



Supplementary Information for

**Room-temperature autonomous self-healing glassy polymers with hyperbranched structure**

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Movies S1

## Supplementary Information Text

### Materials and Methods

**Materials.** *N,N'*-methylene diacrylamide (MBA, 99+%) and Methanol (99.9%) were purchased from Adamas. 1,4-Butanediamine (BDA, 98+%) was purchased from Alfa Aesar. Acetone and deionized water were purchased from Chengdu Kelong Chemical Reagent Factory. All the chemicals were used as received without further purification.

**Synthesis of regular hyperbranched polymers.** The randomly hyperbranched polymers (RHP) were synthesized by one-pot method through Michael addition reaction between MBA and BDA at different molar ratios. The materials were denoted as RHP-x, where x 1, 2, 3 correspond to MBA/BDA molar ratios of 1/1.125, 1/1 and 1/0.875, respectively. In a typical procedure, the synthesis of RHP-1 was described as follows: MBA (12.332 g, 0.08 mol) was added into a round bottom flask equipped with a magnetic stirrer containing mixed solvent of 60 mL methanol and 30 mL deionized water at 30 °C and stirred until it was dissolved totally. Then BDA (8 g, 0.09 mol) was dissolved in a beaker containing mixed solvent of 20 mL methanol and 10 mL deionized water and fed into the flask directly. The mixture was stirred at 30 °C for 24 h. After that, the solution was poured into a beaker containing 1000 mL acetone to precipitate at room temperature. The crude product was washed 5 times with acetone to obtain a solid and then dried in a vacuum oven at 50 °C for 48 h.

**Preparation of RHP films.** In a typical process, 5 g RHP was powdered in a universal crusher, and then the powder was hot pressed under 10 MPa at 100 °C for 30 min in the mold. The resulting yellowish transparent film of RHP-1 was shown in Fig. 1D. We used gel permeation chromatography (GPC) to measure the molecular weight of RHP-2 after processing. The result shows that  $M_n=7286$  g/mol and  $M_w=20182$  g/mol for the hot pressed RHP-2, which are very close to the values of RHP-2 before processing. It indicates that the molecular weight of RHP does not change during processing.

**Characterization.** FTIR spectra were recorded on Nicolet iS10 (Nicolet, America) in the range of 4000-400  $\text{cm}^{-1}$  at room temperature. The FTIR sample of RHP was formed after evaporation of methanol on a piece of KBr plate. The RHP was heated from 20 °C to 150 °C at 1 °C/min, and the temperature-dependent FTIR spectra were collected at the same time. Moreover, we collected 21 spectra (from 20 °C to 40 °C at the heating rate of 1 °C/min) and used 2DCS software (get the open version from <http://muchong.com/t-11812062-1>) to process these data, generating the generalized 2D correlation spectra. To obtain a credible result, the baseline correction was performed in the wavenumber region of 1585-1495  $\text{cm}^{-1}$  before analysis. The  $^1\text{H}$  NMR spectra were measured on a Bruker AV III HD spectrometer operating at 400 MHz in deuterated methanol solution with TMS as reference. Quantitative  $^{13}\text{C}$  NMR spectra were measured by the method of inverse gated  $^1\text{H}$  decoupling on a Bruker AV III HD spectrometer operating at 400 MHz in deuterated dimethyl sulfoxide solvent.  $^{13}\text{C}$ ,  $^1\text{H}$ -HSQC spectra were recorded using the standard pulse sequence provided by Bruker. The  $^1\text{H}$  NMR curves with different temperatures were measured on Bruker AVANCE IIIITM HD 400 MHz spectrometer with deuterated dimethyl sulfoxide as solvent. X-ray diffraction experiments on RHP were carried out on a Rigaku X-ray diffractometer (Ultima IV) equipped with parallel beam optics attachment. X-ray diffraction of RHP at different temperatures was measured with Rigaku X-ray diffractometer (Smartlab). DSC tests were performed on a Perkin Elmer Diamond 3000 with the mass of all samples ranging from 3 mg to 8 mg. Samples were heated from -50 °C to 150 °C at a heating rate of 10 °C/min and then cooling to -50 °C at 10 °C/min. The heating and cooling processes were performed two times. DLS measurements were carried out by a wide angle laser scattering instrument (BI-200SM, Brookhaven) equipped with a cuvette rotation/translation unit (CRTU) and a He-Ne laser (22 mW, wavelength  $\lambda$  of 632.8 nm) to study the mean diameter of RHP. The molecular weight and polydispersity index (PDI) of the RHP were

