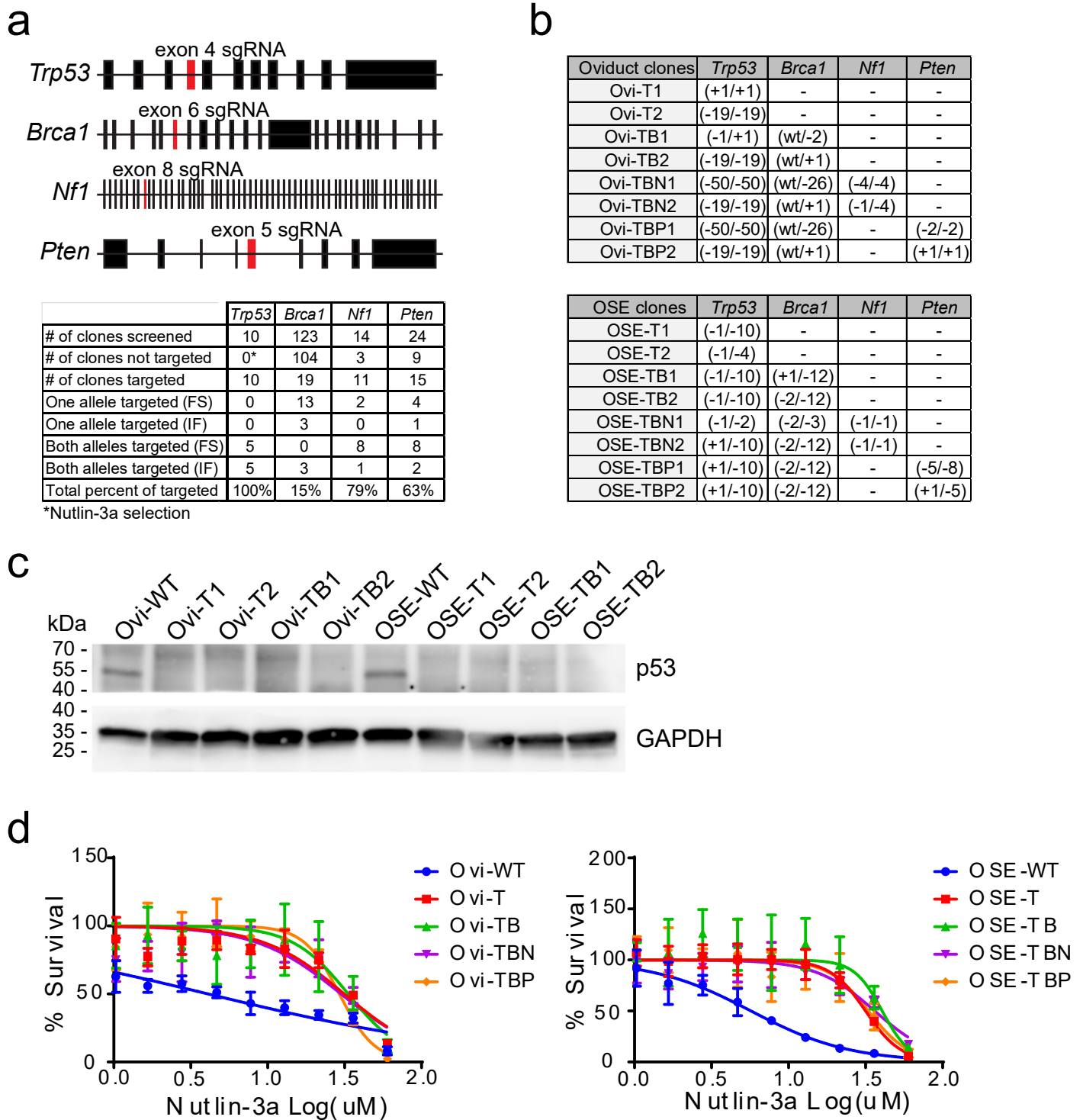


# **Supplementary Information**

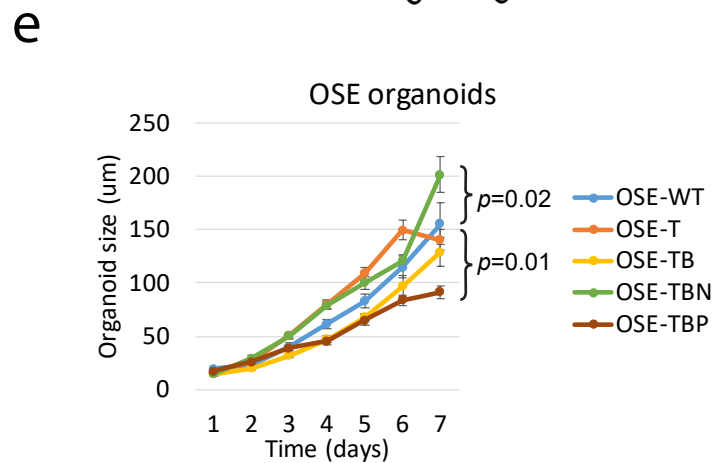
