

Supplementary Information

Peptidomimetic Vinyl-Heterocyclic Inhibitors of Cruzain Effect Anti-Trypanosomal Activity

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Part 1. Molecular Modeling

In addition to compound **9**, we also utilized Schrodinger to develop both covalent and non-covalent models of compounds **7**, **11-13** and **15** bound to cruzain, also based on the crystal structure of **K11777** covalently bound to cruzain as described in the Experimental Section. The covalent and non-covalent models are in general similar in terms of the orientation of the peptide sidechains. The modeled structures differ, however, by virtue of the nature of how the molecular modeling was performed, leading to the distal positioning of the vinyl group in the non-covalent structures from the active-site residues Cys₂₅ and His₁₆₂. Compounds **12** and **15** which each contain an N-methyl-pyridine, indicate that the methyl group with its attending positive charge is directed to His₁₆₂, suggesting that the latter is neutral.

Table S1. Predicted affinity of covalent docking for select structures

Compound	Structure	Cdock Affinity (kcal/mol)	Predicted K_i (μM)
7	Cbz-Phe-Phe-vinyl-2Pyrmd	-7.11	6.11
9	NMePip-Phe-hPhe-vinyl-2Pyrmd	-7.01	7.23
11	Cbz-Phe-Phe-vinyl-2Pyr	-7.17	5.52
12	Cbz-Phe-Phe-vinyl-2PyrNMe	-8.00	1.36
13	Cbz-Phe-hPhe-vinyl-2Pyr	-7.63	2.54
15	Cbz-Phe-hPhe-vinyl-2PyrNMe	-8.32	0.79

