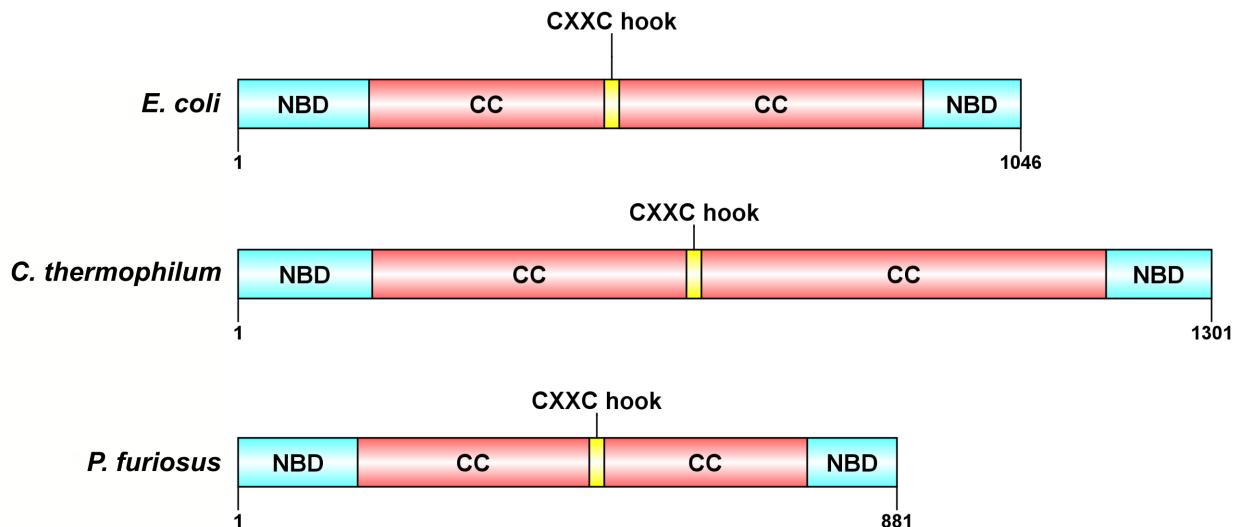


Supplementary Information

Table S1. Mutations cited in the text.