determined by GPC at room temperature on an Agilent PLgel 5um MIXED-C gel columns with a Waters-2414 refractive index detector (DMF as the eluent). DMA tests were measured on Q800 (TA Instrument) in a tension mode with the sample dimension about 20×5×1 mm<sup>3</sup>. Tests were performed in temperature scanning mode in the range of -20 °C to 150 °C at a ramping rate of 3 °C /min and a frequency of 1 Hz with a strain amplitude of 30 μm. Dielectric measurements were carried out through two different patterns on a Novocontrol Concept 50 system with Alpha impedance analyzer and Quatro Cryosystem temperature control. Frequency sweep mode used the frequency range of 10<sup>-1</sup> to 10<sup>7</sup> Hz at each temperature from -60 to 100 °C with 10 °C interval, with temperature stability better than 0.1 K. The disk-shaped film with 1 mm thickness and 20 mm in diameter was placed into two parallel electrodes. Mechanical properties were performed on an Instron 5967 tensile tester at room temperature with a strain rate of 10 mm/min. The dumbbell shaped sheets had a thin strip with central dimensions of 35×2×1 mm<sup>3</sup>. To measure the self-healing efficiency, the dumbbell shaped specimen was cut by a razor blade, and the fresh cut surfaces were recombined by hand and then put into vacuum oven at 25 °C for different time. The healed sample was subjected to on stretching experiment again.

### Theoretical calculation

**Degree of branching (DB).** The degree of branching can be calculated by the following equation (1):

$$DB = \frac{D + T}{D + T + L}$$

where D, T, L represent the fractions of the dendritic unit, terminal unit and linear unit, respectively, which are calculated by the quantitative <sup>13</sup>C NMR spectra.

**Analyses of the dielectric spectra.** Below  $T_g$ , the analyses of the dielectric spectra are made by using Havriliak and Negami (HN) function. In this model, the frequency dependence of the dielectric complex permittivity ( $\epsilon^*$ ) can be described by the following equation (2,3):

$$\epsilon^*(\omega) = \epsilon_\infty + \frac{\Delta\epsilon}{[1 + (i\omega\tau_{HN})^\alpha]^\beta}$$

where  $\Delta\epsilon = \epsilon_s - \epsilon_\infty$  is the dielectric strength,  $\epsilon_s$  and  $\epsilon_\infty$  are the relaxed and unrelaxed values of dielectric constant, and  $\tau_{HN}$  is the characteristic relaxation time. The parameters  $\alpha$  and  $\beta$  ( $0 < \alpha, \alpha\beta \leq 1$ ) define the symmetrical and asymmetrical broadening of the loss peak.

The relation between  $\tau_{HN}$  and average relaxation time  $\tau_{max}$  is given by the following equation:

$$\tau_{max} = \tau_{HN} \left[ \sin \frac{\pi\alpha\beta}{2(1+\beta)} \right]^{\frac{1}{\alpha}} \left[ \sin \frac{\pi\alpha}{2(1+\beta)} \right]^{-\frac{1}{\alpha}} ; f_{max} = \frac{1}{2\pi\tau_{max}}$$

where  $f_{max}$  is the frequency at which  $\epsilon''$  passes through the maximum value.

The relation between the average relaxation time  $\tau_{max}$  and the temperature is Arrhenius-like and can be described by

$$\tau_{max} = \tau_0 \exp\left(\frac{E_a}{RT}\right)$$

where  $E_a$  is the activation energy and  $\tau_0$  is a proportionality constant.

Above  $T_g$ , both real and imaginary parts of dielectric permittivity are large, the relaxation process can be distinguished from the complex dielectric modulus format. Dielectric modulus ( $M^*$ ) is defined as the reciprocal of complex permittivity and can be described by (4)

$$M^*(\omega) = M'(\omega) + iM''(\omega) = \frac{1}{\epsilon^*(\omega)} = \frac{\epsilon'(\omega) + i\epsilon''(\omega)}{[\epsilon'(\omega)]^2 + [\epsilon''(\omega)]^2}$$

We refer to the value obtained from the maximum in the dielectric loss modulus as the average relaxation time  $\tau_{max}$ . This characteristic relaxation time can be correlated with the temperature through Vogel-Fulcher-Tamman (VFT) equation (5,6):

$$\tau_{max} = \tau_0 \exp\left(\frac{B}{T - T_0}\right)$$

where  $\tau_0$  and B are empirical parameters and  $T_0$  is the so-called Vogel temperature.

**Self-healing efficiency.** Healing efficiency ( $\eta$ ) is calculated according to the following equation:

$$\eta = \frac{\sigma_{heal}}{\sigma_{pri}} \times 100\%$$

where  $\sigma_{heal}$  is the tensile strength of the healing samples, and  $\sigma_{pri}$  is the tensile strength for the pristine samples.

