

Adapter Proteins for Opposing Motors Interact Simultaneously with Nuclear Pore Protein Nup358

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SUPPORTING TABLES

Table S1 Molar masses (MW) determined by SEC-MALS for Fig. 5

Protein	Conc. (mg/ml)	MW (kDa) Peak	Conc. at Peak* (mg/ml)	Replicates	Fig. Panel
Nup358-min	5	10.6 ±0.5	1.0 ±0.03	3	A
Nup358-min	1	10.6 ±0.5	0.2 ±0.1	3	
Nup358-min/W2224A/D2225A	5	10.0 ±0.5	1.3 ±0.2	3	B
Nup358-min/W2224A/D2225A	1	11.2 ±0.6	0.3 ±0.1	2	
KLC2 ^{TPR-trunc}	3	37.9 ±1.9	0.1 ±0.004	2	C
Nup358-min + KLC2 ^{TPR}	4	70.2 ±3.5	0.2 ±0.06	4	D
Nup358-KLC2-fusion	4	68.9 ±3.4	0.4 ±0.01	3	E
Nup358-min + KLC2 ^{TPR}	4	70.2 ±3.5	0.2 ±0.06	4	
Nup358-KLC2-fusion	8	72.1 ±3.6	0.7 ±0.01	2	F
Nup358-KLC2-fusion	1	56.3 ±2.8	0.1 ±0.01	2	
Nup358/W2224A/D2225A-KLC2	8	45.1 ±2.3	1.1 ±0.01	2	
Nup358/W2224A/D2225A-KLC2	1	43.2 ±2.2	0.1 ±0.02	3	

* Protein concentration at the apex of the elution peak, determined by the refractive index and averaged from all replicates; standard deviations are shown. Note that in panels D and E the same dataset of Nup358-min + KLC2^{TPR} is shown.

Table S2 Molar masses (MW) determined by SEC-MALS for Fig. 7

Protein	Conc. (mg/ml)	MW (kDa) Peak 1 & 2 & 3	Conc. at Peak 1 & 2 & 3*	Replicates	Fig. Panel
Nup358-min + KLC2 ^{TPR} + BicD2-CTD	5	125.1 ±6.3	0.1 ±0.02	3	A
Nup358-min + KLC2 ^{TPR}	4	70.2 ±3.5	0.2 ±0.06	4	
Nup358-KLC2-fusion + BicD2-CTD	5	117.4 ±5.9	0.1 ±0.02	3	B
Nup358-KLC2-fusion	4	68.9 ±3.4	0.4 ±0.01	3	
Same data as in panels A & B					C
Nup358/W2224A/ D2225A-KLC2 + BicD2- CTD	5	113.4 ±5.7 & 53.7 ±2.7 & 43.2 ±2.2	0.02 ±0.002 & 0.2 ±0.08 & 0.2 ±0.05	2	D
Nup358/W2224A/ D2225A-KLC2	4	44.0 ±2.2	0.6 ±0.1	2	

* Protein concentration at the apex of the elution peaks 1 & 2 & 3 in mg/ml, determined by the refractive index and averaged from all replicates; standard deviations are shown. Elution peaks are numbered from highest to lowest molar mass. Conc.: Protein concentration. Note that for panels A and B the datasets of Nup358-min + KLC2^{TPR} and Nup358-KLC2-fusion from Fig. 5 are shown again.

Table S3 Molar masses (MW) determined by SEC-MALS for Fig. 8

Protein	Conc. injected (mg/ml)	MW (kDa) Peak 1	MW (kDa) Peak 2	Conc. at Peak 1 & 2 (mg/ml)*	Replicates	Fig. Panel
Nup358-KLC2/BicD2-CTD	5	118.4 ±5.9	64.7 ±3.2	0.4 ±0.01 & 0.2 ±0.01	2	A
Nup358-KLC2	5	67.9 ±3.4		0.4 ±0.1	3	
Nup358-KLC2/BicD2-CTD	2.5	119.3 ±6.0	64.9 ±3.2	0.1 ±0.01 & 0.1 ±0.01	4	B
Nup358-KLC2	2.5	63.2 ±3.2		0.2 ±0.01	2	
Nup358/W2224A/D2225A- KLC2/BicD2-CTD	2.5	119.2 ±6.0	51.1 ±2.6	0.02 ±0.01 0.2 ±0.01 &	3	C
Nup358/W2224A/D2225A- KLC2	2.5	43.6 ±2.2		0.4 ±0.01	3	

* Protein concentration at the apex of the elution peaks 1 & 2, determined by the refractive index and averaged from all experiments; standard deviations are shown. Elution peaks are numbered from highest to lowest molar mass. Conc.: protein concentration. For all experiments in this Table, pre-assembled and purified Nup358-KLC2/BicD2-CTD complexes were analyzed by SEC-MALS (rather than mixtures of these proteins).