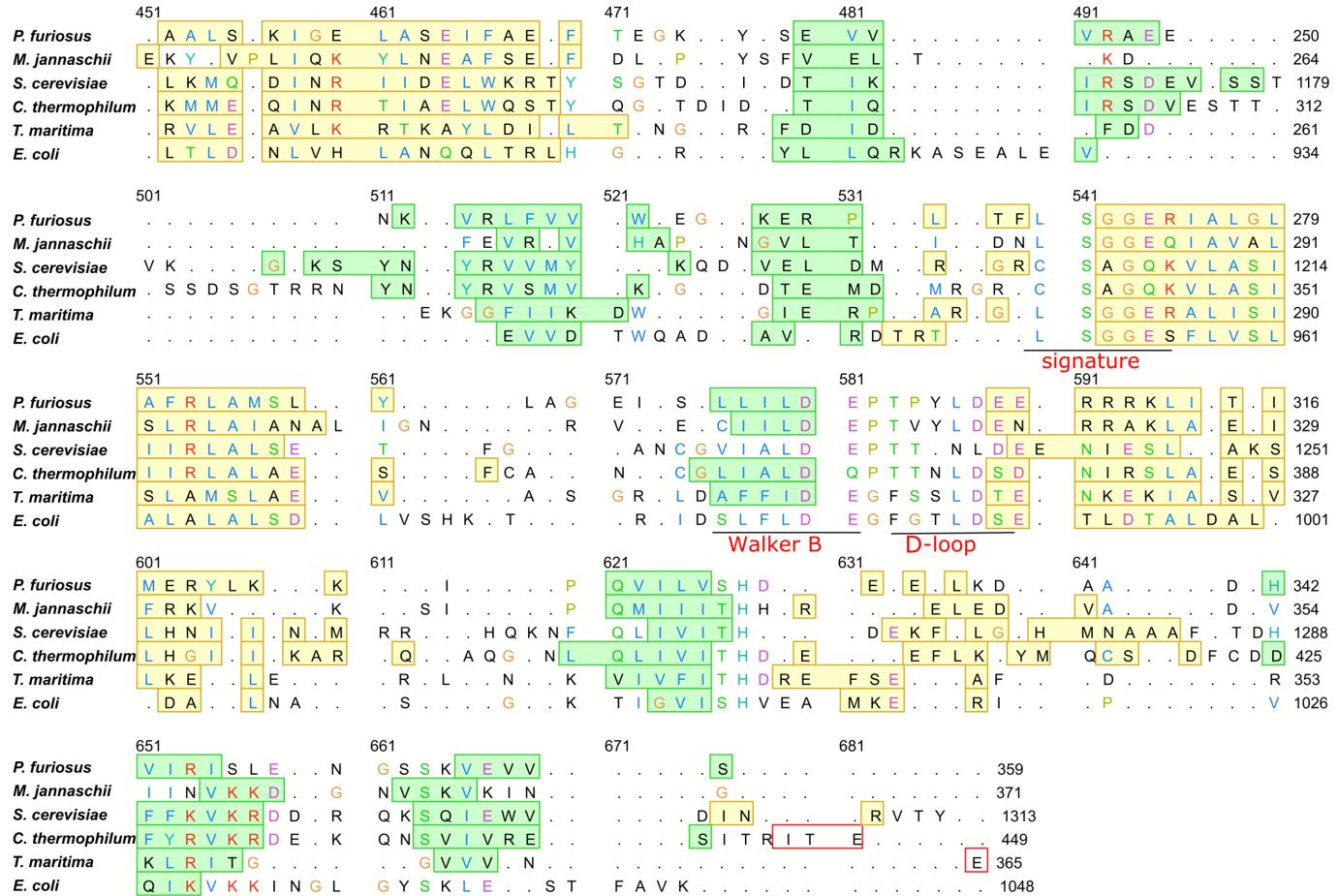
Allele	Mutation	Protein	Organism	Phenotype	Ref.
<i>rad50-48</i>	S679R, P682R	Rad50	<i>H. sapiens</i>	defect in Rad50 dimerization	[1]
	R805E	Rad50	<i>P. furiosus</i>	slow ATP hydrolysis	[2]
	L828F	Rad50	<i>P. furiosus</i>	defect in ATM activation	[3]
	D829N	Rad50	<i>P. furiosus</i>	defect in ATM activation	[3]
	A78T	Rad50	<i>S. cerevisiae</i>	defect in Tel1 activation	[4]
	E115K	Rad50	<i>E. coli</i>	reduced endo/exonuclease and altered DNA binding activity	[5]
<i>rad50S</i>	K6E	Rad50	<i>S. cerevisiae</i>	defective in removing Spo11 from DSB end	[6]
<i>rad50S</i>	R20M	Rad50	<i>S. cerevisiae</i>	defective in removing Spo11 from DSB end	[6]
<i>rad50S</i>	K81I	Rad50	<i>S. cerevisiae</i>	defective in removing Spo11 from DSB end	[6]
	Y328A	Mre11	<i>S. cerevisiae</i>	DNA damage sensitivity	[7]
	N113S	Mre11	<i>S. pombe</i>	defective Mre11 interaction with Nbs1	[8]
	S499P	Mre11	<i>S. cerevisiae</i>	defective Mre11-Rad50 interaction	[4]
	K149E	Mre11	<i>E. coli</i>	reduced endo/exonuclease and altered DNA binding activity	[5]
<i>mre11S</i>	T188I	Mre11	<i>S. cerevisiae</i>	defective Spo11 removal from DSB end	[9]
	R184A	Mre11	<i>S. cerevisiae</i>	DNA damage sensitive in <i>sgs1Δ</i> background	[10]
	H125N	Mre11	<i>S. cerevisiae</i>	nuclease deficient	[11]
	H52S	Mre11	<i>P. furiosus</i>	loss of exonuclease activity	[12]
	Y187C	Mre11	<i>P. furiosus</i>	loss of exonuclease activity	[13]
	R10T	Mre11	<i>S. cerevisiae</i>	<i>sae2Δ</i> DNA damage and resection defect rescue	[14]

