

SUPPORTING MATERIALS

ST2 as checkpoint target for colorectal cancer immunotherapy

Kevin Van der Jeught¹, Yifan Sun¹, Yuanzhang Fang¹, Zhuolong Zhou¹, Hua Jiang², Tao Yu¹, Jinfeng Yang², Malgorzata Maria Kamocka³, Kaman So⁴, Yujing Li¹, Haniyeh Eyvani¹, George E. Sandusky⁵, Michael Frieden¹, Harald Braun^{6,7}, Rudi Beyaert^{6,7}, Xiaoming He^{8,9}, Xinna Zhang^{1,10}, Chi Zhang^{1,4,10}, Sophie Paczesny^{2,10*}, Xiongbin Lu^{1,10*}

¹Department of Medical and Molecular Genetics, Indiana University School of Medicine, Indianapolis, IN, USA; ²Department of Pediatrics, Indiana University School of Medicine, Indianapolis, IN, USA; ³Department of Medicine, Division of Nephrology, Indiana University School of Medicine, Indianapolis, IN, USA; ⁴Center for Computational Biology and Bioinformatics, Indiana University School of Medicine, Indianapolis, IN, USA; ⁵Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, USA; ⁶VIB Center for Inflammation Research, Ghent, Belgium; ⁷Department of Biomedical Molecular Biology, Ghent University, Ghent, Belgium; ⁸Fischell Department of Bioengineering, University of Maryland, College Park, MD, USA; ⁹Marlene and Stewart Greenebaum Comprehensive Cancer Centre, University of Maryland, Baltimore, MD, USA; ¹⁰Melvin and Bren Simon Cancer Center, Indiana University School of Medicine, Indianapolis, IN, USA

*Corresponding authors:

*Sophie Paczesny, MD, PhD

Nora Letzter Professor of Pediatrics

Indiana University School of Medicine

Professor, Microbiology & Immunology

Melvin and Bren Simon Cancer Center

Wells Center for Pediatric Research

1044 W Walnut Street, R4 425

Indianapolis, IN 46202

317-278-5487 (office)

sophpacz@iu.edu

*Xiongbin Lu, PhD

Vera Bradley Foundation Chair in Breast Cancer Innovation

Strategic Research Initiative Distinguished Investigator

Indiana University Melvin and Bren Simon Cancer Center

Professor, Medical and Molecular Genetics

Indiana University School of Medicine

980 W Walnut St, R3.C218D

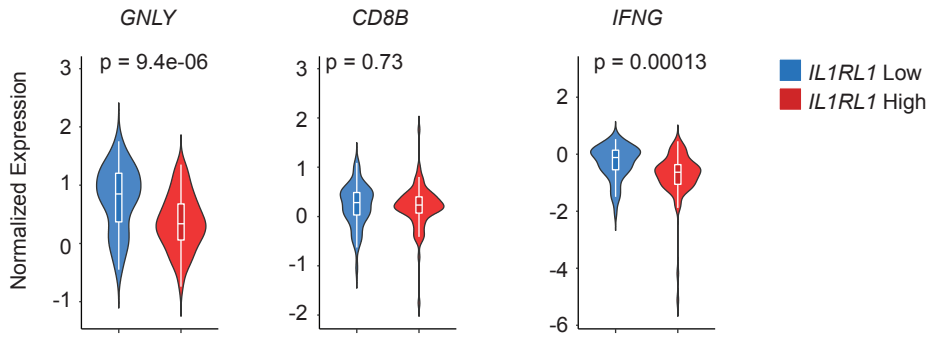
Indianapolis, IN 46202

317-274-4398 (office)

xiolu@iu.edu

SUPPORTING SUPPLEMENTARY FIGURE LEGENDS

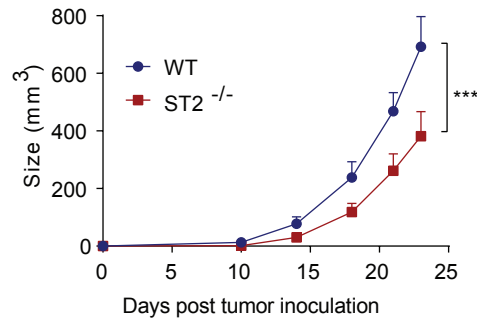
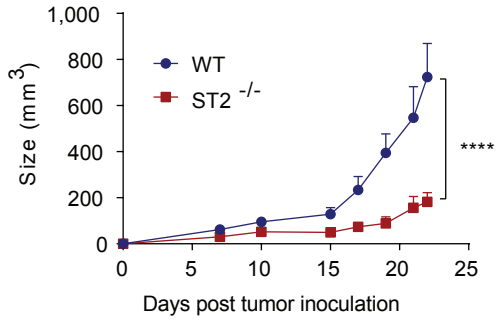
Supplementary Figure 1. Computational gene correlation analysis of T cell anti-tumor immunity. Violin box plots for the correlation of *IL1RL1* expression with the normalized expression of GNLV, CD8B and IFNG genes. Data analysis was conducted using the ICTD algorithm on 93 (*IL1RL1* High, 53 patients; *IL1RL1* Low, 40 patients) biologically independent colorectal cancer patients from the TCGA database. Significance was determined by two-tailed unpaired *t* test.



