Supplementary Information

Pivotal role of CD103 in the development of psoriasiform dermatitis

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Supplementary Table 1

RT-PCR primer	Sequence
Gapdh	F-5'-AAATTCAACGGCACAGTCAAG-3'
	R-5'-TGGTGGTGAAGACACCAGTAG-3'
Illa	F-5'-TCCAGGGCAGAGAGGGAGT-3'
	R-5'-GGAACTTTGGCCATCTTGATTT-3'
Il1b	F-5'-GAAGAAGAGCCCATCCTCTG-3'
	R-5'-TCATCTCGGAGCCTGTAGTG-3'
Il6	F-5'-GACTGATGCTGGTGACAACC-3'
	R-5'-CCTCCGACTTGTGAAGTGG-3'
1110	F-5'-TGCAGCAGCTCAGAGGGTT-3'
	R-5'-TGGCCACAGTTTTCAGGGAT-3'
Il12a	F-5'-AAACCAGACCCGCCCAAGAAC-3'
	R-5'-AAAAAGCCAACCAAGCAGAAGACAG-3'
Il17a	F-5'-CTGCTGAGCCTGGCGGCTAC-3'
	R-5'-CATTGCGGTGGAGAGTCCAGGG-3'
1119	F-5'-GCCAACTCTTTCCTCTGCGT-3'
	R-5'-GGTGGCTTCCTGACTGCAGT-3'
<i>Il20</i>	F-5'-GACCCCTGACCACCATACCC-3'
	R-5'-CCATTGCTTCTTCCCCACAA-3'
<i>Il22</i>	F-5'-CAGCTCCTGTCACATCAGCGGT-3'
	R-5'-AGGTCCAGTTCCCCAATCGCCT-3'
Il23a	F-5'-TCCGTTCCAAGATCCTTCG-3'
	R-5'-GAACCTGGGCATCCTTAAGC-3'
Tnf	F-5'-GCCCACGTCGTAGCAAACCAC -3'
	R-5'-GCAGGGGCTCTTGACGGCAG -3'
Cxcl1	F-5'-GCCTATCGCCAATGAGCTG-3'
	R-5'-ATTCTTGAGTGTGGCTATGA-3'
Cxcl2	F-5'-ATGCCTGAAGACCCTGCCAAG-3'
	R-5'-GGTCAGTTAGCCTTGCCTTTG-3'
S100a7	F-5'-GGGCAGCTGACAAAAACAAG-3'
	R-5'-TGGAACTGGAGATGGTAGTCC-3'
S100a8	F-5'-CCATGCCCTCTACAAGAATG-3'
	R-5'-ATCACCATCGCAAGGAACTC-3'



Supplementary Figure 1. Generation and identification of $Cd103^{+}$ mice. (a) Strategy used to produce the $Cd103^{+}$ mice. (1) Partial restriction map of the WT Cd103 gene. Exons are depicted as black boxes. The restriction site indicated is S: *SpeI* (2) Targeting vector used to introduce the mutations in the Cd103 gene. A *Sal*I site engineered in place of the start codon in exon 1 of the Cd103 gene was used to clone the *FRT-PGK-gb2-Neo-FRT-Stop* cassette. DTa: diphtheria toxin a expression cassette, Stop: Stop cassette, Neo: Neo cassette. The Neo cassette is shown bracketed by FRT sites (filled triangles). (3) Structure of the targeted allele following homologous ESC clones. (4) Structure of the Cd103 allele following expression of FLP recombinase and excision of the Neocassette in mutant mice. The 3' external single-copy probe (a hatched box) and the PCR primers at the 5' end (blackarrows) used to verify proper homologous recombination events are shown. (**b**,**c**) DNA-PCR (**b**) and Southern blot (**c**) analysis of WT and recombinant ESC clones. (**d**) Genotyping of tail DNA from WT mice and homozygous mice for the Cd103 allele by DNA-PCR. The results are representative of at least three independent experiments.

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Supplementary Figure 2. Constituencies of leukocytes in PLNs in $Cd103^{+}$ mice. The frequency of the indicated leukocytes in skin-draining PLNs was analyzed by flow cytometry in WT mice (n=3) and $Cd103^{+}$ mice (n=3). Data are the mean ± s.d. from three individual samples in a single experiment. The results are representative of at least three independent experiments.



Supplementary Figure 3. Constituencies of splenic leukocytes in $Cd103^{\pm}$ mice. The frequency of the indicated leukocytes in Spl was analyzed by flow cytometry in WT mice (n=3) and $Cd103^{\pm}$ mice (n=3). Data are the mean ± s.d. from three individual samples in a single experiment. **P* < .05 compared with WT mice. The results are representative of at least three independent experiments.



Supplementary Figure 4. Absence of CD103 enhances the accumulation of inflammatory leukocytes in skin-draining PLNs. The absolute cell numbers of leukocytes in the skin-draining PLNs at day 6 after topical application of IMQ on the left ear skin every day for 6 days in WT mice (n=5) and *Cd103* mice (n=5). Data are the mean \pm s.d. in three to four individual samples in a single experiment. **P* < .05,***P* < .01 compared with WT mice. The results are representative of at least three independent experiments.



