

Supporting Information – Characterization of the Effect of Drug Metabolism on the Gas-Phase Structures of Drugs Using Ion Mobility-Mass Spectrometry

*Dylan H. Ross, Ryan P. Seguin, and Libin Xu**

Department of Medicinal Chemistry, University of Washington, Seattle, WA 98195

Correspondence:

Libin Xu, Ph.D.
Department of Medicinal Chemistry
University of Washington
Tel: (206) 543-1080
Fax: (206) 685-3252
Email: libinxu@uw.edu

Contents

1 – Experimental Section

1.1 – Synthesis of Benzalkonium Chlorides (BACs) and ω -OH Metabolites

1.2 – Ion Mobility-Mass Spectrometry Electrospray Conditions

1.3 – Scheme S1 – Computational Modeling Workflow

2 – Results and Discussion

2.1 – $^1\text{H-NMR}$ and HRMS Characterization of Synthesized BACs and ω -OH Metabolites

2.2 – Computational Modeling on Sodiated Adduct of Quercetin Glucuronide

3 – Tables and Figures

3.1 – Figure S1 – Initial Characterization of Midazolam Metabolites

3.2 – Table S1 – Experimental CCS Values for Drugs and Observed Metabolites

3.3 – Figure S2 – Expected Metabolites from Literature, Observed Metabolites, and
Fragmentation Data for Drugs

3.4 – Figure S3 – Theoretical CCS of Sodiated Quercetin Glucuronide Isomers

4 – References

1 – Experimental Section

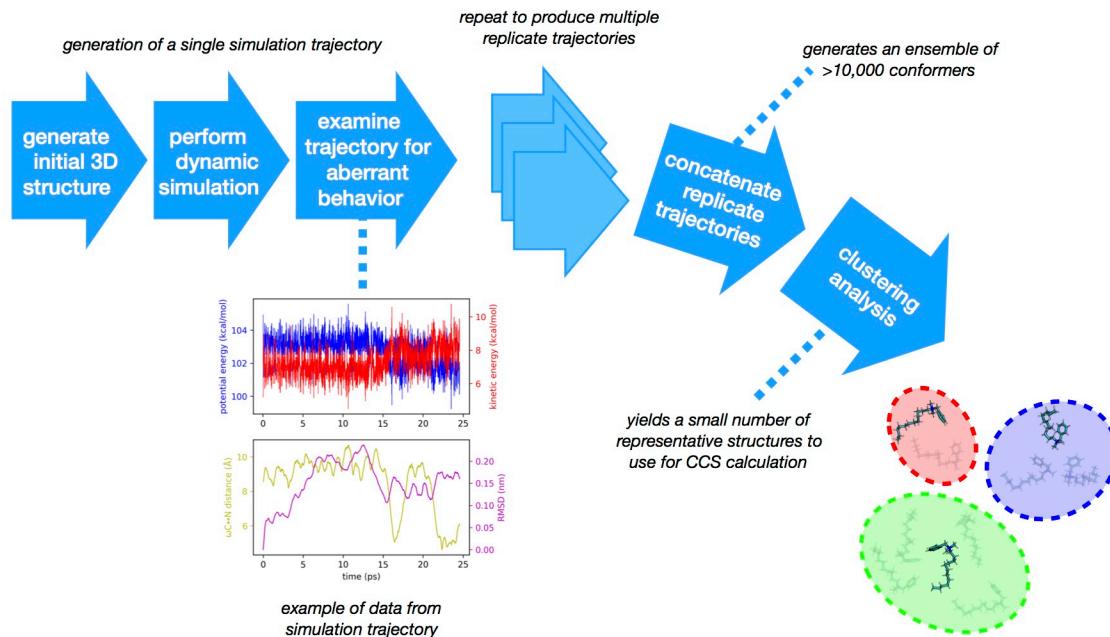
1.1 – Synthesis of Benzalkonium Chlorides (BACs) and ω -OH Metabolites

BACs were synthesized by nucleophilic coupling of N,N-dimethylbenzylamine with 1.4 equivalents of an alkyl chloride of the appropriate alkyl chain length in ethanol heated under reflux¹. The corresponding ω -hydroxy versions of each BAC were prepared in the same manner by substituting the alkyl chloride with the appropriate ω -hydroxy alkyl chloride. Products were recrystallized from hot acetone and rinsed with cold diethyl ether. Chemical identities were confirmed by ¹H-NMR and/or high-resolution mass spectrometry (HRMS).

1.2 – Ion Mobility-Mass Spectrometry Electrospray Conditions

IM-MS analysis was performed on a Waters Synapt G2-Si HDMS (Waters Corp., Milford, MA) equipped with an electrospray ionization (ESI) source using nitrogen as the drift gas. ESI conditions were as follows: capillary, +2.5 kV; sampling cone, 40 V; source temperature, 100 °C; desolvation temperature, 250 °C; cone gas, 50 L/h; and desolvation gas, 600 L/h.