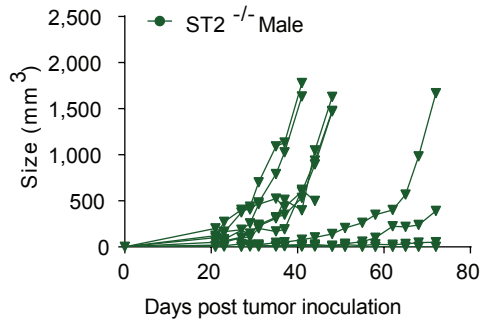
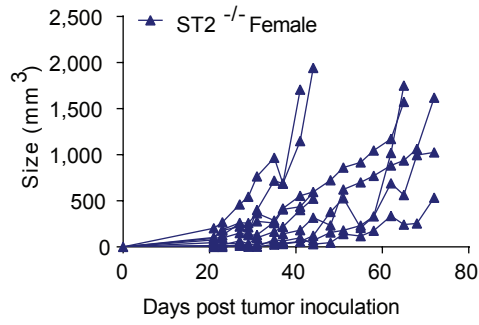
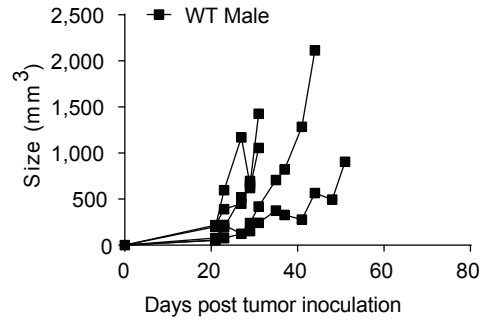
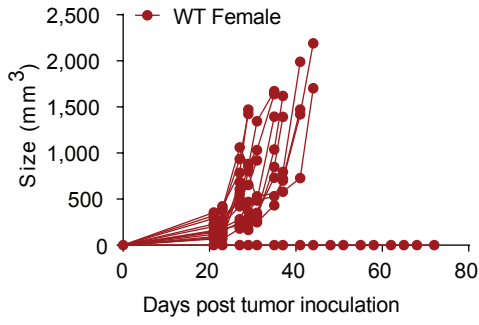
Supplementary Figure 2. Tumor growth of MC38 and CT26 tumors in *WT* and *ST2*^{-/-} mice.

(A) Tumor growth curve of MC38 (left panel) and CT26 (right panel) in *WT* and *ST2*^{-/-} mice (C57BL/6 *WT* *n*=8; C57BL/6 *ST2*^{-/-} *n*=7; BALB/C *WT* *n*=8; BALB/C *ST2*^{-/-} *n*=9; Data displayed as mean + SEM). The data represents one tumor growth experiments out of at least 3 independent experiments. (B) Individual spaghetti plots from both male and female mice of the tumor growth shown in *figure 2B*. Significance was determined by two-way ANOVA (A).

