

Table S1. List of genes used for lignin modification.

Gene	Function	Organism	Kingdom	Locus tag/ Gene symbol	Accession	Reference
<i>F5H</i>	ferulic acid 5-hydroxylase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT4G36220	AEE86636	[1]
<i>3AT2</i>	coumaroyl-CoA:anthocyanidin 3- <i>O</i> -glucoside-6"- <i>O</i> -coumaroyltransferase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT1G03495	AAF86513	[2]
<i>ACT</i>	agmatine coumaroyltransferase	<i>Hordeum vulgare</i>	Viridiplantae	–	BAF97626	[3]
<i>AT208</i>	anthocyanidin 3- <i>O</i> -glucoside 6'- <i>O</i> -acyltransferase	<i>Perilla frutescens</i>	Viridiplantae	–	BAA93475	[4]
<i>Atu1417</i>	enoyl-CoA hydratase	<i>Agrobacterium fabrum</i> C58	Bacteria	Atu1417	AAK87210	[5]
<i>Balat_0669</i>	chlorogenic acid esterase	<i>Bifidobacterium animalis</i> DSM 10140	Bacteria	Balat_0669	ACS47609	[6]
<i>BrRCD</i>	caffeic acid 4,5-dioxygenase	<i>Bradyrhizobium</i> sp. ORS278	Bacteria	BRADO_RS26060	WP_012029184	[7]
<i>C2H</i>	<i>p</i> -coumaroyl 2'-hydroxylase	<i>Ipomoea batatas</i>	Viridiplantae	–	BAL22347	[8]
<i>CA9OMT</i>	coniferyl alcohol 9- <i>O</i> -methyltransferase	<i>Linum nodiflorum</i>	Viridiplantae	–	ABX71750	[9]
<i>CADH</i>	coniferyl alcohol dehydrogenase	<i>Streptomyces</i> sp. NL15-2K	Bacteria	–	BAN09098	[10]
<i>CALipB</i>	lipase B	<i>Moesziomyces antarcticus</i>	Fungi	–	CAA83122	[11]
<i>calA</i>	coniferyl alcohol dehydrogenase	<i>Pseudomonas</i> sp. HR199	Bacteria	–	CAB69495	[12]
<i>calB</i>	coniferyl aldehyde dehydrogenase	<i>Pseudomonas</i> sp. HR199	Bacteria	–	CAA06926	[13]
<i>CASHT</i>	hydroxycinnamoyl-CoA: serotonin <i>N</i> -(hydroxycinnamoyl)transferase	<i>Capsicum annuum</i>	Viridiplantae	LOC107839132	AAK15312	[14]
<i>CCMT1</i>	cinnamic acid/ <i>p</i> -coumaric acid carboxyl methyltransferase	<i>Ocimum basilicum</i>	Viridiplantae	–	ABG75942	[15]
<i>CGT</i>	chlorogenic acid: glucaric acid caffeoyltransferase	<i>Solanum lycopersicum</i>	Viridiplantae	LOC101265588	CBV37053	[16]
<i>CHS5</i>	chalcone synthase	<i>Glycine max</i>	Viridiplantae	GLYMA_08G109200	AAB01004	[17]
<i>couA</i>	hydroxycinnamoyl-CoA hydratase-lyase	<i>Rhodopseudomonas palustris</i> CGA009	Bacteria	RPA1786	CAE27227	[18]
<i>CURS3</i>	curcumin synthase	<i>Curcuma longa</i>	Viridiplantae	–	BAH85781	[19]
<i>CYP84A4</i>	<i>p</i> -coumaraldehyde 3-hydroxylase	<i>Arabidopsis thaliana</i>	Viridiplantae	At5g04330	AED90728	[20]
<i>CYP98A22</i>	<i>p</i> -coumaroyl 2'-hydroxylase	<i>Ruta graveolens</i>	Viridiplantae	CYP98A22	AEG19446	[21]
<i>F6H1</i>	feruloyl-CoA 6'-hydroxylase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT3G13610	AEE75382	[22]
<i>FDC1</i>	phenacrylic acid decarboxylase	<i>Saccharomyces cerevisiae</i> S288C	Fungi	YDR539W	DAA12368	[23]
<i>HCALDH</i>	hydroxycinnamaldehyde dehydrogenase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT3G24503	AEE76907	[24]
<i>HCBT</i>	anthranilic acid <i>N</i> -hydroxycinnamoyl /benzoyltransferase	<i>Dianthus caryophyllus</i>	Viridiplantae	–	CAB06427	[25]
<i>HCT2</i>	malic acid <i>O</i> -hydroxycinnamoyl transferase	<i>Trifolium pratense</i>	Viridiplantae	–	ACI16631	[26]
<i>HHT1</i>	omega-hydroxypalmitic acid <i>O</i> -feruloyl transferase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT5G41040	AED94628	[27]
<i>ligL</i>	β-aryl ether <i>Cα</i> -dehydrogenase	<i>Sphingobium</i> sp. SYK-6	Bacteria	SLG_33660	BAK68041	[28]
<i>OsHCT4</i>	glycerol <i>O</i> -hydroxycinnamoyltransferase	<i>Oryza sativa</i>	Viridiplantae	OSNPB_060185500	BAS96510	[29]
<i>padC</i>	phenolic acid decarboxylase	<i>Bacillus subtilis</i> 168	Bacteria	BSU34400	CAB15445	[30]
<i>pcaHG</i>	protocatechuic acid 3,4-dioxygenase	<i>Stenotrophomonas maltophilia</i> KB2	Bacteria	–	AFH89645 (<i>pcaH</i>) AFH89644 (<i>pcaG</i>)	[31]
<i>pcaH2G2</i>	protocatechuic acid 3,4-dioxygenase	<i>Hydrogenophaga intermedia</i>	Bacteria	BN948_05072 BN948_05073	AAK84297 (<i>pcaH2</i>) AAK84298 (<i>pcaG2</i>)	[32]
<i>pdC</i>	<i>p</i> -coumaric acid decarboxylase	<i>Lactobacillus plantarum</i>	Bacteria	LPU63827	AAC45282	[33]
<i>PFOMT</i>	phenylpropanoid/flavonoid <i>O</i> -methyltransferase	<i>Mesembryanthemum crystallinum</i>	Viridiplantae	–	AAN61072	[34]
<i>PKS1</i>	<i>p</i> -coumaroyl-diketide-CoA synthase	<i>Wachendorfia thyrsoiflora</i>	Viridiplantae	–	AAW50921	[35]
<i>RAS</i>	rosmarinic acid synthase	<i>Melissa officinalis</i>	Viridiplantae	–	CBW35684	[36]
<i>rpaI</i>	acyl-homoserine lactone synthases	<i>Rhodopseudomonas palustris</i> CGA009	Bacteria	RPA0320	CAE25764	[37]
<i>SDT</i>	spermidine disinapoyl acyltransferase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT2G23510	AEC07460	[38]
<i>slr0095</i>	caffeoyl-CoA <i>O</i> -methyltransferase	<i>Synechocystis</i> sp. PCC 6803	Bacteria	SYNGTS_2605	BAK51353	[39]
<i>SNG1</i>	sinapoylglucose:malic acid <i>O</i> -sinapoyltransferase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT2G22990	AEC07390	[40]
<i>SPC4</i>	cationic peroxidase	<i>Sorghum bicolor</i>	Viridiplantae	LOC8074114	EES02106	[41]
<i>STS2</i>	resveratrol synthase	<i>Vitis vinifera</i>	Viridiplantae	VIT_00010554001	ABC84860	[42]
<i>THT10</i>	tyramine hydroxycinnamoyltransferase	<i>Nicotiana tabacum</i>	Viridiplantae	LOC107822189	CAB55502	[43]
<i>tyrB</i>	tyrosine aminotransferase	<i>Escherichia coli</i> K-12	Bacteria	b4054	NP_418478	[44]
<i>UGT72E1</i>	coniferaldehyde UDP-glucosyl transferase	<i>Arabidopsis thaliana col</i>	Viridiplantae	AT3G50740	AEE78703	[45]
<i>UGT84A2</i>	sinapic acid-1-glucosyltransferase	<i>Arabidopsis thaliana</i>	Viridiplantae	AT3G21560	AEE76523	[46]
<i>VpVAN</i>	vanillin synthase	<i>Vanilla planifolia</i>	Viridiplantae	–	AKG47593	[47]
<i>ydiB</i>	quinic acid/shikimic acid 5-dehydrogenase	<i>Escherichia coli</i> K-12	Bacteria	b1692	NP_416207	[48]

