

Supplementary Information for:

**Receptor Characterization and Functional Activity of Pyrokinins on the Hindgut in the Adult Mosquito, *Aedes aegypti*.**

**Aryan Lajevardi and Jean-Paul V. Paluzzi\***

Laboratory of Integrative Vector Neuroendocrinology, Department of Biology, York University, Toronto, Ontario, Canada, M3J1P3

**\* Corresponding author:**

Jean-Paul V. Paluzzi

[paluzzi@yorku.ca](mailto:paluzzi@yorku.ca)

**This file includes:**

Supplementary Table S1

Supplementary Figure S1

Supplementary Figure S2

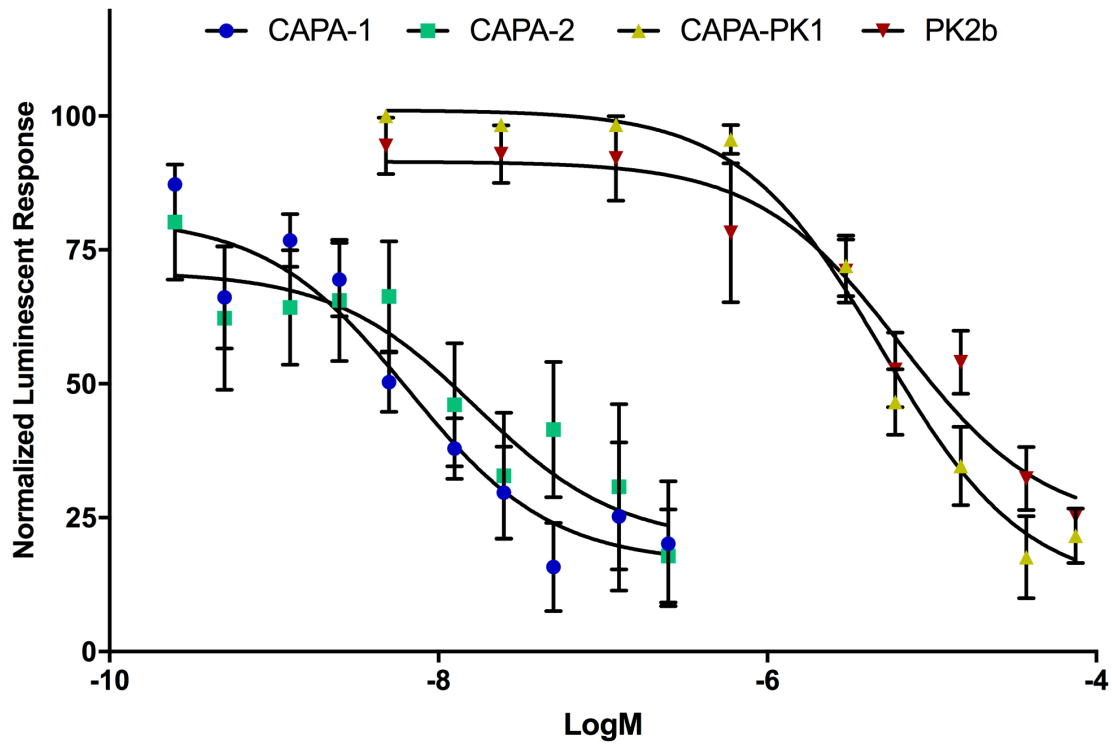
Supplementary Figure S3

Supplementary Video S1 (caption only)

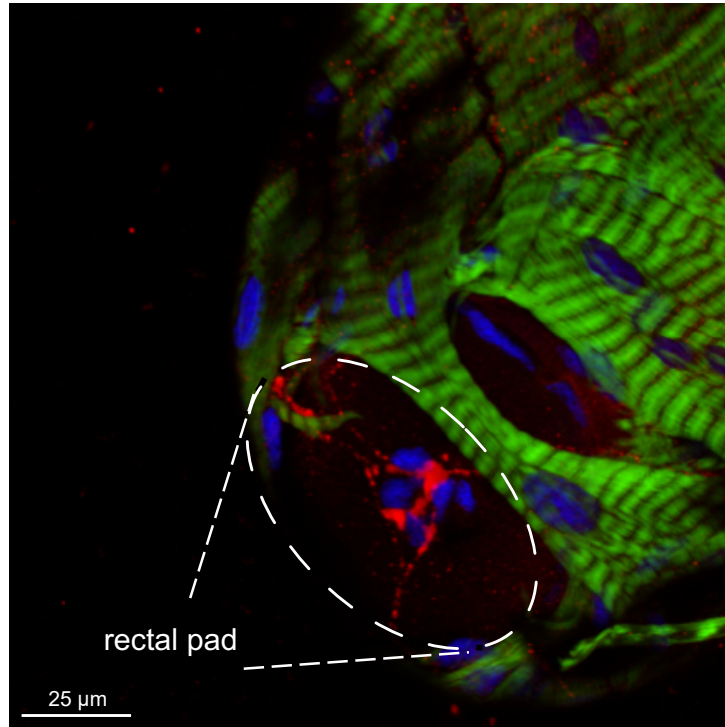
Supplementary Video S2 (caption only)

**Supplementary Table S1.** Sequence characteristics of the various peptides used in the study including endogenous *A. aegypti* neuropeptides and homologous peptides from other insects. Peptides in black font were used in this study whereas peptides in blue font are *A. aegypti* peptides listed for sequence comparison. The family-specific C-terminal motifs and conserved residues required for peptide bioactivity are denoted in red font.