A

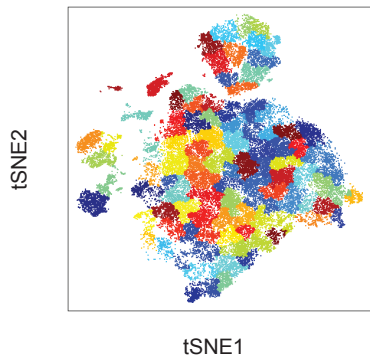


B

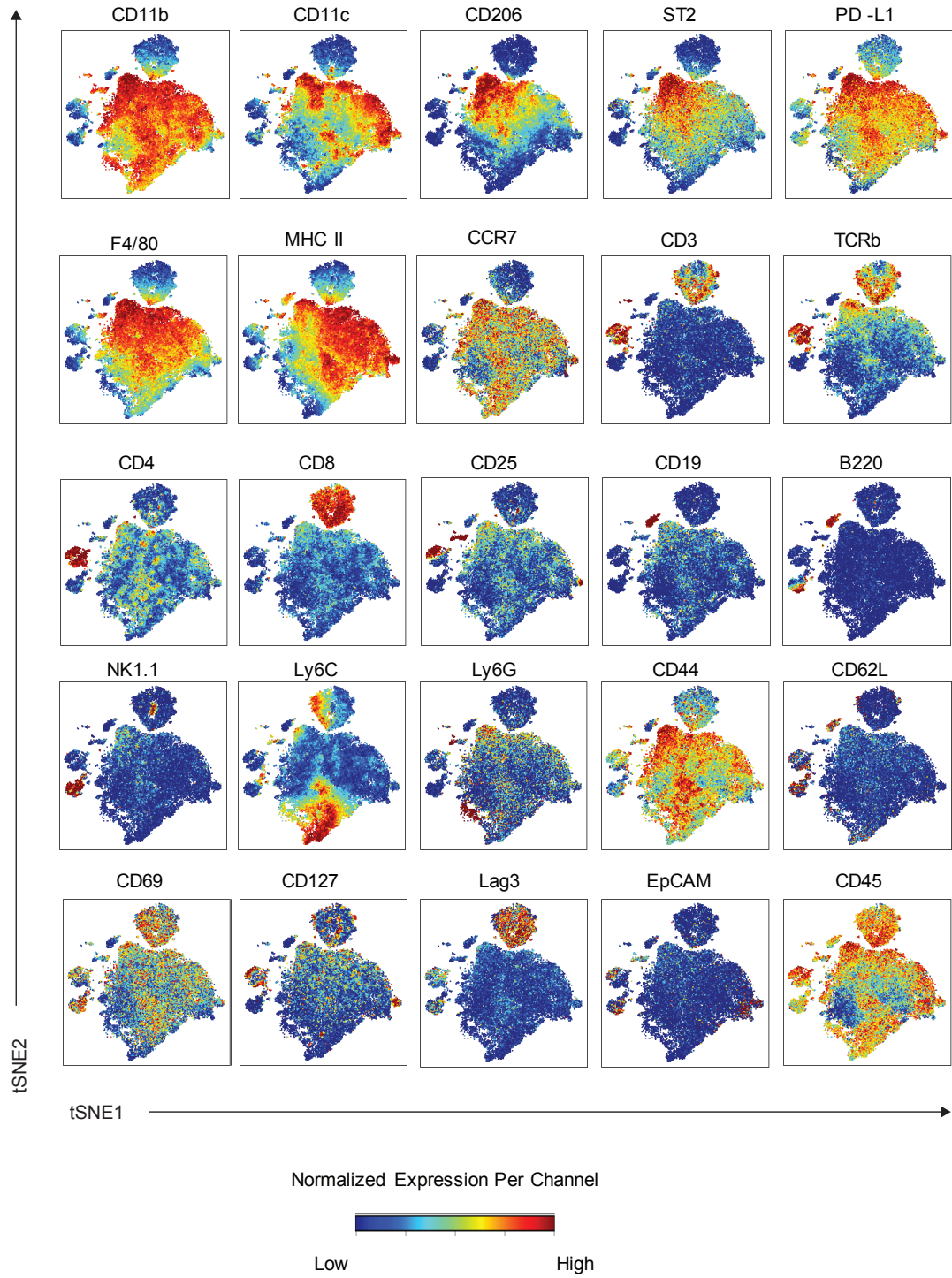


Supplementary Figure 3. CyTOF phenotyping of *WT* and *ST2*^{-/-} mice harboring MC38 tumors. (A, B) Mass cytometry immune cell analysis on MC38 injected tumor cells in *WT* and *ST2*^{-/-} mice ($n=5$ mice per group). **(A)** t-SNE plot of tumor infiltrating lymphocytes after running SPADE on viSNE. The plot shows the clustering on the tSNE1 and tSNE2 axis. **(B)** Normalized intensities on t-SNE plot of immune cells displaying the indicated channel expression levels.

