

## Supplementary Materials for

### **Retinoic acid synthesis by ALDH1A proteins is dispensable for meiosis initiation in the mouse fetal ovary**

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#### **This PDF file includes:**

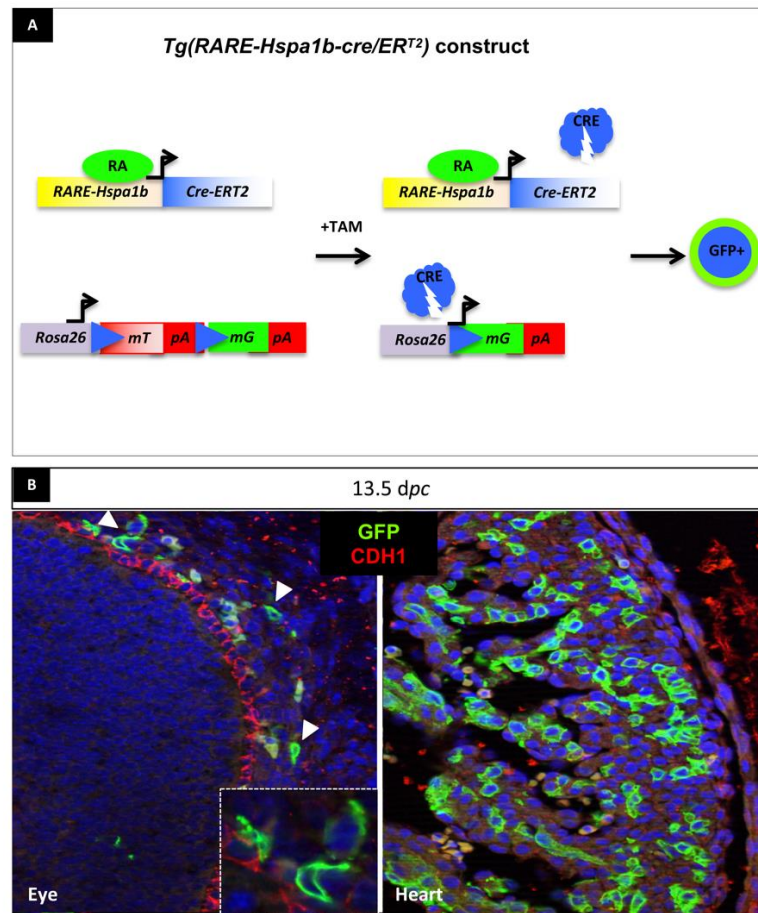
Figs. S1 to S8

## SUPPLEMENTARY MATERIALS

### Fig. S1: Validation of an ATRA-reporter inducible mouse model.

- A. Schematic maps of the *Tg(RARE-Hspa1b-cre/ER<sup>T2</sup>)* construct, in which ATRA-response elements (RARE) coupled to the *Hspa1b* minimal promoter (Yellow rectangles) drive the expression of the tamoxifen (TAM) inducible cre<sup>ERT2</sup> recombinase (blue rectangles), thus allowing ATRA-responsive cell lineage tracing. Blue triangles indicate *loxP* sequences. *RARE-Hspa1b-cre/ER<sup>T2</sup>* allows *Gt(ROSA)26Sor<sup>tm4(ACTB-tdTomato,-EGFP)Luo</sup>* (noted *mTmG*) recombination in ATRA-responsive cells upon Tamoxifen (TAM) administration. The *Gt(ROSA)26Sor<sup>tm4(ACTB-tdTomato,-EGFP)Luo</sup>* reporter line triggers membrane GFP-staining in the cells where the CRE recombinase is active. mG: membrane GFP.
- B. Immunodetection of CDH1 (epithelial cells) (red) and GFP (cells where the cre<sup>ERT2</sup> is active) (green) at 13.5 *dpc* in the developing eye and heart of *Tg(RARE-Hspa1b-cre/ER<sup>T2</sup>)* embryos. DAPI (blue): nuclei.