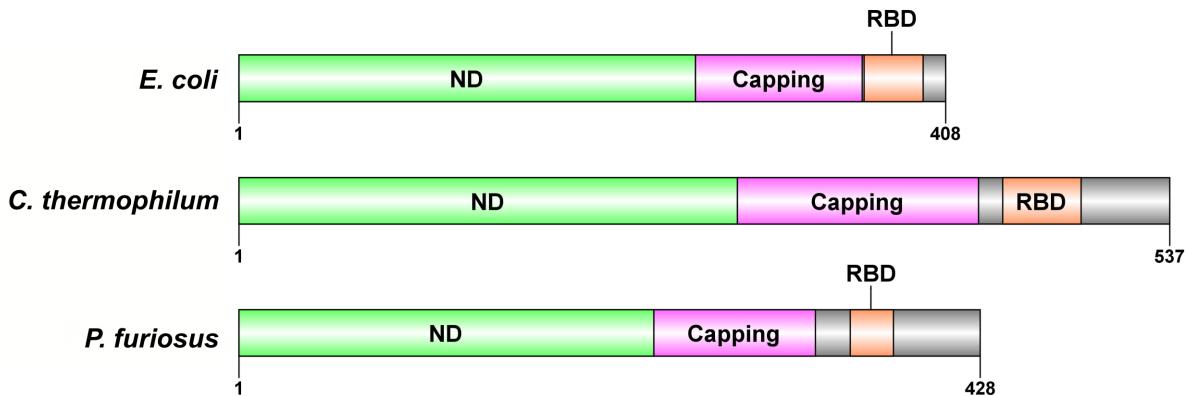
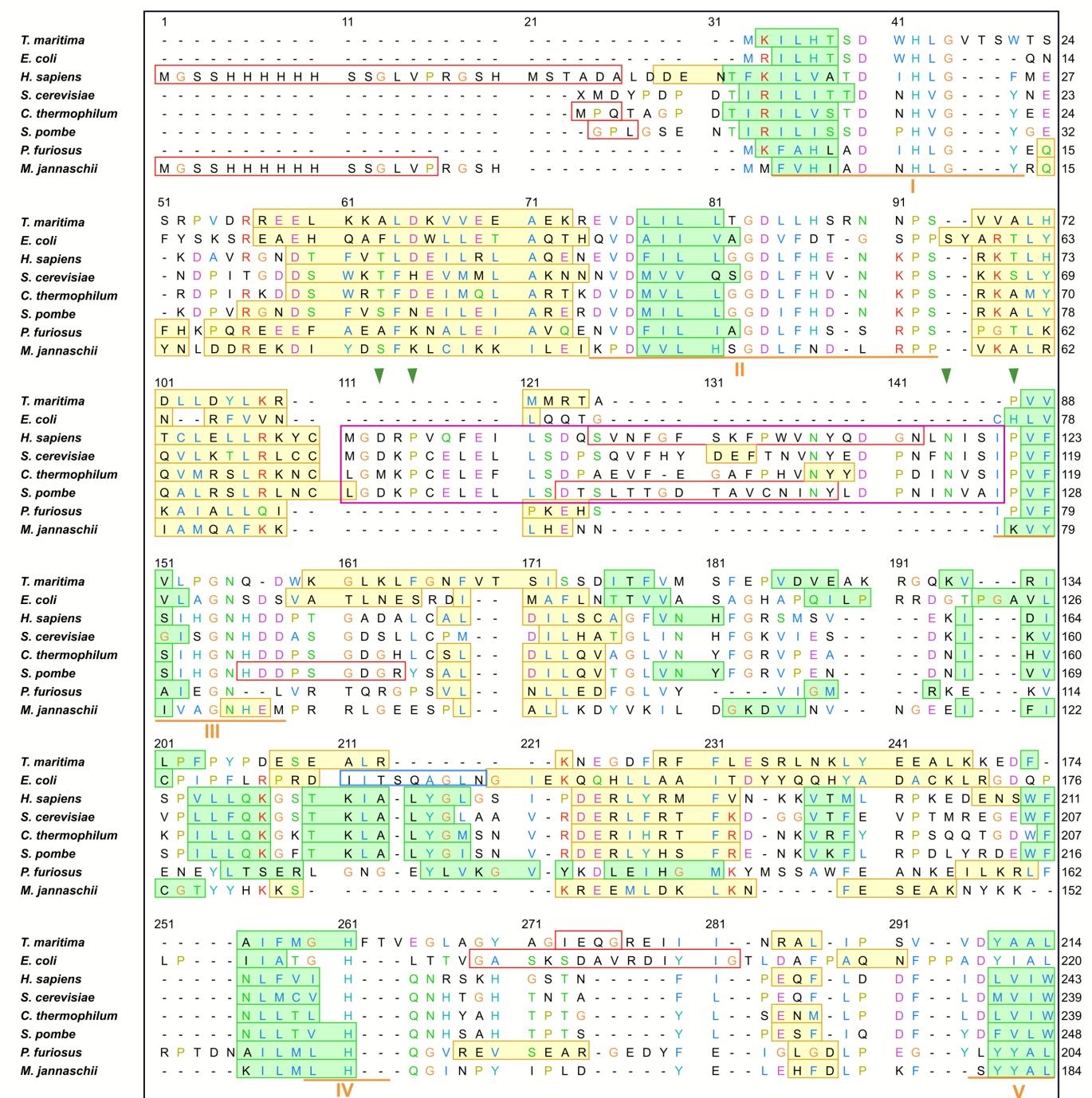
A**B**