A

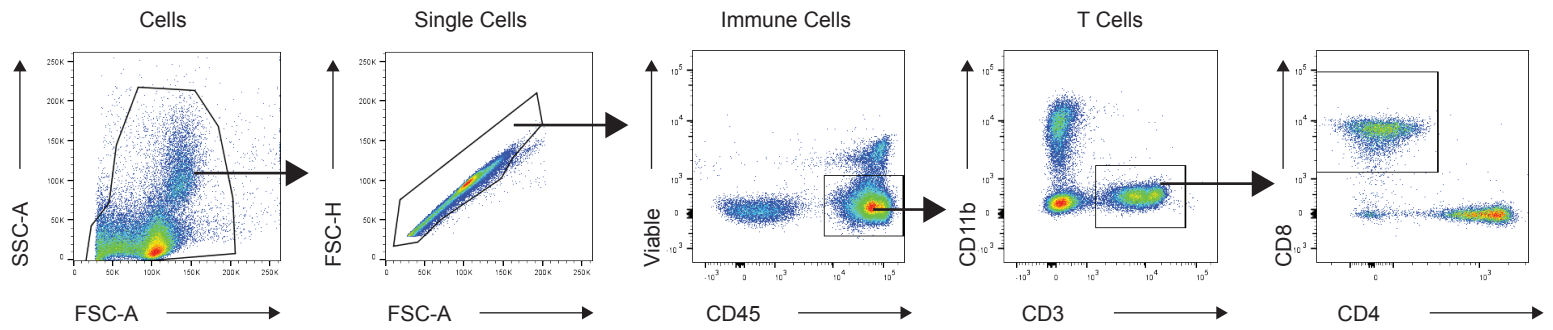


B

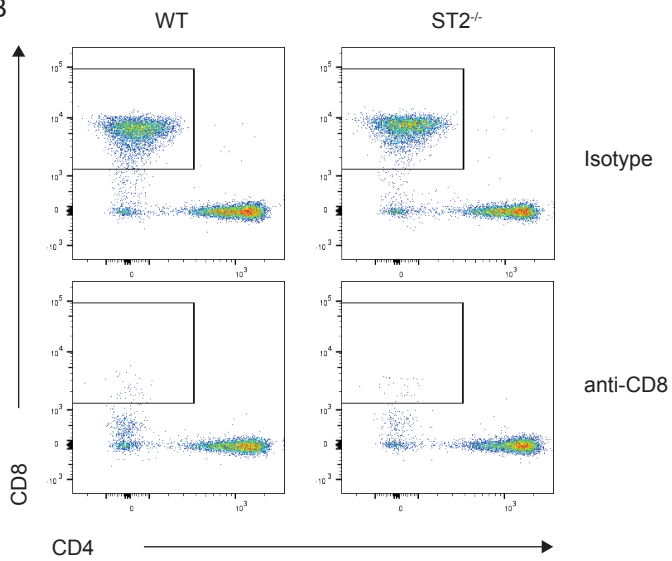


Supplementary Figure 4. Validation of CD8⁺ T cell depletion. (A) Flow cytometry gating strategy to monitor CD8⁺ T-cell depletion from the mouse blood. (B) Representative flow cytometry plots from *WT* and *ST2^{-/-}* mice upon treatment with the isotype or anti-CD8 antibody. (C) Quantification of the CD8⁺ T-cell depletion from *WT* and *ST2^{-/-}* mice upon treatment with the isotype or anti-CD8 antibody. (WT isotype, *n*=4; *ST2^{-/-}* isotype, *n*=5; WT anti-CD8, *n*=4; *ST2^{-/-}* anti-CD8, *n*=5; Data displayed as mean ± SD). Significance was determined by one-way ANOVA (C).