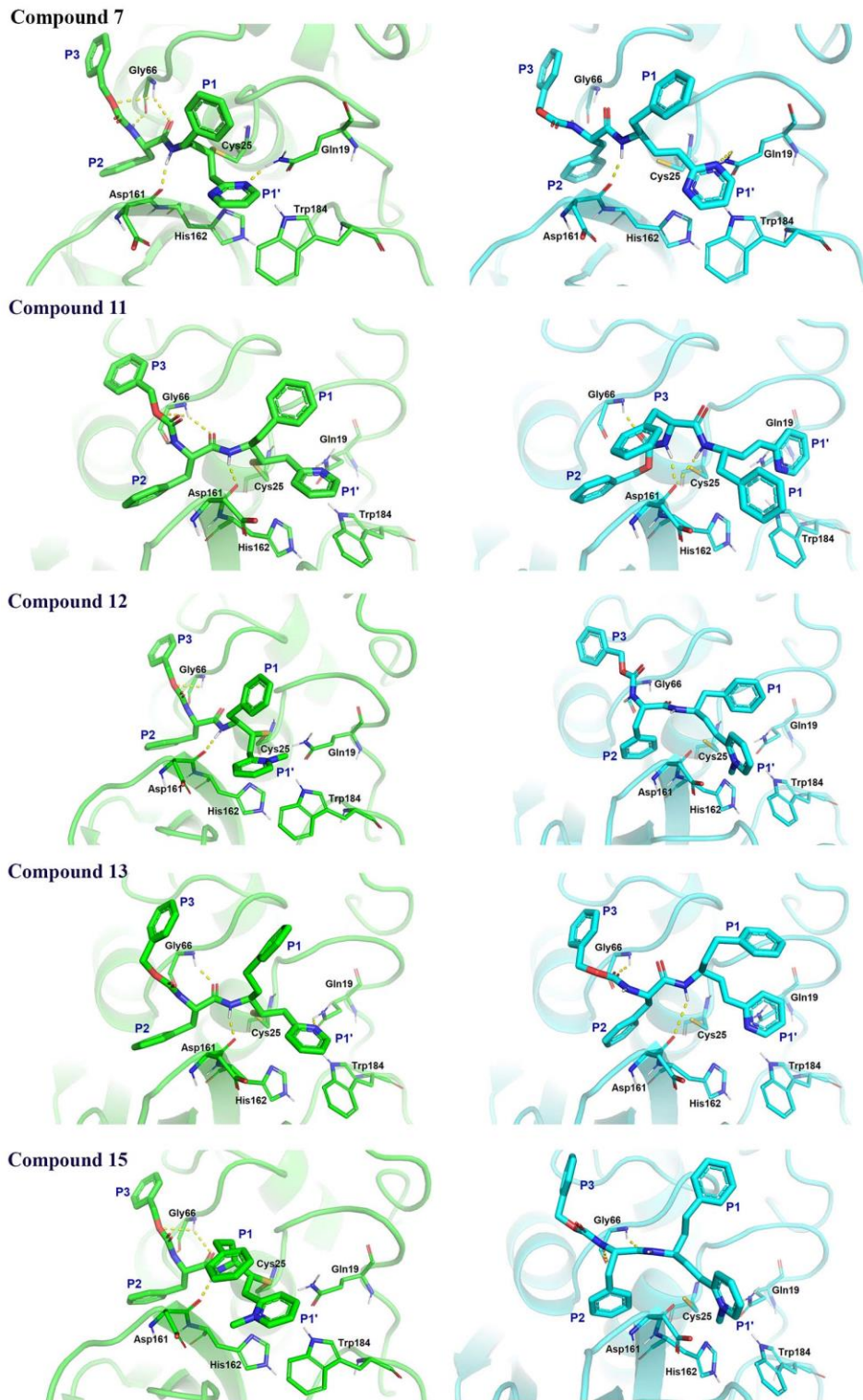
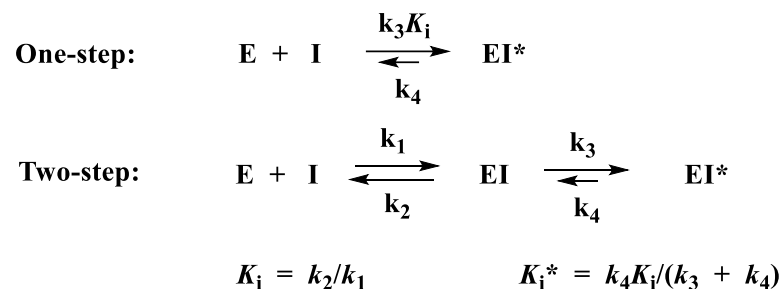


Figure S1. Molecular models of compound 7, 11-13, and 15 bound to cruzain created using Schrodinger. Left panels: binding poses in which a covalent bond is formed between the β -carbon of the vinyl group of the inhibitor. Right panels: binding poses in which no covalent bond is formed with Cys25.

Part 2. Fitting of Time-Course Inhibition Data using Kintek Explorer®

In addition to data fitting as described in the Experimental Section, time courses for PVH inhibitors found in **Table 4** and elsewhere were fitted using Kintek Explorer, using both a single-step and two-step binding model (**Scheme S1**).