Fig. S1



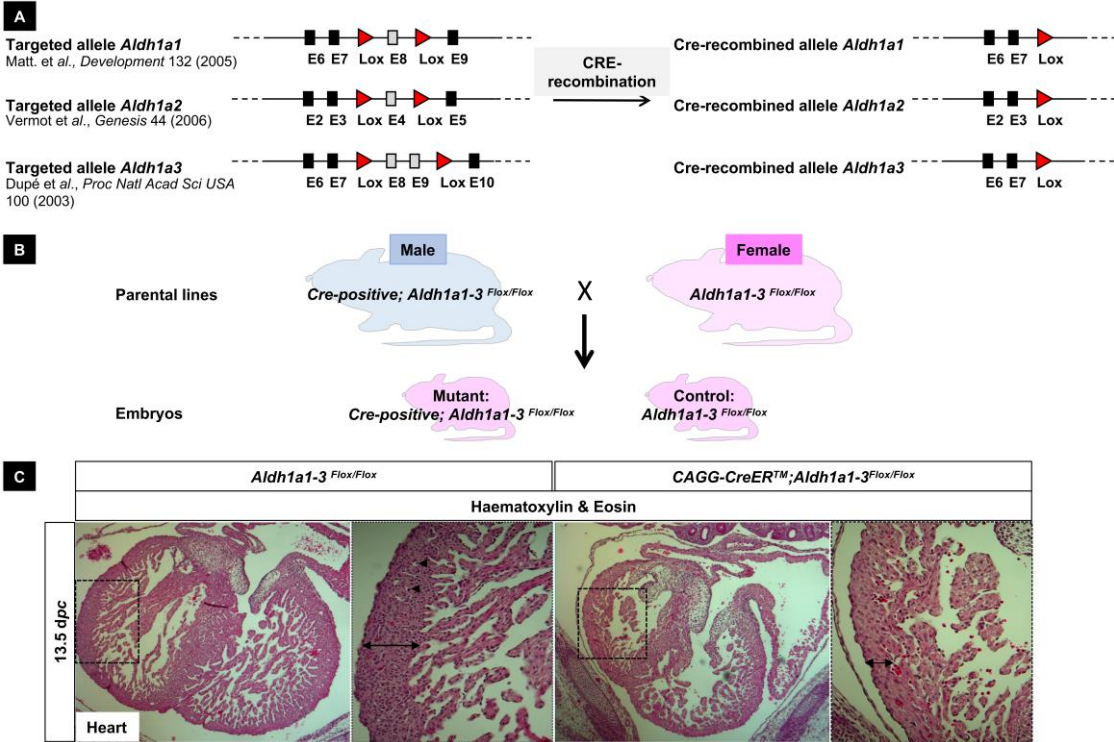
**Fig. S2: Construction of a stable loss-of-function of *Aldh1a1-3* in mouse embryos.**

A. Schematic maps of the targeted *Aldh1a1*, *Aldh1a2* and *Aldh1a3* alleles (left panel) and the CRE-recombined resultant alleles (right panel). Exons are shown as black (not recombined) or grey (recombined exons) boxes, whereas intronic sequences are shown as solid lines. Red triangles sandwiching exons in grey indicate *loxP* sequences.

B. Schematic representation of the experimental mating strategy for obtention of triple conditional mutant mice. Both parents are carrying the *Aldh1a1-3* triple floxed alleles, and the father only is carrying the *Cre*-encoding gene (upper panel), which allows collecting 50% of *Cre*-positive; *Aldh1a1-3* triple floxed embryos (25% being females).

C. Haematoxylin and Eosin staining at 13.5 *dpc* in control (left panel) and *CAGG-CreER<sup>TM</sup>; Aldh1a1-3<sup>flox/flox</sup>* (right panel) hearts.