1.3 – Scheme S1 – Computational Modeling Workflow



2 – Results and Discussion

2.1 – $^1\text{H-NMR}$ and HRMS Characterization of Synthesized BACs and ω -OH Metabolites

C₄ BAC: HRMS [M-Cl⁺] (C₁₃H₂₂N): *observed*, 192.1747; *theoretical*: 192.1752.

C₆ BAC: HRMS [M-Cl⁺] (C₁₅H₂₆N): *observed*, 220.2063; *theoretical*: 220.2065.

C₈ BAC: HRMS [M-Cl⁺] (C₁₇H₃₀N): *observed*, 248.2375; *theoretical*: 248.2378.

C₁₀ BAC: HRMS [M-Cl⁺] (C₁₉H₃₄N): *observed*, 276.2694; *theoretical*: 276.2691.

ω -OH C₄ BAC: HRMS [M-Cl⁺] (C₁₃H₂₂NO): *observed*, 208.1694; *theoretical*: 208.1701.

ω -OH C₆ BAC: HRMS [M-Cl⁺] (C₁₅H₂₆NO): *observed*, 236.2010; *theoretical*: 236.2014.

ω -OH C₈ BAC: HRMS [M-Cl⁺] (C₁₇H₃₀NO): *observed*, 264.2327; *theoretical*: 264.2327.

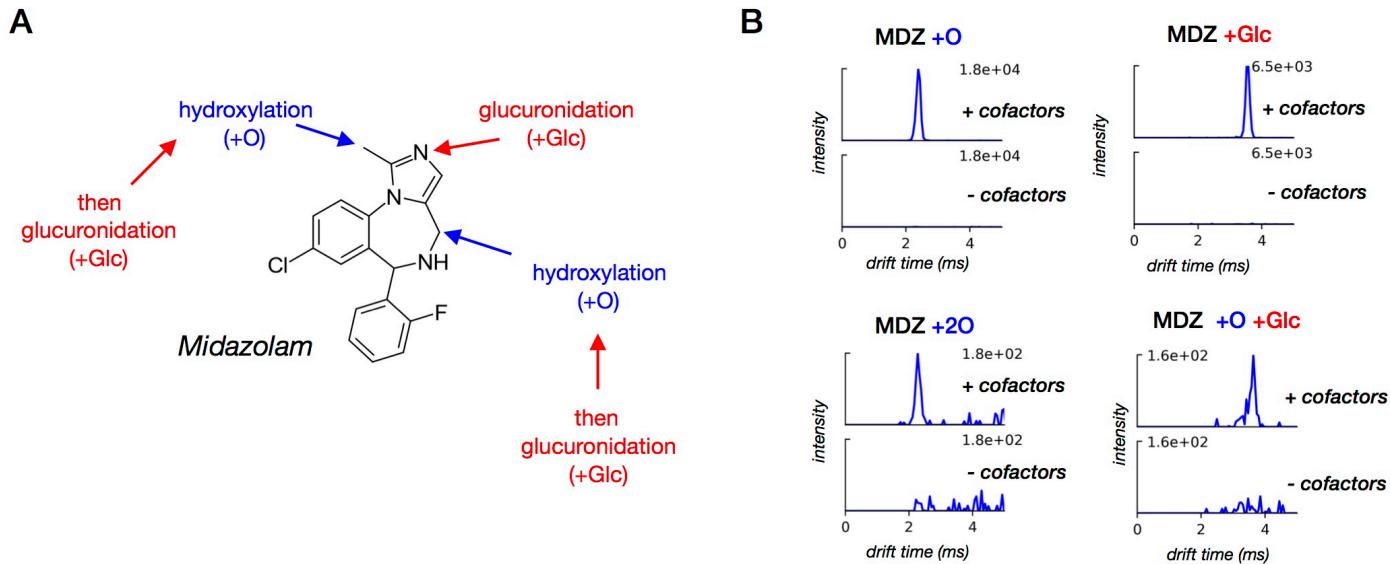
ω -OH C₁₀ BAC: $^1\text{H-NMR}$ (CDCl₃, 500 MHz): 1.29 and 1.36 (*br s*, 12H), 1.56 (*m*, 2H), 1.81 (*m*, 2H), 3.30 (*s*, 6H), 3.53 (*m*, 2H), 3.64 (*q*, 2H, *J* = 6.1 Hz), 5.03 (*s*, 2H), 7.47 (*m*, 3H), 7.64 (*d*, 2H, *J* = 7.0 Hz); MS [M-Cl⁺] (C₁₉H₃₄NO): *observed*, 292.2639; *theoretical*: 292.2640.

2.2 – Computational Modeling on Sodiated Adduct of Quercetin Glucuronide

Both protonated and sodiated MS adducts of quercetin glucuronide displayed bimodal ATDs (Figure S1A), indicating conformational heterogeneity and/or isomerism. Sodiated adducts of all potential positional isomers of quercetin glucuronide (3-, 3'-, 4'-, 5-, 7-) were modeled using the same computational methodology as was applied to the protonated isomers. Representative minimum energy structures are presented in Figure S1B, and their corresponding theoretical CCS values in Figure S1C (measured values as dotted lines). The theoretical CCS values follow the same trend as was observed for the protonated isomers: the higher CCS peak likely corresponds to the 7-isomer (due to lack of formation of the 5-isomer in human hepatic metabolism²) while the lower CCS peak likely has contributions from the 3-, 3'-, and/or 4'-isomers.