	1	11	21	31	41	
<i>P. furiosus</i>	- - M K L E R V T V	- - - K N F R S H S	D T - V V E F K E -	- - - - G I N L I I G Q N G S G K S S	38	
<i>M. jannaschii</i>	M S M I L K E I R M	- - - N N F K S H V	G N S - R I K F E K -	- - - - G I V A I I G E N G S G K S S	40	
<i>S. cerevisiae</i>	- M S A I Y K L S I	Q G I R S F D S N D	R E - T I E F G K -	- - - - P L T L I V Q M N G S G K T T	42	
<i>C. thermophilum</i>	- M S K I E K L S I	L G V R S F G P H H	P E - T I A F N T -	- - - - P L T L I V G Y N G S G K T T	42	
<i>T. maritima</i>	- - M R P E R L T V	- - - R N F L G L K	N V - D I E F Q S -	- - - - G I T V V E G P N G A G K S S	38	
<i>E. coli</i>	- - M K I L S L R L	- - - K N L N S L K	G E W K I D F T R E	P F A S N G L F A I T	45	G P T G A G K T T
	51	61	71	81	91	Walker A (P-loop)
<i>P. furiosus</i>	L L D A I L V G L Y	W P L R I K D - - -	- - - I K K D E F T K	V G A R D T - Y I D L I F - E K D G T K	81	
<i>M. jannaschii</i>	I F E A V F F A L F	G A G S N F N - - -	- - - Y D T I I T	K G K K S V - Y V E L D F I - E V N G N N	81	
<i>S. cerevisiae</i>	I I E C L K Y A T T	G D L P P N S - K G	G V F I H D P K I T	G E K D I R A Q V K L A F T S A N G L N	91	
<i>C. thermophilum</i>	V I E C L K Y A T T	G E L P P N S T R N	G A F I H D P D L V	G E K E V R A Q V K L S F R S T I G E S	92	
<i>T. maritima</i>	L F E A I S F A L F	G N G I R Y P - - -	- - - N S Y D Y V N R	N A V D G T A R L V F Q F - E R G G K R	82	
<i>E. coli</i>	L L D A I C L A L Y	H E T P R L S - - N	V S Q S Q N D L M T	R D T A E C - L A E V E F - E V K G E A	91	
	101	111	121	131	141	
<i>P. furiosus</i>	Y R I T R R F - - L	K G Y S S S G E I H A	- - - - M K R L V G N E W K H V T E	P S S - - K A I S A	121	
<i>M. jannaschii</i>	Y K I I R E Y D - -	S G R G G A K L Y K	- - - - S G D R S T L S T R	I S A V N K A V N E	117	
<i>S. cerevisiae</i>	M I V T R N I Q L L	M K K T T T T F K T	L E G Q L V A I N N	S L E L D A Q V P L	141	
<i>C. thermophilum</i>	Y V V T R N I Q L L	V Q R N N K R T Q K	T L E G S L L L R N	V A E L D K L V S E	142	
<i>T. maritima</i>	Y E I I R E I N A L	Q R K H N A K L S E	- - - - I L E	P T S V K Q E V E K	125	
<i>E. coli</i>	Y R A F W S Q N R A	R N Q P D G N L Q V	- - - P R V E L A R	C A D G K I L A D K	138	
	151	161	171	181	191	
<i>P. furiosus</i>	F M E K L I P Y N I	F L N A I Y I R Q G	Q I D A I L E - S D	E A R E K V V R E V	L N L D K - F E T A	169
<i>M. jannaschii</i>	I L G - - V D R N M	F L N S I Y I K Q G	E I A K F L S L K P	S E K L E T V A K L	L G I D E - F E K C	164
<i>S. cerevisiae</i>	Y L G - - V P K A I	L E Y V I F C H Q E	D - S L W P L S E P	S N L K K K F D E I	F Q A M K - F T K A	187
<i>C. thermophilum</i>	K L G - - V P P A I	L D A V I F C H Q D	D - S L W P M S E P	A A L K K R F D E I	F E A Q K - Y T K V	188
<i>T. maritima</i>	I L G - - I E H R T	F I R T V F L P Q G	E I D K L L I S P P	S E I T E I I S D V	F Q S K E T L E K L	173
<i>E. coli</i>	L T G - - L D Y G R	F T R S M L L S Q G	Q F A A F L N A K P	K E R A E L L E E L	T G T E I - Y G Q I	185
	201	211	221	231	241	
<i>P. furiosus</i>	Y K K L S E L K K T	I N N R - - - I K E	Y G G S - - - -	- - - - G G I K	D L E K A K D F T E	204
<i>M. jannaschii</i>	Y Q K M G E I V K E	Y E K R - - - L E R	I E G E L N Y K E E	S L K A R L K E M S	N L E K E K E K L T	211
<i>S. cerevisiae</i>	L D N L K S -	-	-	-	-	194
<i>C. thermophilum</i>	J E N I R L L K K K	K G D E L K I L K E	R E V Q D K A N K E	R A E K V D G G A G	G A G G E L D L K D	238
<i>T. maritima</i>	E K L L K E K M K K	L E N E - - - I S S	G G A G - - - -	- - - - G A G G	S L E K K L K E M S	208
<i>E. coli</i>	S A M V F E Q H K -	- - - -	- - - -	- - - -	- - - -	194

