>, 393–398.</p> <p>Paupy, C. et al. Comparative role of <i>Aedes albopictus</i> and <i>Aedes aegypti</i> in the emergence of Dengue and Chikungunya in central Africa. <i>Vector Borne Zoonotic Dis.</i> 2010, <i>10</i>, 259–266.</p>
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Table S7. Continued

Mosquito-virus pairs
<i>Culex pipiens</i>*WNV (continued)
Vanlandingham, D.L.; McGee, C.E.; Klinger, K.A.; Vessey, N.; Fredregillo, C.; Higgs, S. Short report: Relative susceptibilities of South Texas mosquitoes to infection with West Nile virus. <i>Am. J. Trop. Med. Hyg.</i> 2007 , <i>77</i> , 925–928.
Vaidyanathan, R.; Fleisher, A.E.; Minnick, S.L.; Simmons, K.A.; Scott, T.W. Nutritional Stress Affects Mosquito Survival and Vector Competence for West Nile Virus. <i>Vector-Borne Zoonotic Dis.</i> 2008 , <i>8</i> , 727–732.
<i>Haemagogus leucocelenu</i>*YFV
Couto-Lima, Di.; Madec, Y.; Bersot, M.I.; Campos, S.S.; Motta, M.D.A.; Dos Santos, F.B.; Vazeille, M.; Da Costa Vasconcelos, P.F.; Lourenço-De-Oliveira, R.; Failloux, A.B. Potential risk of re-emergence of urban transmission of Yellow Fever virus in Brazil facilitated by competent <i>Aedes</i> populations. <i>Sci. Rep.</i> 2017 , <i>7</i> , 4848.

Figure S1. Analysis of the host feeding patterns of *Ae. albopictus* for the different species of domestic animals without taking into account the host availability. Black line: median.

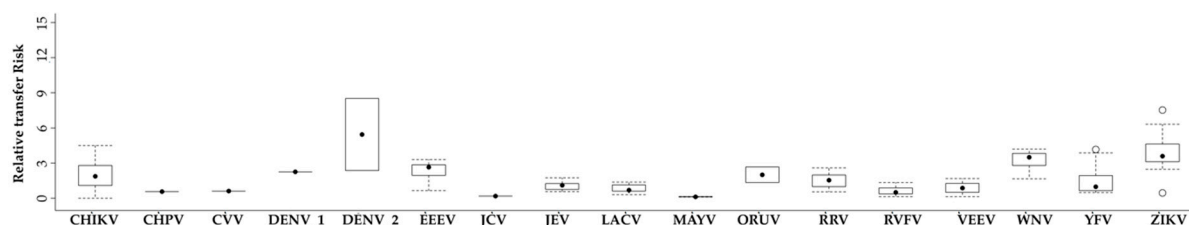


Supplementary Material and Methods.

Estimating the relative virus transfer risk mediated by *Ae. albopictus*.

We calculated the relative transfer risk of viruses mediated by *Ae. albopictus* from their natural host to humans by adapting Kilpatrick's equation: $Risk = A \times P \times Fm \times Cv$, where A corresponds to the mosquito abundance, P is the virus infection prevalence, Fm is the fraction of blood meals taken from mammals, and Cv corresponds to the fraction of infected mosquitoes that can transmit the virus in a subsequent bite. In our study and for all viruses, information on A and P were lacking, but as these two parameters are directly proportional fractions of the formula (i.e. the more we increase their value, the greater the risk is) we assumed that the absence of information should not affect the result of the risk assessment. We decomposed Fm in FAi (i.e. the mean relative frequency *Ae. albopictus* bites in vertebrate host families) and FHi (i.e. the mean relative frequency of *Ae. albopictus* bites in humans). We then calculated the average FAi and FHi using the information on *Ae. albopictus* feeding behavior extracted from the articles identified in our literature search. For such purpose, we listed all the natural vertebrate hosts of each virus using the information available in the CDC catalogue and then classified hosts according to their family. We calculated the mean frequency of *Ae. albopictus* biting for each family, when data were available. Cv is equal to the Transmission efficiency (TRi). In conclusion, for each virus in the present study, the risk assessment was calculated as $TRi \times FAi \times FHi$. The results are presented in Figure S2, but are not included in the main text due to the uncertainties in the calculation because of the limitations of the representative data on feeding behaviour, arbovirus hosts and vector competence to produce quantitative estimations and extrapolations

Figure S2. Relative virus transfer risk by *Ae. albopictus* for 13 enzootic viruses experimentally proven to be transmitted by *Ae. albopictus* and for four epidemic arboviruses (DENV-1, DENV-2, ZIKAV, and CHIKV).



CHIKV, Chikungunya virus; CHPV, Chandipura virus; CVV, Cache Valley virus; DENV-1, Dengue virus serotype 1; DENV-2, Dengue virus serotype 2; EEEV, Eastern equine encephalomyelitis virus; JCV, Jamestown Canyon virus; JEV, Japanese encephalitis virus; LACV, La Crosse virus; MAYV, Mayaro virus; ORUV, Orungo virus; RRV, Ross River virus; RVFV, Rift Valley fever virus; VEEV, Venezuelan equine encephalitis virus; WNV, West Nile virus; YFV, Yellow fever virus; and ZIKV, Zika virus.

