

## Supplementary Materials:

# The Dark Side of Orchid Symbiosis: Can *Tulasnella calospora* Decompose Host Tissues?

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## Supplementary Figures

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**Table S1.** Species and experimental conditions of RNA-seq studies considered for gene expression meta-analysis.

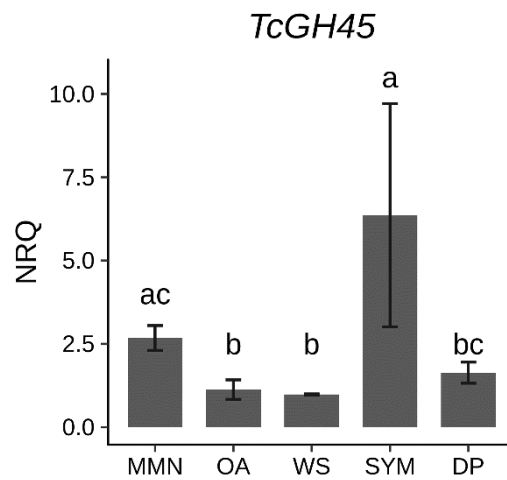
**Table S2.** Reads number and mapping details of selected RNA-seq libraries from short read archive (SRA).

**Table S3.** Reference JGI fungal transcriptomes used for mapping.

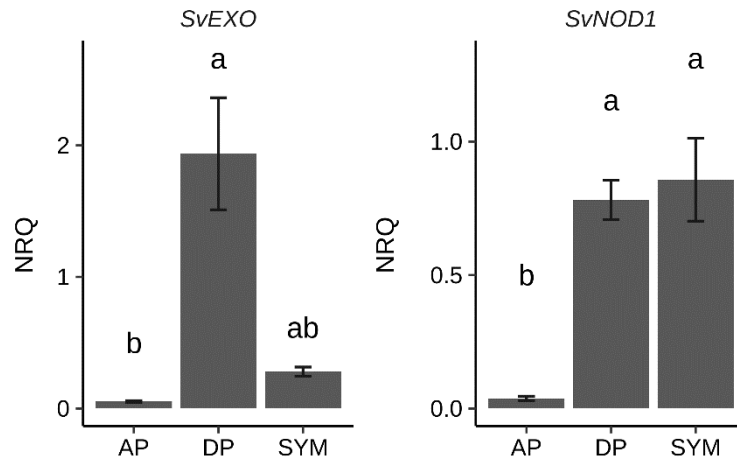
**Table S4.** Selected primers used in RT-qPCR experiments.



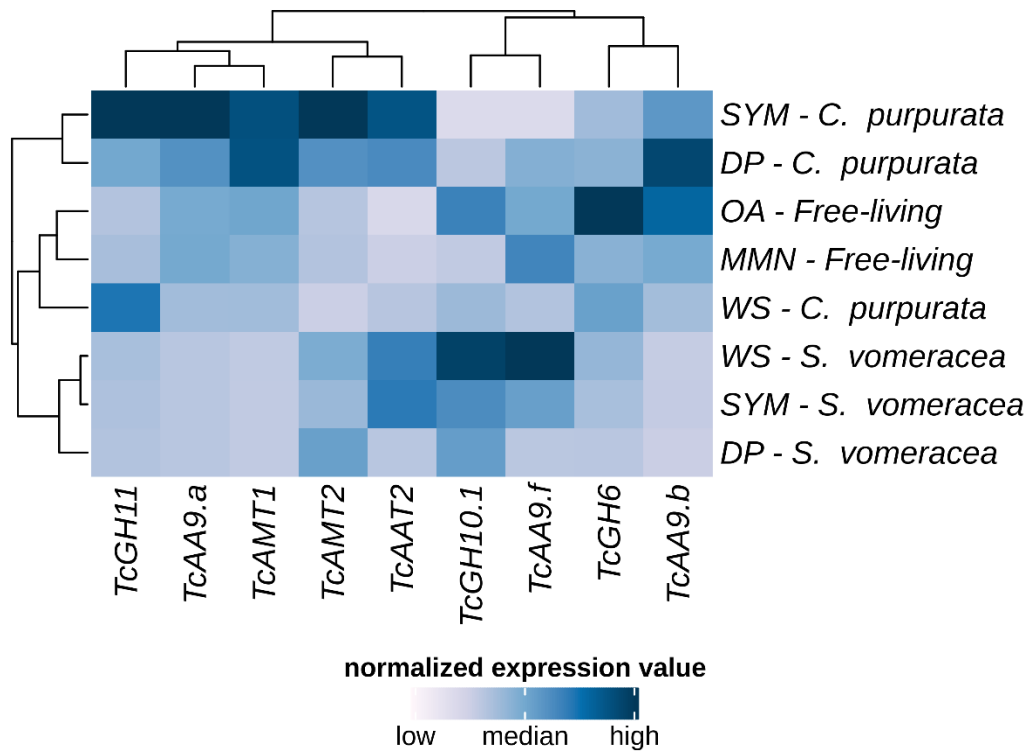
**Figure S1.** *Cattleya purpurata* seedlings colonized by *Tulasnella calospora*. **a)** Detail of a peloton (yellow arrow) inside the orchid root cells. Scale bar = 100  $\mu$ m; **b)** dead orchid tissues covered by fungal hyphae. Fungal inoculum is indicated by a yellow arrow. Scale bar = 2 mm; **c)** young dead seedling where the saprotrophic behavior of the fungus seems to prevail. Scale bar = 3 mm.