Fig. S2



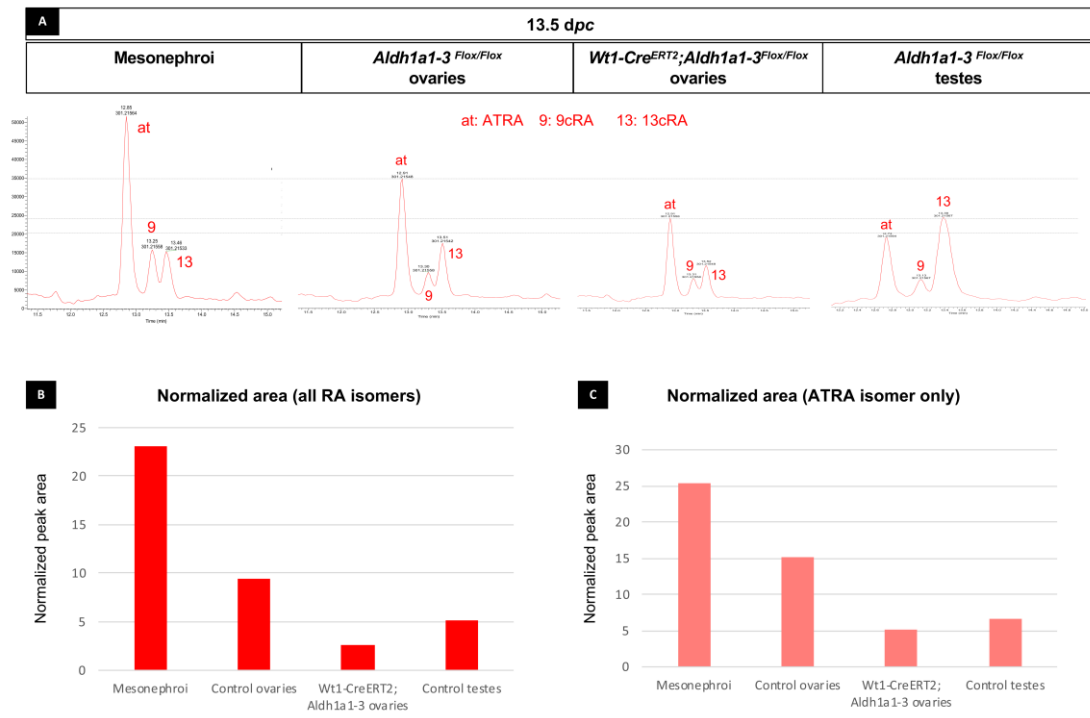
**Fig. S3: Retinoic Acid isomer separation by mass spectrometry in control and *Wtl-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* embryos.**

A. Mass spectrometry chromatograms of All-*Trans* Retinoic Acid (ATRA, at), 9-*Cis* Retinoic Acid (9cRA, 9) and 13-*Cis* Retinoic Acid (13cRA, 13) isomers in *Aldh1a1-3<sup>lox/lox</sup>* mesonephroi, *Aldh1a1-3<sup>lox/lox</sup>* ovaries, *Wtl-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* ovaries and *Aldh1a1-3<sup>lox/lox</sup>* testes from 13.5 dpc littermate embryos. n = 12 individual gonads or mesonephroi.

B. Histograms: quantification of all three RA isomers using total peak area normalized by the protein concentration of each sample (*Aldh1a1-3<sup>lox/lox</sup>* mesonephroi, *Aldh1a1-3<sup>lox/lox</sup>* ovaries, *Wtl-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* ovaries and *Aldh1a1-3<sup>lox/lox</sup>* testes).

C. Histograms: quantification of ATRA isomer only using ATRA (at) peak area normalized by the protein concentration of each sample (*Aldh1a1-3<sup>lox/lox</sup>* mesonephroi, *Aldh1a1-3<sup>lox/lox</sup>* ovaries, *Wtl-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* ovaries and *Aldh1a1-3<sup>lox/lox</sup>* testes).