SUPPORTING FIGURES

Drosophila melanogaster Nup358

MFTTRKEVDAHVHJKLMQLQGRERDKGLAVALRMLYMKVQEYPKAIYEYLNGYLRVRDDAVGHNMIATCYSRLNPPDVTEALQHYQRSIQIDPRQSEVVIDA
 CELLVKENNASCITECARYWLDQANSQDLSGNKQVFNLRMVRVLNADSNGERDDTSGGDGEQNTLEILMYKELQARPQDVNIRIQLRSYVEKMKIDQAFNYA
 LKTELESKNCTSQSNEWYEQIWMLFKIEMAKDVKKNWRFWHFALHTLDRVQLSLEGSGLADSSKQLFRLDQYLFKFSTSIERSGDAQPQRDLHQACIDHF
 TGQLLLHATLIFKREVLANKNKWMSTLRSALPLLLLGYQVRPIDDSTSNTQWIKHCDAEQKQLIQMWRPQGAFRCAQLGRTLLGCLDRSQMEIKNDRENAE
 FDENKNSGSNSMPGLFADSEELLASAHQQCLDKWSRSQIYQQFLTHAEHKLKDTSVHLVRNRLQLPLFEWPMLNLAHIENYELQALVLPPLSLAQHVYALGT
 DPNKLGDAAPRVVFYEGFQRDVQKQNLYNCGQDSISQVDVLYLATTIQTRRKQLQIREVYDSSNLGNRNAARPHHMPFANLVQLGAPEQSNWWDLVV
 RLNSNQLITEGNRAEQRAQLQHGLEYAVRGVNGPKADAIIFQLGKILNSRSRDRSSLEARIDTLYRQGFSILRHQHNQQMESYVRFKYGSAQSTAAWQDLQ
 SLAEEAVTYFSEKMFRIQYEQFLDEVRGLHLPMAFLQSEACHHLEESSKLPRTSRDRYERRRECLQTKLQKLIKNDDKHPLIAAMHRHQQDRNSRGIDN
 SFGSPDVHNNSAYEDAEDDFYSHAAFSANSRRQLEVTPTIVMAQPSQEMEQAVKQISKSLCVLKDDSVGMEAMRQDQVLTTEKFTGLEDDLKKIKI
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 AQAQAQFLRTPPAPGSIPPPNMFGPURNPGLSMFPPPTVPSVAPYIDAMGNFTQPPPSLIPPAQPAAPPAPLNILESKPVVAALPTPGFFNTTPVFGAS
 PIQVQSKPLTVPTVPIPSTAPAPIGVNPPATTAVPPPVHIPQVAPSVAQPPAPAVSVPSMFNRALNNQPVKEEPANVITSSDPLPKTTASVQPT
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 VTLERFLVRFKTGELAEERVFAFTKASEAAKSKEVTKPTVNTAEKGSTATAPAFAKSFVTPAANSLINKPQEQTQKPNPDPPATAAKSLFGTLSVSAAP
 ATSAPASATPFASFSTPTNGSSGFTSTASPFGNLSFGTASAVGSGNNNTLFTTALIKDNTVQGKTLQQESQNLNSNSDAAEEYYVPTAQFVPIALPDIVE
 VVTGEENEDVLVFEHRKLLRWDEKEANEWKERGLGNMKLRRDTPNKRLLMRREQVHKLCCNQRLLPETKFTYATTNKAATVWTGAQDYSDEELTALL
 AVRFBKSQDICQQFLEAVQKAQQSINEPKKEEVPSAAGEKEPKIKGFDAPFKKAGSWNCQACYTNNGQDQLYCLACQEPKDATVPPKQSGLDQGNALN
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 APKNDTVPQKEKSLGSGLNLPPTSKSFSGFGAAAAGDKDQAGDGFNFNFAAMPAAVAPTTSIGSSFTSMTKPKPDQQQPNSTAEEKEDENDSQEVEE
 EENNTYFSPVILPKDVIDVKTGEEDDEELLYVHKAKLYRNLNESDWKERGLGDVVKLRRQTKKLRVVMREQVFKICLNLHVNLENVYREKTETSWMFAVHD
 FSEGESVLERFTLRFKNKEVAQGFMEAINKNALNETAKPIEDSPVVGVSQSSTEANKPSQKNDGAKSRRGESEVLVKGKTTSSVRPTTHEVIPPLPMTLPLT
 LPQPLAKPNDYQTPATILFKGSSLSSRNNSASEASKTPSSAFIGGSTDKSEPGKDAGPLANLQKLASGEQQGNVLGSIFRGSSNENSDGSVKFFFGGGN
 KAAEQQKKDSSSESVFGGNKADQSXPATQEAKPLAFGGIAAPVFGDANPFGGHKVNLQKSDGKEEPKSIIGGTPLLFGGSNSAFQKPIETQSPAKDFVFGSA
 PAFGQMATFSFTAAKNEKEKDELTSNNTDLKAEGKEKKEVLPETTSTFADLAKTGSTFADLASNPGGTFADLANKTGNDFAFNLSANSQGTTVGFNKSAGG
 GFYNLTHQNAFKNFESPQATEECDDDGADTTDDNYDPHYDAIVELPDEIVTTGEENETKLGERAKLYRYDAESKWKERGVIEKVLHPELQFTRLIM
 RQEIQHKVLVNMNISASLQMDYMAQMKSFWAGYNYAVDAEGKVDTEGVLERLACRFAKEEIASEFLNTVNSCIKRAKALQGDEENKNDDAPEEQASS