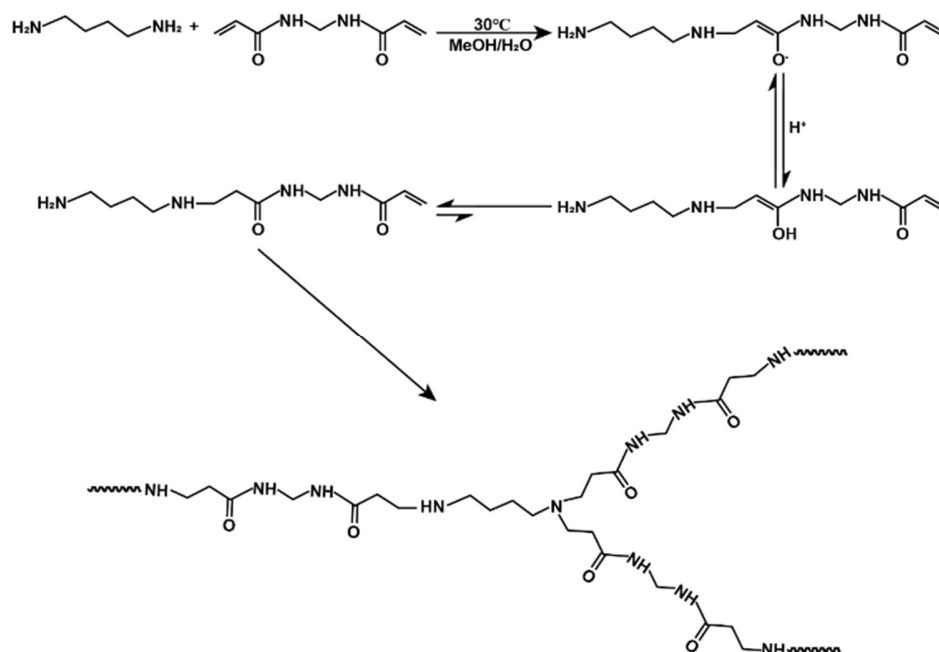
### **Noda's rule for the generalized 2D correlation spectra**

If the correlation intensity  $\Phi(v1, v2)$  in synchronous spectra has the same symbol (positive or negative) as the correlation peak  $\Psi(v1, v2)$  in asynchronous spectra, then the movement of band  $v1$  is prior to or earlier than that of band  $v2$ , and vice versa. Besides, if the correlation intensity in synchronous spectra is not zero (or blank), but zero in asynchronous one, then the movements of bands at  $v1$  and  $v2$  are simultaneous.

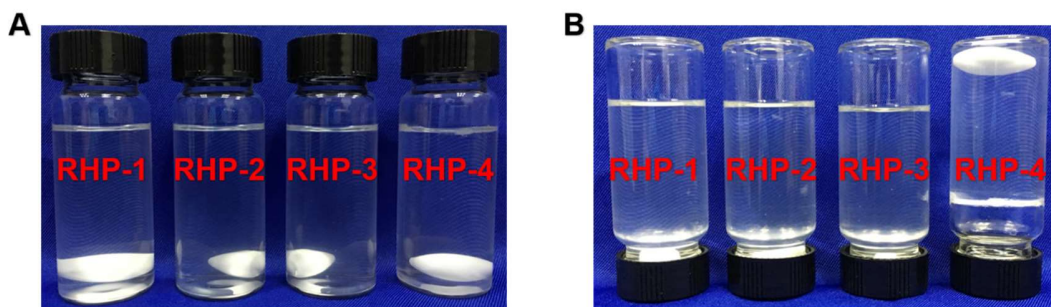
Noda's rules are summarized as follows:

- (1) If  $\Phi(v1,v2) > 0, \Psi(v1,v2) > 0$  or  $\Phi(v1,v2) < 0, \Psi(v1,v2) < 0$ , then the movement of  $v1$  is before than that of  $v2$ .
- (2) If  $\Phi(v1,v2) > 0, \Psi(v1,v2) < 0$  or  $\Phi(v1,v2) < 0, \Psi(v1,v2) > 0$ , then the movement of  $v1$  is after than that of  $v2$ .
- (3) If  $\Phi(v1,v2) > 0, \Psi(v1,v2) = 0$  or  $\Phi(v1,v2) < 0, \Psi(v1,v2) = 0$ , then the movements of  $v1$  and  $v2$  are simultaneous.

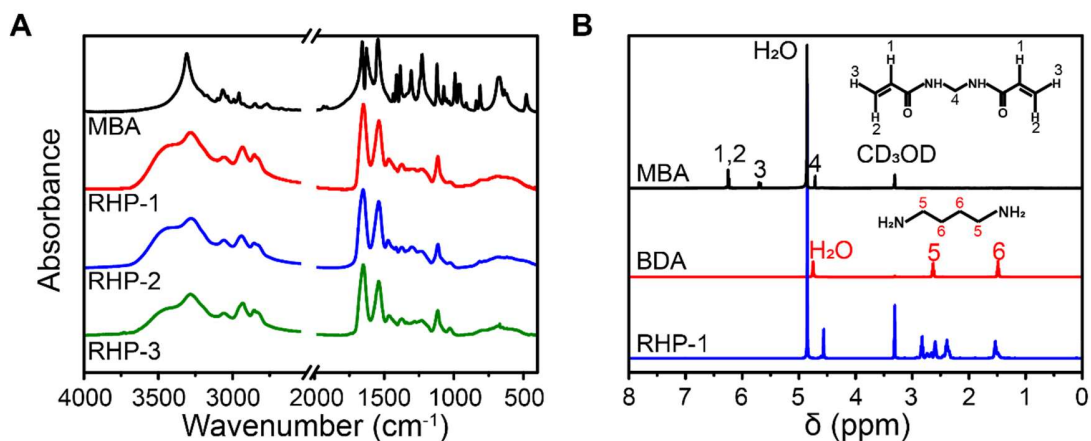
$\Phi(v1,v2)$  and  $\Psi(v1,v2)$  represent the correlation peaks in synchronous and asynchronous spectra, respectively.