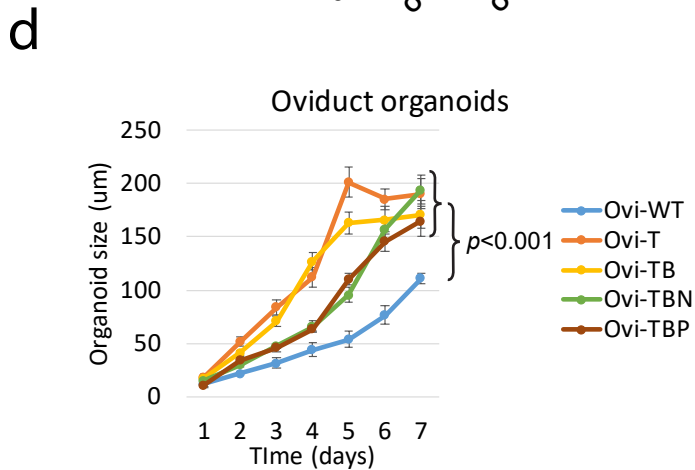
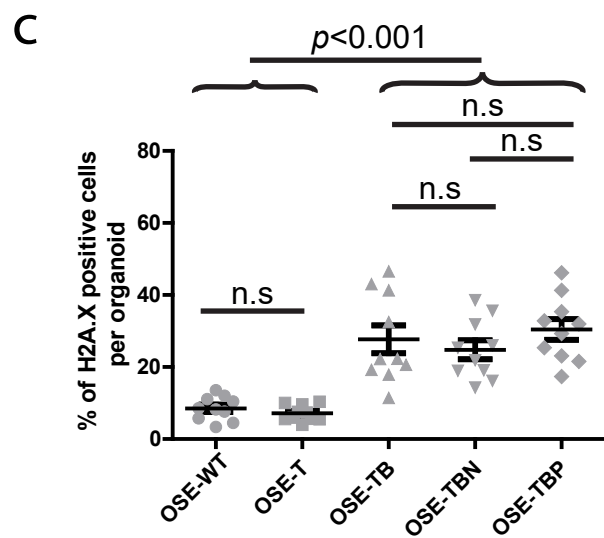
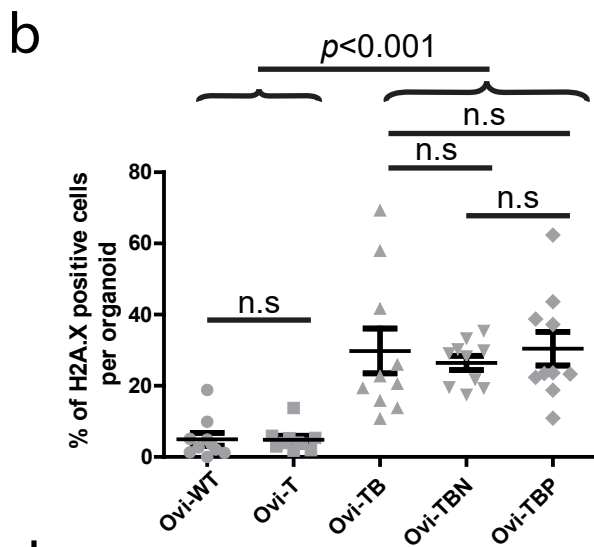
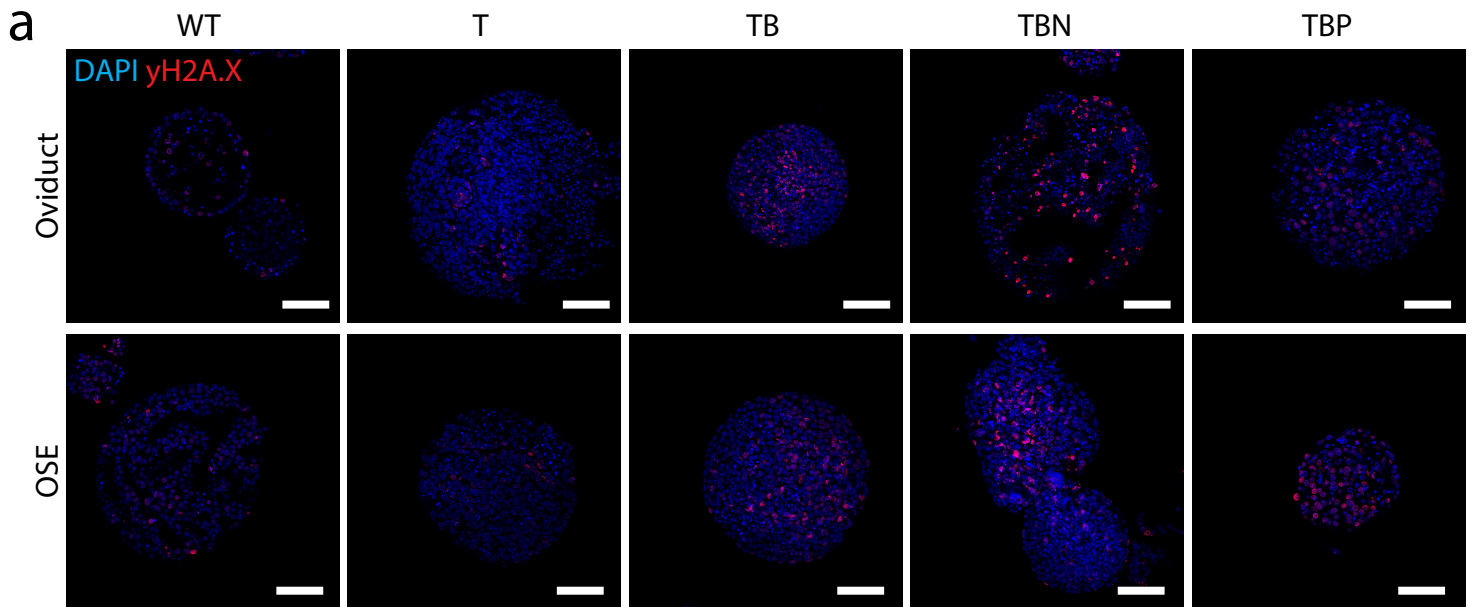
## **Assessing the origin of high-grade serous ovarian cancer using CRISPR-modification of mouse organoids**