3 – Tables and Figures

3.1 – Figure S1 – Initial Characterization of Midazolam Metabolites



A Expected metabolism for midazolam^{3,4} (MDZ). **B** Arrival time distributions (ATDs) for observed primary and secondary metabolites of MDZ showing cofactor-dependent formation.

3.2 – Table S1 – Experimental CCS Values for Drugs and Observed Metabolites

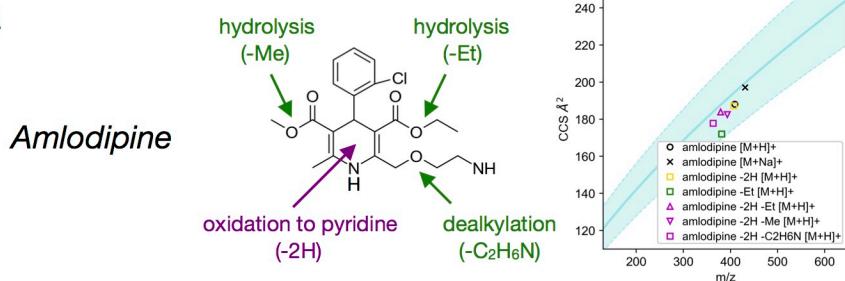
compound	metabolite	adduct	m/z	CCS (\AA^2)	CCS RSD (%)	Compaction factor (C)
C ₄ BAC		[M] ⁺	192.1838	144.96	0.52	
C ₄ BAC	ω -OH	[M] ⁺	208.1795	148.19	0.22	1.032
C ₆ BAC		[M] ⁺	220.2167	157.07	0.51	
C ₆ BAC	ω -OH	[M] ⁺	236.2128	158.92	0.26	1.036
C ₈ BAC		[M] ⁺	248.2505	170.09	0.55	
C ₈ BAC	ω -OH	[M] ⁺	264.2128	170.14	0.31	1.042
C ₁₀ BAC		[M] ⁺	276.2869	182.24	0.60	
C ₁₀ BAC	ω -OH	[M] ⁺	292.2815	178.99	0.23	1.057
C ₁₂ BAC		[M] ⁺	304.3187	193.19	0.21	
C ₁₂ BAC	ω -OH	[M] ⁺	320.3119	187.55	0.20	1.066
C ₁₄ BAC		[M] ⁺	332.3427	203.67	0.27	
C ₁₄ BAC	ω -OH	[M] ⁺	348.3444	196.33	0.24	1.070
C ₁₆ BAC		[M] ⁺	360.3843	212.76	0.16	
C ₁₆ BAC	ω -OH	[M] ⁺	376.3800	205.00	0.53	1.068
amlodipine		[M+H] ⁺	409.1586	188.14	0.53	
amlodipine		[M+Na] ⁺	431.1457	197.21	0.24	
amlodipine	-2H	[M+H] ⁺	407.1475	187.24	0.04	1.002
amlodipine	-2H, -Me	[M+H] ⁺	393.1390	182.61	0.11	1.003
amlodipine	-2H, -Et	[M+H] ⁺	379.1376	183.95	0.10	0.972
amlodipine	-2H, -C ₂ H ₆ N	[M+H] ⁺	363.1180	177.84	0.08	0.977
amlodipine	-Et	[M+H] ⁺	381.0955	172.06	0.30	1.043
amlodipine	-Et	[M+Na] ⁺	403.1226	207.14	0.20	0.910
bupropion		[M+H] ⁺	240.1217	157.37	0.24	