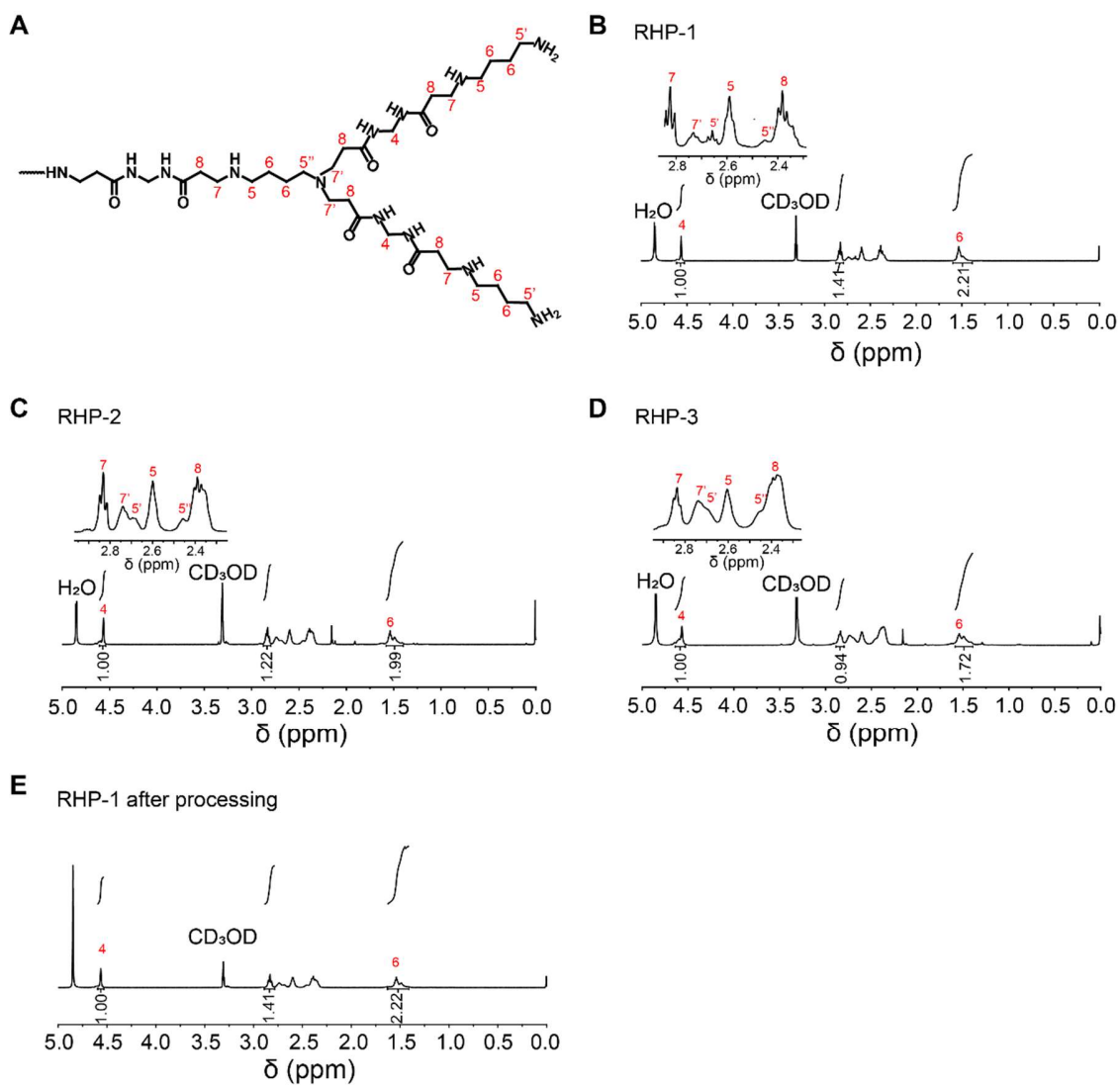
**Fig. S1.** Schematic diagram of synthetic reaction mechanism of RHP. The -NH- of BDA can react with a CH<sub>2</sub>=CH- on MBA under a mild condition to fabricate RHP molecules. And the reaction can form new secondary amines and tertiary amines.



**Fig. S2.** Photographs showing the dissolution state of RHP in methanol/water solvent after reacting for 0 minutes (A) and 24 h (B). After reaction for 24 h, RHP-1, RHP-2 and RHP-3 with MBA to BDA ratio from 1/1.125 to 1/0.875 are soluble in the solvent, as when the vials are upside down, the solutions flow down. However, when the MBA to BDA reaches 1:0.75 in RHP-4, gelation takes place. Therefore, even when the bottle is upside down, the system cannot flow and the stirrer bar is still on the top.