Scheme S1

Fitted parameters from the one-step model: Sigma: 91.608; $k_3 K_i = 0.0246 \pm 0.000045 \mu\text{M}^{-1} \text{s}^{-1}$, $k_4 = 0.00175 \pm 0.00005 \text{s}^{-1}$, $k_4/k_3 K_i = 0.071 \pm 0.001 \mu\text{M}$. Fitted parameters from the two-step model: Sigma: 89.323; $k_1 = 0.02 \pm 0.02 \mu\text{M}^{-1} \text{s}^{-1}$, $k_2 = 0.07 \pm 0.099 \text{s}^{-1}$, $k_3 = 0.0058 \pm 0.001 \mu\text{M}^{-1} \text{s}^{-1}$, $k_4 = 0.0011 \pm 0.0008 \text{s}^{-1}$, $K_i = 0.40 \pm 0.02 \mu\text{M}$, and $K_i^* = 0.06 \mu\text{M}$. The lower value of Sigma for the two-step indicates a slightly better fit, albeit with more poorly determined individual rate constants. Note that when $k_4 \ll k_3$, $K_i^* \sim k_4 K_i / k_3$, as is reflected in the values calculated from both models above: $k_4/k_3 K_i \sim K_i^* = 0.071 \pm 0.001 \mu\text{M}$ and $K_i^* = 0.06 \mu\text{M}$. These values of K_i^* compared favorably with the value of $K_i^* = 0.126 \pm 0.004 \mu\text{M}$ determined by global fitting as reported in **Table 2**. Application of fitting by Kintek Explorer for potent cruzain inhibitors **11-13** provided respective values for the two models of: $k_4/k_3 K_i \sim K_i^* = 0.23 \pm 0.02 \mu\text{M}$ and $K_i^* = 0.6 \mu\text{M}$, $k_4/k_3 K_i \sim K_i^* = 0.474 \pm 0.002 \mu\text{M}$ and $K_i^* = 0.4 \mu\text{M}$, and $k_4/k_3 K_i \sim K_i^* = 0.233 \pm 0.008 \mu\text{M}$ and $K_i^* = 0.2 \mu\text{M}$, which again are comparable to their respective values of K_i^* obtained by global data fitting as reported in **Table 2**.