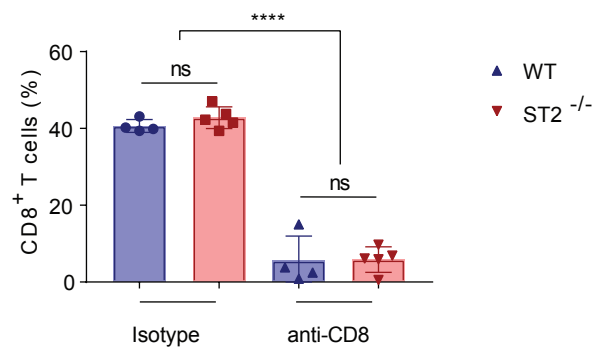
A



B

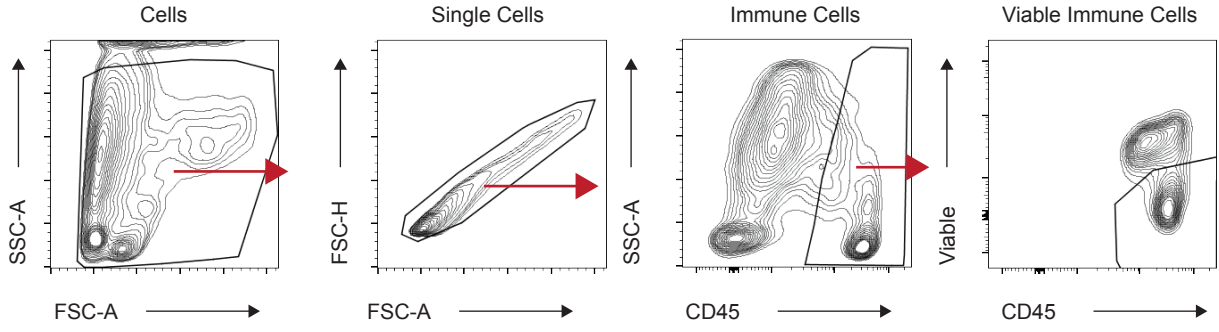


C

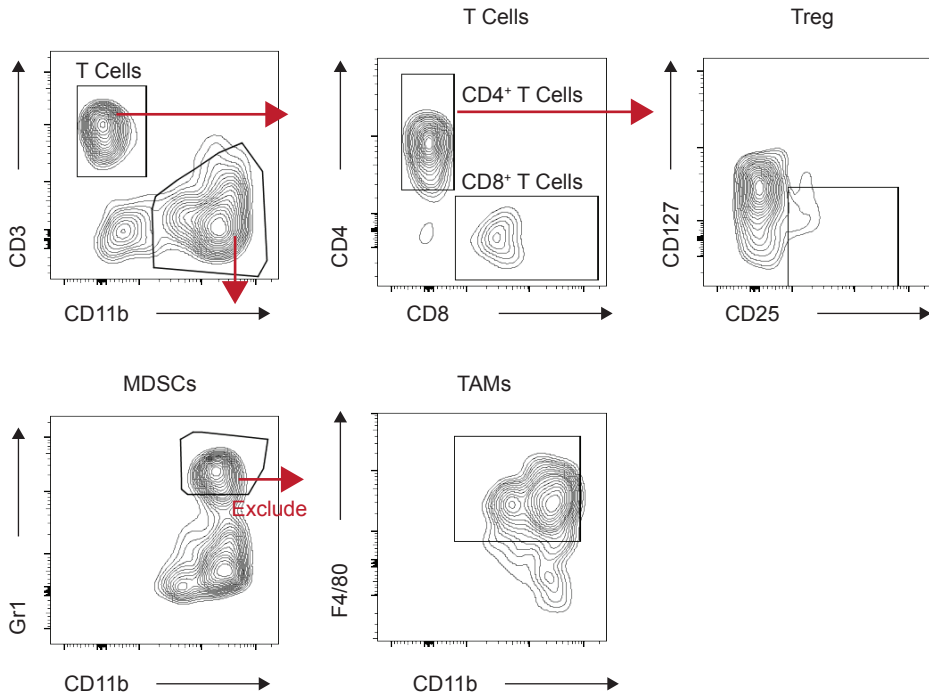


Supplementary Figure 5. Gating Strategy ST2 expression in the CT26 Tumor Microenvironment. (A) Gating strategy to assess viable immune cells in the CT26 TME. (B) Gating strategy to define the immune cell subtypes.

A

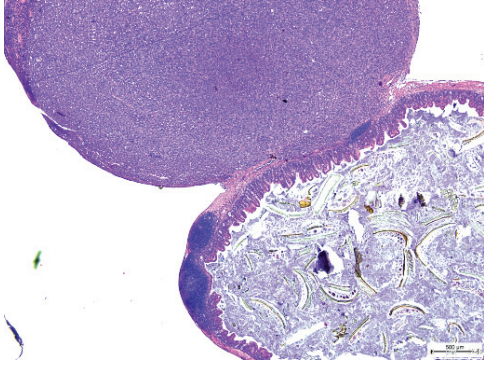


B



Supplementary Figure 6. Orthotopic mouse colorectal cancer model. (A) Representative H&E staining of orthotopic cecal wall tumors. (B) Representative confocal microscopy images from at 20x magnification stained with the indicated markers on orthotopic cecal wall tumor of injected MC38 cells (Tumor) and from the healthy cecal wall (Healthy). Scale bar represents 50 μm .

A



B

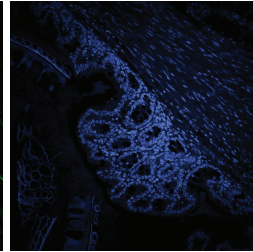
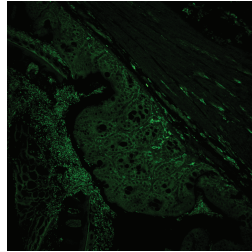
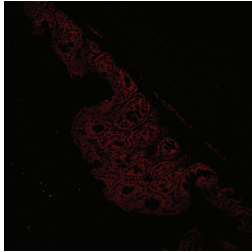
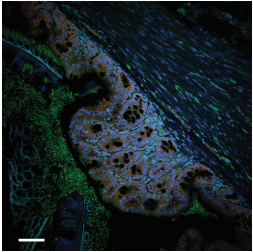
Merged

ST2

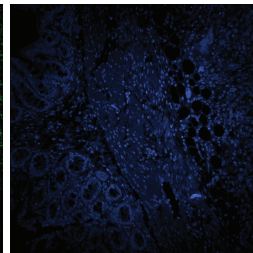
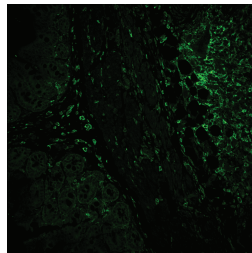
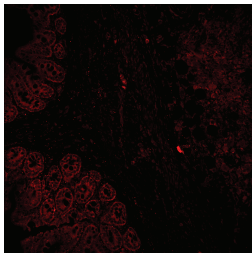
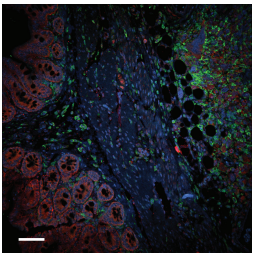
F4/80