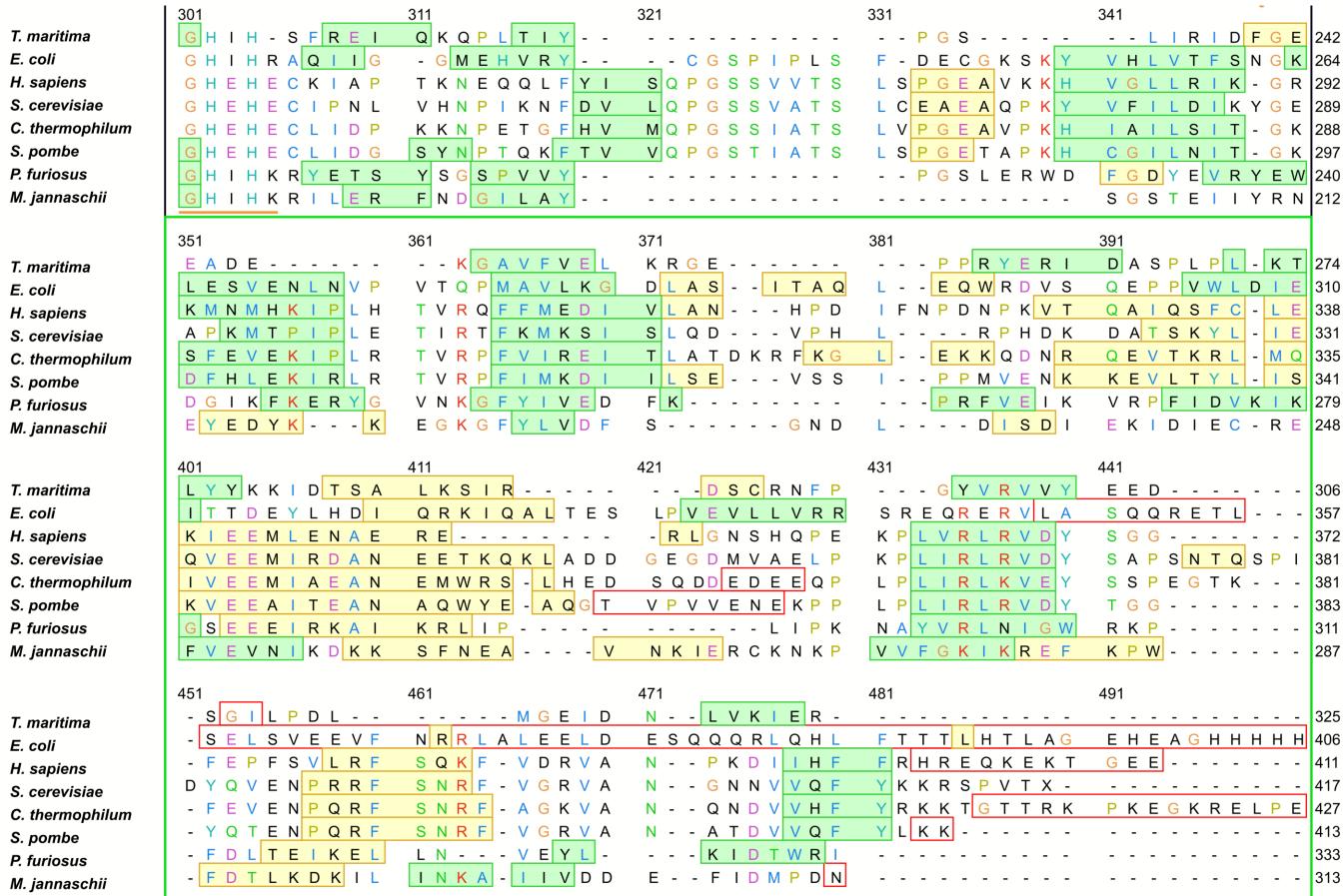
Supplementary Figure S1

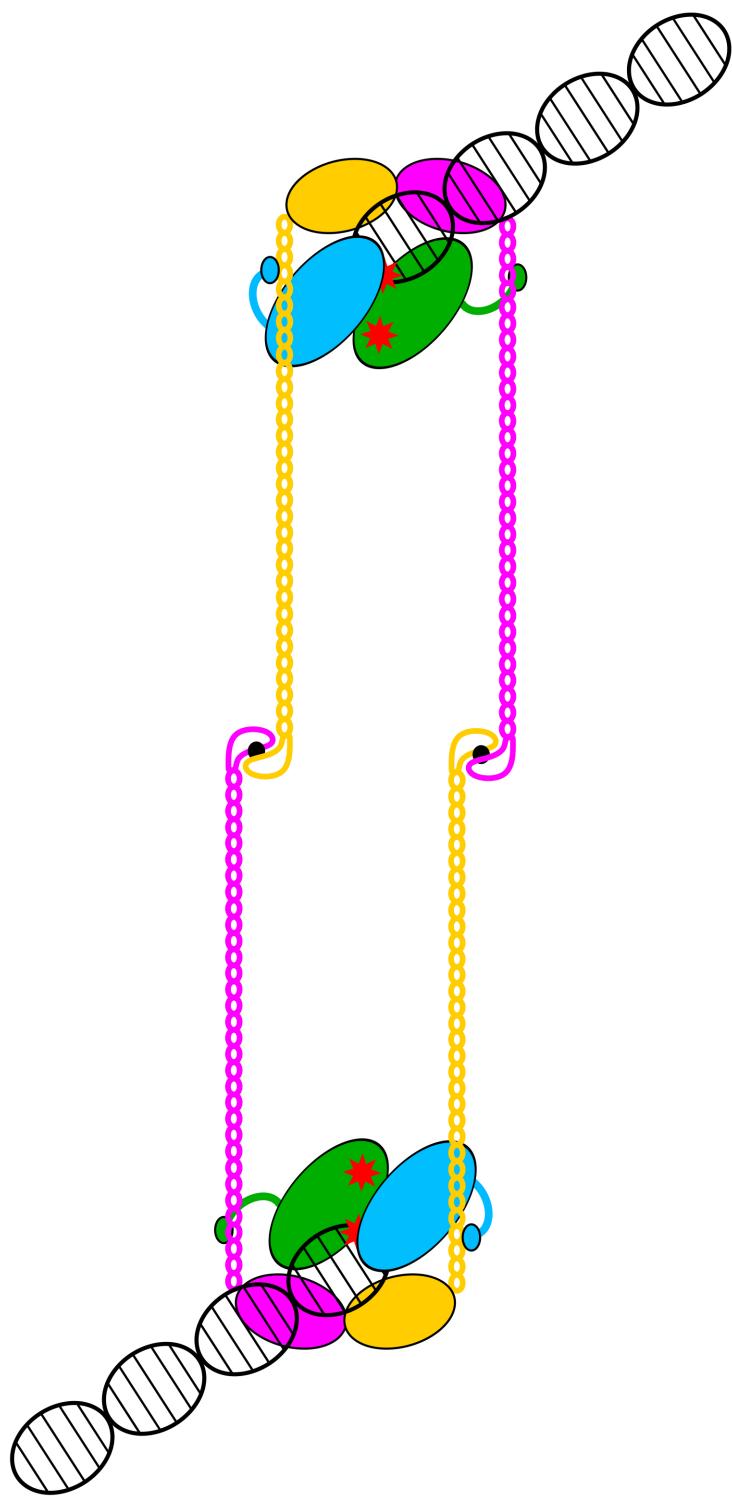
C



A**B**

B - continued





Supplementary figure legends

Supplementary Figure S1. Structural features and structural alignment of Rad50 orthologs. (A) Scheme depicting the conserved domains present in Rad50 orthologues from the indicated organisms. (B,C) Structural alignment of the lobe I (B) and lobe II (C) of Rad50 orthologues from eubacteria *T. maritima* (PDB ID: 4W9M) and *E. coli* (PDB ID: 6S6V), eukaryotes *S. cerevisiae* (model from reference [4]) and *C. thermophilum* (PDB ID:5DAC), and archaea *P. furiosus* (PDB ID:3QKU) and *M. jannaschii* (PDB ID:3AV0). Secondary structures are indicated, wherever the structure is available (otherwise the sequence is marked by a red rectangle), as follows: solid yellow rectangles, alpha helix; solid green rectangles, beta sheets. Alignment residues coloring is according to ClustalW style. Conserved functionally relevant motifs are underlined and labelled.

Supplementary Figure S2. Structural features and structural alignment of Mre11 orthologs. A. Scheme depicting the conserved domains present in Mre11 orthologs from the indicated organisms. (B) Structural alignment of the N-terminal regions of Mre11 orthologues from eubacteria *T. maritima* (PDB ID: 4NZV) and *E. coli* (PDB ID: 6S6V), eukaryotes *S. cerevisiae* (model from reference [15]), *H. sapiens* (PDB ID:3T1I), *C. thermophilum* (PDB ID:4KYE), and *S. pombe* (PDB ID:4FCX), and archaea *P. furiosus* (PDB ID:1II7) and *M. jannaschii* (PDB ID:3AV0). Secondary structures are indicated, wherever the structure is available (otherwise the sequence is marked by a red rectangle), as follows: solid yellow rectangles, alpha helix; solid green rectangles, beta sheets. Alignment residues coloring is according to ClustalW style. Functional regions of the proteins are indicated by open boxes: black box, endonuclease domain; green box, capping domain; purple box, latching loop; blue box, fastener loop. Orange bars highlight conserved phosphoesterase signature motifs (I to V). Green triangles indicate the residues identified for Nbs1 binding in *S. pombe* [8].

Supplementary Figure S3. DNA tethering can be achieved by bridging the DSB ends through an intercomplex dimerization at the Zn-hook interface of two distinct MR complexes.

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