Fig. S3



**Fig. S4: Genetic ablation of all three *Aldh1a1*, *Aldh1a2* and *Aldh1a3* does not impair germ cell differentiation *in vivo*.**

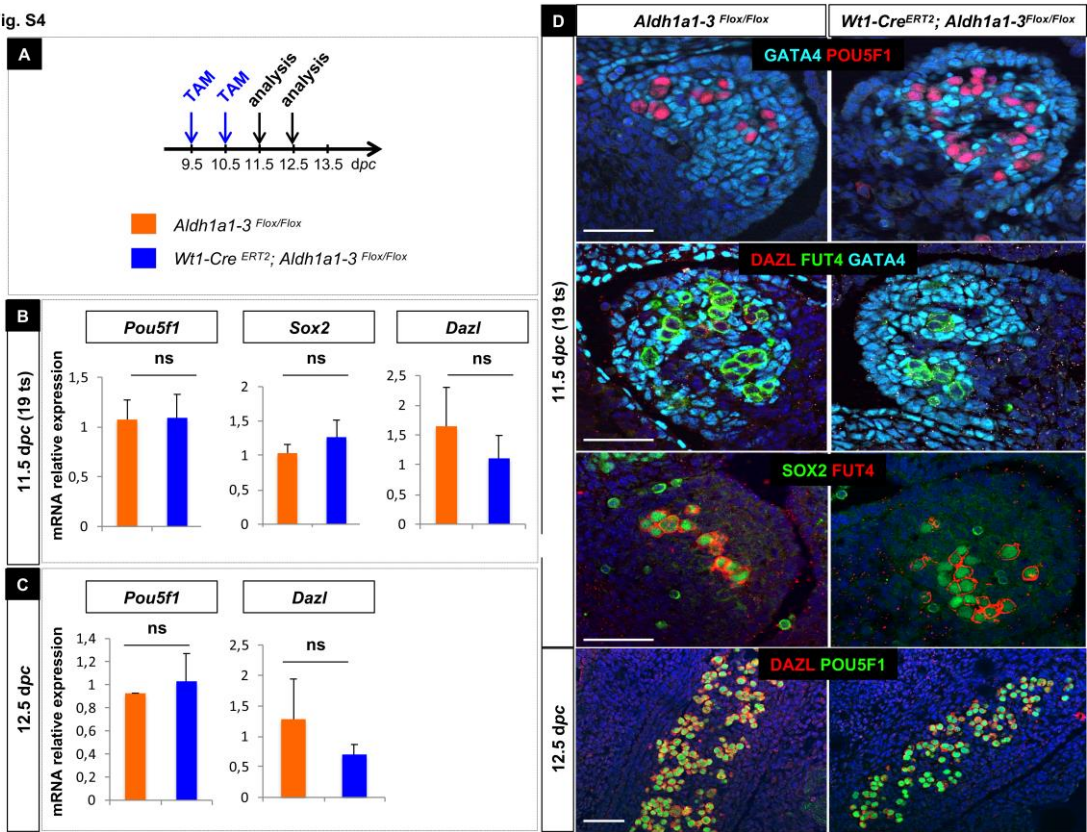
A- Protocol of induction of *Aldh1a1-3* deletion (9.5 dpc onwards). TAM: Tamoxifen.

B- RT-qPCR analysis of *Pou5f1*, *Sox2* and *Dazl* expression in 11.5 and, C- 12.5 dpc control (orange) and *Wtl-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* (blue) gonads. Student's *t*-test, unpaired. Bars represent mean +SEM, n = 10 individual gonads. ns: not significant.

D. Immunodetection of GATA4 (cyan, gonadal somatic cells) and POU5F1 (pluripotent germ cells, red); Middle panels: GATA4 (cyan), DAZL (germ cells, red) and FUT4 (or SSEA1, germ cells, green); SOX2 (pluripotent germ cells, green) and FUT4 (germ cells, red); Lower panel: DAZL (germ cells, red) and POU5F1 (pluripotent germ cells, green) in 11.5 and 12.5 dpc control and *Wtl-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* ovaries. DAPI (blue): nuclei. Scale bars (white): 50 micrometers.