Apis mellifera Nup358

MFRSKKDVRHVVKDFIKLKSSEEKKLRCYNIAKLYYQVGDNESAKKYVSNYLEIRDKSAGAHKLLGQALEGLGQKEAFTQYKISLELEPKQDDLLLKVCEL
 LSNMDINIANDKIKHLVERADKKFPHPIVFLKEKLITLEKPNGNDDDELEKLIISELSVRQTDVNLQVKLLKHYIGNHRLEDAYNKATGIEATHSHRNNIWYQS
 LSELLMKCKESKQSDWTFWIFYISVLERYALCLKEQGNIIKKSITDVTQAVAFNFDQSLFEFKSKNFSNHLQAFIEHMLHMWGQLHFHLACLILRKIKAGEDS
 WSEAGRLCGPLLLTALHVTPIPDPTAWWTMHLKDRLKQNQTHIWYREGSYRCSSQAGHVLQDYARDNTKLLDKIDKFCCTSSWRERIYQRFIGRLYQDGKKT
 FFTNSLISNPPLRLCSYNEKLKRFDEISEEWWDPSLHHQWVWGLRARPQGSQNKNGLHPNQTSHVYELQFSVYNLHQAPDLSLSRLDIAFLNAACITSSII
 EEQHGGLLNPEKLPPLPADLTNTLCQAEQWVSYAYKVYSMDKQKPLDDLGELRLELQRGLEVVRCLGNHGLHPVLLVHLARIFHRAETLKINQEN
 SDIPYLEARSEMYWSAAIPLLRLQNNQVIRVTNSKFFNYQKGKDMNNAELIKALEEGKLLAQRFRVRSKQYEKAINALQALKCPEASFQQGQMYKELADEIIS
 SIAKENLTSEMRSQLQHIIMLSARECFYLTLDRLRSPERNPKHPLNSELSTYIADIEKLKLRIDPDLRSRGLSRNECDGMSDESYSSAHSADVQAVTKIALPTLT
 GLNTSVNVLSTPKNIIHRTPKQSSTPYKLHQDIDLDSRNRSEARPSPERLDAQIRAINQMIHSKDNMIMQSIAEQTKTILELNKTVMEIVERLAKEMTELKEI
 QKQRTQTNVNPNEEDLCILSEDDYNDLNYNANQSGASSISGNMFQSSHRHPYSHLVPSATFQGYYPTGMSFNDPNAQIPSLYPPNIYSMPLVYPNT
 RSKIPENILQQLFMPPTNQPLQIPLQKLETSKPESTIKDAPVNKIPPNVVAITTSDELPPTVPIQPTLSVTIPPFRQGSSTPTVTEQSAPHCYQISMPSQA
 TIPTTVNLPPLSNTFTTTPANLSTSETSKQNTTICSTGQNSSSDHEHDPIDPDFVPIVPLPAIEKVTGEEGQETLFCARAKLYRFVDNEWKERGIGNVKKLNE
 EGKVRLMRREQVLKVCANHLYLVPDMELTAKSNEKAWFVVAHDGFADGELKLEKFCIRFKTVEEGISFKEHFDKAKASLTFEKVKIENVDKSNKDNIAISKK
 TEKIKSEQKIEQTIQPLFISGQQTQKPIDVTASTDSIIEKLQNNITQVTPVIGGFSSTPIQKAITTESSKISTKSEISPFAGFTNKTVTNAENFITTQKSTTIGTITT
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 DCIIAENLTKLETIQVQTHKSENLSSEMFPPEGSWECNGCYIRNNVSDVKCIACEKSRSSESDEIISIPSQNSQISLSQMSKPSTGSWCKKCCCNIFNAAES
 NYCVTCDSKPKDPSLPPKPKIDGFQINSNSGITSTFTFGIPQDTTKKGTNVFSFLRLPTSNDVSKSASLVALKSDAKTDASNAKFVFGTPGKSFGNFVTKSS
 PTKSGGEENSEEEVENDDIHFSPTIPLPDKIEVKTGEEEEVLYSHRAKLFYRNKPAEWKERGIGDIKLRLHNETGKLRVLMREQILKCLNHFVLPNFE
 LNSKDERTWNAADYSEGEIEPTLACRFKTSIDIANFKEIIDESKALPPIEKNTDNTKIEFTEVNTSISQDIEVYEMKVTPEEKQALQKLPNKFYAYQKQP
 DCPGCRGCKESKSLFTDESSEKSKISAENRKSLLTSTMQFLPKSTTFSNASNEVTPPTISSTTSTIVSSIFKPSLYGTNTLQFATNSITTTISFGNVDISITSS
 DKKTGFSVMPQISSNEIQKSMIDKQNISISENLKICSPITSQVTSTCATSFSTTSTIPTSSIFGMAKLDNTGKNGIFFGISKKDSSLKKPSSINVCTNSE
 LVFSNNFTQPKTITNTTITSVLSNATPIFGSSVLKANSENTNTGSLEVSVKSTNSVFPNTSVPFSYNLFGSVRTNTSSFTISNPVTTNAFETITPTTSDSQKES
 DVAFLPMTDLSFSTLAAKSSQSAFKIDPNFSFAGAGTAVFGTKSNTITNKCTEKSKEVKEKKDEDEEEEDDQVDEDNDHDYDPHFEIIPMPDIVEVHTGEEEE
 EKIFSERAKLYRYDSDTREWKERGVGEMKILHHAKYNRYRLLRRDQVYKVVCNFLLTPDITFSRLRTSDRAWWAGMNHAEEQPCLESLSVFKFKSPVLA
 TFKFDTIDKIQQTLSSESREKNVKDSPVIEEHGDEENEDCEEVGAVIDIKEEDEEEEDDDDDDDDDDDDDDDDDDDDDDDDDQNSITFEK
 RATLLTRTNKDSQWEPVALGNLIYDSNIFGERIILKADKTDEIVSNTIISMTKMEVNEKECVWTайдYALTPTPKRTLCAVFSSVHVAREMHKFQEGVEY
 AYQADISEPFYEE