**Figure S2.** Expression pattern of *TcGH45* in *C. purpurata* model system and under free-living conditions. Letters indicate significant differences after Kruskal-Wallis test and Dunn's post-hoc test ( $p < 0.05$ ). NRQ, normalized relative quantities; mean  $\pm$  standard error (SE) is plotted.



**Figure S3.** Symbiosis marker genes in *S. vomeracea*. Letters indicate significant differences after Kruskal-Wallis test and Dunn's post-hoc test ( $p < 0.05$ ). NRQ, normalized relative quantities; mean  $\pm$  standard error (SE) is plotted.



**Figure S4.** *T. calospora* gene expression clustering in *C. purpurata*, *S. vomeracea* and free-living condition. Heatmaps show z-score scaled NRQ values by gene from RT-qPCR experiments. Hierarchical clustering dendrograms show Pearson correlation of expression values across conditions and genes.

**Table S1.** Species and experimental conditions of RNA-seq studies considered for gene expression meta-analysis. Fungal species with different ecology were considered: orchid mycorrhizal (OM), ecto-mycorrhizal (ECM), white rot (WR), pathogens.

Species	Strain	Ecology	Conditions	Plant Host	Instrument and Layout	SRA Study	References
<i>Tulasnella calospora</i>	AL13	OM	free-living mycelium, protocorm	<i>Serapias vomeracea</i>	HiSeq2000, PE	SRP050561	Kohler et al. (2015)
<i>Serendipita vermifera</i>	MAFF 30583	OM	free-living mycelium, roots	<i>Arabidopsis thaliana</i>	HiSeq2000, PE	SRP050866	Kohler et al. (2015)5
<i>Hebeloma cylindrosporum</i>	TV98 IV3	ECM	free-living mycelium, roots	<i>Pinus pinaster</i>	HiSeq2000, SE	SRP050559	Kohler et al. (2015)
<i>Piloderma croceum</i>	F 1598	ECM	free-living mycelium, roots	<i>Quercus robur</i>	HiSeq2000, PE	SRP050868	Kohler et al. (2015)
<i>Phanerochaete chrysosporium</i>	RP78	WR	vegetative mycelium	-	HiSeq2500, PE	SRP179770	Krizsán et al. (2019)
<i>Schizophyllum commune</i>	H4-8	WR	vegetative mycelium	-	HiSeq2500, PE	SRP179769	Krizsán et al. (2019)
<i>Puccinia graminis f.sp. tritici</i>	99KS76A-1	Pathogen	infected leaf (4 dpi*)	<i>Triticum aestivum</i>	Illumina Genome Analyzer, PE	SRP121504	Salcedo et al. (2017)
<i>Ustilago maydis</i>	FB1 x FB2	Pathogen	infected leaf (4 dpi*)	<i>Zea mays</i>	HiSeq2000, PE	SRP117697	Lanver et al. (2018)

\* days post infection.