Löhmussaar et al.

# Supplementary Figures



**Supplementary Figure 1. Derivation of mutant clones via CRISPR-Cas9.** (a) The sgRNA targeting exons and their targeting efficiency in the indicated genes. Asterisk - followed by Nutlin-3a selection 100% of the clones were targeted in *Trp53* gene. (b) Summary tables of all the established clones and their exact mutations from oviductal (top) and OSE (bottom) origin. (c) Western blot analysis of p53 expression in wild-type, T- and TB-mutant organoids from both lineages. GAPDH is shown as a loading control. Representative from n=2 independent experiments. Uncropped images of the blots are provided in the Supplementary Figure 6. (d) Nutlin-3a sensitivity assay of mutants and respective wild-types from oviductal (left graph) and OSE (right graph) lineages. Dots and error bars represent the mean and  $\pm$ SEM of technical quadruplicates (n=4), respectively, over two independent experiments. (c-d) Ovi - oviduct; WT - wild-type; T - *Trp53* mutant; TB - *Trp53*, *Brca1* mutant; TBN - *Trp53*, *Brca1*, *Nf1* mutant; TBP - *Trp53*, *Brca1*, *Pten* mutant.

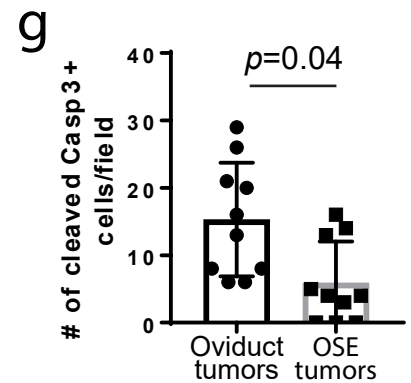
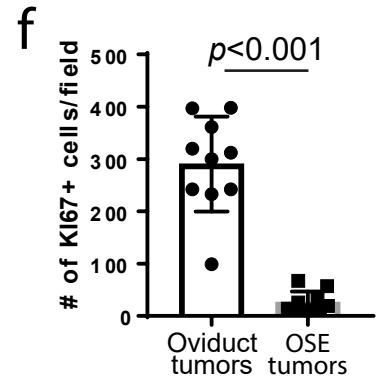
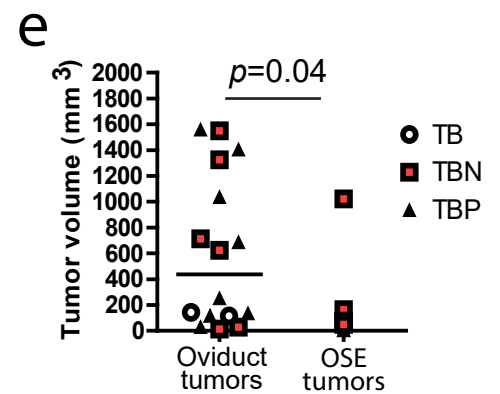
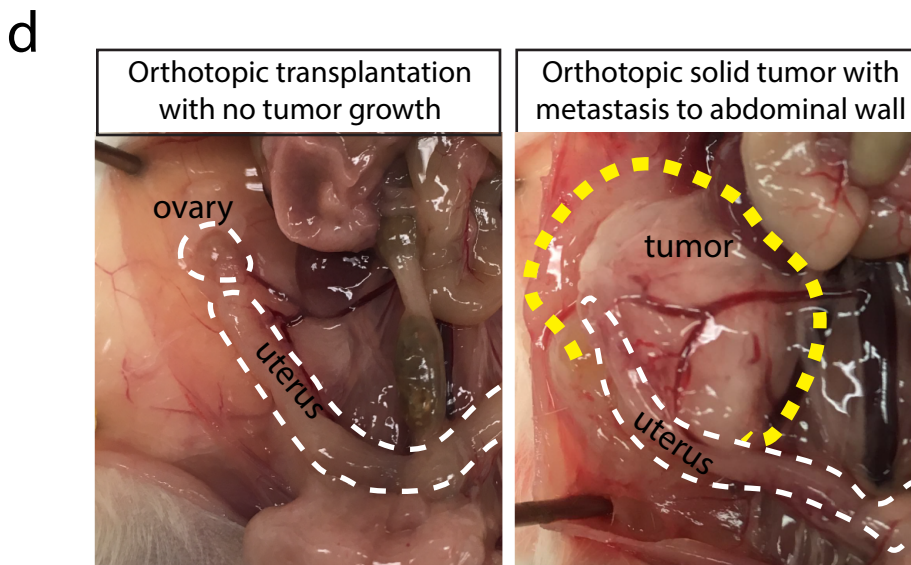
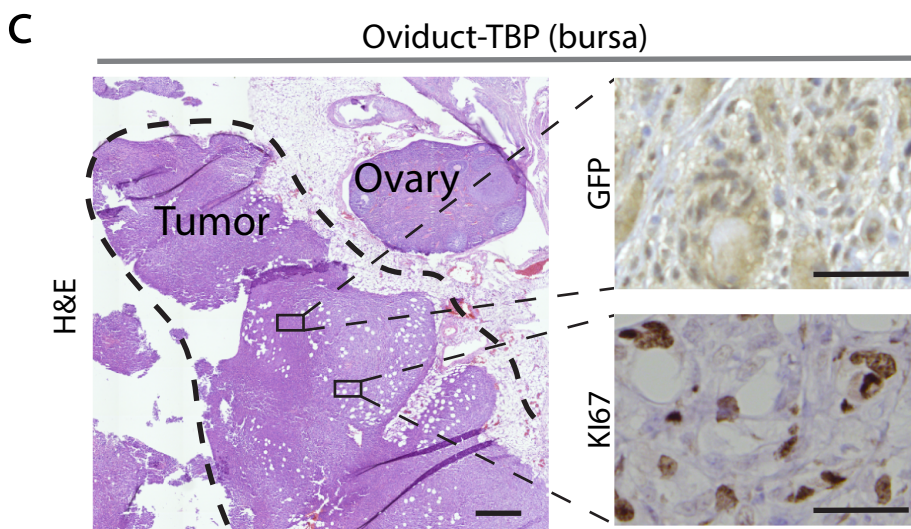
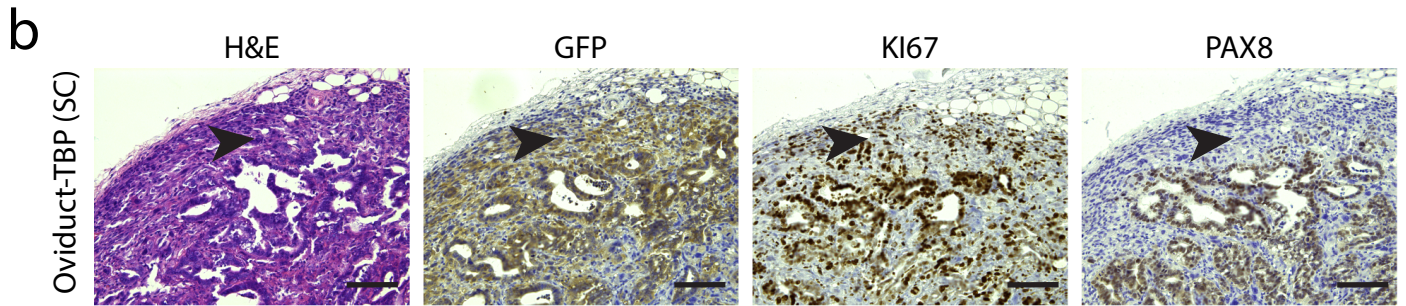
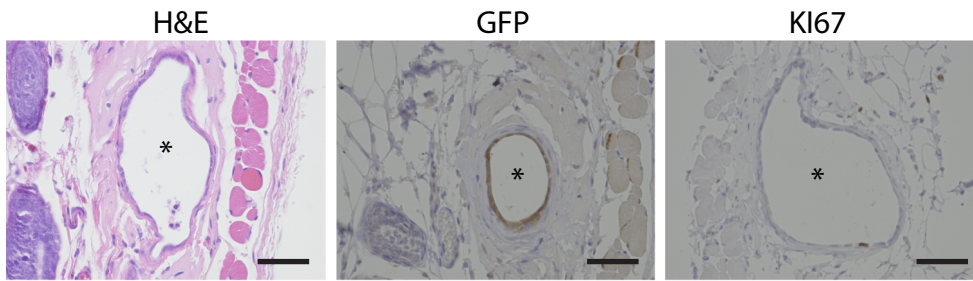