Figure S1. The W-acidic motif with the sequence LEWD in Nup358 is conserved in vertebrates, but lost in insects. Protein sequences of Nup358 from *Drosophila melanogaster* (*Dm*) (Uniprot database identifier A0A0B4K7J2-1) and *Apis mellifera* (Uniprot database identifier A0A088AHI9-1). Tryptophan residues are highlighted in cyan. Note that the sequences lack a W-acidic LEWD motif. It should be noted that the sequence conservation between human Nup358 and *Dm* Nup358 is very low, therefore we were despite performing various sequence alignments unable to identify any region in *Dm* Nup358 that would be homologous to the minimal BicD2 and kinesin-1 binding site in human Nup358 (residues 2147-2240).

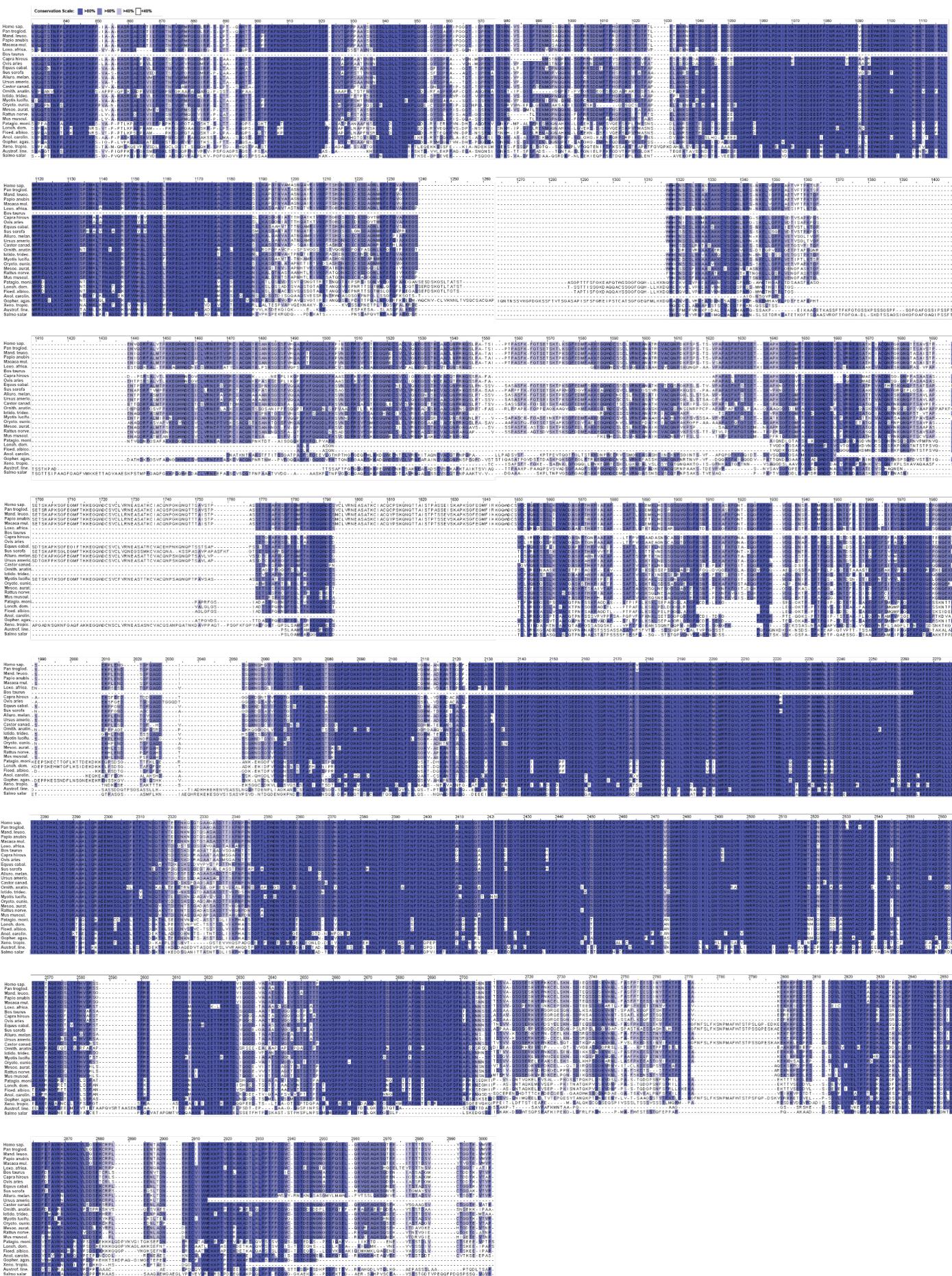


Figure S2. The sequence alignment of Nup358 from **Fig. 2** is shown for a larger portion of Nup358.