**Table S2.** Reads number and mapping details of selected RNA-seq libraries from short read archive (SRA).

SRA Run	Species	Average Read Length	Condition	Raw	Filtered	Mapped	Mapped (%)
SRR1693601	<i>Tulasnella calospora</i>	200	Free-living mycelium	16,495,698	14,788,396	7,537,645	50.97
SRR1693602	<i>Tulasnella calospora</i>	200	Free-living mycelium	29,808,337	26,571,893	19,400,139	73.01
SRR1693603	<i>Tulasnella calospora</i>	200	Free-living mycelium	23,835,798	21,445,992	16,663,536	77.7
SRR1693604	<i>Tulasnella calospora</i>	200	Mycorrhizal protocorm	24,757,364	22,887,687	2,085,068	9.11
SRR1693605	<i>Tulasnella calospora</i>	200	Mycorrhizal protocorm	26,783,635	24,835,443	2,200,420	8.86
SRR1693606	<i>Tulasnella calospora</i>	200	Mycorrhizal protocorm	28,131,331	26,299,612	2,422,194	9.21
SRR1695524	<i>Serendipita vermifera</i>	202	Free-living mycelium	40,981,314	34,350,150	26,645,411	77.57
SRR1695525	<i>Serendipita vermifera</i>	202	Free-living mycelium	42,985,842	35,737,581	28,297,017	79.18
SRR1695526	<i>Serendipita vermifera</i>	202	Free-living mycelium	15,118,764	12,620,290	10,050,799	79.64

SRR1695527	<i>Serendipita vermifera</i>	202	Mycorrhizal roots	6,047,711	4,973,626	72,118	1.45
SRR1695528	<i>Serendipita vermifera</i>	202	Mycorrhizal roots	7,767,250	6,429,432	76,510	1.19
SRR1695529	<i>Serendipita vermifera</i>	202	Mycorrhizal roots	6,304,164	5,200,967	46,809	0.9
SRR1693517	<i>Hebeloma cylindrosporum</i>	150	Free-living mycelium	36,570,502	36,097,183	26,751,622	74.11
SRR1693518	<i>Hebeloma cylindrosporum</i>	150	Free-living mycelium	35,884,216	35,583,533	24,826,631	69.77
SRR1693519	<i>Hebeloma cylindrosporum</i>	150	Free-living mycelium	33,590,534	33,229,737	23,024,885	69.29
SRR1693520	<i>Hebeloma cylindrosporum</i>	150	Ectomycorrhiza	104,061,378	101,762,901	36,807,641	36.17
SRR1693521	<i>Hebeloma cylindrosporum</i>	150	Ectomycorrhiza	90,962,146	89,468,417	27,529,432	30.77
SRR1693522	<i>Hebeloma cylindrosporum</i>	150	Ectomycorrhiza	99,771,880	98,001,116	32,085,565	32.74
SRR1695538	<i>Piloderma croceum</i>	202	Free-living mycelium	59,973,882	55,067,234	45,815,939	83.2
SRR1695539	<i>Piloderma croceum</i>	202	Free-living mycelium	16,176,073	12,951,125	10,821,960	83.56
SRR1695540	<i>Piloderma croceum</i>	202	Free-living mycelium	24,212,031	19,310,902	16,157,432	83.67
SRR1695541	<i>Piloderma croceum</i>	202	Ectomycorrhiza	22,510,865	19,224,485	2,868,293	14.92
SRR1695542	<i>Piloderma croceum</i>	202	Ectomycorrhiza	21,303,775	18,468,413	2,478,461	13.42
SRR1695543	<i>Piloderma croceum</i>	202	Ectomycorrhiza	21,366,078	18,418,760	2,562,050	13.91
SRR8448307	<i>Phanerochaete chrysosporium</i>	249	vegetative mycelium	23,244,895	23,141,396	17,064,465	73.74
SRR8448308	<i>Phanerochaete chrysosporium</i>	249	vegetative mycelium	48,638,574	48,414,251	36,020,203	74.4
SRR8448309	<i>Phanerochaete chrysosporium</i>	249	vegetative mycelium	27,240,214	27,127,794	20,299,728	74.83
SRR8448292	<i>Schizophyllum commune</i>	249	vegetative mycelium	23,834,135	23,668,839	18,594,240	78.56
SRR8448293	<i>Schizophyllum commune</i>	249	vegetative mycelium	26,519,052	26,318,179	20,915,057	79.47
SRR8448294	<i>Schizophyllum commune</i>	249	vegetative mycelium	25,878,484	25,711,912	20,096,430	78.16
SRR6218744	<i>Puccinia graminis</i>	202	infected leaf (4 dpi)	39,258,693	35,329,192	936,224	2.65
SRR6218745	<i>Puccinia graminis</i>	202	infected leaf (4 dpi)	50,449,094	42,319,457	922,564	2.18
SRR6218772	<i>Puccinia graminis</i>	202	infected leaf (4 dpi)	25,880,263	23,943,494	474,081	1.98
SRR6039731	<i>Ustilago maydis</i>	202	infected plant (4 dpi)	30,838,063	29,406,524	349,938	1.19
SRR6039732	<i>Ustilago maydis</i>	202	infected plant (4 dpi)	38,837,342	37,108,931	890,614	2.4
SRR6039733	<i>Ustilago maydis</i>	202	infected plant (4 dpi)	35,511,412	32,985,380	1,576,701	4.78