Fig. S4

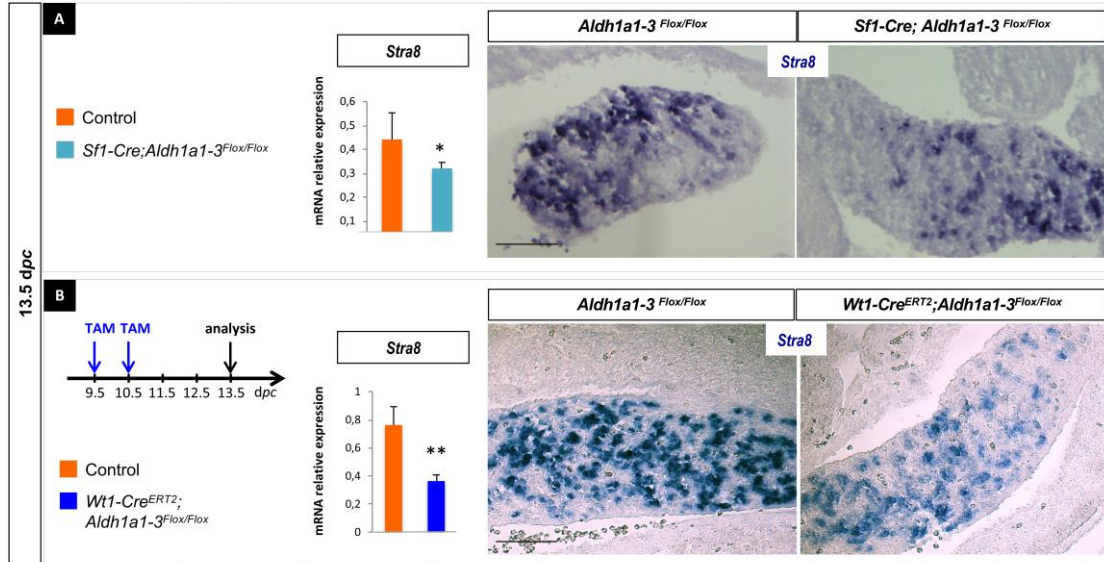


**Fig. S5: Genetic ablation of all three *Aldh1a1*, *Aldh1a2* and *Aldh1a3* genes using either *Sfl-cre* or *Wtl-cre*<sup>ERT2</sup> induced at 9.5 dpc does not impair germ cell entry into meiosis.**

A. Left panel: RT-qPCR analysis of *Stra8* expression in 13.5 dpc control (orange) and *Sfl-Cre;Aldh1a1-3*<sup>lox/lox</sup> (pale blue) ovaries. Student's *t*-test, unpaired. Bars represent mean +SEM, n = 10 individual gonads. \* p<0.05. Right panel: *In situ* hybridization using *Stra8* riboprobe at 13.5 dpc in control (left panel) and *Sfl-Cre;Aldh1a1-3*<sup>lox/lox</sup> (right panel) ovaries. Scale bars (black): 50 micrometers.

B. Left panel: Protocol of induction of *Aldh1a1-3* deletion (9.5 dpc onwards). TAM: tamoxifen. RT-qPCR analysis of *Stra8* expression in 13.5 dpc control (orange) and *Wtl-Cre*<sup>ERT2</sup>;*Aldh1a1-3*<sup>lox/lox</sup> (blue) gonads. Student's *t*-test, unpaired. Bars represent mean +SEM, n = 10 individual gonads. \*\*p<0.01. Right panel: *In situ* hybridization using *Stra8* riboprobe at 13.5 dpc in control (left panel) and *Wtl-Cre*<sup>ERT2</sup>;*Aldh1a1-3*<sup>lox/lox</sup> (right panel) gonads. Scale bars (black): 50 micrometers.