bupropion	+2H	[⁽³⁷ Cl)M+H] ⁺	244.1346	156.07	0.31	1.020
chlorpromazine		[M+H] ⁺	319.1169	169.58	0.32	
chlorpromazine		[M+Na] ⁺	341.1305	170.83	0.42	
chlorpromazine	-Me	[M+H] ⁺	305.0947	167.66	0.47	0.982
chlorpromazine	+O	[M+H] ⁺	335.1066	170.74	0.55	1.026
clomifene		[M+H] ⁺	406.2054	203.68	0.60	
clomifene	+O	[M+H] ⁺	422.1923	209.10	0.11	0.999
clomifene	-Et	[M+H] ⁺	378.1691	197.10	0.11	0.985
clomifene	+O, -Et	[M+H] ⁺	394.1640	200.20	0.12	0.997
clopido <u>grel</u>		[M+H] ⁺	322.0720	167.32	0.27	
clopido <u>grel</u>	+O	[M+H] ⁺	338.0700	172.49	0.26	1.002
clozapine		[M+H] ⁺	327.1458	177.67	0.27	
clozapine		[M+Na] ⁺	349.1254	178.03	0.17	
clozapine	+O	[M+H] ⁺	343.1388	179.97	0.23	1.019
clozapine	+GSH	[M+H] ⁺	632.2207	238.91	0.11	1.154
dextromethorphan		[M+H] ⁺	272.2093	164.54	0.38	
dextromethorphan		[M+Na] ⁺	294.2091	174.13	0.91	
dextromethorphan	-Me	[M+H] ⁺	258.1915	159.93	0.43	0.993
diclofenac		[M+H] ⁺	296.0769	159.05	0.45	
diclofenac		[M+Na] ⁺	318.0157	159.32	1.19	
diclofenac	+O	[M+Na] ⁺	334.0125	167.11	1.07	0.985
diclofenac	+Glc	[M+Na] ⁺	494.0488	200.79	0.50	1.064
diclofenac	+O, +Glc	[M+Na] ⁺	510.0204	202.54	0.38	1.078
midazolam		[M+H] ⁺	326.0963	171.68	0.32	
midazolam	+O	[M+H] ⁺	342.0881	175.13	0.26	1.012
midazolam	+Glc	[M] ⁺	502.1203	211.85	1.45	1.081
progesterone		[M+H] ⁺	315.2368	178.43	0.26	
progesterone		[M+Na] ⁺	337.2199	202.53	0.16	
progesterone		[M+H-H ₂ O] ⁺	297.2274	172.96	0.58	
progesterone	+O	[M+H] ⁺	331.2334	181.35	0.15	1.017
progesterone	+2H	[M+H] ⁺	317.2529	178.82	0.17	1.002
progesterone	+4H, +Glc	[M+H] ⁺	495.2497	215.41	0.22	1.119
quercetin		[M+H] ⁺	303.0568	162.96	0.44	
quercetin		[M+Na] ⁺	325.1876	173.17	1.16	
quercetin (peak 1)	+Glc	[M+H] ⁺	479.0892	204.10	0.20	1.084
quercetin (peak 2)	+Glc	[M+H] ⁺	479.0892	209.87	0.02	1.054
quercetin (peak 1)	+Glc	[M+Na] ⁺	501.0968	207.30	0.21	1.114
quercetin (peak 2)	+Glc	[M+Na] ⁺	501.0968	216.62	0.43	1.067
quinidine		[M+H] ⁺	325.2060	174.69	0.29	
quinidine		[M+Na] ⁺	347.1915	177.11	0.30	
quinidine	+O	[M+H] ⁺	341.1923	177.48	0.31	1.016
terfenadine		[M+H] ⁺	472.3376	227.04	0.13	
terfenadine		[M+Na] ⁺	494.3174	223.48	0.07	
terfenadine	+O	[M+H] ⁺	488.3268	222.82	0.12	1.042
thioridazine		[M+H] ⁺	371.1743	184.60	0.09	
thioridazine		[M+Na] ⁺	393.1823	188.74	0.13	
thioridazine	+O	[M+H] ⁺	387.1683	185.91	0.36	1.021
thioridazine	-Me	[M+H] ⁺	357.1578	184.20	0.23	0.977
thioridazine	+O, -Me	[M+H] ⁺	373.1677	184.83	0.16	1.002
thioridazine	+2O	[M+H] ⁺	403.1605	190.50	0.19	1.024
thioridazine	+2O, -Me	[M+H] ⁺	389.1668	186.11	0.35	1.024
triclosan		[M+H] ⁺	288.9481	153.08	0.30	
triclosan		[M+Na] ⁺	310.9798	163.04	0.28	
triclosan	+Glc	[M+H] ⁺	464.9557	179.03	0.28	1.174
triclosan	+O, +Glc	[M+Na] ⁺	502.9555	203.88	0.10	1.102

3.3 – Figure S2 – Expected Metabolites from Literature, Observed Metabolites, and Fragmentation