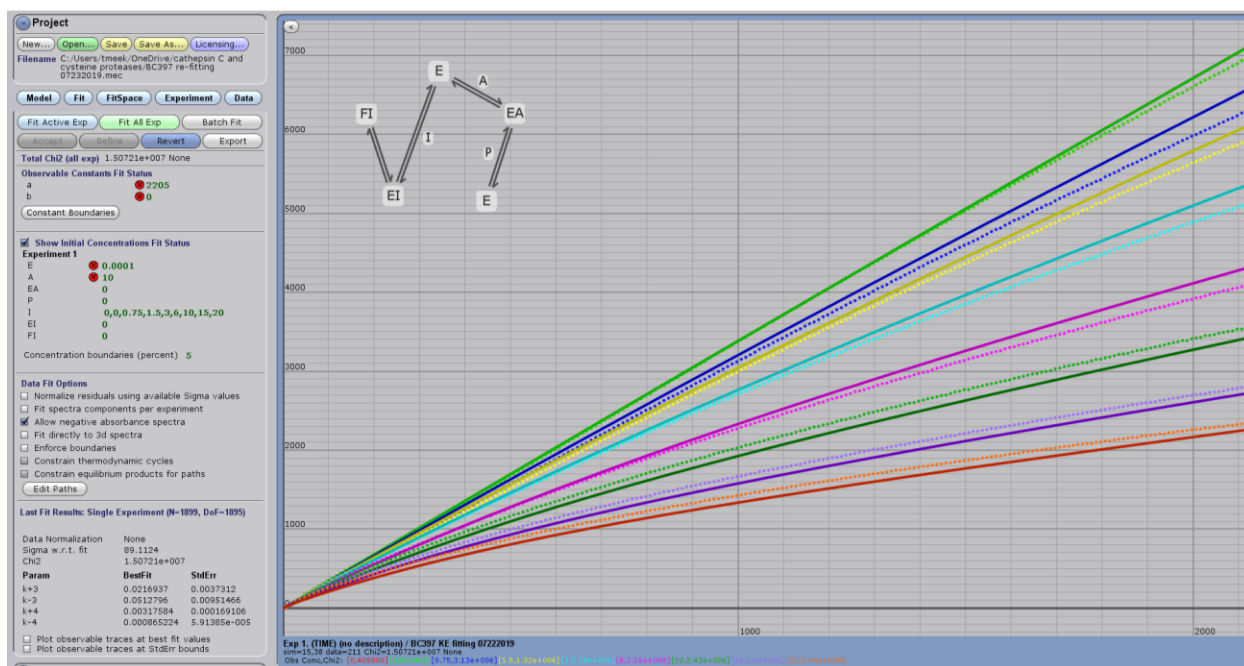
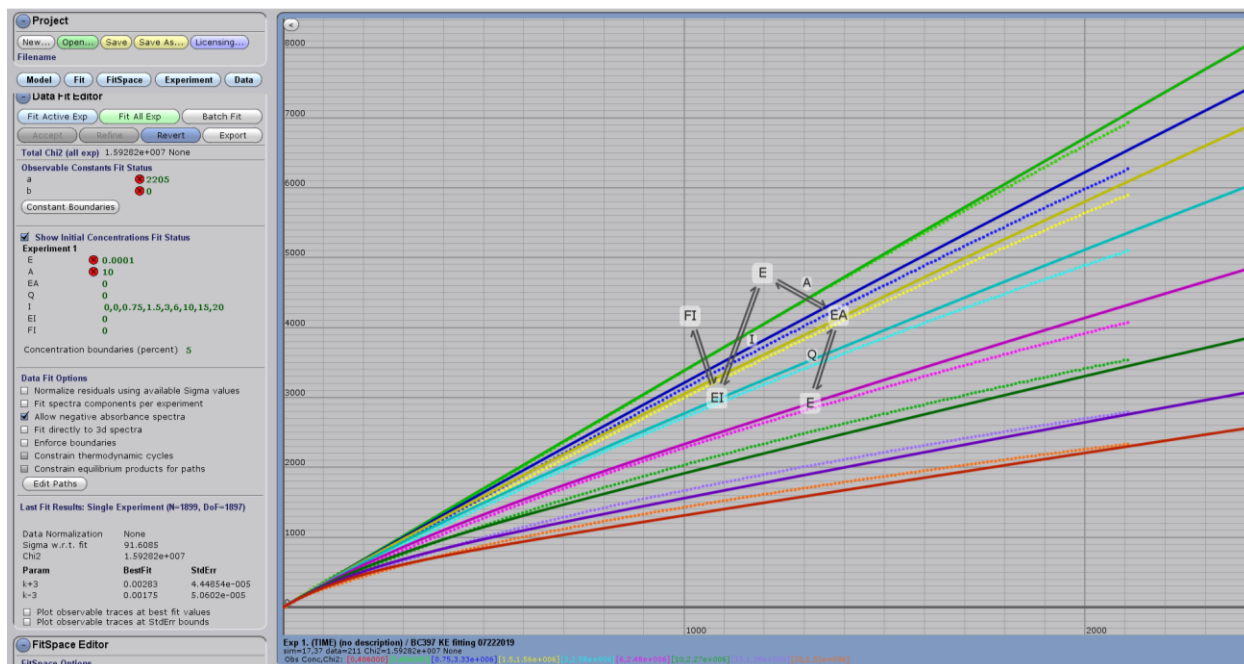


Figure S2. Time-dependent inhibition of cruzain by **15** depicting formation of product AMC vs. time for 0-2100 sec using the one-step model (top) and two-step model (bottom) at micromolar concentrations of **15** of 0 (green), 0.75 (blue), 1.5 (yellow), 3.0 (cyan), 6.0 (magenta), 10.0 (dark green), 15.0 (purple) and 20.0 (orange). Fitting of time course using Kintek Explorer with resulting rate constants obtained from fitting of the 2-step model of **Scheme S1**.

Part 3. Thiolation of Cruzain Inhibitors by Glutathione

Table S2. Kinetic Constants of Thiolation of Cruzain Inhibitors^a

Compound	Rate of Thiolation, k ($\text{mM}^{-1}\text{min}^{-1}$)	K_{eq} (M^{-1})
K11777	0.00028 ± 0.0004	NA
7	Negligible	NA
11	Negligible	NA
12	0.037 ± 0.002	7400
15	0.054 ± 0.004	2400
17^b	Negligible	NA
25	Negligible	NA
26	0.015 ± 0.005	930

^a 1 mM glutathione was mixed with 0.5 mM K11777 and compounds in Tris (pH 8.0), 10% DMSO (v/v) at room temperature. Aliquots were analyzed by LCMS as described; ^b 5mM glutathione was used for **17**; NA, not applicable.