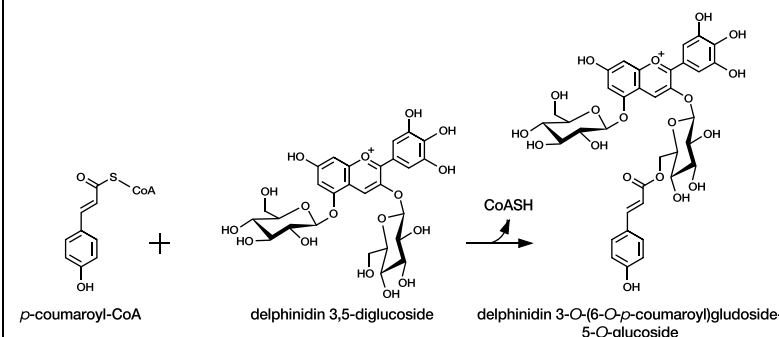
Table S2. Summary of enzyme properties for the conversion of metabolites in the shikimate, general phenylpropanoid, flavonoid, or monolignol biosynthetic pathways.

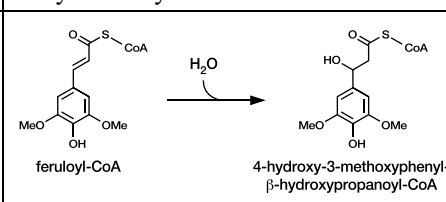
Reaction scheme, coenzyme requirement, enzyme activity (kinetic parameters, specific activity, or substrate range), and reference(s) are shown.

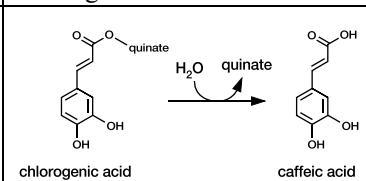
Gene	<i>F5H</i>
Enzyme	ferulic acid 5-hydroxylase
Reaction	<p>coniferaldehyde $\xrightarrow{\text{NADPH, H}^+, \text{O}_2}$ 5-hydroxy-coniferaldehyde</p>
Coenzyme	NADPH
Kinetics V_{\max} and K_m	coniferaldehyde, 5 $\text{pkat}\cdot\text{mg}^{-1}$ and 1 (μM) coniferyl alcohol, 6 $\text{pkat}\cdot\text{mg}^{-1}$ and 3 (μM) ferulic acid, 4 $\text{pkat}\cdot\text{mg}^{-1}$ and 1,000 (μM)
Reference	[1]

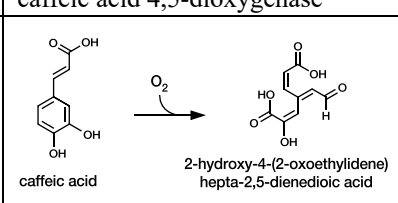
Gene	<i>3AT2</i>
Enzyme	anthocyanin acyltransferase
Reaction	<p><i>p</i>-coumaroyl-CoA + cyanidin 3,5-diglucoside $\xrightarrow{\text{CoASH}}$ cyanidin 3-O-(6-O-<i>p</i>-coumaroyl)glucoside-5-O-glucoside</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	<i>p</i> -coumaroyl-CoA, 6.9 sec^{-1} and 6.6 μM feruloyl-CoA, 7.7 sec^{-1} and 9.4 μM caffeoyl-CoA, 8.5 sec^{-1} and 4.9 μM sinapoyl-CoA, not determined
Reference	[2]

Gene	<i>ACT</i>
Enzyme	agmatine coumaroyltransferase
Reaction	<p><i>p</i>-coumaroyl-CoA + agmatine $\xrightarrow{\text{CoASH}}$ <i>p</i>-hydroxycinnamoylagmatine</p>
Coenzyme	unnecessary
Specific activity and K_m	<i>p</i> -coumaroyl-CoA, 9.9 $\text{kat}\cdot\text{M}^{-1}\cdot\text{g}^{-1}$ and 1.0 μM feruloyl-CoA, 1.4 $\text{kat}\cdot\text{M}^{-1}\cdot\text{g}^{-1}$ and 6.1 μM caffeoyl-CoA, 5.9 $\text{kat}\cdot\text{M}^{-1}\cdot\text{g}^{-1}$ and 5.8 μM
Reference	[3]

Gene	<i>AT208</i>
Enzyme	anthocyanidin 3- <i>O</i> -glucoside 6'- <i>O</i> -acyltransferase
Reaction	 <p><i>p</i>-coumaroyl-CoA + delphinidin 3,5-diglucoside $\xrightarrow{\text{CoASH}}$ delphinidin 3-<i>O</i>-(6-<i>O</i>-<i>p</i>-coumaroyl)glucoside-5-<i>O</i>-glucoside</p>
Coenzyme	unnecessary
Kinetics V_{max} and K_m	<i>p</i> -coumaroyl-CoA, 138 nkat·mg ⁻¹ and 24 μM caffeoyl-CoA, 168 nkat·mg ⁻¹ and 45 μM
Reference	[49]

Gene	<i>Atu1417</i>
Enzyme	enoyl-CoA hydratase
Reaction	 <p>feruloyl-CoA + H₂O \rightarrow 4-hydroxy-3-methoxyphenyl-β-hydroxypropanoyl-CoA</p>
Coenzyme	unnecessary
Substrate range	feruloyl-CoA, <i>p</i> -coumaroyl-CoA, caffeoyl-CoA
Reference	[5]

Gene	<i>Balat_0669</i>
Enzyme	chlorogenic acid esterase
Reaction	 <p>chlorogenic acid + H₂O \rightarrow caffeic acid + quinate</p>
Coenzyme	unnecessary
Specific activity	chlorogenic acid, 2.5 μmol·min ⁻¹ ·mg ⁻¹
Reference	[6]