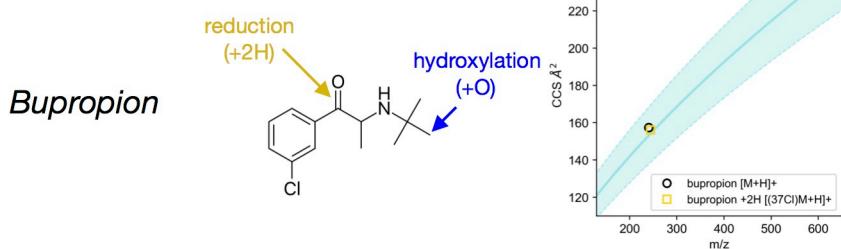
Data for Drugs

A



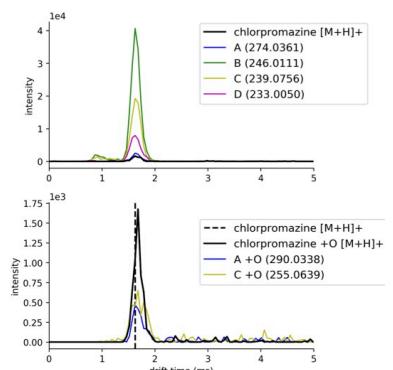
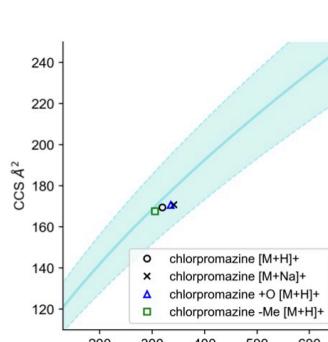
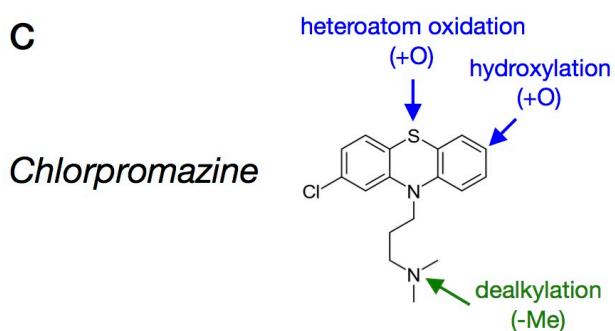
Expected metabolism for amlodipine⁵ (left) and observed metabolites (right).

B



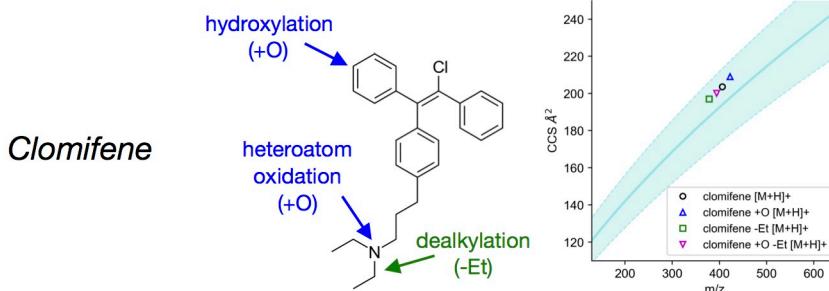
Expected metabolism for bupropion⁶ (left) and observed metabolites (right).

C



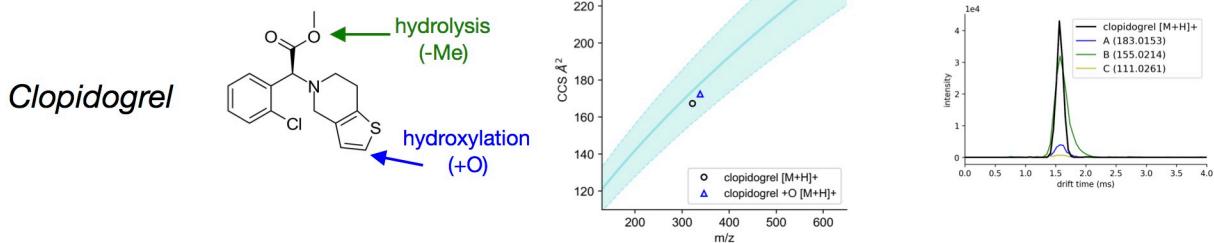
Expected metabolism for chlorpromazine⁷ (left), observed metabolites (center), and drift time aligned fragmentation data (right).

D



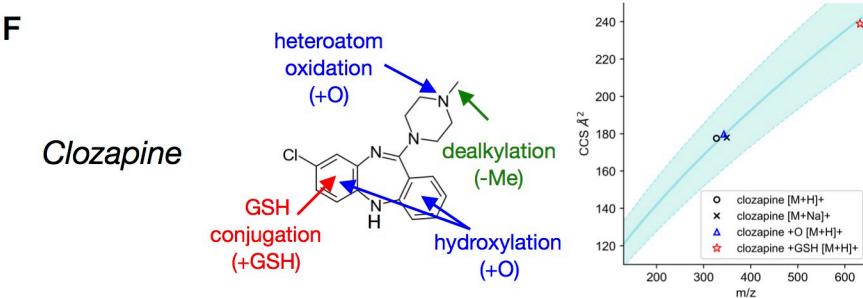
Expected metabolism for clomifene⁸ (left) and observed metabolites (right).

E



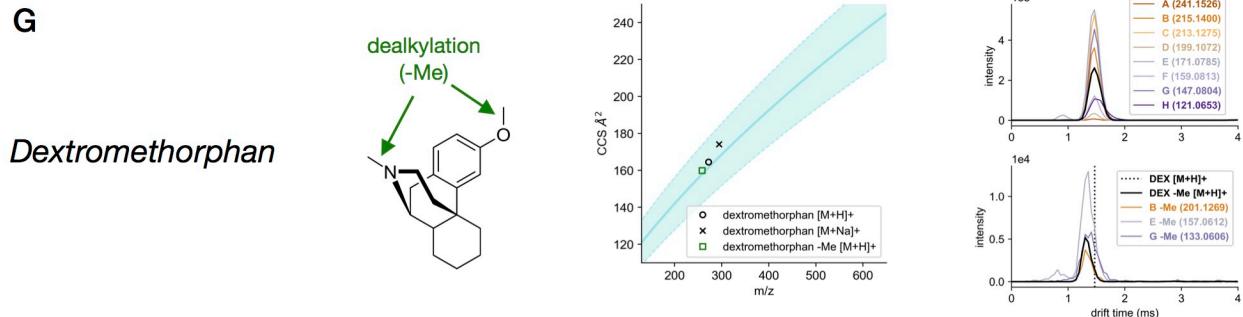
Expected metabolism for clopidogrel⁹ (left), observed metabolites (center), and drift time aligned fragmentation data (right).