Part 4. Synthesis and Characterization of AMC-Peptide Substrates

Boc-L-homophenylalanine-AMC: *tert-butyl (S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)carbamate*. A round bottom flask charged with Boc-L-homophenylalanine (1.5 g, 5.64 mmol) and 7-amino-4-methylcoumarin (627.2 mg, 3.58 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (50 ml) and tetrahydrofuran (15 mL) was added followed by the addition of N,N-diisopropylethylamine (915 μ L, 5.25 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (3.2 mL, 5.37 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 30 hours and was monitored by TLC (3:1 hexanes:ethyl acetate) or LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~60 mL) and washed with water (25 mL, 4x) and brine (25 mL, 2x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield an off white solid. The crude amide directly used for the next reaction without purification. LC-MS Rt: 5.59 min, *m/z* 437.3 ([M+H]⁺, C₂₅H₂₈N₂O₅⁺ Calcd 437.21

H₂N-L-homophenylalanine-AMC-TFA: *(S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)aminium-TFA*. A round bottom flask charged with Boc-L-homophenylalanine-AMC (60 mg, 0.137 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (6 mL) was added followed by the dropwise addition of trifluoroacetic acid (2 mL) at 0°C. The reaction was monitored via TLC (3:1 hexanes:ethyl acetate) and was complete following 1.5 hours. The solvents were removed under reduced pressure and co-evaporated with chloroform (15 mL, 4x) and ether (15 mL, 2x) to yield a tan solid. LC-MS analysis verified the formation of the desired amine. The product was directly used for the next reaction without purification.

(S2) *Cbz-L-phenylalanine-L-homophenylalanine-AMC*: *Benzyl ((S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate*. A round bottom flask charged with ³H₃N-L-homophenylalanine-TFA (60 mg, 0.138 mmol) and Cbz-phenylalanine-OH (49.5 mg, 0.166 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (4 mL) was added followed by the addition of N,N-diisopropylethylamine (120 μ L, 0.69 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (164 μ L, 0.276 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes or until no starting material remained via TLC (3:1 hexanes:ethyl acetate v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~20 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid. The crude was purified via preparative HPLC to yield the desired amide (50.1 mg, 59% yield) as a white powder. ¹H NMR (400 MHz, DMSO) δ 1.91 – 2.17 (m, 2H), 2.41 (d, *J* = 1.2 Hz, 3H), 2.56 – 2.89 (m, 3H), 3.07 (dd, *J* = 13.7, 4.1 Hz, 1H), 4.30 – 4.60 (m, 2H), 4.84 – 5.11 (m, 2H), 6.28

(d, $J = 1.5$ Hz, 1H), 7.10 – 7.39 (m, 14H), 7.45 – 7.60 (m, 2H), 7.68 – 7.85 (m, 2H), 8.40 (d, $J = 7.7$ Hz, 1H), 10.46 (s, 1H). LC-MS Rt: 5.88 min, m/z 618.4 ($[M+H]^+$), $C_{37}H_{35}N_3O_6^+$ Calcd 618.3.