Gene	<i>BrRCD</i>
Enzyme	caffeic acid 4,5-dioxygenase
Reaction	 <p>caffeic acid + O₂ \rightarrow 2-hydroxy-4-(2-oxoethylidene)hepta-2,5-dienedioic acid</p>
Coenzyme	unnecessary
Substrate range	caffeic acid, homoprotocatechuic acid, protocatechuic acid, catechol
Reference	[7]

Gene	<i>C2H</i>
Enzyme	<i>p</i> -coumaroyl 2'-hydroxylase
Reaction	<p><i>p</i>-coumaroyl-CoA + 2-oxoglutaric acid $\xrightarrow{O_2}$ 2,4-dihydroxycinnamoyl-CoA + succinic acid + CO₂</p>
Coenzyme	ascorbate
Kinetics k_{cat} and K_m	<i>p</i> -coumaroyl-CoA, 0.64 sec ⁻¹ and 15.25 μM feruloyl-CoA, 0.55 sec ⁻¹ and 14.06 μM
Reference	[8]

Gene	<i>CA9OMT</i>
Enzyme	coniferyl alcohol 9- <i>O</i> -methyltransferase
Reaction	<p>coniferyl alcohol + S-adenosyl-L-methionine \rightarrow coniferyl alcohol 9-methyl ether + S-adenosyl-L-homocysteine</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	coniferyl alcohol, 0.029 sec ⁻¹ and 7.23 μM
Reference	[9]

Gene	<i>CADH</i>
Enzyme	coniferyl alcohol dehydrogenase
Reaction	<p>coniferyl alcohol + NAD⁺ + H₂O $\xrightarrow{\quad}$ coniferaldehyde + NADH + H⁺</p>
Coenzyme	NAD ⁺
Kinetics k_{cat} and K_m	coniferyl alcohol, 69.7 sec ⁻¹ and 130 μM coniferyl alcohol, 116 sec ⁻¹ and 1700 μM
Reference	[10, 50]

Gene	<i>CALipB</i>
Enzyme	lipase B
Reaction	<p>ferulic acid + glycerol $\xrightarrow{-H_2O}$ feruloyl glycerol</p>
Coenzyme	unnecessary
Substrate range	ferulic acid, <i>p</i> -coumaric acid, caffeic acid
Reference	[51, 52]

Gene	<i>calA</i>
Enzyme	coniferyl alcohol dehydrogenase
Reaction	<p>coniferyl alcohol $\xrightarrow{\text{NAD}^+, \text{H}_2\text{O}}^{\text{NADH}, \text{H}^+}$ coniferaldehyde</p>
Coenzyme	NAD ⁺
Substrate	Only reported for coniferyl alcohol
Reference	[12]

Gene	<i>calB</i>
Enzyme	coniferyl aldehyde dehydrogenase
Reaction	<p>coniferaldehyde $\xrightarrow{\text{NAD}^+, \text{H}_2\text{O}}^{\text{NADH}, \text{H}^+}$ ferulic acid</p>
Coenzyme	NAD ⁺
Specific activity and K_m	coniferaldehyde, 36.5 $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{g}^{-1}$ and 7–12 μM sinapaldehyde, 28.1 $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{g}^{-1}$
Reference	[13]

Gene	<i>CASHT</i>
Enzyme	hydroxycinnamoyl-CoA: serotonin <i>N</i> -(hydroxycinnamoyl)transferase
Reaction	<p><i>p</i>-coumaroyl-CoA + serotonin $\xrightarrow{\text{CoASH}}$ <i>p</i>-coumaroylserotonin</p>
Coenzyme	unnecessary
Kinetics V_{max} and K_m	<i>p</i> -coumaroyl-CoA, 35 nkat·mg ⁻¹ and 23.2 μM feruloyl-CoA, 7 nkat·mg ⁻¹ and 3.5 μM caffeoyl-CoA, 31 nkat·mg ⁻¹ and 12.7 μM
Reference	[14, 53]

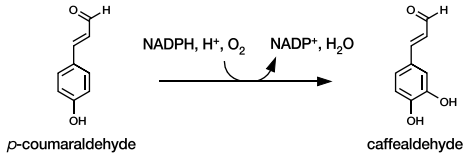
Gene	<i>CCMT1</i>
Enzyme	cinnamic acid/ <i>p</i> -coumaric acid carboxyl methyltransferase
Reaction	<p>cinnamic acid + S-adenosyl-L-methionine \longrightarrow methylcinnamate + S-adenosyl-L-homocysteine</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	cinnamic acid, 0.188 sec ⁻¹ and 124 μM <i>p</i> -coumaric acid, 116 sec ⁻¹ and 70.4 μM
Reference	[15]

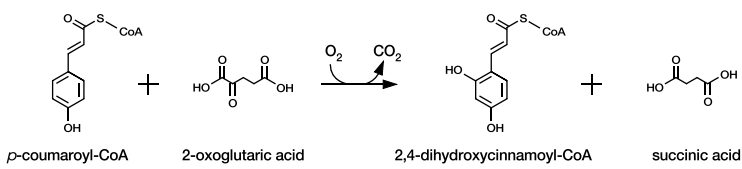
Gene	<i>CGT</i>
Enzyme	chlorogenic acid: glucaric acid caffeoyltransferase
Reaction	<p>caffeoyl-D-quininate + D-glucarate → caffeoylglucarate + L-quininate</p>
Coenzyme	unnecessary
Kinetics V_{\max} and K_m	D-glucaric acid, 624 nkat·mg ⁻¹ and 400 μM D-galactaric acid, 310 nkat·mg ⁻¹ and 1,700 μM
Reference	[16, 54]

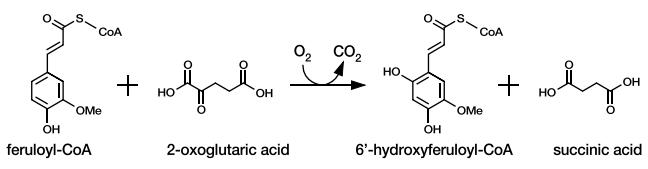
Gene	<i>CHS5</i>
Enzyme	chalcone synthase
Reaction	<p><i>p</i>-coumaroyl-CoA + 3 malonyl-CoA → naringenin chalcone + 4 CoASH, 3 CO₂</p>
Coenzyme	unnecessary
Substrate range	<i>p</i> -coumaroyl-CoA, caffeoyl-CoA, feruloyl-CoA
Reference	[17, 55]

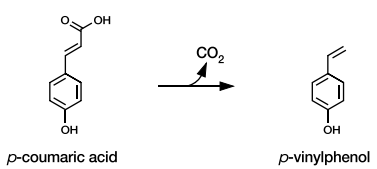
Gene	<i>couA</i>
Enzyme	hydroxycinnamoyl-CoA hydratase-lyase
Reaction	<p><i>p</i>-coumaroyl-CoA + H₂O → 3-(4-hydroxyphenyl)-3-hydroxypropionyl-CoA → <i>p</i>-hydroxybenzaldehyde + acetyl-CoA</p>
Coenzyme	unnecessary
Substrate range	<i>p</i> -coumaroyl-CoA, caffeoyl-CoA, feruloyl-CoA
Reference	[18]