**f**

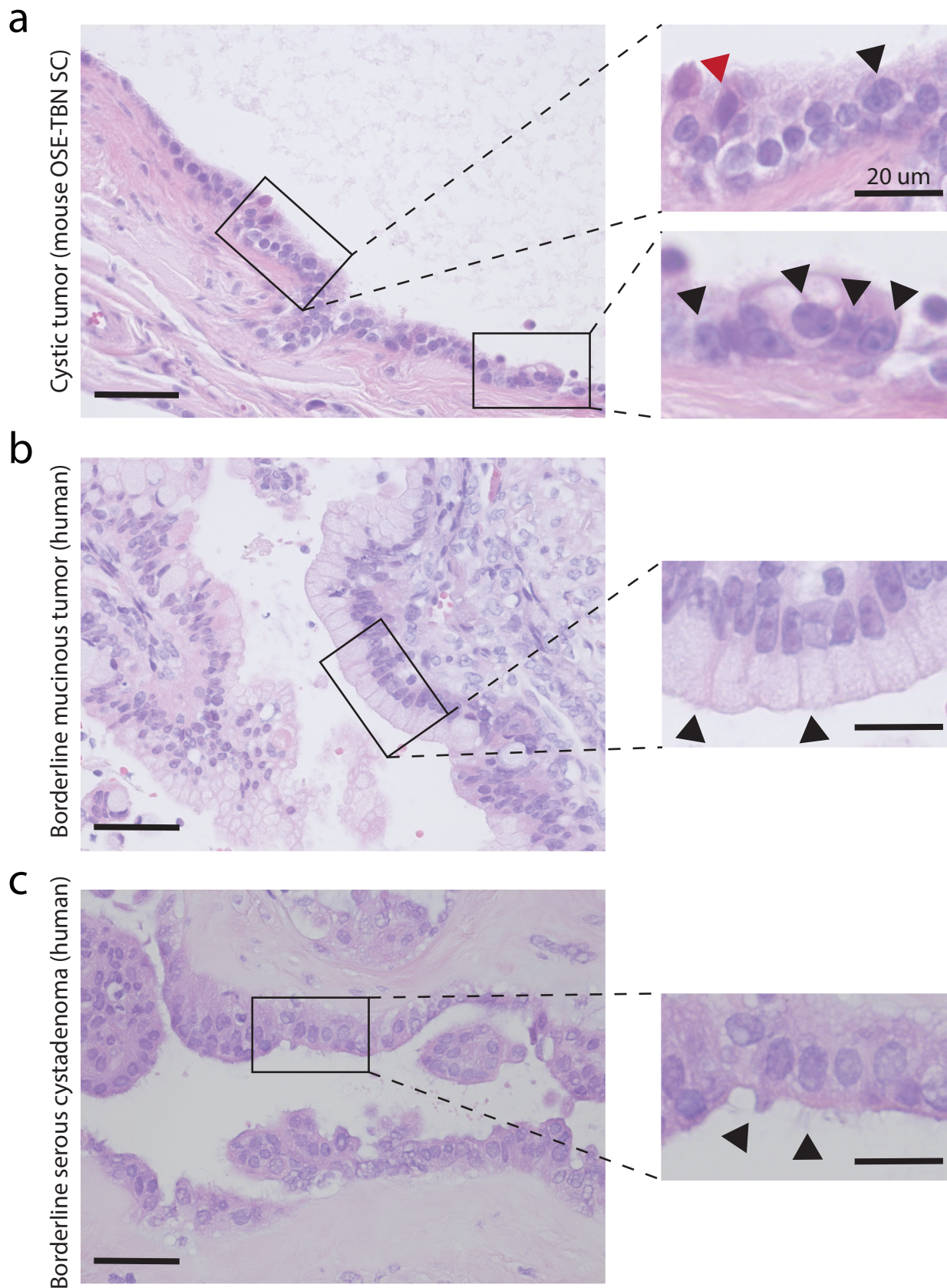
Clone	Live	Early apoptotic	Late apoptotic
	(Annexin V-/PI-)	(Annexin V+/PI-)	(Annexin V+/PI+)
Ovi-WT	53.4%	38.2%	4.2%
Ovi-T	70.2%	24.4%	2.3%
Ovi-TB	66.7%	27.3%	2.8%
Ovi-TBN	79.8%	16.7%	1.3%
Ovi-TBP	80.1%	16.2%	1.2%
OSE-WT	53.7%	27.6%	8.9%
OSE-T	87.1%	7.3%	1.0%
OSE-TB	76.7%	17.6%	1.7%
OSE-TBN	71.8%	21.7%	2.9%
OSE-TBP	43.6%	48.1%	6.3%

**Supplementary Figure 2. Additional characterization of mutant clones.** **(a)** Representative images of DNA damage induction in clones from both lineages after overnight treatment with Mitomycin C, measured by  $\gamma$ H2A.X immunofluorescence (n=2 independent experiments). Scale bar, 100  $\mu$ m. **(b-c)** Quantification of **(a)**: Percentage of nuclei positive for  $\gamma$ H2A.X in Mitomycin C-treated organoids. Error bars represent  $\pm$ SEM (n=10 organoids/line). Statistical significance was calculated by two-tailed Student's t-test, *p*-values were not adjusted for multiple comparisons, n.s - not significant. **(d-e)** Organoid growth assay measured by daily increase in organoid sizes in oviductal **(d)** and OSE **(e)** lineages for a week (n=2 independent experiments). Statistical significance was calculated by two-tailed Student's t-test, *p*-values were not adjusted for multiple comparisons. Error bars represent  $\pm$ SEM (n=12 organoids/line). **(f)** Percentages of cells stained for Annexin V and PI and analysed by flow cytometry to evaluate apoptosis in all clones (n=2 independent experiments). **(a-f)** Ovi - oviduct; WT – wild-type, T – *Trp53* mutant; TB – *Trp53*, *Brca1* mutant; TBN – *Trp53*, *Brca1*, *Nf1* mutant; TBP – *Trp53*, *Brca1*, *Pten* mutant.