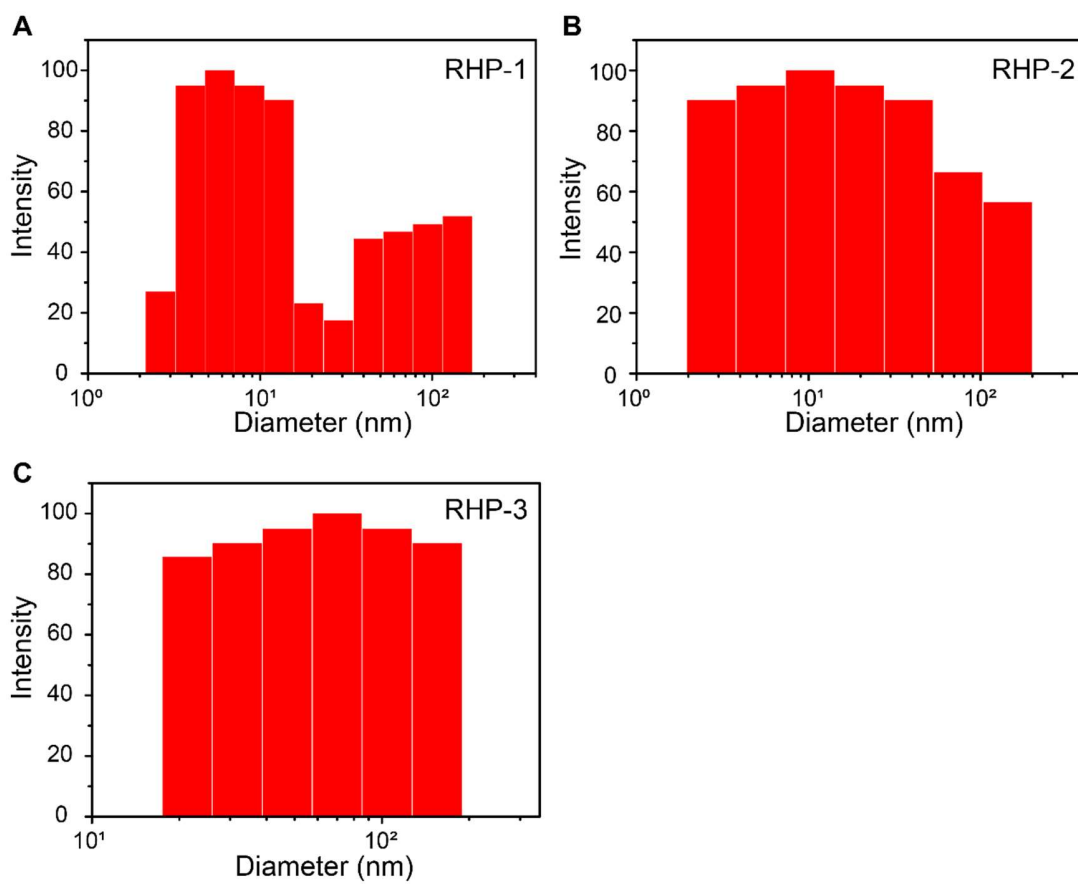


**Fig. S3.** FTIR and NMR analysis. (A) FTIR spectra of MBA, RBP-1, RBP-2 and RBP-3, respectively. In the FTIR spectrum of MBA, the characteristic peaks at 3305 cm<sup>-1</sup>, 1658 cm<sup>-1</sup>, 1543 cm<sup>-1</sup> are belonged to the strong stretching vibration of N-H group, C=O stretching vibration peak and N-H bending vibration peak, respectively, while the peaks at 992 cm<sup>-1</sup> and 967 cm<sup>-1</sup> are assigned to the bending vibration peaks of vinyl groups (7). In RHP, the characteristic amide I band at around 1648 cm<sup>-1</sup>, amide II band at around 1536 cm<sup>-1</sup> and disappeared vinyl group bending vibration peak indicate that the Michael addition reaction consumes all double bonds and hyperbranched networks are successfully formed. (B) <sup>1</sup>H NMR spectra of MBA, BDA and RHP-1. The typical proton signals at 6.25 ppm and 5.70 ppm corresponding to vinyl groups disappear after reaction, while new signals appear at 2-3 ppm belonging to the methylene of -CH<sub>2</sub>-CH<sub>2</sub>-CONH- in RHP-1. It indicates that all the vinyl groups have been consumed (8).