Gene	<i>CURS3</i>
Enzyme	curcumin synthase
Reaction	<p><i>p</i>-coumaroyl-CoA + <i>p</i>-coumaroyldiketide-CoA → bisdemethoxycurcumin + 2 CoASH, CO₂</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	<i>p</i> -coumaroyl-CoA, 0.006 sec ⁻¹ and 3.4 μM (<i>p</i> -coumaroyl-CoA + cinnamoyldiketide- <i>N</i> -acetylcysteamine) feruloyl-CoA, 0.003 sec ⁻¹ and 2.2 μM (feruloyl-CoA + cinnamoyldiketide- <i>N</i> -acetylcysteamine)
Reference	[19]

Gene	<i>CYP84A4</i>
Enzyme	<i>p</i> -coumaraldehyde 3-hydroxylase
Reaction	 <p><i>p</i>-coumaraldehyde → caffealdehyde</p>
Coenzyme	NADPH
Kinetics V_{\max} and K_m	<i>p</i> -coumaraldehyde, 0.69 $\mu\text{kat}\cdot\text{mg}^{-1}$ and 41 μM
Reference	[20]

Gene	<i>CYP98A22</i>
Enzyme	<i>p</i> -coumaroyl 2'-hydroxylase
Reaction	 <p><i>p</i>-coumaroyl-CoA + 2-oxoglutaric acid → 2,4-dihydroxycinnamoyl-CoA + succinic acid</p>
Coenzyme	ascorbate
Kinetics k_{cat} and K_m	<i>p</i> -coumaroyl-CoA, 0.71 sec^{-1} and 50 μM feruloyl-CoA, 0.46 sec^{-1} and 37 μM
Reference	[21]

Gene	<i>F6H1</i>
Enzyme	feruloyl-CoA 6'-hydroxylase
Reaction	 <p>feruloyl-CoA + 2-oxoglutaric acid → 6'-hydroxyferuloyl-CoA + succinic acid</p>
Cofactor	ascorbate
Kinetics k_{cat} and K_m	feruloyl-CoA, 11.0 sec^{-1} and 36 μM <i>p</i> -coumaroyl-CoA, trace reaction
Reference	[22]

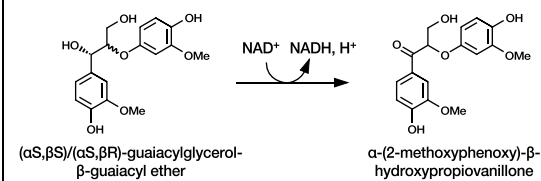
Gene	<i>FDC1</i>
Enzyme	phenacrylic acid decarboxylase
Reaction	 <p><i>p</i>-coumaric acid → <i>p</i>-vinylphenol</p>
Cofactor	prenylated FMNH ₂
Kinetics k_{cat} and K_m	<i>p</i> -coumaric acid, 1.5 sec^{-1} and 110 μM cinnamic acid, 4.6 sec^{-1} and 180 μM ferulic acid, 3.8 sec^{-1} and 180 μM
Reference	[56]

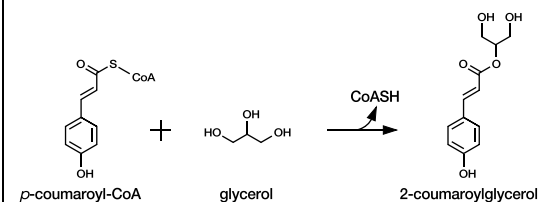
Gene	<i>HCALDH</i>
Enzyme	hydroxycinnamaldehyde dehydrogenase
Reaction	<p>sinapaldehyde → sinapic acid</p>
Coenzyme	NADP ⁺
Kinetics V_{\max} and K_m	coniferaldehyde, 560 pkat·mg ⁻¹ and 43 μM sinapaldehyde, 1700 pkat·mg ⁻¹ and 150 μM <i>p</i> -coumaraldehyde, 310 pkat·mg ⁻¹ and 19 μM
Reference	[57]

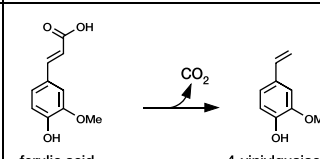
Gene	<i>HCBT</i>
Enzyme	anthranilic acid <i>N</i> -hydroxycinnamoyl/benzoyltransferase
Reaction	<p><i>p</i>-coumaroyl-CoA + anthranilic acid → <i>p</i>-coumaroyl-anthranilic acid</p>
Coenzyme	unnecessary
Substrate range	<i>p</i> -coumaroyl-CoA, caffeoyl-CoA, feruloyl-CoA, sinapoyl-CoA
Reference	[25, 58]

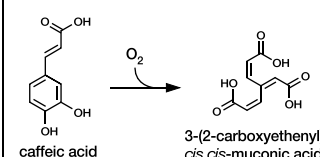
Gene	<i>HCT2</i>
Enzyme	malic acid <i>O</i> -hydroxycinnamoyl transferase
Reaction	<p><i>p</i>-coumaroyl-CoA + malic acid → <i>p</i>-coumaroyl-malic acid</p>
Coenzyme	unnecessary
Specific activity	<i>p</i> -coumaroyl-CoA, 34 nkat·mg ⁻¹ caffeoyl-CoA, 5.1 nkat·mg ⁻¹
Reference	[26]

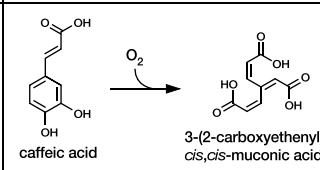
Gene	<i>HHT1</i>
Enzyme	omega-hydroxypalmitic acid <i>O</i> -feruloyl transferase
Reaction	<p><i>p</i>-coumaroyl-CoA + 16-hydroxypalmitic acid → 16-<i>p</i>-coumaroyloxy palmitic acid</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	<i>p</i> -coumaroyl-CoA, 0.0136 sec ⁻¹ and 36.7 μM feruloyl-CoA, 0.0173 sec ⁻¹ and 9.7 μM
Reference	[27]

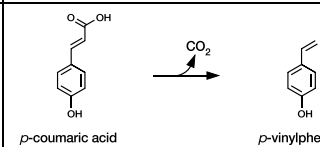
Gene	<i>ligL</i>
Enzyme	β -aryl ether C α -dehydrogenase
Reaction	 <p>(αS,βS)/(αS,βR)-guaiacylglycerol-β-guaiacyl ether</p> <p>α-(2-methoxyphenoxy)-β-hydroxypropiovanillone</p>
Coenzyme	NAD ⁺
Substrate range	β -O-4 dimers and oligomers with guaiacyl and syringyl structure
Reference	[28, 59]

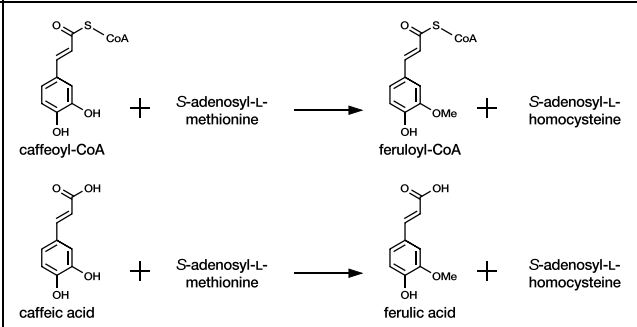
Gene	<i>OsHCT4</i>
Enzyme	glycerol <i>O</i> -hydroxycinnamoyltransferase
Reaction	 <p><i>p</i>-coumaroyl-CoA</p> <p>glycerol</p> <p>2-coumaroylglycerol</p>
Coenzyme	unnecessary
Kinetics <i>k</i> _{cat} and <i>K</i> _m	<i>p</i> -coumaroyl-CoA, 0.037 sec ⁻¹ and 60 μ M feruloyl-CoA, 0.071 sec ⁻¹ and 72 μ M caffeoyl-CoA, 0.082 sec ⁻¹ and 102 μ M
Reference	[29]