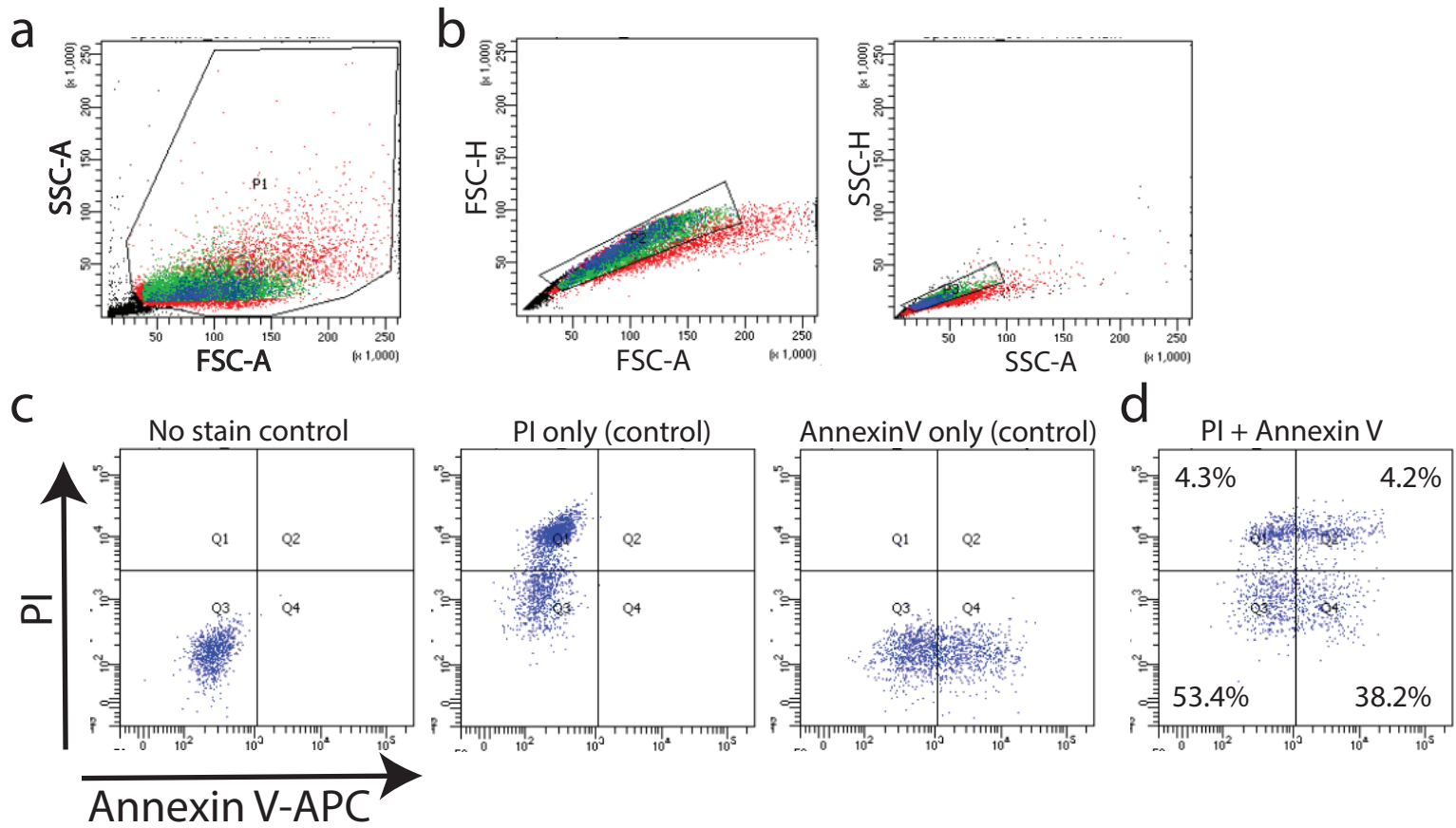
**a** Non-proliferative remnant cysts



**Supplementary Figure 3. Additional characterization of organoid-derived tumors.** (a) Representative histological stainings of non-proliferative remnant cysts (asterisks) observed in subcutaneous transplantation with the wild-type organoids (n=4 injections). Scale bar, 50  $\mu$ m. (b) Histological example of an oviduct-TBP clone-derived subcutaneous tumor showing epithelio-mesenchymal phenotype. H&E, GFP, KI67 and PAX8 stainings are shown (n=4 mice observed). Arrow heads point to the GFP-positive mesenchymal-like cells that have lost the expression of PAX8. Scale bar 100  $\mu$ m. (c) Representative histological stainings of orthotopic solid tumor derived from oviductal TBP clone (n=8 tumors). H&E, GFP and KI67 stainings are shown. H&E staining scale bar, 500  $\mu$ m; GFP/KI67 image scale bar, 50  $\mu$ m. (d) Representative images of orthotopic transplantations with oviductal clones which yielded no tumor (left, n=23 injections) or solid tumor (right, n=15 injections) growth with abdominal wall metastases. Uterus horn and ovary – white dashed line, tumor – yellow dashed line. (e) The distribution and mean of the tumor volumes derived from oviduct (n=16) and OSE (n=7). Statistical significance was calculated by one-sided unpaired Student's t-test. (f) Number of KI67-positive cells per 20x magnification image fields in oviduct- and OSE-derived tumors (5 fields per tumor, 2 tumors/origin, n=10). Error bars represent  $\pm$ SEM. Statistical significance was calculated by two-tailed Student's t-test. (g) Number of cleaved Caspase-3 positive cells per 20x magnification image fields in oviduct- and OSE-derived tumors (5 fields per tumor, 2 tumors/origin, n=10). Error bars represent  $\pm$ SEM. Statistical significance was calculated by two-tailed Student's t-test.