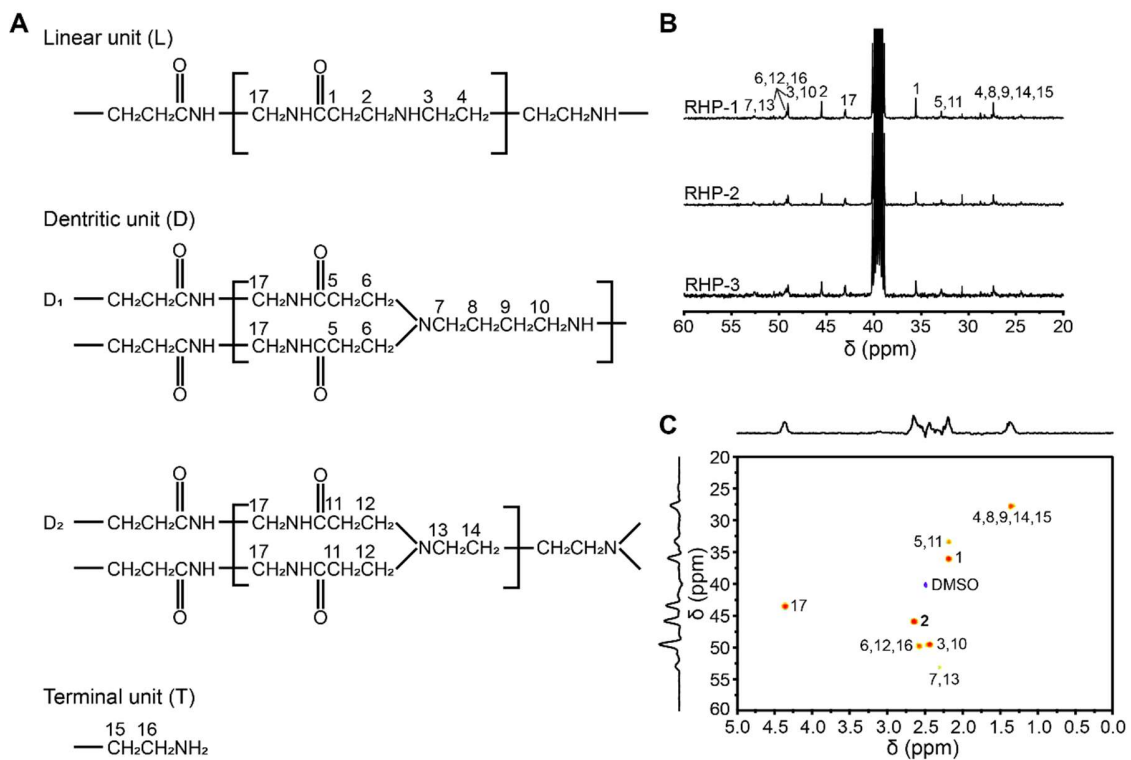


**Fig. S4.** Structure characterization of RHP. (A) Chemical structure of RHP.  $^1\text{H}$  NMR spectrum of RHP-1 (B), RHP-2 (C) and RHP-3 (D) (400 MHz, in  $\text{CD}_3\text{OD}$ ). With the molar ratio between MBA and BDA increases, the percentage of primary amine units decreases and the tertiary amine units increases (details are shown in Table S2). (E)  $^1\text{H}$  NMR spectrum of RHP-1 after processing (400 MHz, in  $\text{CD}_3\text{OD}$ ). The  $^1\text{H}$  NMR spectrum of RHP-1 after processing is essentially the same as that before processing (Fig S4 B), indicating that the hyperbranched structure do not change after processing.