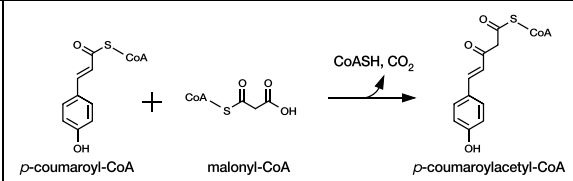
Gene	<i>padC</i>
Enzyme	phenolic acid decarboxylase
Reaction	 <p>ferulic acid</p> <p>4-vinylguaiacol</p>
Coenzyme	prenylated FMNH ₂ ?
Kinetics <i>V</i> _{max} and <i>K</i> _m	ferulic acid, 280 μ mol·min ⁻¹ ·mg ⁻¹ and 1100 μ M <i>p</i> -coumaric acid, 265 μ mol·min ⁻¹ ·mg ⁻¹ and 1300 μ M caffeic acid, 180 μ mol·min ⁻¹ ·mg ⁻¹ and 2600 μ M
Reference	[30]

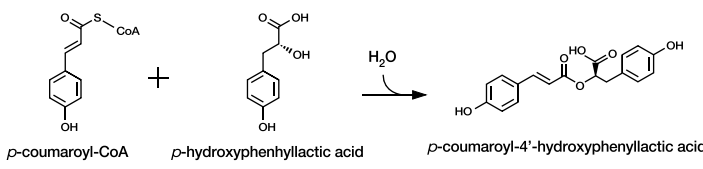
Gene	<i>pcaG</i> + <i>pcaH</i>
Enzyme	protocatechuic acid 3,4-dioxygenase
Reaction	 <p>caffeic acid</p> <p>3-(2-carboxyethenyl)-<i>cis,cis</i>-muconic acid</p>
Coenzyme	unnecessary
Substrate range	caffeic acid and dihydrocaffeic acid
Reference	[31]

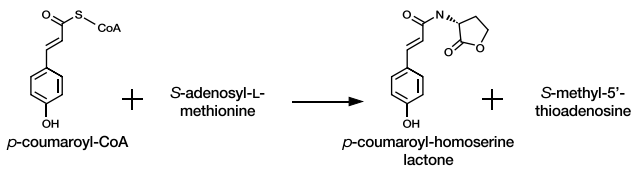
Gene	<i>pcaG2 + pcaH2</i>
Enzyme	protocatechuic acid 3,4-dioxygenase
Reaction	 <p>caffeic acid</p> <p>3-(2-carboxyethenyl)- <i>cis,cis</i>-muconic acid</p>
Coenzyme	unnecessary
Substrate range	caffeic acid and 3,4-dihydroxyphenylacetic acid
Reference	[32]

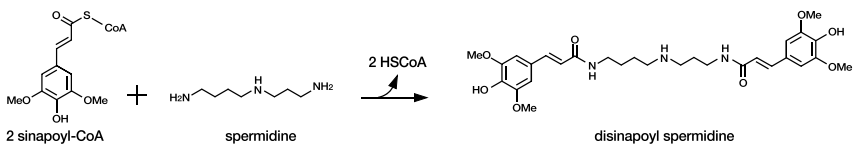
Gene	<i>pdc</i>
Enzyme	<i>p</i> -coumaric acid decarboxylase
Reaction	 <p><i>p</i>-coumaric acid</p> <p><i>p</i>-vinylphenol</p>
Coenzyme	prenylated FMNH ₂
Kinetics <i>V</i> _{max} and <i>K</i> _m	<i>p</i> -coumaric acid, 766 μmol·min ⁻¹ ·mg ⁻¹ and 1400 μM
Reference	[33]

Gene	<i>PFOMT</i>
Enzyme	phenylpropanoid/flavonoid <i>O</i> -methyltransferase
Reaction	 <p>caffeoyl-CoA</p> <p><i>S</i>-adenosyl-L-methionine</p> <p>feruloyl-CoA</p> <p><i>S</i>-adenosyl-L-homocysteine</p> <p>caffeic acid</p> <p><i>S</i>-adenosyl-L-methionine</p> <p>ferulic acid</p> <p><i>S</i>-adenosyl-L-homocysteine</p>
Coenzyme	unnecessary
Kinetics <i>V</i> _{max} and <i>K</i> _m	caffeoyl-CoA, 0.025 μmol·min ⁻¹ ·mg ⁻¹ and 25 μM caffeic acid, 0.005 μmol·min ⁻¹ ·mg ⁻¹ and 44 μM quercetin, 0.034 μmol·min ⁻¹ ·mg ⁻¹ and 6.4 μM
Reference	[34]

Gene	<i>PKS1</i>
Enzyme	<i>p</i> -coumaroyl-diketide-CoA synthase
Reaction	 <p><i>p</i>-coumaroyl-CoA</p> <p>malonyl-CoA</p> <p>CoASH, CO₂</p> <p><i>p</i>-coumaroylacetyl-CoA</p>
Coenzyme	unnecessary
Kinetics <i>k</i> _{cat} and <i>K</i> _m	<i>p</i> -coumaroyl-CoA, 0.00048sec ⁻¹ and 85 μM phenylpropionyl-CoA, 0.0025 sec ⁻¹ and 40 μM cinnamoyl-CoA, 0.00053 sec ⁻¹ and 28 μM
Reference	[35]

Gene	<i>RAS</i>
Enzyme	rosmarinic acid synthase
Reaction	 <p><i>p</i>-coumaroyl-CoA + <i>p</i>-hydroxyphenyllactic acid $\xrightarrow{-H_2O}$ <i>p</i>-coumaroyl-4'-hydroxyphenyllactic acid</p>
Coenzyme	unnecessary
Kinetics V_{max} and K_m	<i>p</i> -coumaroyl-CoA (+ <i>p</i> -hydroxyphenyllactic acid), 1076 pkat·mg ⁻¹ and 9.4 μM <i>p</i> -coumaroyl-CoA (+3,4-dihydroxyphenyllactic acid), 294 pkat·mg ⁻¹ and 3.2 μM caffeoyl-CoA (+ <i>p</i> -hydroxyphenyllactic acid), 283 pkat·mg ⁻¹ and 2.3 μM caffeoyl-CoA (+3,4-dihydroxyphenyllactic acid), 103 pkat·mg ⁻¹ and 3.3 μM
Reference	[37]

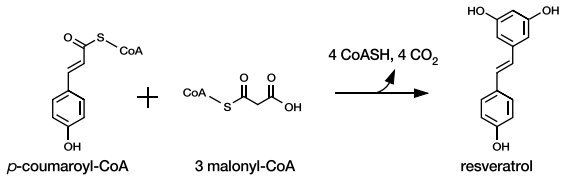
Gene	<i>rpal</i>
Enzyme	acyl-homoserine lactone synthases
Reaction	 <p><i>p</i>-coumaroyl-CoA + S-adenosyl-L-methionine \rightarrow <i>p</i>-coumaroyl-homoserine lactone + S-methyl-5'-thioadenosine</p>
Coenzyme	unnecessary
Reference	[36]