F



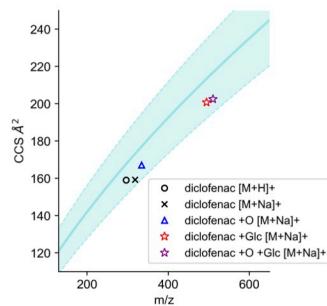
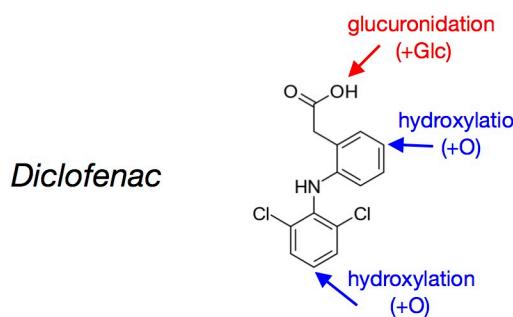
Expected metabolism for clozapine¹⁰ (left) and observed metabolites (right).

G



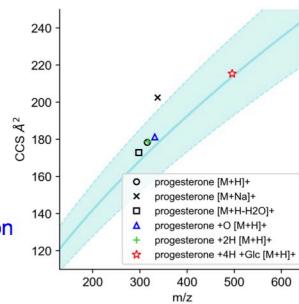
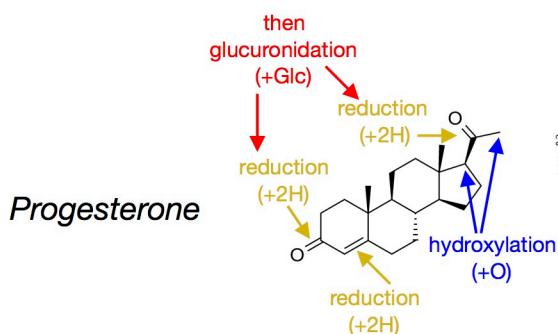
Expected metabolism for dextromethorphan¹¹ (left), observed metabolites (center), and drift time aligned fragmentation data (right).

H



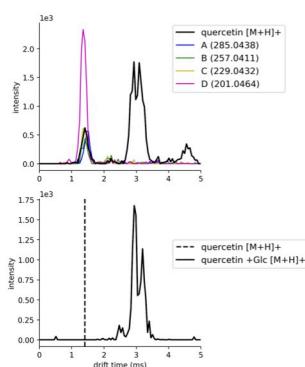
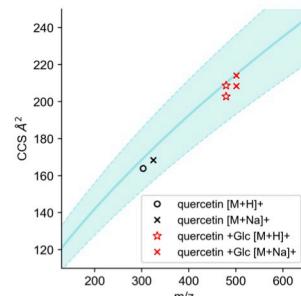
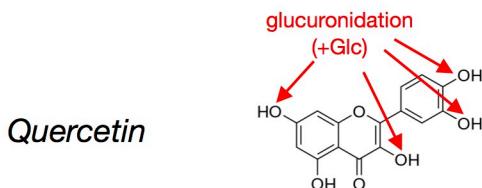
Expected metabolism for diclofenac^{12,13} (left) and observed metabolites (right).

I



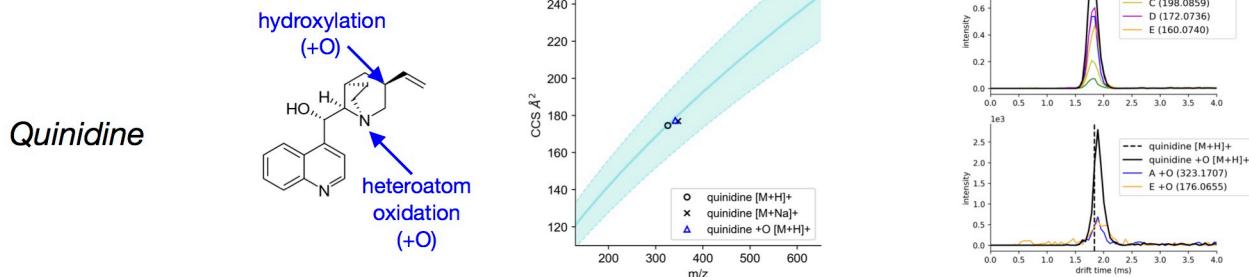
Expected metabolism for progesterone¹⁴ (left) and observed metabolites (right).

