

Supplemental Information

Astrocytic *trans*-Differentiation Completes

a Multicellular Paracrine Feedback Loop

Required for Medulloblastoma Tumor Growth

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Supplemental Table 1. Related to STAR Methods section “EXPERIMENTAL MODEL AND SUBJECT DETAILS, Mice”, and to the first section in “RESULTS, Establishment of MADM-based and other genetic models for medulloblastoma”. Medulloblastoma models used in this paper.

Model#	Genotype	Genetic components and their roles	Uniqueness of the model and its usage in this paper
A	Ptch1 KO/wildtype; MADM-p53KO; Math1-Cre	Ptch1 KO: Driver mutation for Shh-subtype medulloblastoma MADM-p53KO: Increase the tumor penetrance to 100% upon Cre-mediated knockout Math1-Cre: Induce inter-chromosomal recombination to generate sporadic p53-null GFP+ GNPs	This model enables single-cell resolution analysis of the entire course of tumorigenesis, and was used in Figures 1E-R, 2B, and Supplemental Figures 1F, 2A.
B	Ptch1 KO/wildtype; p53flox/p53flox; Math1-Cre; ROSA26-LSL-tdTomato	Ptch1 KO: Driver mutation for Shh-subtype medulloblastoma p53flox/p53flox: Increase the tumor penetrance to 100% upon Cre-mediated knockout Math1-Cre: Induce loxP-mediated recombination to knockout p53 and turn on Cre reporter tdTomato in all GNPs ROSA26-LSL-tdTomato: Cre reporter used for lineage tracing of all cells derived from Math1-Cre+ cells	This model enables lineage tracing from tumor GNPs even after they differentiate into other cells types, and was used in Figure 7J-K, Supplemental Figures 1G, 2E-H, 3E-G, 7G-H.
C	Ptch1 KO/wildtype; p53flox/p53flox; Math1-Cre; Math1-GFP	Ptch1 KO: Driver mutation for Shh-subtype medulloblastoma p53flox/p53flox: Increase the tumor penetrance to 100% upon Cre-mediated knockout Math1-Cre: Induce loxP-mediated recombination to knockout p53 in all GNPs Math1-GFP: Transgenic reporter for wildtype and tumor GNPs, which turns off after they differentiate	This model enables one to distinguish GFP+ tumor from GFP- TME cells, and was used in Figures 4F-G, 6J-K, 7C-I, and Supplemental Figures 3A-D, 4A-C, 7C-F.
D	Ptch1 KO/wildtype; p53KO/p53KO; Math1-GFP	Ptch1 KO: Driver mutation for Shh-subtype medulloblastoma p53KO/p53KO: Increase the tumor penetrance to 100% in the absence of Cre Math1-GFP: Transgenic reporter for wildtype and tumor GNPs, which turns off after they differentiate	This model frees up Cre for other purposes, such as to label TuAstrocytes with GLAST-CreER and Cre reporter in Supplemental Figure 2B-D and Figure 3, to knock out IGF1R after tumor formation with Math1-CreER in Figure 4H-J and Supplemental Figure 4E-I, to knock out IGF1 in TAMs with CSF1R-Cre in Figure 5H-J, and to trace the lineage of TAMs with CX3CR1-CreER in Figure 6A-E and Supplemental Figure 6.

Supplemental Table 2. Related to Figure 7. Luminex profiling of cytokines and chemokines.

Tumor tissues were dissected out based on Math1-GFP expression and lysed in Tissue Extraction Reagent I (Invitrogen) supplemented with protease inhibitors. Normal cerebelli from age-matched mice were processed in the same fashion as control samples. Protein concentration of the lysates was determined by BCA protein quantification method. All lysates were carefully adjusted to 1.5mg/ml with lysis buffer, and 25 μ l each was applied to mouse 32-plex cytokine/chemokine luminex panel. The absolute quantity of cytokine/chemokine present in cell lysate was expressed as picogram per 10ug of tissue lysate. Four tumors and three normal cerebella were used for this study.