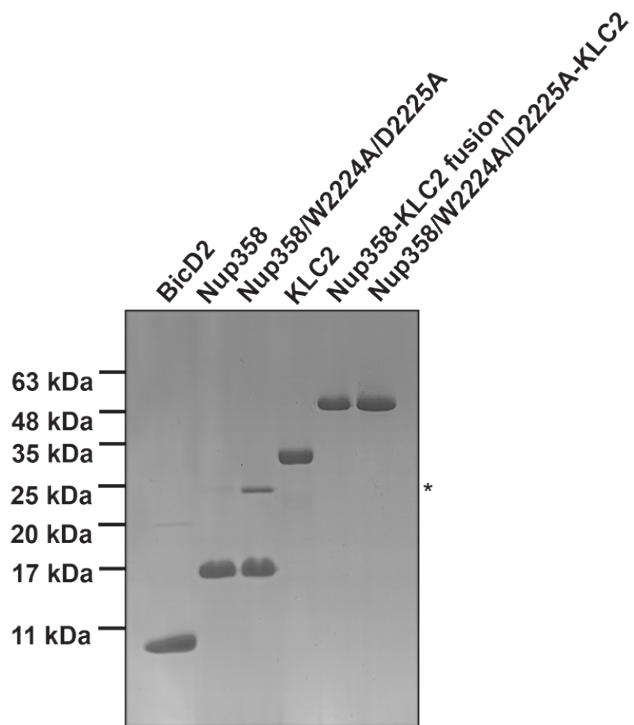


Figure S3. SDS-PAGE analysis of all purified proteins, stained by Coomassie Blue. Molar masses of standards are depicted on the left. From left to right: BicD2-CTD, Nup358-min, Nup358-min/W2224A/D2225A, KLC2^{TPR}, Nup358-KLC2-fusion protein, Nup358/W2224A/D2225A-KLC2-fusion protein. The asterisk indicates the location of GST bands. Multiple batches of purified proteins were used for this study, this SDS-PAGE shows a representative result. See also **Fig. 3**.

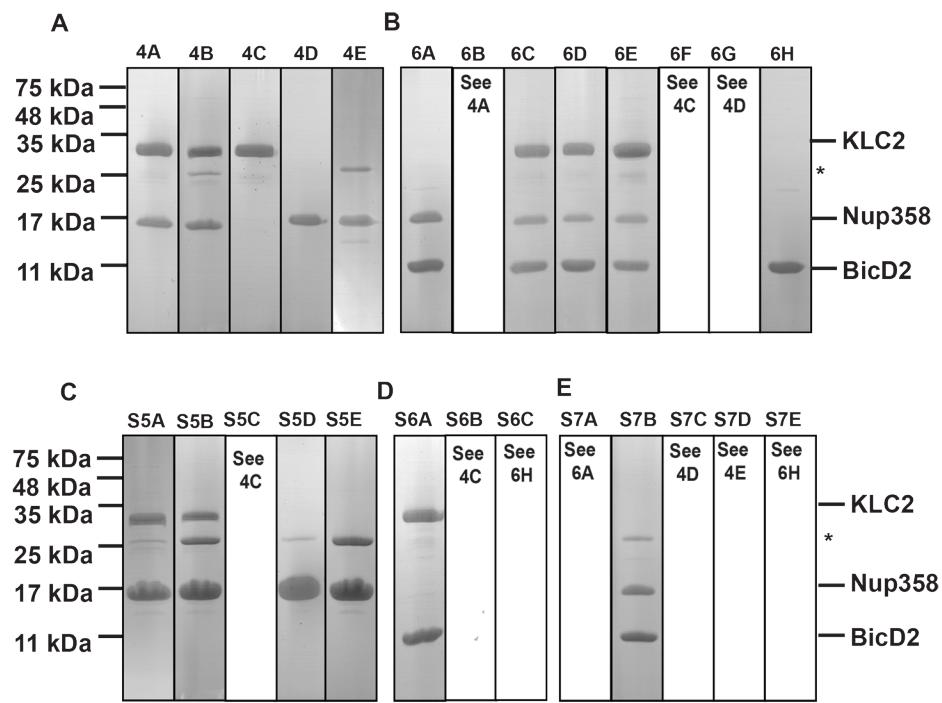


Figure S4. Load controls for all analytical size exclusion chromatography experiments (see **Figs. 4, 6, S5, S6** and **S7**). Coomassie-stained SDS-PAGE analyses of 10 μ L of the samples that were injected onto the column are shown, which is the same amount as the elution fractions that were analyzed. Gel lanes are grouped by figures and labeled on top with the figure panel showing the corresponding experiment. The locations of the bands of the molecular weight standards are indicated on the left. The asterisk indicates the location of the GST band. Multiple batches of purified proteins were used for the analytical size exclusion chromatography experiments in this study, the figure shows a SDS-PAGE analysis of representative samples. See also **Fig. S3**.