Supplementary Figure 5. CD103 deficiency enhances the TLR7-mediated activation of cDCs. (**a**) Splenic cDCs derived from WT mice and $Cd103^{\circ}$ mice were not stimulated (None) or stimulated with IMQ (IMQ). The transcriptional expressions of cytokines in cDCs. Data are the mean ± s.d. from three to six individual samples in a single experiment. **P < .01 compared with WT mice. (**b**) CD8[,] T cells were cultured with cDCs obtained from the skin-draining PLNs in WT mice and $Cd103^{\circ}$ mice for 5 days under T_c17-polarized culture conditions. The frequency of IL-17A- and IFN- γ -producing cells among CD8[,] T cells. Data are presented as a contour plot, and numbers mean the proportion of the indicated cell populations in each gate (left panel). Data are the mean ± s.d. in three individual samples in a single experiment (right panel). *P < .05 compared with WT mice. The results are representative of at least three independent experiments.



Supplementary Figure 6. Staining of IL-17A-prodcing lymphocytes with isotype control mAb. WT mice (n=3) and $Cd103^{-1}$ mice (n=3) were treated topically with IMQ on the left ear skin every day for 6 days. Data are presented as a contour plot for isotype control staining (cont. Ig) of innate lymphocytes (**a**) and CD8⁺ T cells (**b**) in the skin-

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draining PLNs. The results are representative of at least three independent experiments.

b а □ WT ,■ *Cd103*^{,,}-□ WT ■ *Cd103^{/-}* Lin-CD90+IL-17A⁺ cells (\times 10⁵) $\gamma \delta T C R^{+} I L^{-} 17 A^{+} T$ cells ($\times 10^{5})$ 0.25 2.0 0.20 1.5 0.15 1.0 0.10 0.5 0.05 0 0 None None INO INO d С □ WT 0.51 **□ Cd103**^{/-} □ WT ■ *Cd103*⁴⁻ 2.0 CD4+IL-17A+T cells (\times 10⁵) CD8+IL-17A+T cells (\times 10⁵) 0.4 1.5 0.3 1.0 0.2 0.5 0.1 0 0 INO None None INO f е □ WT ■ *Cd103*^{/-} □ WT ■ *Cd103*^{-/-} $\gamma \delta TCR^{+}IL^{-}22^{+}T$ cells ($\times 10^{5}$) 0.6 Lin-CD90+IL-22+ cells ($\times 10^5$) 1.5 0.4 1.0 0.2 0.5 INO n 0 INO None None g h WT *Cd103*≁ □ WT ■ *Cd103*^{-/-} 0.4 0.4 CD4+IL-22+T cells (imes 10⁵) CD8+IL-22+T cells (\times 10⁵) 0.3 0.3 0.2 0.2 0.1 0.1 0┘ 0 INO None INO None

Supplementary Figure 7. CD103 deficiency enhances the generation of IL-17A-prodcing lymphocytes in the skin-draining PLNs in the progression of psoriasiform skin

inflammation. WT mice (n=3) and $Cd103^{+}$ mice (n=3) were treated topically with IMQ on the left ear skin every day for 6 days. The absolute cell numbers of IL-17A-producing cells (**a-d**) and IL-22-producing cells (**e-h**) among innate lymphocytes (**a,e**), $\gamma\delta$ TCR⁺ T cells (**b,f**), CD4⁺ T cells (**c,g**), and CD8⁺ T cells (**d,h**) in the skin-draining PLNs at days 0 and 6. Data are the mean \pm s.d. in three individual samples in a single experiment. **P* < .05, ***P* < .01compared with WT mice. The results are representative of at least three independent experiments.



Supplementary Figure 8. CD103 deficiency promotes the generation of IL-17Aprodcing lymphocytes in the dermal tissues in the development of psoriasiform skin inflammation. WT mice (n=3) and $Cd103^{\circ}$ mice (n=3) were treated topically with IMQ on the left ear skin every day for 6 days. The frequencies of IL-17A-producing cells and IL-22-producing cells CD45.2 leukocytes in the dermal tissues at days 0 and 6. Data are the mean \pm s.d. in three individual samples in a single experiment. **P* < .05, ***P* < .01compared with WT mice. The results are representative of at least three independent experiments.



Supplementary Figure 9. Influence of CD103 deficiency on the generation of T_c17 cells. CD8[•] T cells derived from WT mice and *Cd103⁺* mice were cultured for 5 days under T_c17 -polarized culture conditions. The frequency of IL-17A- and IFN- γ -producing cells among CD8⁺ T cells. Data are presented as a contour plot, and numbers mean the proportion of the indicated cell populations in each gate (left panel). Data are the mean \pm s.d. in three to five individual samples in a single experiment (right panel). ***P* < .01 compared with WT mice. The results are representative of at least three independent experiments.





Figure S1c



Figure S1d



Supplementary Figure 10. Full-length gels and blots of Supplementary Figure 1b-d.