Gene name	Normal cerebellum (Means \pm SD, pg/10 μ g)	Tumor mass (Means \pm SD, pg/10 μ g)	p value
G-CSF	4.47 \pm 1.10	6.49 \pm 2.46	
GM-CSF	14.98 \pm 1.80	20.02 \pm 3.57	
M-CSF	4.32 \pm 0.96	5.21 \pm 1.84	
IFN γ	3.13 \pm 0.16	3.20 \pm 2.0	
IL1a	31.04 \pm 7.45	20.53 \pm 9.28	
IL1b	92.74 \pm 13.23	77.68 \pm 58.19	
IL2	90.09\pm4.0	13.81\pm8.31	p<0.001
IL3	Below detection	Below detection	
IL4	5.97\pm1.37	9.66\pm1.49	p<0.05
IL5	1.98 \pm 0.12	1.83 \pm 0.61	
IL6	8.76 \pm 2.97	6.44 \pm 0.54	
IL7	2.02 \pm 0.86	1.68 \pm 0.29	
IL9	868.44\pm68.9	546.4\pm68.19	p<0.01
IL10	13.47 \pm 0.69	30.08 \pm 20.36	
IL12	4.06 \pm 0.32	4.06 \pm 0.50	
IL13	635.6 \pm 49.22	857.7 \pm 571.1	
IL15	60.50\pm12.03	33.50\pm7.74	p<0.05
IL17A	4.92\pm0.61	1.01\pm0.41	p<0.01
LIF	Below detection	1.33 \pm 0.52	
LIX	61.3 \pm 5.77	49.74 \pm 44.2	
CXCL10	17.01\pm2.21	102.87\pm25.70	p<0.01
KC	15.69 \pm 2.57	106.60 \pm 128.97	
MCP1	28.94 \pm 4.97	39.15 \pm 22.62	
MIP-1a	26.79\pm2.85	81.56\pm33.99	p<0.05
MIP-1b	62.24\pm15.45	24.08\pm4.31	p<0.05
MIP-2	17.21 \pm 0.55	19.18 \pm 3.30	
MIG	12.40\pm2.94	29.11\pm5.66	p<0.05
RANTES	2.45 \pm 0.12	3.72 \pm 0.96	
Eotaxin	8.98\pm1.85	88.17\pm38.09	p<0.05
TNF α	2.25 \pm 0.25	2.01 \pm 0.88	

Supplemental Table 3. Related to “Key Resources Table, Oligonucleotides section”.
Oligonucleotides used in this paper. This is a list of oligonucleotides that were used for mouse genotyping, qRT-PCR, and probes for in situ hybridization.