Fig. S5

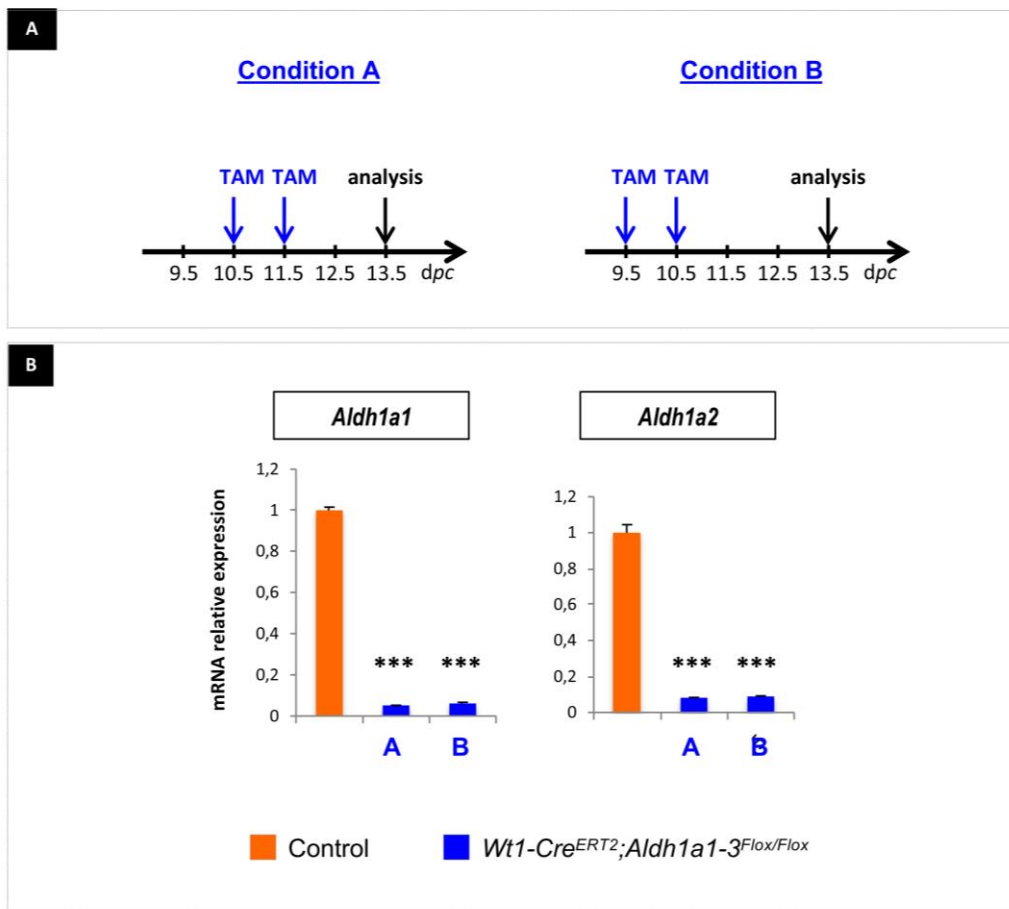


**Fig. S6: Comparison of two experimental conditions for *Aldh1a1* and *Aldh1a2* deletion**

A- Protocol of induction of *Aldh1a1-3* deletion 10.5 (condition A) or 9.5 (condition B) *dpc* onwards. TAM: tamoxifen.

B- RT-qPCR analysis of *Aldh1a1* and *Aldh1a2* expression in 13.5 *dpc* control (orange) and *Wt1-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* (blue) gonads in experimental conditions A and B. Student's *t*-test, unpaired. Bars represent mean +SEM, n = 10 individual gonads. \*\*\*p<0.001.