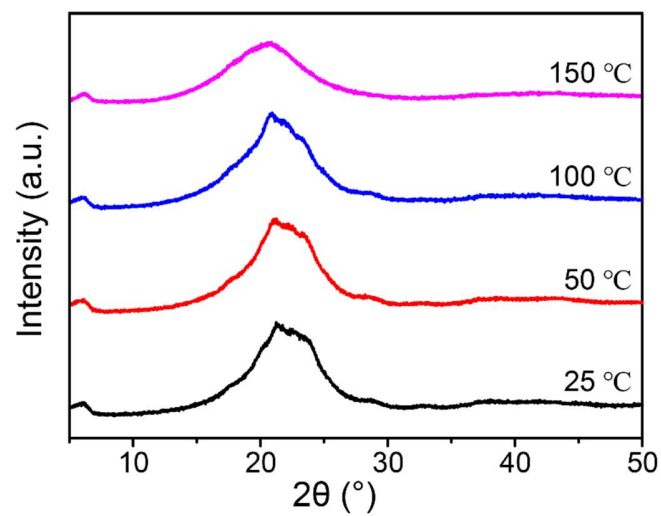




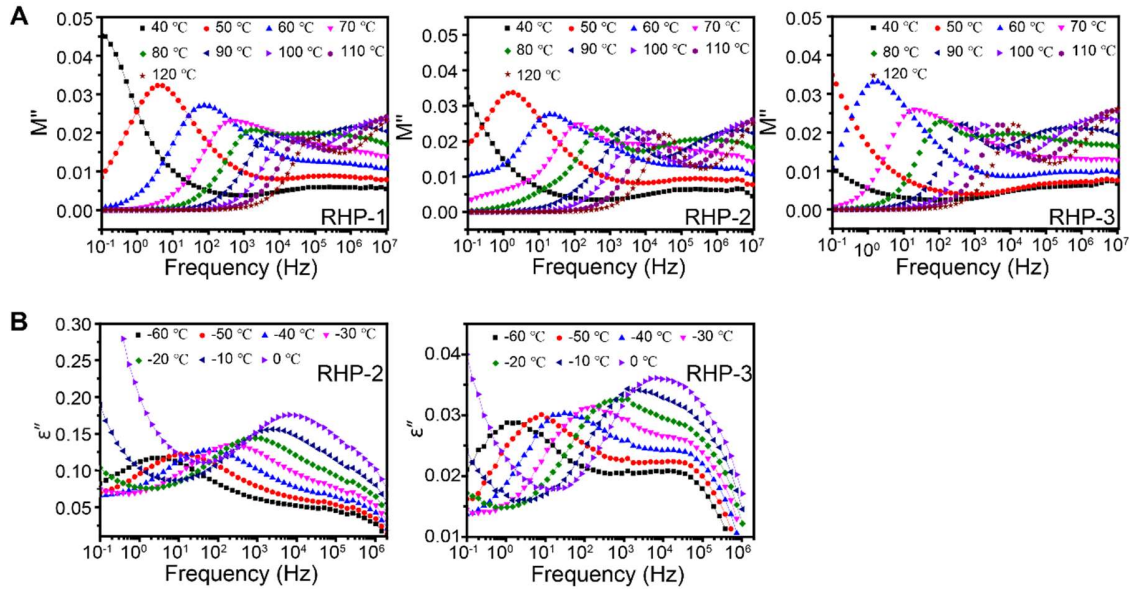
**Fig. S5.** DLS measurements of RHP. Size distribution of RHP-1 (A), RHP-2 (B) and RHP-3 (C) measured by DLS (details are shown in Table S3).



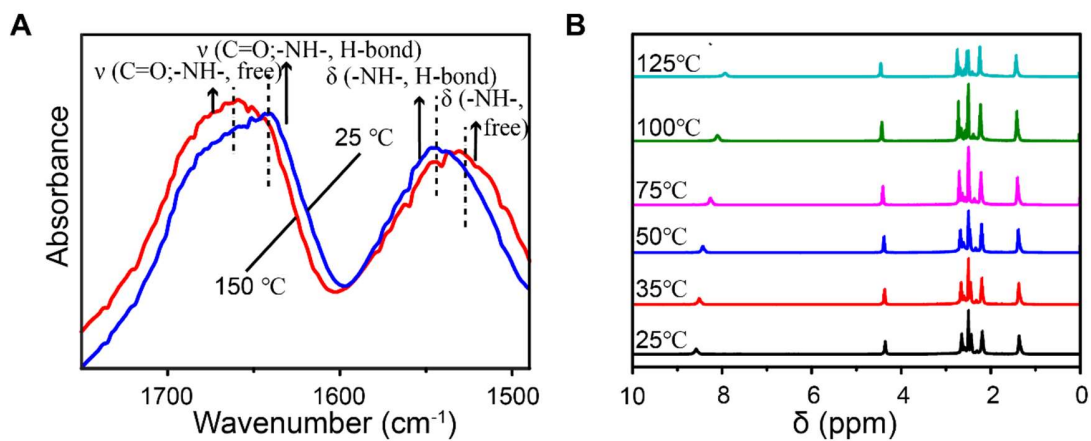
**Fig. S6.** Characterization of the degree of branching of RHP. (A) Structure units of the RHP. (B) Quantitative  $^{13}\text{C}$  NMR spectra of RHP and (C)  $^{13}\text{C}$ ,  $^1\text{H}$ -HSQC spectra of RHP-1. The  $^{13}\text{C}$ ,  $^1\text{H}$ -HSQC spectra reveal the assignment of the structure units, and the quantitative  $^{13}\text{C}$  NMR spectra are used to calculate the degree of branching of RHP (details are shown in Table S4).



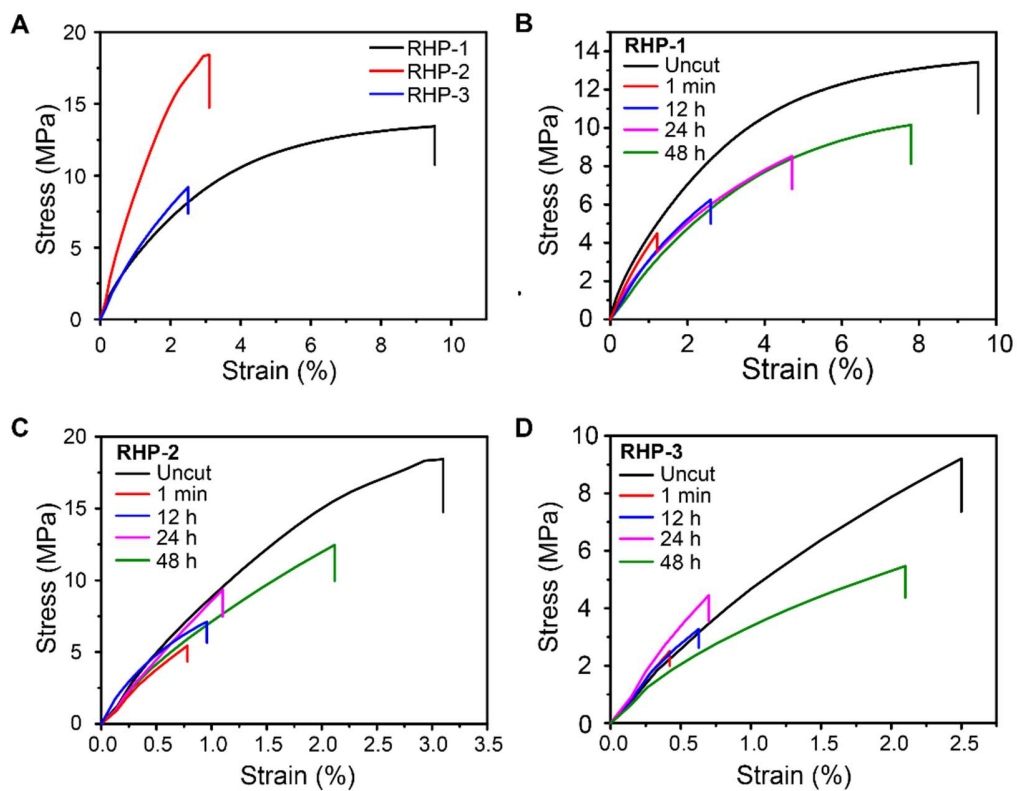
**Fig. S7.** Temperature-dependent XRD profile of RHP-1. The XRD profile of RHP-1 shows several weak sharp peaks in addition to the broad diffusion-like peak at room temperature, indicating the existence of crystals. These sharp peaks are stable up to 100 °C, while disappear at 150 °C which is above the melting point of RHP-1, suggesting the melting of the crystals between 100 °C and 150 °C.