DAPI

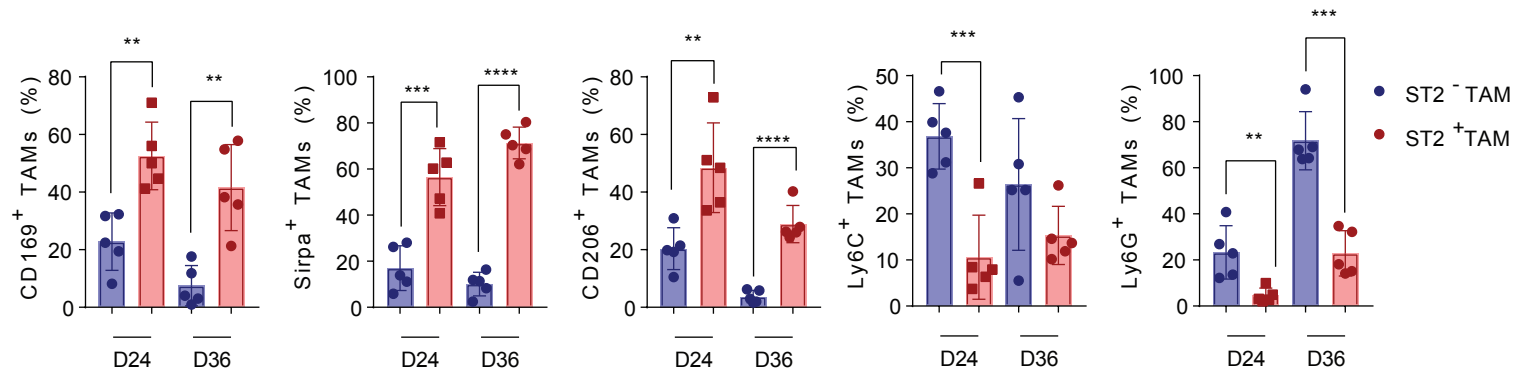
Healthy



Tumor

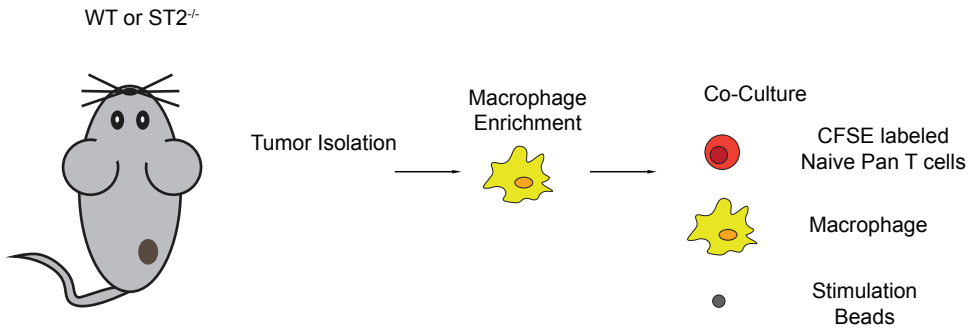


Supplementary Figure 7. Kinetic analysis of ST2⁺TAMs in MC38 tumors. Flow cytometry analysis of ST2⁺TAMs and ST2⁻TAMs from MC38 tumors at the indicated time point ($n=5$; Data displayed as mean \pm SD). Significance was determined by two-tailed unpaired t test.