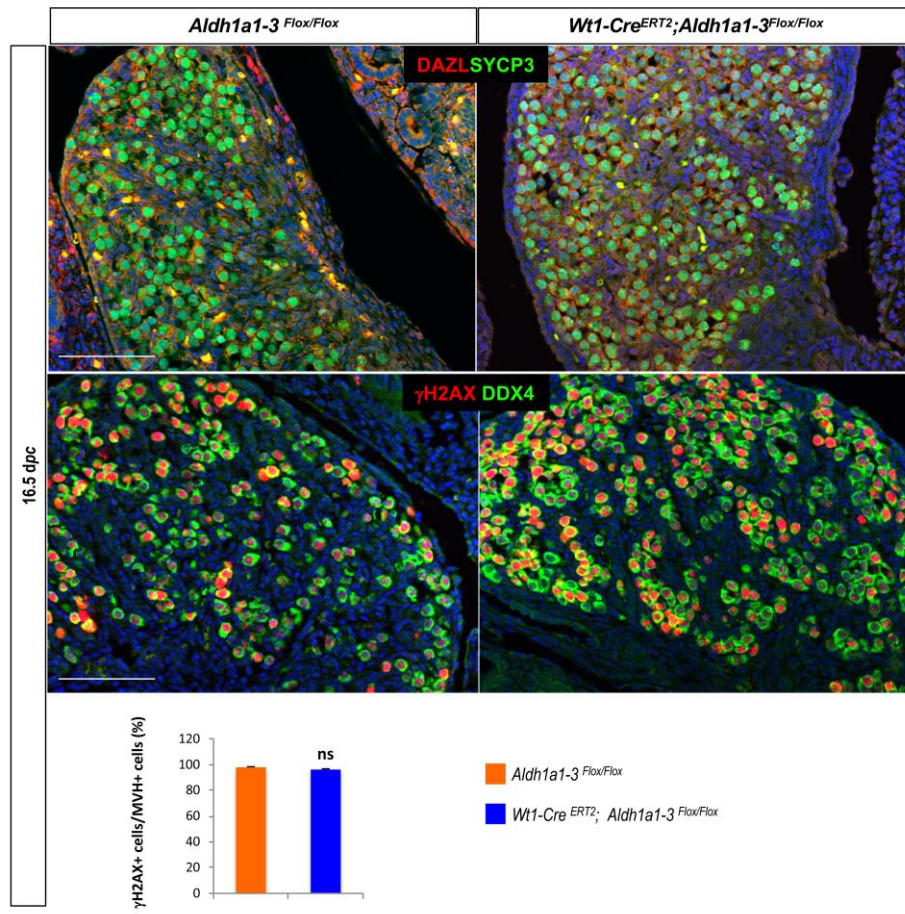
Fig. S6



**Fig. S7: Genetic ablation of all three *Aldh1a1*, *Aldh1a2* and *Aldh1a3* genes does not impair germ cell entry into meiosis.**

Upper panel: Immunodetection of DAZL (red) and SYCP3 (green) in 16.5 *dpc* control and *Wt1-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* ovaries. Middle panel: Immunodetection of Phospho Histone  $\gamma$ -H2AX (red) and DDX4 (green) in 16.5 *dpc* control and mutant ovaries. DAPI (blue): nuclei. Scale bars (white): 50 micrometers. Lower panel, Histograms: quantification of the percentage of both Phospho Histone  $\gamma$ H2AX- and DDX4-positive cells 16.5 *dpc* in control (orange) and *Wt1-Cre<sup>ERT2</sup>;Aldh1a1-3<sup>lox/lox</sup>* (blue) gonads. Student's *t*-test, unpaired. Bars represent mean +SEM, ns: not significant.

Fig. S7



**Fig. S8: Genetic ablation of all three *Aldh1a1*, *Aldh1a2* and *Aldh1a3* genes in the whole gonad failed to impair meiosis *in vivo*.**

A- RT-qPCR analysis of *Stra8*, *Mei1*, *Dmc1*, *Rec8*, *Syce 1* and *Spo11* expression in 13.5 dpc control (orange) and *CAGG-CreER<sup>TM</sup>; Aldh1a1-3<sup>flox/flox</sup>* (blue) gonads. Student's *t*-test, unpaired. Bars represent mean +SEM, n = 5 individual gonads. \* p<0.05; ns: not significant.

B- *In situ* hybridization using *Stra8* riboprobe at 13.5 dpc in control (left panel) and *CAGG-CreER<sup>TM</sup>; Aldh1a1-3<sup>flox/flox</sup>* (right panel) gonads. Scale bars (black): 50 micrometers.

C. Immunodetection of Phospho Histone  $\gamma$ H2AX (red) in 16.5 dpc control and *CAGG-CreER<sup>TM</sup>; Aldh1a1-3<sup>flox/flox</sup>* ovaries. DAPI (blue): nuclei. Scale bars (white): 50 micrometers.



Fig. S8