**Fig. S8.** Dielectric measurements of RHP above and below  $T_g$ . (A) Dielectric modulus  $M''$  as a function of frequency for RHP-1, RHP-2 and RHP-3 from 40 °C to 120 °C, respectively. The average relaxation time  $\tau_{max}$  of  $\alpha$  process at 25 °C was calculated by VFT equation. The  $\tau_{max}$  values were in the range of  $10^5$  s (RHP-1) to  $10^9$  s (RHP-3) (details were show in Table S5). (B) Dielectric loss  $\epsilon''$  as a function of frequency for RHP-2 and RHP-3 from -60 °C to 0 °C, respectively.



**Fig. S9.** Temperature-dependent FTIR and <sup>1</sup>H NMR measurement of RHP. (A) Temperature-dependent FTIR spectra of the RHP-1 upon heating at 25 °C (blue) and 150 °C (red). (B) <sup>1</sup>H NMR spectral change of RHP-1 in deuterated dimethyl sulfoxide with an increase in temperature from 25 °C to 125 °C.



**Fig. S10.** Mechanical properties and self-healing tests of RHP. (A) The stress-strain curve of RHP. The representative stress-strain curves of the RHP-1 (B), RHP-2 (C) and RHP-3 (D) self-healed at 25 °C for different time.

**Table S1.** The feed ratio and yield of RHP.

RHP	MBA/BDA <sup>a</sup>	Yield (wt%) <sup>b</sup>
RHP-1	1:1.125	75.3%
RHP-2	1:1	84.7%
RHP-3	1:0.875	92.1%

<sup>a</sup> The molar ratio of MBA and BDA.

<sup>b</sup> Weight of the RHP obtained/weight of the monomer used.

**Table S2.** The mole percentage of primary amine, secondary amine and tertiary amine units.

RHP	MBA/BDA	primary amine (%)	secondary amine (%)	tertiary amine (%)
RHP-1	1:1.125	22.8	63.8	13.4
RHP-2	1:1	19.1	61.3	19.6
RHP-3	1:0.875	14.6	54.6	30.8



**Table S3.** The mean diameter,  $T_g$  and molecular weight of RHP.

RHP	Mean Diameter (nm) <sup>a</sup>	$T_g$ (°C) <sup>b</sup>	$M_n^c$ (g/mol)	$M_w^c$ (g/mol)	PDI <sup>c</sup>
RHP-1	35.6	37	8199	14547	1.77
RHP-2	42.5	42	8091	16173	2.00
RHP-3	71.6	49	9943	23319	2.35

<sup>a</sup> Determined by DLS (dissolved in methanol, 0.5 g/L).

<sup>b</sup> Determined by DSC at a heating rate of 10 °C/min.

<sup>c</sup> Determined by GPC (dissolved in DMF at room temperature).

**Table S4.** The DB values of RHP.

RHP	DB
RHP-1	0.16
RHP-2	0.24
RHP-3	0.33

**Table S5.** The VFT Parameters for Segmental Process.

RHP	$\tau_0$	B (K)	$T_0$ (K)	$\tau_{\max}$ (s) <sup>a</sup>
RHP-1	$4.76 \times 10^{-9}$	1011	265.8	$1.7 \times 10^5$
RHP-2	$4.76 \times 10^{-9}$	1098	266.2	$3.8 \times 10^6$
RHP-3	$4.76 \times 10^{-9}$	1213	268.9	$4.9 \times 10^9$

<sup>a</sup>The average relaxation time of segmental motion at 25 °C.

**Movie S1 (separate file).** The instantaneous self-healing behavior of RHP.

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