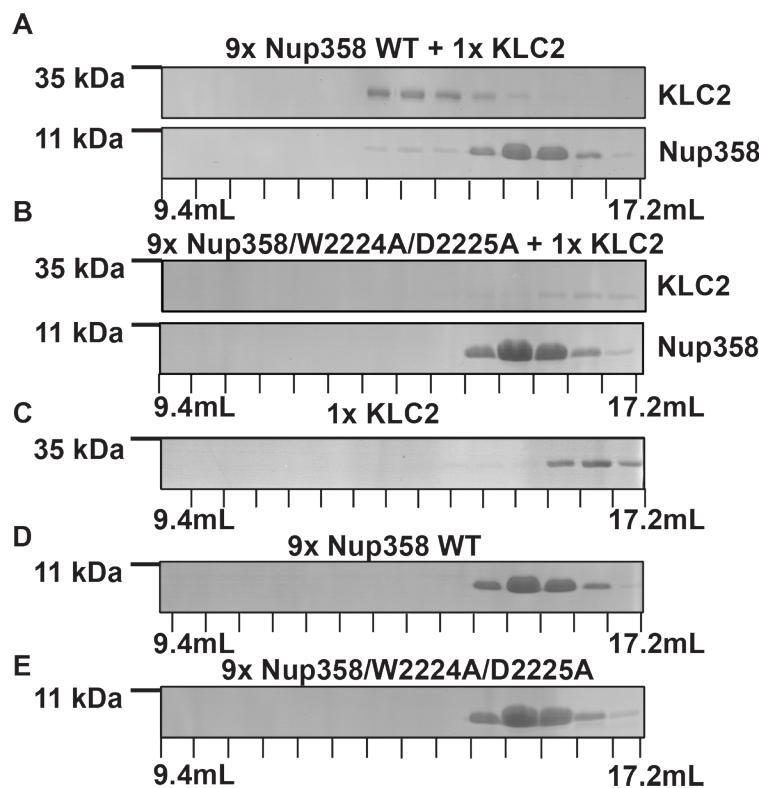


Figure S5. A very weak interaction with KLC2^{TPR} is observed with a large excess of the Nup358-min/W2224A/D2225A mutant. (A-E) To assess binding, purified Nup358-min (WT or W2224A/D2225A mutant) and KLC2^{TPR} were mixed in 9:1 molar ratio and analyzed using gel filtration chromatography. The elution fractions were separated by SDS-PAGE. The individual proteins were analyzed as well. (A) KLC2^{TPR} and Nup358-min. (B) KLC2^{TPR} and Nup358-min/W2224A/D2225A mutant. (C) KLC2^{TPR}. (D) Nup358-min. (E) Nup358-min/W2224A/D2225A mutant. Note that with addition of the large excess of Nup358-min/W2224A/D2225A, KLC2^{TPR} is shifted slightly towards higher mass compared to the individual KLC2^{TPR}, suggesting a weak interaction between these proteins. The interaction is however much weaker compared to Nup358-min WT. The experiments were repeated and very similar results were obtained. The number of replicates for experiments shown in each figure panel were: (A) 2, (B) 2, (C) 3, (D) 3, (E) 2. See also Figs. S3 and S4. Note that KLC2 has a higher solubility when mixed with Nup358-min (resulting in thicker gel bands) and that solubility of KLC2 is decreased when mixed with Nup358-min/W2224/D2225A.