J



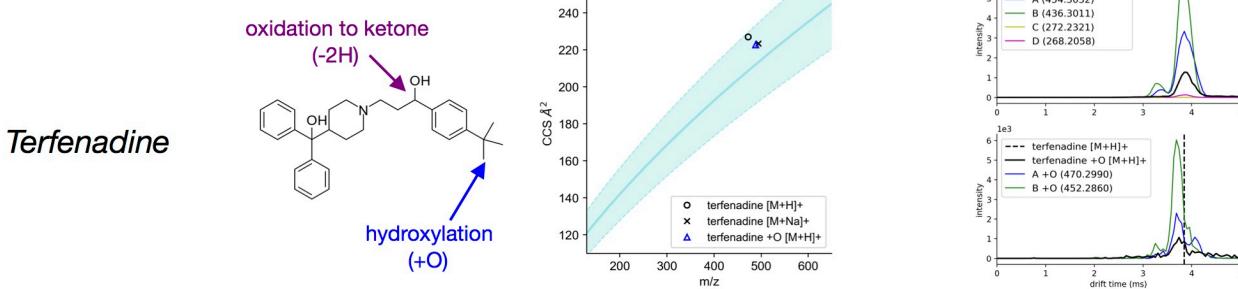
Expected metabolism for quercetin² (left), observed metabolites (center), and drift time aligned fragmentation data (right).

K



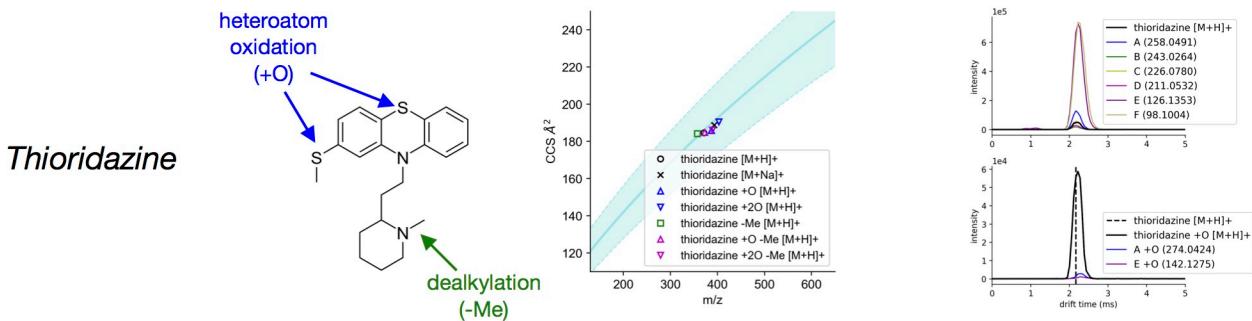
Expected metabolism for quinidine¹⁵ (left), observed metabolites (center), and drift time aligned fragmentation data (right).

L



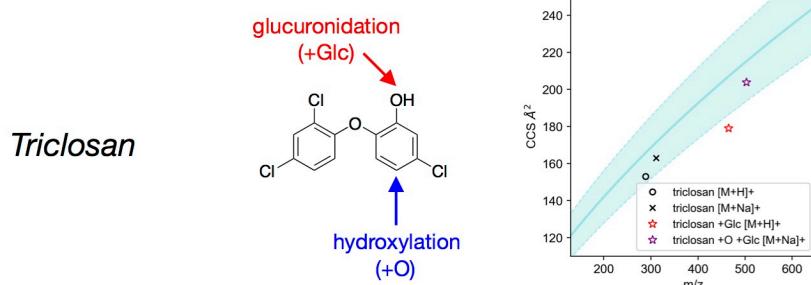
Expected metabolism for terfenadine¹⁶ (left), observed metabolites (center), and drift time aligned fragmentation data (right).

M



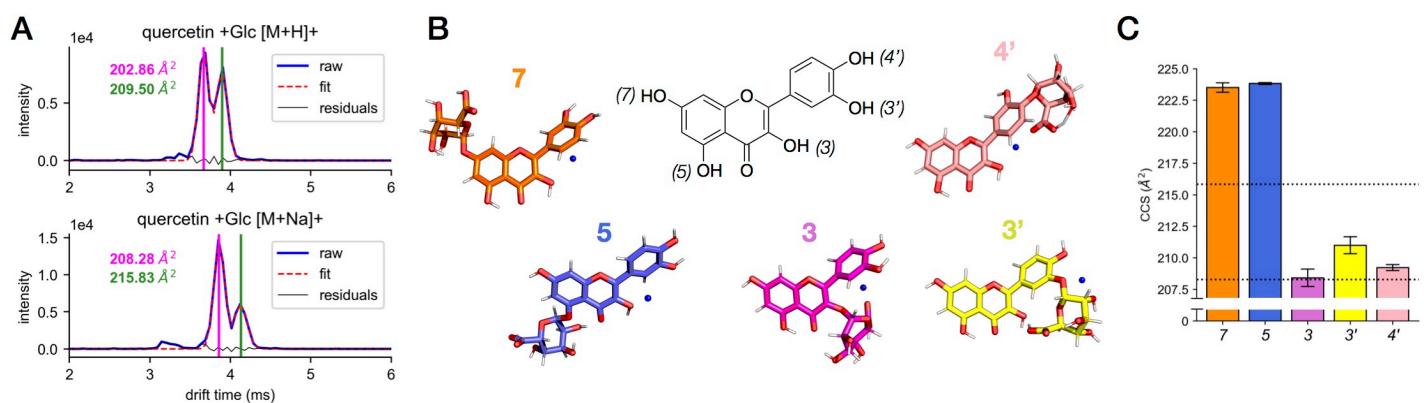
Expected metabolism for thioridazine¹⁷ (left), observed metabolites (center), and drift time aligned fragmentation data (right).