Mouse genotyping	Source	Reference
TG11ML, GT11ML mouse genotyping: Chr11_CS1: 5'-TGGAGGAGGACAAACTGGTCAC-3' Rosa4: 5'-TCAATGGGCGGGGGTCGTT-3' Chr11_CS2: 5'- TTCCCTTTCTGCTTCATCTTGC-3'	This paper	N/A
IGF1R flox mouse genotyping: 10451: 5'-CTTCCCAGCTTGCTACTCTAGG-3' 10452: 5'-CAGGCTTGCAATGAGACATGGG-3' IGF1R-rec-F: 5'-TGAGACGTAGCGAGATTGCTGTA-3'	This paper	N/A
IGF1 flox mouse genotyping: 10663 5'-CACTAAGGAGTCTGTATTTGGACC-3' 10664 5'-AAACCACACTGCTCGACATTG-3' 11736 5'-GGCAAATGGAAATCCTATGTCT-3'	This paper	N/A
CSF1R-iCre mouse genotyping: Cre-F: 5'-CACCCCTGTTACGTATAGCCG-3' Cre-R: 5'-GAGTCATCCTTAGCGCCGTA-3'	This paper	N/A
CX3CR1-CreER ¹² mouse genotyping: Cre-F: 5'-CACCCCTGTTACGTATAGCCG-3' Cre-R: 5'-GAGTCATCCTTAGCGCCGTA-3'	This paper	N/A
Ptch1-KO mouse genotyping: Ptc-Neo-F3: 5'-TGGGGTGGGATTAGATAAATGCC-3' Ptc-R3: 5'-TGTCTGTGTGTGCTCCTGAATCAC-3' Ptc-WT3: 5'-CTGCGGCAAGTTTTTGGTTG-3' Ptc-WT4: 5'-AGGGCTTCTCGTTGGCTACAAG-3'	This paper	N/A
p53-KO mouse genotyping: neo tail: 5'-ACCGCTATCAGGACATAGCGTTGG-3' p53 TJW5: 5'-CACAGCGTGGTGGTACCTTATG-3' p53 TJW3: 5'- GGTATACTCAGAGCCGGCCTG-3'	This paper	N/A
p53-flox mouse genotyping: T008: 5'-CACAAAAACAGGTTAAACCCAG-3' T009: 5'-AGCACATAGGAGGCAGAGAC-3'	This paper	N/A
Math1-Cre mouse genotyping: Cre-F: 5'-CACCCCTGTTACGTATAGCCG-3' Cre-R: 5'-GAGTCATCCTTAGCGCCGTA-3'	This paper	N/A
Math1-CreER ¹² mouse genotyping: Cre-F: 5'-CACCCCTGTTACGTATAGCCG-3' Cre-R: 5'-GAGTCATCCTTAGCGCCGTA-3'	This paper	N/A
Math1-GFP mouse genotyping: FACS G5': 5'-CTTCAAGTCCGCCATGCCCCGA-3' GFP2-Hermie: 5'-TCCAGCAGGACCATGTGATCGC-3' IMR0015 (IC): 5'-CAAATGTTGCTTGTCTGGTG-3' IMR0016 (IC): 5'-GTCAGTCGAGTGCACAGTTT-3'	This paper	N/A
IL4-GFP reporter mouse genotyping: FACS G5': 5'-CTTCAAGTCCGCCATGCCCCGA-3' GFP2-Hermie: 5'-TCCAGCAGGACCATGTGATCGC-3' IMR0015 (IC): 5'-CAAATGTTGCTTGTCTGGTG-3' IMR0016 (IC): 5'-GTCAGTCGAGTGCACAGTTT-3'	This paper	N/A