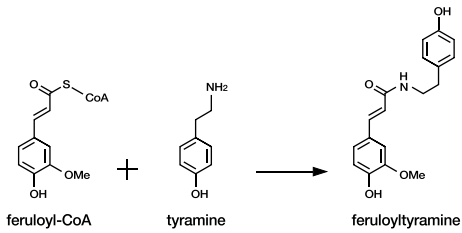
Gene	<i>SDT</i>
Enzyme	spermidine disinapoyl acyltransferase
Reaction	 <p>2 sinapoyl-CoA + spermidine $\xrightarrow{-2 HSCoA}$ disinapoyl spermidine</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	sinapoyl-CoA, 5.1 sec ⁻¹ and 8.3 μM
Reference	[38]

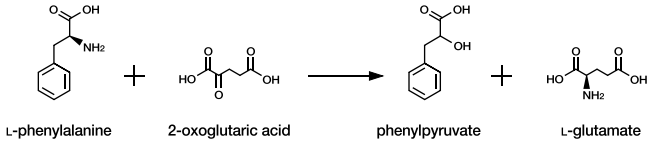
Gene	<i>slr0095</i>
Enzyme	caffeoyl-CoA <i>O</i> -methyltransferase
Reaction	<p>3,4,5-trihydroxycinnamic acid + S-adenosyl-L-methionine → 5-hydroxyferulic acid + 3,5-dihydroxy-4-O-methoxycinnamic acid + 5-hydroxy-3,4-O-dimethoxycinnamic acid</p> <p>5-hydroxyferulic acid + S-adenosyl-L-methionine → sinapic acid</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	3,4,5-trihydroxycinnamic acid, 0.0104 liter·sec ⁻¹ and 20.7 μM caffeoyl-CoA, 0.0245 liter·sec ⁻¹ and 32.9 μM caffeic acid, 0.0055 liter·sec ⁻¹ and 69.3 μM 5-hydroxyferulic acid, 0.0250 liter·sec ⁻¹ and 74.0 μM
Reference	[39]

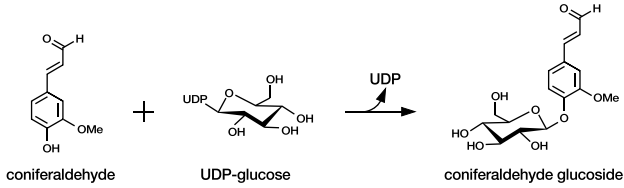
Gene	<i>SNG1</i>
Enzyme	sinapoylglucose:malic acid <i>O</i> -sinapoyltransferase
Reaction	<p>sinapoylglucose + malic acid → sinapoyl-malic acid + glucose</p>
Coenzyme	unnecessary
Kinetics V_{max} and K_m	sinapoylglucose, 0.528 μmol·min ⁻¹ ·mg ⁻¹ and 460 μM
Reference	[40]

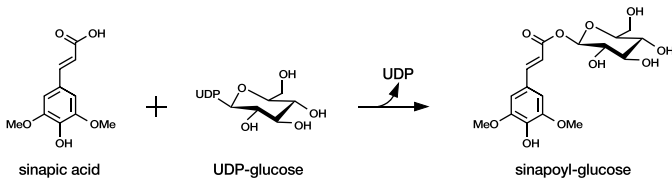
Gene	<i>SPC4</i>
Enzyme	cationic peroxidase
Reaction	<p>ferulic acid + H₂O₂ → ferulic acid radical + H₂O</p>
Coenzyme	unnecessary
Kinetics V_{max} / K_m	ferulic acid, 0.92 M ⁻¹ ·s ⁻¹ <i>p</i> -coumaric acid, 0.23 M ⁻¹ ·s ⁻¹
Reference	[41]

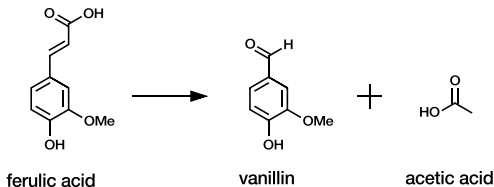
Gene	<i>STS2</i>
Enzyme	resveratrol synthase
Reaction	 <p><i>p</i>-coumaroyl-CoA + 3 malonyl-CoA $\xrightarrow{4 \text{ CoASH}, 4 \text{ CO}_2}$ resveratrol</p>
Coenzyme	unnecessary
Reference	[42]

Gene	<i>THT10</i>
Enzyme	tyramine hydroxycinnamoyltransferase
Reaction	 <p>feruloyl-CoA + tyramine \rightarrow feruloyltyramine</p>
Coenzyme	unnecessary
Kinetics V_{\max} and K_m	feruloyl-CoA, 69960 pkat·mg ⁻¹ and 0.6 μM sinapoyl-CoA, 76000 pkat·mg ⁻¹ and 1 μM <i>p</i> -coumaroyl-CoA, 19200 pkat·mg ⁻¹ and 2 μM cinnamoyl-CoA, 120320 pkat·mg ⁻¹ and 4.7 μM
Reference	[43]

Gene	<i>tyrB</i>
Enzyme	tyrosine aminotransferase
Reaction	 <p>L-phenylalanine + 2-oxoglutaric acid \rightarrow phenylpyruvate + L-glutamate</p>
Coenzyme	unnecessary
Kinetics V_{\max} and K_m	L-phenylalanine, 9.1 μmol·min ⁻¹ ·nmol protein ⁻¹ and 333 μM L-tyrosine, 20.0 μmol·min ⁻¹ ·nmol protein ⁻¹ and 625 μM
Reference	[44]

Gene	<i>UGT72E1</i>
Enzyme	coniferaldehyde UDP-glucosyl transferase
Reaction	 <p>coniferaldehyde + UDP-glucose $\xrightarrow{\text{UDP}}$ coniferaldehyde glucoside</p>
Coenzyme	unnecessary
Kinetics k_{cat} and K_m	coniferaldehyde, 1.22 sec ⁻¹ and 270 μM sinapaldehyde, 1.43 sec ⁻¹ and 460 μM
Reference	[45]

Gene	<i>UGT84A2</i>
Enzyme	sinapic acid-1-glucosyltransferase
Reaction	 <p>sinapic acid + UDP-glucose $\xrightarrow{\text{UDP}}$ sinapoyl-glucose</p>
Coenzyme	unnecessary
Kinetics V_{max} and K_m	sinapic acid, 13.44 nkat·mg ⁻¹ and 250 μM
Reference	[46]

Gene	<i>VpVAN</i>
Enzyme	vanillin synthase
Reaction	 <p>ferulic acid \rightarrow vanillin + acetic acid</p>
Coenzyme	unnecessary
Substrate range	ferulic acid and ferulic acid glucoside
Reference	[47]

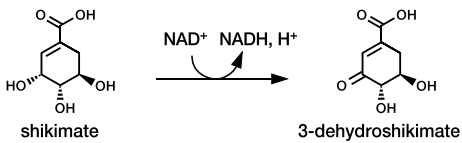
Gene	<i>ydiB</i>
Enzyme	quinic acid/shikimic acid 5-dehydrogenase
Reaction	 <p>shikimate $\xrightarrow{\text{NAD}^+ \rightarrow \text{NADH}, \text{H}^+}$ 3-dehydroshikimate</p>
Coenzyme	NAD ⁺
Kinetics k_{cat} and K_m	shikimic acid, 91000 sec ⁻¹ and 2.9 μM quinic acid, 113000 sec ⁻¹ and 9.1 μM
Reference	[48]

Table S3. List of plants generated and mixed *Agrobacterium* strains harboring different gene constructs used for seven independent inoculation procedures.