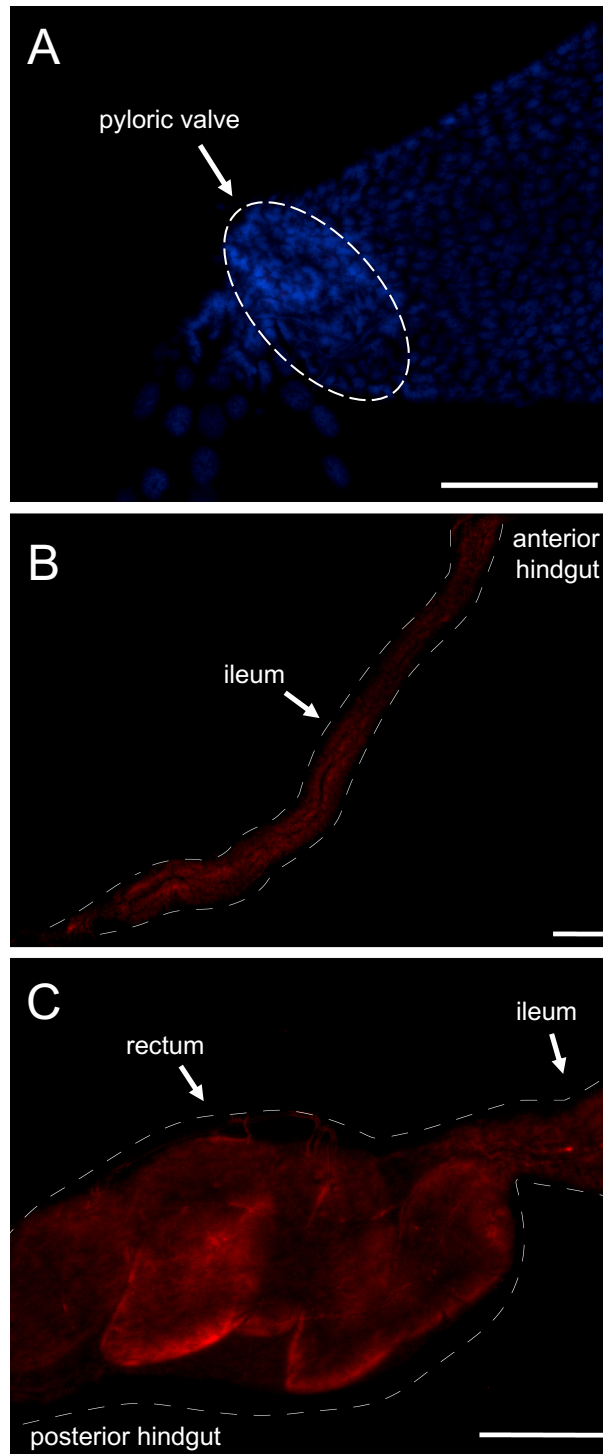
Peptide	Sequence	Reference
<i>Aedae</i> CAPA-PK1	AGNSGANSGM <b>WFGPRL</b> -NH <sub>2</sub>	Predel et al., 2010
<i>Aedae</i> PK1	AAAM <b>WFGPRL</b> -NH <sub>2</sub>	Predel et al., 2010
<i>Aedae</i> PK2	DASSSNENNSRPP <b>FAPRL</b> -NH <sub>2</sub>	Predel et al., 2010
<i>Aedae</i> PK3	NLP <b>FSPRL</b> -NH <sub>2</sub>	Predel et al., 2010
<i>Rhopr</i> PK2a	NTVN <b>FSPRL</b> -NH <sub>2</sub>	Paluzzi and O' Donnell, 2012
<i>Rhopr</i> PK2b	SPP <b>FAPRL</b> -NH <sub>2</sub>	Paluzzi and O' Donnell, 2012
<i>Aedae</i> CAPA-1	GPTVGLFA <b>FPRV</b> -NH <sub>2</sub>	Predel et al., 2010
<i>Aedae</i> CAPA-2	pQGLVP <b>FPRV</b> -NH <sub>2</sub>	Predel et al., 2010
<i>Droso</i> -leucokinin	NSVVLGKKQR <b>FHSWG</b> -NH <sub>2</sub>	Zandawala et al., 2018
<i>Aedae</i> -leucokinin 1	NSKYVSKQK <b>FYSWG</b> -NH <sub>2</sub>	Veenstra, 1994
<i>Aedae</i> -leucokinin 2	NP <b>FHAWG</b> -NH <sub>2</sub>	Veenstra, 1994
<i>Aedae</i> -leucokinin 3	NNPNV <b>FYPWG</b> -NH <sub>2</sub>	Veenstra, 1994
<i>Rhopr</i> MIP-7	AWNSLHGG <b>W</b> -NH <sub>2</sub>	Paluzzi et al., 2015
<i>Aedae</i> MIP1	TWKNLQGG <b>W</b> -NH <sub>2</sub>	Kim et al., 2010
<i>Aedae</i> MIP2	AWNKINGG <b>W</b> -NH <sub>2</sub>	Kim et al., 2010
<i>Aedae</i> MIP3	VNAGPAQ <b>WNKFRGSW</b> -NH <sub>2</sub>	Kim et al., 2010
<i>Aedae</i> MIP4	EPG <b>WNNLKGLW</b> -NH <sub>2</sub>	Kim et al., 2010
<i>Aedae</i> MIP5	SEK <b>WNKLSSSW</b> -NH <sub>2</sub>	Kim et al., 2010