IL4-KO mouse genotyping: oIMR0077 5'-GCACAGAGCTATTGATGG -3' GTCIMR0078 5'-GCTGTGAGGACGTTTGGC-3' oIMR0079 5'-TCAGGACATAGCGTTGGC-3'	This paper	N/A
Rosa26-LSL-tdTomato mouse genotyping: oIMR9020 5'-AAGGGAGCTGCAGTGGAGTA-3' oIMR9021 5'-CCGAAAATCTGTGGGAAGTC-3' oIMR9103 5'-GGCATTAAAGCAGCGTATCC-3' oIMR9105 5'-CTGTTCCTGTACGGCATGG-3'	This paper	N/A
Aldh1L1-GFP reporter mouse genotyping: FACS G5': 5'-CTTCAAGTCCGCCATGCCCGA-3' GFP2-Hermie: 5'-TCCAGCAGGACCATGTGATCGC-3' IMR0015 (IC): 5'-CAAATGTTGCTTGTCTGGTG-3' IMR0016 (IC): 5'-GTCAGTCGAGTGCACAGTTT-3'	This paper	N/A
qRT-PCR		
Mouse IGF1 qPCR Forward: 5'-GGCATTGTGGATGAGTGTTG-3'	This paper	N/A
Mouse IGF1 qPCR Reverse: 5'-TCTCCTTTGCAGCTTCGTTT-3'	This paper	N/A
Mouse CSF1R qPCR Forward: 5'-TCCACCGGGACGTAGCA-3'	This paper	N/A
Mouse CSF1R qPCR Reverse: 5'-CCAGTCCAAAGTCCCCAATCT-3'	This paper	N/A
mouse GAPDH qPCR Forward: 5'-CAGGTTGTCTCCTGCGACTT-3'	This paper	N/A
mouse GAPDH qPCR Reverse: 5'-ATGTAGGCCATGAGGTCCAC-3'	This paper	N/A
mouse PDGFA qPCR Forward: 5'-GAGATACCCCGGGAGTTGAT-3'	This paper	N/A
mouse PDGFA qPCR Reverse: 5'-TCTTGCAAACCTGCAGGAATG-3'	This paper	N/A
mouse PDGFB qPCR Forward: 5'-GATCTCTCGGAACCTCATCG-3'	This paper	N/A
mouse PDGFB qPCR Reverse: 5'-GGCTTCTTTCGCACAATCTC-3'	This paper	N/A
mouse HGF qPCR Forward: 5'-TTCCCAGCTGGTCTATGGTC-3'	This paper	N/A
mouse HGF qPCR Reverse: 5'-TGGTGCTGACTGCATTTCTC-3'	This paper	N/A
Mouse FGF2 qPCR Forward: 5'-AGCGGCTCTACTGCAAGAAC-3'	This paper	N/A
Mouse FGF2 qPCR Reverse: 5'-GCCGTCCATCTTCCTTCATA-3'	This paper	N/A
mouse EGF qPCR Forward: 5'-CCCAGGCAACGTATCAAAGT-3'	This paper	N/A
mouse EGF qPCR Reverse: 5'-GGTCATACCCAGGAAAGCAA-3'	This paper	N/A
mouse NT3 qPCR Forward: 5'-GATCCAGGCGGATATCTTGA-3'	This paper	N/A
mouse NT3 qPCR Reverse: 5'-AGCGTCTCTGTTGCCGTAGT-3'	This paper	N/A

mouse KitL qPCR Forward: 5'-TGGTGGCAAATCTTCCAAAT-3'	This paper	N/A
mouse KitL qPCR Reverse: 5'-TTCTTCGGTGCCTTTTCTTC-3'	This paper	N/A
mouse Math1 qPCR Forward: 5'-GCCAATGCAGAGCAACTTTT-3'	This paper	N/A
mouse Math1 qPCR Reverse: 5'-AATGGGTCCAAATACGCAAT-3'	This paper	N/A
mouse Aldh1L1 qPCR Forward: 5'-GCCTTCCAACCTTCTGTTGC-3'	This paper	N/A
mouse Aldh1L1 qPCR Reverse: 5'-CGCCACCGAGGGAACTTAAA-3'	This paper	N/A
Probes for in situ hybridization		
mouse IGF1 in situ Forward: 5'-ACACAAGTAGAGGAAGTGCAGGA-3'	Allen brain	http://mouse.brain-map.org
mouse IGF1 in situ Reverse: 5'-GCAGCCAAAATTCAGAGAGAA-3'	Allen brain	http://mouse.brain-map.org
mouse IL4 in situ Forward: 5'-TCACTGACGGCACAGAGC-3'	Allen brain	http://mouse.brain-map.org
mouse IL4 in situ Reverse: 5'-TGCTCTTTAGGCTTTCCAGG-3'	Allen brain	http://mouse.brain-map.org
Arg1-TaqMan Probe	Mm00475988_m1	N/A
Chil3-TaqMan Probe	Mm00657889_mH	N/A
Retnla-TaqMan Probe	Mm00445109_m1	N/A
IGF1-TaqMan Probe	Mm00439560_m1	N/A
GAPDH-TaqMan Probe	Mm99999915_g1	N/A