Gene set	Gene	Number of plants	Efficiency (%)	
1	Single-gene introduction	<i>CASHT</i>	9	18.4
		<i>pdv</i>	7	14.3
		<i>CCMT1</i>	7	14.3
		<i>tyrB</i>	6	12.2
		<i>THT10</i>	5	10.2
		<i>FDC1</i>	4	8.2
		<i>couA</i>	2	4.1
		<i>Atu1417</i>	0	0
	Multiple-genes introduction	9	-	
Subtotal	49	-		
2	Single-gene introduction	<i>rpaI</i>	11	26.2
		<i>STS2</i>	7	16.7
		<i>C2H</i>	6	14.3
		<i>CURS3</i>	2	4.8
		<i>PKS1</i>	2	4.8
		<i>AT208</i>	2	4.8
		<i>CHS5</i>	1	2.4
	Multiple-genes introduction	11	-	
Subtotal	42	-		
3	Single-gene introduction	<i>3AT2</i>	11	22.4
		<i>HCBT</i>	8	16.3
		<i>ACT</i>	6	12.2
		<i>HCT2</i>	5	10.2
		<i>OsHCT4</i>	5	10.2
		<i>RAS</i>	2	4.1
		<i>SDT</i>	2	4.1
		<i>CYP84A4</i>	2	4.1
	Multiple-genes introduction	8	-	
Subtotal	49	-		
4	Single-gene introduction	<i>CALipB</i>	8	16.3
		<i>SPC4</i>	8	16.3
		<i>UGT84A2</i>	6	12.2
		<i>BrRCD</i>	4	8.2
		<i>F5H</i>	4	8.2
		<i>PFOMT</i>	3	6.1
		<i>slr0095</i>	2	4.1
		<i>VpVAN</i>	2	4.1
	Multiple-genes introduction	12	-	
Subtotal	49	-		

Table S3. continued

Gene set	Gene	Number of Plants	Efficiency (%)	
5	Single-gene introduction	<i>Balat_0669</i>	8	16.3
		<i>calB</i>	8	16.3
		<i>HHT1</i>	5	10.2
		<i>HCALDH</i>	5	10.2
		<i>CGT</i>	4	8.2
		<i>F6H1</i>	4	8.2
		<i>UGT72E1</i>	4	8.2
		<i>padC</i>	0	0
		Multiple-genes introduction	11	-
		Subtotal	49	-
6	Single-gene introduction	<i>CYP98A22</i>	7	16.3
		<i>CA9OMT</i>	6	14
		<i>CADH</i>	3	9.3
		<i>calA</i>	4	9.3
		<i>ligL</i>	2	4.7
		<i>ydiB</i>	2	4.7
		<i>SNG1</i>	2	4.7
		Multiple-genes introduction	16	-
Subtotal	43	-		
7	Single-gene introduction	<i>pcaG2 + pcaH2</i>	8	53.3
		<i>pcaG + pcaH</i>	7	46.7
	Multiple-genes introduction	0	-	
	Subtotal	15	-	

The number of transgenic plants generated with a single gene and with multiple transgenes are indicated in the third column. Efficiency refers to the relative proportions of plants harboring a particular gene among all of the plants generated with the same mixture of *Agrobacterium* strains.

Table S4. Relative abundance of pyrolysis products within AIR-sample from wild-type and transgenic inflorescence stem