**Table S3.** Reference JGI fungal transcriptomes used for mapping.

<b>Species</b>	<b>Version</b>	<b>Reference</b>
<i>Tulasnella calospora</i>	Tulca1 AL13/4D v1.0	Kohler et al. (2015)
<i>Serendipita vermifera</i>	Sebve1 MAFF 305830 v1.0	Kohler et al. (2015)
<i>Hebeloma cylindrosporum</i>	Hebcy2 h7 v2.0	Kohler et al. (2015)
<i>Piloderma croceum</i>	Pilcr1 F 1598 v1.0	Kohler et al. (2015)
<i>Phanerochaete chrysosporium</i>	Phcr2 RP-78 v2.2	Ohm et al. (2014)
<i>Schizophyllum commune</i>	Schco3 H4-8 v3.0	Ohm et al. (2010)
<i>Puccinia graminis f.sp. tritici</i>	Pucgr2 f. sp. tritici v2.0	Duplessis et al. (2011); Cuomo et al. (2017)
<i>Ustilago maydis</i>	Ustma2 521 v2.0	Kamper et al. (2006)

**Table S4.** Selected primers used in RT-qPCR experiments. Gene description corresponds to the main activity of the gene. In the reference column, the original studies for primers sequences are reported. Reference genes primer names are reported in bold.