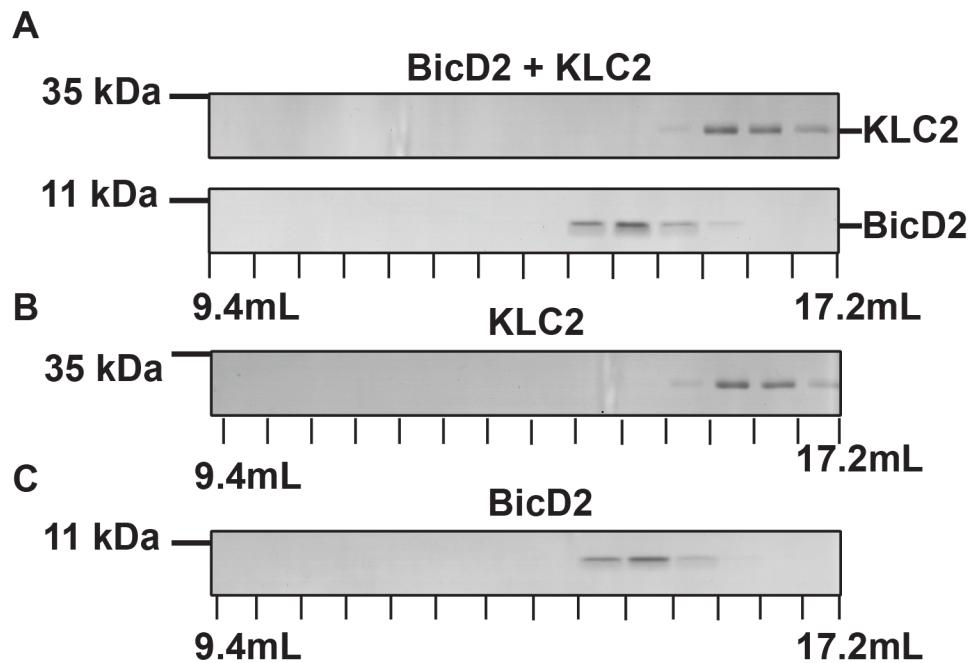


Figure S6 BicD2-CTD and KLC2^{TPR} do not interact. (A-C) To assess binding, purified BicD2-CTD and KLC2^{TPR} were mixed (1:1 molar ratio) and analyzed by gel filtration chromatography. The elution fractions were separated by SDS-PAGE. BicD2-CTD and KLC2^{TPR} were also analyzed individually. (A) BicD2-CTD and KLC2^{TPR}. (B) KLC2^{TPR}. (C) BicD2-CTD. Note that BicD2-CTD and KLC2^{TPR} elute at the same elution volumes when mixed as when individually analyzed, suggesting that they do not interact. The experiments were repeated and very similar results were obtained. The number of replicates for experiments shown in each figure panel were: (A) 3, (B) 3 (C) 5. See also Figs. S3 and S4.

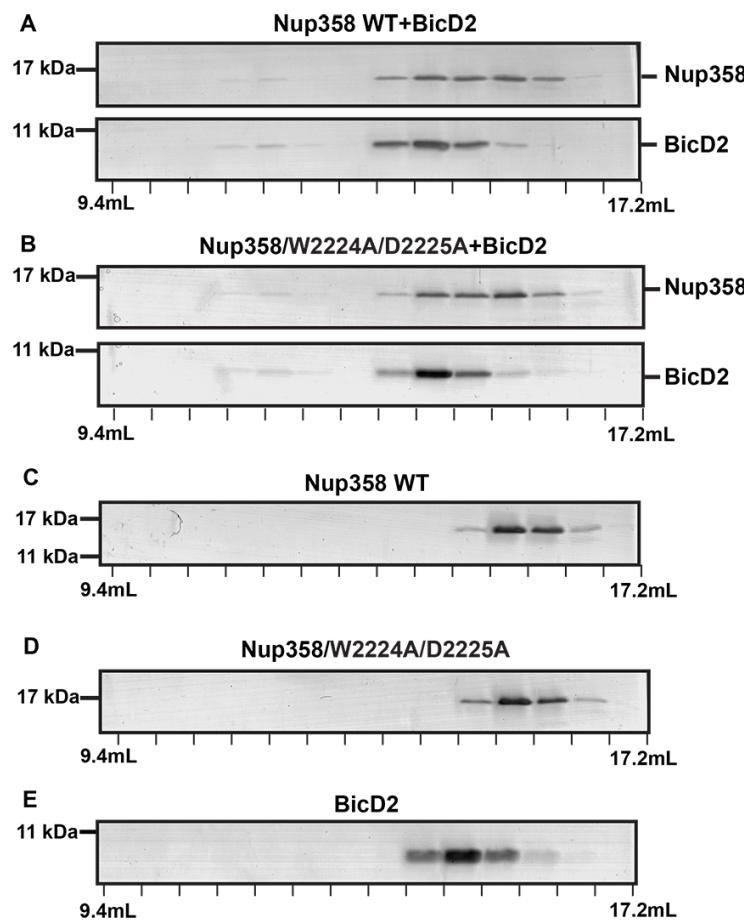


Figure S7. The W-acidic motif is not required for the interaction of Nup358-min with BicD2-CTD. (A-E) To assess binding, purified Nup358-min (WT or W2224A/D2225A mutant) and BicD2-CTD were mixed (1:1 molar ratio) and separated by gel filtration chromatography. The elution fractions were separated by SDS-PAGE. The individual proteins were analyzed as well. (A) Nup358-min and BicD2-CTD. (B) Nup358-min/W2224A/D2225A mutant and BicD2-CTD. (C) Nup358-min WT. (D) Nup358-min/W2224A/D2225A mutant. (E) BicD2-CTD. Note that both Nup358-min WT and the W2224A/D2225A mutant coelute with BicD2-CTD. Furthermore, the elution peaks are shifted towards higher mass when mixed compared to the individual proteins. The effect is comparable for Nup358-min WT and the W2224A/D2225A mutant, suggesting that the W2224A/D2225A mutation does not affect the interaction of Nup358-min with BicD2-CTD. The experiments were repeated and very similar results were obtained. The number of replicates for experiments shown in each figure panel were: (A) 3, (B) 2 (C) 4, (D) 2, (E) 5. See also Figs. S3 and S4.