(S3) Cbz-L-leucine-L-homophenylalanine-AMC: benzyl ((S)-4-methyl-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxopentan-2-yl)carbamate. A round bottom flask charged with ^+H_3N -L-homophenylalanine • TFA (64 mg, 0.142 mmol) and Cbz-Leucine-OH (49.1 mg, 0.185 mmol) was purged with N_2 gas yielding positive pressure. Anhydrous dichloromethane (4 mL) and tetrahydrofuran (4 mL) was added followed by the addition of N,N-diisopropylethylamine (124 μ L, 0.71 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (164 μ L, 0.276 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes and no starting material remained via TLC (3:1 hexanes:ethyl acetate v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~20 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na_2SO_4 and solvents removed under reduced pressure to yield a yellow solid. The crude amide was purified via preparative HPLC to yield (48.9 mg, 59%) as a white powder. 1H NMR (400 MHz, DMSO) δ 0.90 (t, $J = 6.9$ Hz, 6H), 1.50 (t, $J = 7.3$ Hz, 2H), 1.68 (t, $J = 7.0$ Hz, 1H), 2.02 (d, $J = 32.8$ Hz, 2H), 2.41 (s, 3H), 2.54 – 2.79 (m, 3H), 4.15 (q, $J = 7.8$ Hz, 1H), 4.47 (s, 1H), 5.06 (s, 2H), 6.27 (s, 1H), 7.09 – 7.41 (m, 9H), 7.49 (t, $J = 8.9$ Hz, 2H), 7.64 – 7.86 (m, 2H), 8.25 (d, $J = 7.7$ Hz, 1H), 10.42 (s, 1H). LC-MS Rt: 5.88 min, m/z 584.3 ($[M+H]^+$), $C_{34}H_{37}N_3O_6^+$ Calcd 584.3

Boc-L-4-pyridylalanine-AMC: tert-butyl (S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-3-(pyridin-4-yl)propan-2-yl)carbamate. A round bottom flask charged with Boc-L-4-pyridylalanine-OH (325 mg, 1.22 mmol) and 7-amino-4-methylcoumarin (164.5 mg, 0.939 mmol) was purged with N_2 gas yielding positive pressure. Anhydrous dichloromethane (6 mL) and tetrahydrofuran (14 mL) was added followed by the addition of N,N-diisopropylethylamine (600 μ L, 3.44 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (1.125 mL, 1.89 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 15 hours and product formation was monitored via TLC (1% MeOH:DCM v/v) and LC-MS analysis. Solvents were removed under reduced pressure and the yellow oil was solubilized in DCM (~30 mL) and washed with water (15 mL, 4x) and brine (15 mL, 2x). The organic layer was dried over Na_2SO_4 and solvents were removed under reduced pressure. The resulting yellow solid was deemed pure enough (~94% via LC-MS, 425 mg) to proceed directly to the next step. LC-MS Rt: 2.84 min, m/z 424.26 ($[M+H]^+$), $C_{23}H_{25}N_3O_5^+$ Calcd 424.2

NH_2 -L-4-pyridylalanine-AMC • TFA: (S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-3-(pyridin-4-yl)propan-2-aminium • TFA. A round bottom flask charged with Boc-L-4-pyridylalanine-AMC (150 mg, 0.353 mmol) was purged with N_2 gas yielding positive pressure. Anhydrous dichloromethane (10 mL) was added followed by the dropwise addition of trifluoroacetic acid (3 mL) at 0°C. The reaction was monitored

via TLC (1% MeOH:DCM v/v) and was complete following 1.5 hours. The solvents were removed under reduced pressure and co-evaporated with chloroform and ether to yield a yellow solid. LCMS analysis verified the formation of the product. The product was directly used for the next reaction without purification.