<b>Primer Name</b>	<b>Target Species</b>	<b>Description</b>	<b>Sequence (5'–3')</b>	<b>Transcript ID<sup>†</sup></b>	<b>Reference</b>
<i>TcGH6.03</i>	<i>Tulasnella calospora</i>	CAZyme - cellobiohydrolase	F - GCCACCCAGAGCAGTTCTAC R - GCAAGGTAATCGCCGAGAGT	69053	This study
<i>TcGH10a</i>	<i>Tulasnella calospora</i>	CAZyme - endo-1,4- $\beta$ -xylanase/endo-1,3- $\beta$ -xylanase	F - CGGAGCCTTCGCAGAACTCG R - GCTCGCGCTCGTGATAGACG	14789	This study
<i>TcGH11*</i>	<i>Tulasnella calospora</i>	CAZyme - endo-1,4- $\beta$ -xylanase/endo-1,3- $\beta$ -xylanase	F - CGCTACTGCTGCCGTCTTGG R - GCCGTCCGTCCAGAAGGAGT	80414	This study
<i>TcGH11new</i>	<i>Tulasnella calospora</i>	CAZyme - endo-1,4- $\beta$ -xylanase/endo-1,3- $\beta$ -xylanase	F - CTACAAGGCCATCCGAACCA R - GTTCCGCCAGTACGATGACT	80414	This study
<i>TcGH45*</i>	<i>Tulasnella calospora</i>	CAZyme - endoglucanase	F - CAGCAACCTTCCTGCGGCTC R - AGAAGCGGGGCAAGTGACCT	224031	This study
<i>TcAA9a</i>	<i>Tulasnella calospora</i>	CAZyme - Lytic Polysaccharide Monooxygenase (LPMO)	F - CGTTAGCATCTTCTGGGGCA R - CGATCTTGGTCGGGTCAGTC	75481	This study
<i>TcAA9b</i>	<i>Tulasnella calospora</i>	CAZyme - Lytic Polysaccharide Monooxygenases (LPMO)	F - ATCAGGCGAACCTTCTGTCC R - ACTTGGCGTCGAGAGAGTTG	6298	This study
<i>TcAA9c<sup>§</sup></i>	<i>Tulasnella calospora</i>	CAZyme - Lytic Polysaccharide Monooxygenases (LPMO)	F - CGGTCTGGTTCAAAGTTGCC R - CGTGCAGAGCAATGTGTTCG	115568	This study
<i>TcAA9d<sup>§</sup></i>	<i>Tulasnella calospora</i>	CAZyme - Lytic Polysaccharide Monooxygenases (LPMO)	F - CTCTGATCGTCAACGGCACT R - AAGAAACGACTGTGACGGGG	27439	This study
<i>TcAA9e<sup>§</sup></i>	<i>Tulasnella calospora</i>	CAZyme - Lytic Polysaccharide Monooxygenases (LPMO)	F - CAAGCCAACAATCCGCAACT R - CTGATTGACTGGGGGTTGGG	641723	This study
<i>TcAA9f</i>	<i>Tulasnella calospora</i>	CAZyme - Lytic Polysaccharide Monooxygenases (LPMO)	F - GGAGACTGAAGGGATTGGGC R - CGTCAGCCATGTTGCGTATG	4643	This study
<i>TcAMT1</i>	<i>Tulasnella calospora</i>	Ammonium transporter	F - AGGACCAACTACCGCCACCA R - CATCCGTAAGGAGCGCCATC	241330	Fochi et al. (2017a)
<i>TcAMT2</i>	<i>Tulasnella calospora</i>	Ammonium transporter	F - ATTCGCGAGCTGCTTCCATC R - GCAAGGTGGGGCTGAACATC	183841	Fochi et al. (2017a)

<i>TcAAT2</i>	<i>Tulasnella calospora</i>	Aminoacid transporter	F - AAAGGCGGTATTGCGCTTCA R - GGCAGAGAAAGCACCGAGGA	81605	Fochi et al. (2017a)
<i>TcEF1α</i>	<i>Tulasnella calospora</i>	Elongation factor (fungal reference gene)	F - CAAGCCTATGTGCGTTGAGA R - ACGAGAATGCGAAGGAAGAA	241957	Perotto et al. (2014)
<i>SvNod1</i>	<i>Serapias vomeracea</i>	Early nodulin 55-2, putative	F - TGACGGCATCCGCCGAGTTCT R - ACTTGGAATCTCATACGCTCCGCC	DN89686_c0_g1_i1	Perotto et al. (2014)
<i>SvEXO</i>	<i>Serapias vomeracea</i>	Exocyst subunit exo70 family protein H2	F - CTGCTTCGCCGATGTTACCC R - TCGGAGAGGGCTTGGTGAAG	DN73752_c2_g2_i1	Fochi et al. (2017b)
<i>SvUBI</i>	<i>Serapias vomeracea</i>	Ubiquitin	F - TCTATCTATTCTTGCGGGGG R - CACATGGCGCAGTCAATAAA	DN70641_c1_g2_i1	Perotto et al. (2014)
<i>SvEF1α</i>	<i>Serapias vomeracea</i>	Elongation factor (plant reference gene)	F - CGATCTCGTAGTGATCTGAGCAAG R - GAGACAGAAATAACAACCAGCAACA	DN66666_c0_g1_i1	Fochi et al. (2017a)

**Notes:** \* oligonucleotides that worked only in the *Cattleya-Tulasnella* model. § oligonucleotides tested, but not included in results due to low or no amplification. † *T. calospora* transcripts IDs refers to the AL13/D transcriptome deposited in JGI portal (Kohler et al. 2015) while *S. vomeracea* IDs refers to the assembled transcriptome (Fochi et al. 2017a) available at NCBI (accession GSE87120). ‡ MYC reads and FLM reads refer to the *T. calospora* RNA-seq transcriptome analysis (Kohler et al. 2015; Fochi et al. 2017a). MYC reads and FLM reads columns show the mean number of reads observed in symbiotic *T. calospora* and free-living *T. calospora* respectively.



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