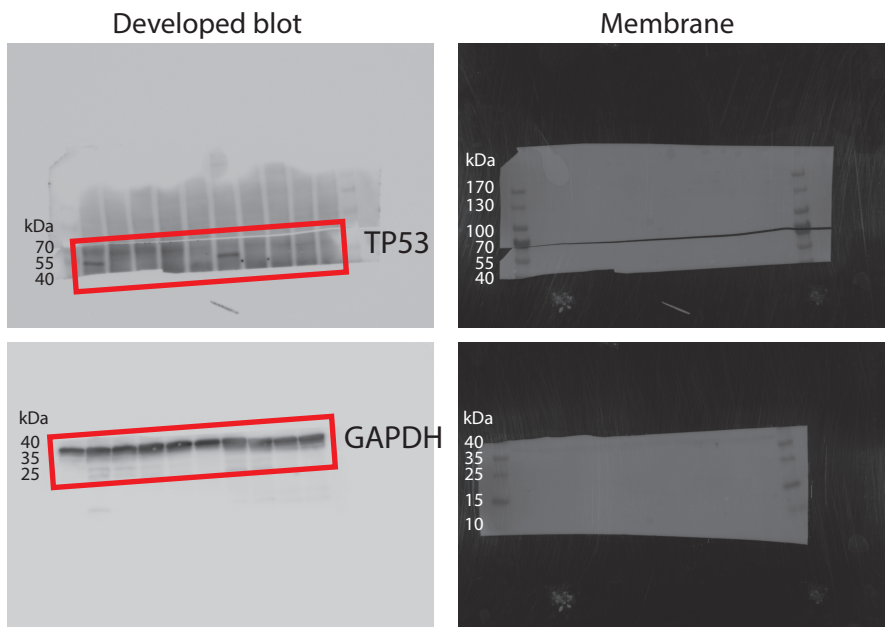


**Supplementary Figure 4. Comparative histological properties of a murine organoid-derived cystic tumor and two distinct human benign ovarian tumors. (a)** Representative image of a murine organoid-derived cystic tumor (n=9 mice observed). As an example, OSE-TBN (*Trp53*, *Brca1*, *Nf1* mutant) clone-derived subcutaneous (SC) tumor is shown. Upper inset: multiple nucleoli (black arrowhead) and nuclear atypia (red arrowhead). Bottom inset: abundant mitotic figures (black arrowheads). **(b)** Human borderline mucinous tumor from a patient. Inset: mucinous glands (arrowheads). **(c)** Human borderline serous cystadenoma from a patient. Inset: cilium (arrowheads). **(a-c)** Large image scale bar, 50  $\mu\text{m}$ ; inset scale bar, 20  $\mu\text{m}$ .



**Supplementary Figure 5. FACS gating strategy.** (a) In this sample gating, the cells were first gated for forward- and side-scatter area (FSC-A vs SSC-A) to select the cell population of interest and exclude the debris. (b) Next, a sequential gating was performed to obtain single cells. The cells were first gated for forward-scatter area and height (FSC-A vs FSC-H) followed by gating for side-scatter area and height (SSC-A vs SSC-H), which allows for higher sensitivity in doublet exclusion. (c) No stain, “PI only” and “Annexin V only” samples were used to set up the gates for the assay. (d) Subsequently, PI and Annexin V-APC double-stained clones were analysed for apoptotic events. Q1: PI-positive and Annexin V-negative necrotic cell fraction, Q2: PI and Annexin V double-positive late apoptotic cell fraction; Q3: PI- and Annexin V-negative live cell fraction; Q4: PI-negative and Annexin V-positive early apoptotic cells. The main results are shown in the Supplementary Figure 2f.





**Supplementary Figure 6. Uncropped images of Western blots.** The panels circled with red rectangles are displayed in the Supplementary Figure 1c.