No.	Products	Structural feature	Wild type	<i>PkC4Hpro:: F5H</i>	<i>PkC4Hpro:: calB</i>	<i>PkC4Hpro:: couA</i>	<i>PkC4Hpro:: F6H1</i>
1	Phenol	H, unsub	0.8 (0.36)	0.49 (0.26)	0.65 (0.21)	1.36 (0.24)*	0.81 (0.3)
2	2-methylphenol	H, unsub	0.19 (0.06)	0.08 (0.05)*	0.16 (0.05)	0.23 (0.04)	0.15 (0.07)
3	4-Methylphenol	H, unsub	0.41 (0.14)	0.21 (0.12)*	0.36 (0.07)	0.65 (0.1)*	0.42 (0.15)
4	Guaiacol	G, unsub	2.87 (0.56)	0.56 (0.49)***	2.73 (0.2)	2.79 (0.31)	2.56 (0.38)
5	hydroquinone	H, misc	1.4 (0.44)	0.85 (0.52)	1.4 (0.36)	1.98 (0.48)	1.55 (0.5)
6	4-ethylphenol	H, ethyl	ND	ND	ND	0.01 (0.01)	0.01 (0.02)
7	2,4-dimethylphenol	H, methyl	1.12 (0.14)	0.22 (0.25)***	1.13 (0.15)	1.1 (0.17)	1.26 (0.22)
8	4-methylguaiacol	G, methyl	1.36 (0.14)	0.25 (0.3)***	1.35 (0.16)	1.29 (0.18)	1.52 (0.27)
9	4-Ethylguaiacol	G, ethyl	0.15 (0.08)	ND	0.15 (0.09)	0.31 (0.05)**	0.15 (0.09)
10	4-Vinylguaiacol	G, vinyl	12.13 (2.15)	2.34 (2.26)***	11.79 (0.74)	15.45 (1.71)*	11.43 (1.74)
11	4-vinylphenol	H, vinyl	8.09 (1.43)	1.56 (1.51)***	7.86 (0.49)	10.3 (1.14)*	7.62 (1.16)
12	3-methoxy-5-methylphenol	G, methyl	0.02 (0.02)	0.03 (0.02)	0.04 (0.02)	0.08 (0.05)*	0.03 (0.03)
13	4-propenylphenol	H, methyl	0.02 (0.01)	0.02 (0.01)	0.01 (0.01)	0.03 (0.01)	0.02 (0.02)
14	syringol	S, unsub	1.38 (0.26)	3.51 (0.7)***	1.96 (0.32)**	1.99 (0.39)**	1.79 (0.35)*
15	Eugenol	G, misc	0.61 (0.08)	0.13 (0.14)***	0.59 (0.07)	0.41 (0.07)**	0.64 (0.12)
16	4-hydroxybenzaldehyde	H, C α -O	0.03 (0.01)	0.02 (0.01)	0.02 (0.02)	0.06 (0.04)	0.03 (0.03)
17	4-Propylguaiacol	G, misc	0.27 (0.05)	0.05 (0.07)***	0.27 (0.05)	0.33 (0.04)*	0.28 (0.05)
18	Vanillin	G, C α -O	1.01 (0.11)	0.21 (0.17)***	0.93 (0.12)	0.88 (0.07)	0.98 (0.13)
19	cis-isoeugenol	G, misc	0.49 (0.1)	0.11 (0.11)***	0.47 (0.07)	0.38 (0.05)	0.5 (0.1)
20	4-methylsyringol	S, methyl	0.71 (0.13)	1.46 (0.29)***	1.01 (0.15)**	1.08 (0.12)***	1.11 (0.14)***
21	trans-Isoeugenol	G, misc	3.4 (0.64)	0.76 (0.76)***	3.32 (0.37)	2.6 (0.31)*	3.5 (0.67)
22	4-propyneguaiacol	G, misc	0.35 (0.06)	0.16 (0.05)***	0.31 (0.06)	0.29 (0.06)	0.32 (0.08)
23	4-alleneguaiacol	G, misc	0.31 (0.06)	0.14 (0.04)**	0.27 (0.06)	0.25 (0.05)	0.28 (0.06)
24	acetovanilone	G, C α -O	1.08 (0.03)	0.25 (0.2)***	0.99 (0.05)**	1.34 (0.15)**	1.06 (0.13)
25	4-ethylsyringol	S, ethyl	0.05 (0.01)	0.12 (0.04)**	0.08 (0.02)*	0.11 (0.02)***	0.07 (0.02)*
26	Guaiacylacetone	G, C β -O	1.21 (0.11)	0.25 (0.27)***	1.19 (0.04)	1.1 (0.06)	1.34 (0.21)
27	4-vinylsyringol	S, vinyl	5.29 (0.73)	13.16 (2.94)***	7.22 (1.28)**	9.26 (0.82)***	6.87 (1.05)**
28	4-allylsyringol	S, misc	0.42 (0.06)	1.2 (0.35)***	0.6 (0.1)**	0.65 (0.09)***	0.62 (0.08)***
29	homosyringaldehyde	S, C β -O	0.08 (0.01)	0.18 (0.05)***	0.11 (0.02)*	0.13 (0.02)***	0.11 (0.02)**
30	cis-4-propenylsyringol	S, misc	0.29 (0.05)	0.8 (0.25)***	0.41 (0.08)**	0.49 (0.07)***	0.42 (0.07)**
31	Syringaldehyde	S, C α -O	0.9 (0.13)	2.8 (0.34)***	1.29 (0.15)***	1.05 (0.22)	1.29 (0.18)***
32	cis-Coniferyl-alcohol	G, C α -O	3.45 (0.18)	0.69 (0.8)***	3.14 (0.27)*	2.45 (0.24)***	3.15 (0.27)*
33	4-propynesyringol	S, misc	0.22 (0.03)	0.76 (0.27)***	0.31 (0.05)***	0.34 (0.05)***	0.29 (0.05)**
34	4-allenesyringol	S, misc	0.16 (0.02)	0.52 (0.18)***	0.22 (0.04)**	0.24 (0.04)***	0.2 (0.03)*
35	trans-4-propenylsyringol	S, misc	2 (0.35)	5.55 (1.71)***	2.85 (0.53)**	3.36 (0.47)***	2.89 (0.44)**
36	acetosyringone	S, C α -O	0.11 (0.15)	1.06 (0.32)***	0.25 (0.19)	0.36 (0.23)*	0.35 (0.15)*
37	Coniferylaldehyde	G, C γ -O	1.12 (0.88)	ND	0.54 (0.78)	0.16 (0.3)	1.27 (0.74)
38	trans-coniferyl-alcohol	G, C γ -O	30.77 (2.46)	7.07 (5.66)***	26.41 (3.03)*	19.46 (1.83)***	24.87 (3.24)**
39	Syringylacetone	S, C β -O	0.17 (0.02)	0.54 (0.12)***	0.24 (0.04)***	0.32 (0.04)***	0.25 (0.05)***
41	Coniferylacetate	S, C γ -O	0.01 (0)	ND	0.01 (0)	0.01 (0.01)*	0.02 (0)*
42	cis-Sinapyl alcohol	S, C γ -O	1.22 (0.19)	3.72 (0.49)***	1.54 (0.11)*	1.45 (0.25)	1.6 (0.25)**
43	trans-sinapaldehyde	S, C γ -O	0.08 (0.09)	0.65 (0.49)*	0.08 (0.12)	0.02 (0.03)	0.13 (0.09)
44	trans-sinapyl-alcohol	S, C γ -O	14.26 (3.74)	47.49 (11.44)***	15.73 (1.78)	13.81 (2.95)	15.61 (4.53)

Peak molar areas were calculated for the lignin degradation products with relative response factor reported by van Erven (2017). □

The summed areas were normalized, and the amount of each compound is expressed as percentages.

Numbers in parentheses are 95% confidential intervals.

Table S5. carbohydrates composition in transgenic lines supporting Fig.6

Lines	Carbohydrates ($\mu\text{g}/\text{mgAIR}$)										Total carbohydrates ($\mu\text{g}/\text{mgAIR}$)
	Glc	Xyl	GalA	Rha	Man	Gal	Ara	GlcA	mGlcA	Fuc	
Wild-type	377 (16)	145 (7)	119 (9)	44.9 (2.9)	30.5 (1.1)	22.1 (2)	12.7 (2.2)	5.92 (0.43)	2.03 (0.27)	1.07 (0.11)	685 (26)
<i>PkC4Hpro::F5H</i>	381 (9)	154 (6)*	120 (11)	46 (3.7)	31.8 (1.4)	23.4 (1.9)	14.5 (1.7)	6.48 (0.78)	2.09 (0.86)	1.08 (0.21)	702 (41)
<i>PkC4Hpro::calB</i>	377 (10)	151 (5)	130 (9)	48 (2.9)	31 (1.4)	24.2 (1.3)*	15.1 (1.6)	5.91 (0.52)	5.92 (2.28)***	1.11 (0.19)	710 (34)
<i>PkC4Hpro::couA</i>	326 (10)***	143 (6)	140 (6)**	63.9 (5.1)***	31.2 (1.1)	36 (3.7)***	31.8 (6.3)***	6.8 (0.55)**	7.53 (2.75)***	1.21 (0.09)*	708 (23)
<i>PkC4Hpro::F6H1</i>	352 (18)*	133 (8)*	128 (7)	46.5 (2.2)	28.3 (1.4)*	23.5 (1.2)	14.2 (1.2)	5.71 (0.55)	1.87 (1.15)	1.23 (0.11)**	661 (50)

Numbers in parentheses are 95% confidential intervals.

Table S6. sugar yield by enzymatic saccharification supporting to Fig.7

Line	Glucose yield ($\mu\text{g}/\text{mgAIR}$)		Xylose yield ($\mu\text{g}/\text{mgAIR}$)	
	Without pretreatment	Pretreatment with 100 mM NaOH	Without pretreatment	Pretreatment with 100 mM NaOH
Wild-type	126.2 (16)	136 (11)	3.9 (2)	87 (5)
<i>PkC4Hpro::F5H</i>	126.1 (12)	178 (32)*	0.9 (1)*	127 (16)***
<i>PkC4Hpro::calB</i>	133.5 (17)	164 (25)	4.5 (2)	108 (12)**
<i>PkC4Hpro::couA</i>	144.7 (13)	210 (17)***	17.9 (7)**	134 (16)***
<i>PkC4Hpro::F6H1</i>	118.8 (7)	140 (13)	0.8 (1)*	91 (10)

Numbers in parentheses are 95% confidential intervals.

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