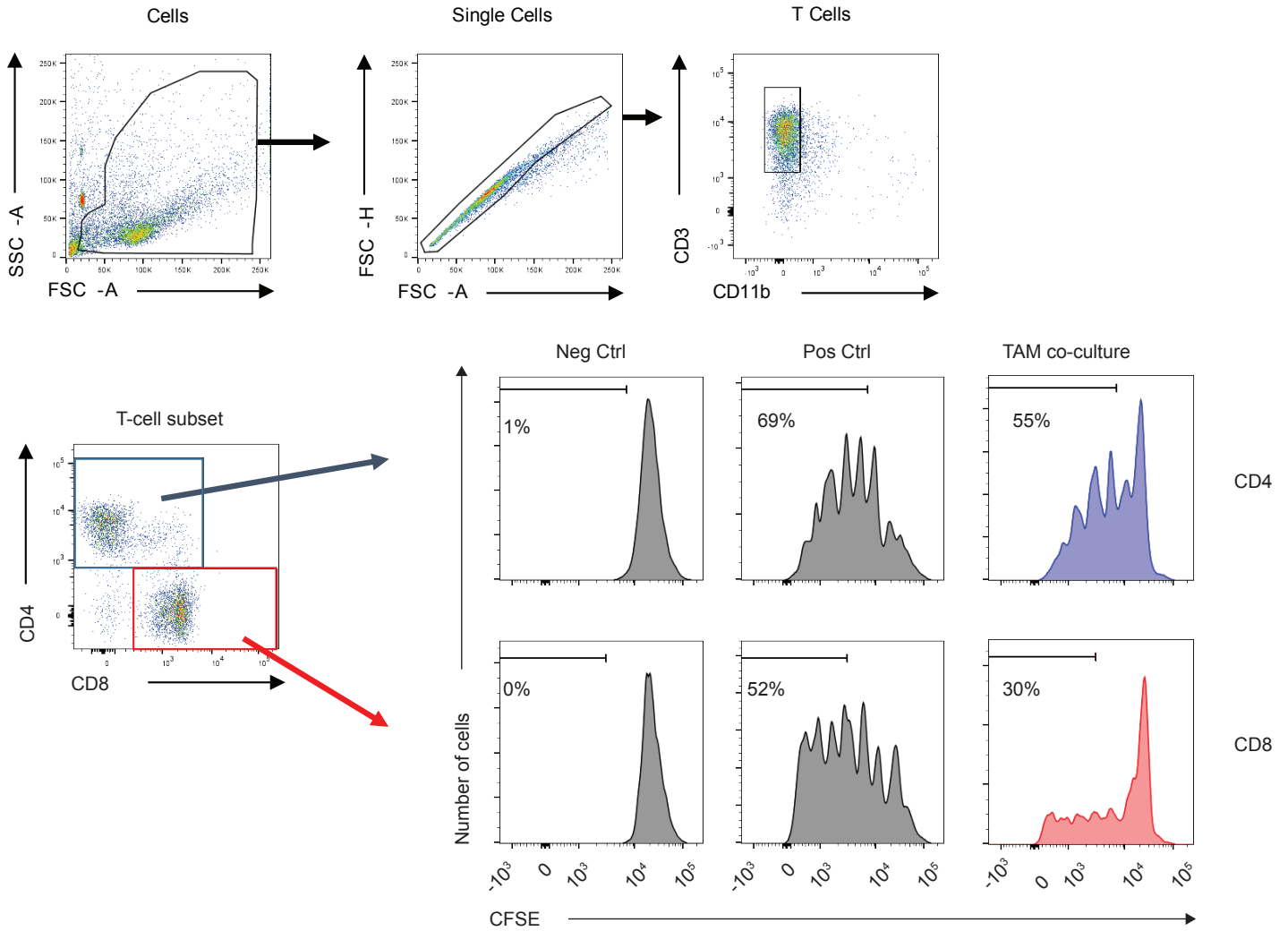


Supplementary Figure 8. Schematic representation of the macrophage suppression assay and gating of T cells. (A) Tumors are processed into single cells; the macrophages are then enriched using MACS sorting and co-cultured with CFSE-labeled pan T cells isolated from naïve mice. (B) The inhibition of T-cell proliferation was assessed 5 days post co-culture. The proliferation of T cells was assessed using the following gating strategy. (C) Quantification of CD4⁺ T-cell proliferation upon TAM co-culture ($n=6$; Data displayed as mean \pm SEM). (D) Representative F4/80 stained tumor immunohistochemistry images from control and anti-CSF1R treated mice. Scale bar represents 50 μ m. Significance was determined by two-tailed unpaired t test (C).

