

**Supplementary Table 1. Oligonucleotides primers/probes used in this study.**

qPCR primers/probe	Sequence (5'- 3')	Target	Annealing (°C)	Reference
Forward	GAAGGC GG CCTACTGGGCAC	<i>Faecalibacterium</i>	60	[1]
Reverse	GTGCAGGCGAGTTGCAGCCT			
Forward	KGGGCTCAACMCMGTATTGCGT	Fusobacteria	51	[2]
Reverse	TCGC GTTAGCTTGGCGCTG			
Forward	TCTGATGTGAAAGGCTGGGGCTTA	<i>Blautia</i>	56	[2]
Reverse	GGCTTAGCCACCCGACACCTA			
Forward	CCTACGGGAGGCAGCAGT	Universal Bacteria	59	[3]
Reverse	ATTACCGCGGCTGCTGG			
Forward	CAGACGGGGACAACGATTGGA	<i>Turicibacter</i>	63	[2]
Reverse	TACGCATCGTCGCCCTGGTA			
Forward	GTTAACCTTGCTCATTGA	<i>E. coli</i>	55	[4]
Reverse	ACCAGGGTATCTAACCTCTGT			
Forward	AGTAAGCTCTGATACTGTCT	<i>C. hiranonis</i>	50	[5]
Reverse	AGGGAAAGAGGGAGATTAGTCC			
Forward	TTATTGAAAGGGCAATTGCT	<i>Streptococcus</i>	54	[6]
Reverse	GTGAACTTCCACTCTCACAC			
Forward	AACTATAGGAGAACAAAATACAATAG	<i>C. perfringens</i> enterotoxin gene	55	[7]
Reverse	TGCATAAACCTTATAATACATATT			
Probe	FAM-TCTGTATCTACAAC TGCTGGTCCA-TAMURA			
Forward	CGCATAACGTTGAAAGATGG	<i>C. perfringens</i> 16S rRNA gene	58	[8]
Reverse	CCTTGGTAGGCCGTTACCC			
Probe	FAM-TCATCATTCAACCAAAGGAGCAATCC-TAMURA			
Forward	AACAATATGTACAGGTATAACT	<i>C. perfringens</i> <i>netF</i> toxin gene	55	[9]
Reverse	TTGATAGGTATAATATGGTTCT			
Forward	TTGAGCGATTACTCGGTAAAGA	<i>C. difficile</i>	61	[10]
Reverse	TGTACTGGCTCACCTTGATATTCA			
Forward	5'-GAGCTTCAAGCCCTATATCC-3'	<i>C. jejuni</i>	62.5	[11]
Reverse	5'-AAGAACACCGCGAAGTTATTT-3'			

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