N



Expected metabolism for triclosan¹⁸ (left) and observed metabolites (right).

3.4 – Figure S3 – Theoretical CCS of Sodiated Quercetin Glucuronide Isomers



A Bimodal arrival time distributions (ATDs) for protonated (top) and sodiated (bottom) adducts of quercetin glucuronide. **B** Minimum energy structures for sodiated isomers of quercetin glucuronide. **C** Theoretical CCS of sodiated isomers of quercetin glucuronide, compared against experimental values (dotted lines).

4 – References

- (1) Kuca, K.; Marek, J.; Stodulka, P.; Musilek, K.; Hanusova, P.; Hrabinova, M.; Jun, D. *Molecules* **2007**, *12* (10), 2341–2347.
- (2) Boersma, M. G.; van der Woude, H.; Bogaards, J.; Boeren, S.; Vervoort, J.; Cnubben, N. H. P.; van Iersel, M. L. P. S.; van Bladeren, P. J.; Rietjens, I. M. C. M. *Chem. Res. Toxicol.* **2002**, *15* (5), 662–670.
- (3) Wandel, C.; Böcker, R.; Böhrer, H.; Browne, A.; Rügheimer, E.; Martin, E. *British Journal of Anaesthesia* **1994**, *73* (5), 658–661.
- (4) Klieber, S.; Hugla, S.; Ngo, R.; Arabeyre-Fabre, C.; Meunier, V.; Sadoun, F.; Fedeli, O.; Rival, M.; Bourrie, M.; Guillou, F.; Maurel, P.; Fabre, G. *Drug Metab. Dispos.* **2008**, *36* (5), 851–862.
- (5) Zhu, Y.; Wang, F.; Li, Q.; Zhu, M.; Du, A.; Tang, W.; Chen, W. *Drug Metab. Dispos.* **2013**, *42* (2), 245–249.
- (6) Jefferson, J. W.; Pradko, J. F.; Muir, K. T. *Clinical Therapeutics* **2005**, *27* (11), 1685–1695.
- (7) Hartmann, F.; Gruenke, L. D.; Craig, J. C.; Bissell, D. M. *Drug Metabolism and Disposition* **1983**, *11* (3), 244–248.
- (8) Mürdter, T. E.; Kerb, R.; Turpeinen, M.; Schroth, W.; Ganchev, B.; Böhmer, G. M.; Igel, S.; Schaeffeler, E.; Zanger, U.; Brauch, H.; Schwab, M. *Human Molecular Genetics* **2011**, *21* (5), 1145–1154.
- (9) Clarke, T. A.; Waskell, L. A. *Drug Metabolism and Disposition* **2003**, *31* (1), 53–59.
- (10) Pirmohamed, M.; Williams, D.; Madden, S.; Templeton, E.; Park, B. K. *J Pharmacol Exp Ther* **1995**, *272* (3), 984–990.
- (11) Kerry, N. L.; Somogyi, A. A.; clinical, F. B. B. J. O.; 1994. *Br. J. Clin. Pharmacol.* **1994**, *38* (3), 243–248.
- (12) Kuehl, G. E.; Lampe, J. W.; Potter, J. D.; disposition, J. B. D. M. A.; 2005. *ASPET*.
- (13) Bort, R.; Macé, K.; Boobis, A.; Gómez-Lechón, M.-J.; Pfeifer, A.; Castell, J. *Biochemical Pharmacology* **1999**, *58* (5), 787–796.
- (14) Aufrère, M. B.; Benson, H. *Journal of Pharmaceutical Sciences* **1976**, *65* (6), 783–800.
- (15) Nielsen, T. L.; Rasmussen, B. B.; Flinois, J.-P.; Beaune, P.; Brøsen, K. *J Pharmacol Exp Ther* **1999**, *289* (1), 31–37.
- (16) Jurima-Romet, M.; Crawford, K.; Cyr, T.; Inaba, T. *Drug Metabolism and Disposition* **1994**, *22* (6), 849–857.
- (17) Eap, C. B.; Guentert, T. W.; Loidl, M. S.; Stabl, M.; Koeb, L.; Powell, K.; Baumann, P. *Clinical Pharmacology & Therapeutics* **1996**, *59* (3), 322–331.
- (18) Wu, Y.; Chitranshi, P.; Loukotková, L.; da Costa, G. G.; Beland, F. A.; Zhang, J.; Fang, J.-L. *Archives of Toxicology* **2016**, *91* (6), 2405–2423.