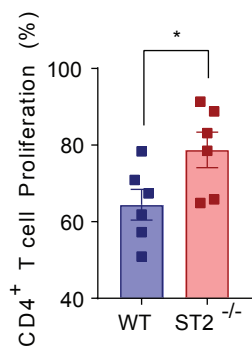
A



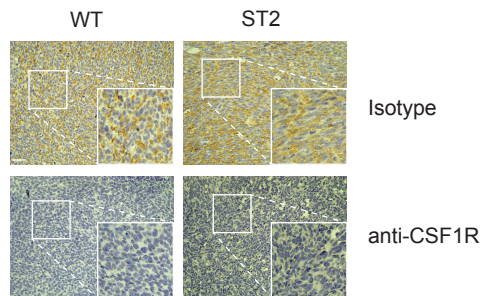
B



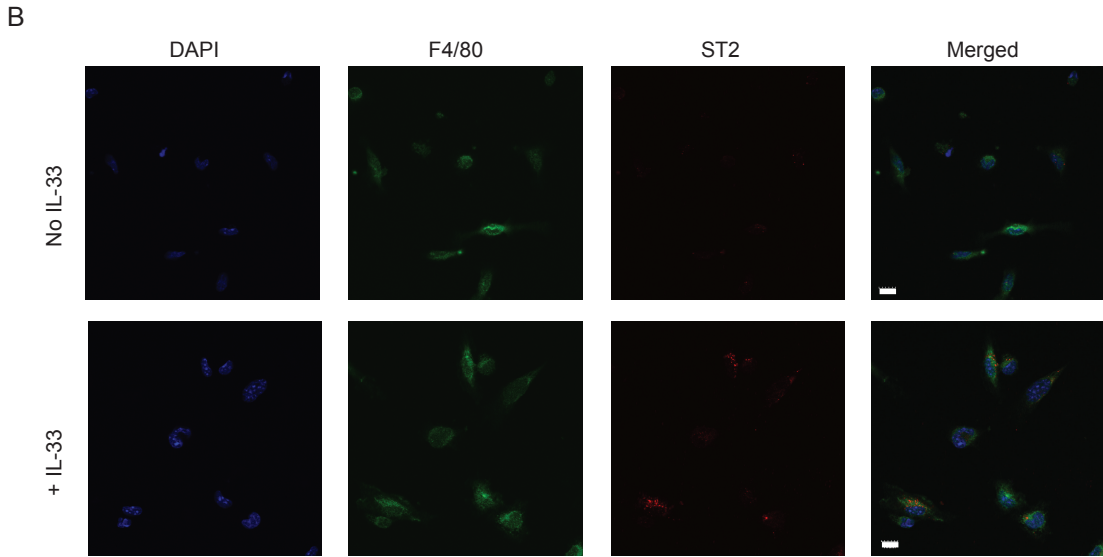
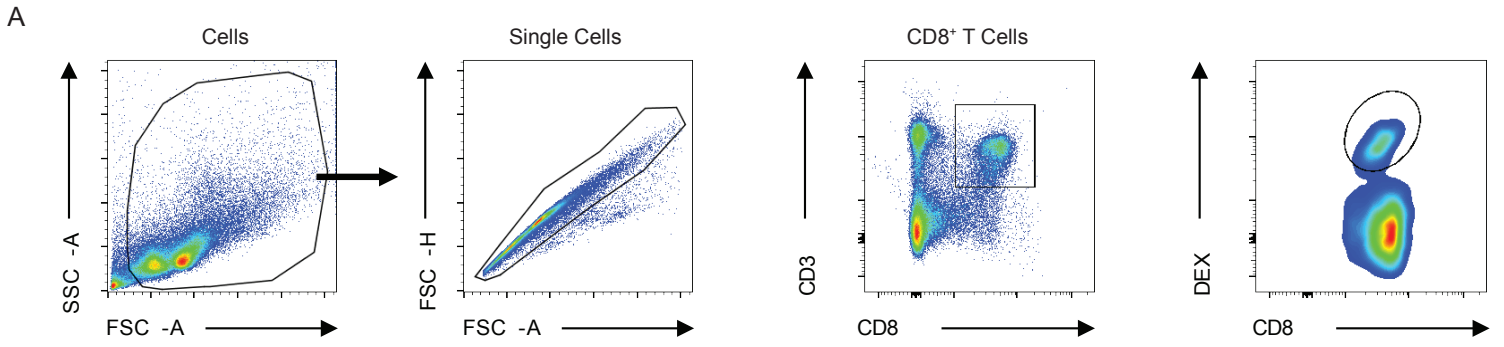
C



D



Supplementary Figure 9. Gating Strategy for OVA-specific CD8⁺ T cells, IL-33 enhances TAM ST2 expression. (A) Flow cytometry gating strategy for the quantification of blood and splenic OVA-specific CD8⁺ T cells. (B) Representative confocal images from MACS F4/80 sorted macrophages from *WT* mice bearing MC38 tumors and treated or not with IL-33. Scale bar represents 10 μ m.



Supplementary Figure 10. Normal phenotype of IL-33trap treated tumor-bearing mice and T-cell infiltration. (A) Body weight of mice treated with PBS (Control, Ctrl) or with IL-33trap (Ctrl, $n=7$, IL-33trap, $n=5$; Data displayed as mean \pm SEM). (B) H&E pictures from the indicated organs in MC38-tumor bearing mice, treated with PBS only (Control) or IL-33trap ($n=3$). Scale bar represents: Heart (control 1x: 4 mm; control 15x: 200 μ m; IL-33trap 1x: 3 mm; IL-33trap 15x: 200 μ m), Kidney (control 1x: 3 mm; control 15x: 200 μ m; IL-33trap 1x: 4 mm; IL-33trap 15x: 200 μ m), Liver (control 1x: 4 mm; control 15x: 200 μ m; IL-33trap 1x: 5 mm; IL-33trap 15x: 200 μ m), Lung (control 1x: 3 mm; control 15x: 200 μ m; IL-33trap 1x: 3mm; IL-33trap 15x: 200 μ m), Spleen (control 1x: 3 mm; control 15x: 200 μ m; IL-33trap 1x: 2mm; IL-33trap 15x: 200 μ m). (C) Determination of the number of CD4⁺ T cells and CD8⁺ T cells from ctrl and IL-33trap treated mice ($n=5$; Data displayed as mean \pm SD). Significance was determined by two-way ANOVA (A), and by two-tailed unpaired t test (C).