**Supplementary Figure S1.** Competitive ELISA used to confirm cross-reactivity and binding affinity of CAPA2-targeted antibody to other structurally related peptides. The custom-synthesized antibody designed to target the antigen sequence EGGFISFPRV-NH<sub>2</sub> selectively targets *Aedae*CAPA-1 (IC<sub>50</sub> = 6.57nM) and *Aedae*CAPA-2 (IC<sub>50</sub> = 16.93nM), but was also able to recognize and bind pyrokinins, including *Aedae*CAPA-PK1 (IC<sub>50</sub> = 4.99μM) and *Rhopr*PK2b (IC<sub>50</sub> = 6.43μM). Normalized responses represent mean ± SEM (n = 4).



**Supplementary Figure S2.** PRXa-like immunoreactive processes (red) terminating in close association to cells within the rectal pad (one of six shown) of adult female mosquitoes. No co-localization with phalloidin-stained F-actin (green) was detected.



**Supplementary Figure S3.** Pre-incubation of primary antibody with 5  $\mu$ M *AedaeCAPA-PK1* abolishes immunoreactivity along the pyloric valve (A), ileum (B) and rectum (C). All microscope acquisition exposures are equalized to settings used for experimental treatments. Scale bars, 100  $\mu$ m.

Videos:

**Supplementary Video S1.** Sample recording of mosquito ileal motility in response to saline (vehicle control), *RhoprPK2b* and 5-HT (stimulatory control). While *RhoprPK2b* decreased contraction frequency, 5-HT reversed this effect and notably increased activity above baseline levels. All video speeds are increased to 300%.

**Supplementary Video S2.** Sample recording of mosquito ileal motility in response to saline (vehicle control), *RhoprPK2b* and *RhoprMIP-7* (inhibitory control). Both *RhoprPK2b* and *RhoprMIP-7* treatments reduced contractile activity of dissected ilea. All video speeds are increased to 300%.

## References

- Kim, Y.J., Bartalska, K., Audsley, N., Yamanaka, N., Yapici, N., Lee, J.Y., et al. (2010). MIPs are ancestral ligands for the sex peptide receptor. *Proc. Natl. Acad. Sci. U. S. A.* 107, 6520–6525. doi: 10.1073/pnas.0914764107
- Paluzzi, J.-P. V., Haddad, A. S., Sedra, L., Orchard, I., and Lange, A.B. (2015). Functional characterization and expression analysis of the myoinhibiting peptide receptor in the Chagas disease vector, *Rhodnius prolixus*. *Mol. Cell. Endocrinol.* 399, 143–153. doi: 10.1016/j.mce.2014.09.004
- Paluzzi, J.-P. V., and O'Donnell, M. J. (2012). Identification, spatial expression analysis and functional characterization of a pyrokinin-1 receptor in the Chagas' disease vector, *Rhodnius prolixus*. *Mol. Cell. Endocrinol.* 363, 36–45. doi: 10.1016/j.mce.2012.07.007
- Predel., R., Neupert, S., Garczynski, S., Crim, J.W., Brown, M. R., Russell, W.K., et al. (2010). Neuropeptidomics of the mosquito *Aedes aegypti*. *J. Proteome Res.* 9, 2006–2015. doi: 10.1021/pr901187p
- Veenstra, J.A. (1994). Isolation and identification of 3 leucokininins from the mosquito *Aedes aegypti*. *Biochem. Biophys. Res. Commun.* 202, 715–719. doi: 10.1006/bbrc.1994.1989
- Zandawala, M., Marley, R., Davies, S.A., and Nässel, D.R. (2018). Characterization of a set of abdominal neuroendocrine cells that regulate stress physiology using colocalized diuretic peptides in *Drosophila*. *Cell. Mol. Life Sci.* 75, 1099–1115. doi: 10.1007/s00018-017-2682-y