(S5) Cbz-L-phenylalanine-L-4-pyridylalanine-AMC: Benzyl ((S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-3-(pyridin-4-yl)propan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate. A round bottom flask charged with ^+H_3N -L-4-pyridylalanine • TFA (154 mg, 0.352 mmol) and Cbz-phenylalanine-OH (137.3 mg, 0.459 mmol) was purged with N_2 gas yielding positive pressure. Anhydrous dichloromethane (10 mL) and tetrahydrofuran (2 mL) was added followed by the addition of N,N-diisopropylethylamine (367.9 μ L, 2.11 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (419.1 μ L, 0.704 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes or until no starting material remained via TLC (1% MeOH:DCM v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~30 mL) and washed with water (15 mL, 4x) and brine (15 mL, 1x). The organic layer was dried over Na_2SO_4 and solvents removed under reduced pressure to yield a yellow solid. 100 mg of the crude was purified via preparative HPLC to yield (32.94 mg, 39%) as a white powder. 1H NMR (400 MHz, DMSO) δ 2.33 – 2.45 (m, 3H), 2.72 (dd, J = 13.9, 10.5 Hz, 1H), 2.85 – 3.09 (m, 2H), 3.14 (dd, J = 13.9, 5.4 Hz, 1H), 4.14 – 4.37 (m, 1H), 4.68 – 4.89 (m, 1H), 4.96 (s, 2H), 6.29 (d, J = 1.5 Hz, 1H), 7.25 (tt, J = 22.3, 14.7, 13.7, 6.4 Hz, 12H), 7.39 – 7.54 (m, 2H), 7.75 (d, J = 8.7 Hz, 2H), 8.25 – 8.56 (m, 3H), 10.52 (s, 1H). LC-MS Rt: 3.57 min, m/z 605.3 ($[M+H]^+$), $C_{35}H_{32}N_4O_6^+$ Calcd 605.23.

N-methyl-piperazine-L-phenylalanine-OH: (4-methylpiperazine-1-carbonyl)-L-phenylalanine. A round bottom flask charged with methyl (4-methylpiperazine-1-carbonyl)-L-phenylalanine (500 mg, 1.64 mmol) was purged with N_2 gas yielding positive pressure. Anhydrous methanol (8.5 mL) and tetrahydrofuran (8.5 mL) was added followed by the addition of 0.5 M LiOH in H_2O (17 mL, 8.5 mmol). The reaction was monitored by TLC (5% MeOH:DCM v/v) and after 3.5 hours the reaction was stopped and solvents were removed under reduced pressure. The reaction mixture was dissolved in acidified water (0.5 M HCl, 10 mL) to neutralize the unreacted LiOH and the solvent was removed under reduced pressure. The crude compound was then transferred to the next reaction step without purification.

(S7) N-methyl-piperazine-L-phenylalanine-L-homophenyl-alanine-AMC: 4-methyl-N-((S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)piperazine-1-carboxamide. A round bottom flask charged with ^+H_3N -L-homophenylalanine • TFA (105 mg, 0.313 mmol) and N-methyl-piperazine-phenylalanine-OH (160 mg, 0.55 mmol) was purged with N_2 gas yielding positive pressure. Anhydrous dichloromethane (10 mL) was added followed by the addition of

N,N-diisopropylethylamine (274 μ L, 1.57 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (377 μ L, 0.626 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes and no starting material remained via TLC (1:1 ethyl acetate: hexanes v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~20 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid. The crude amide was purified via preparative HPLC to yield (21.1 mg, 11%) of a white powder. ¹H NMR (400 MHz, CDCl₃) δ 1.94 – 2.16 (m, 6H), 2.31 (s, 4H), 2.32 – 2.41 (m, 5H), 2.42 (d, *J* = 1.2 Hz, 4H), 2.73 (t, *J* = 7.4 Hz, 2H), 2.85 – 3.04 (m, 2H), 3.25 (ddd, *J* = 12.7, 10.6, 4.9 Hz, 3H), 3.41 (ddt, *J* = 12.5, 5.6, 2.9 Hz, 3H), 4.36 (dt, *J* = 8.7, 5.3 Hz, 1H), 4.60 (td, *J* = 8.5, 4.5 Hz, 1H), 4.78 (d, *J* = 4.4 Hz, 1H), 6.20 (d, *J* = 1.4 Hz, 1H), 6.60 (d, *J* = 8.2 Hz, 1H), 7.12 – 7.40 (m, 10H), 7.52 (d, *J* = 8.5 Hz, 1H), 7.68 – 7.82 (m, 2H), 9.16 (s, 1H). LC-MS Rt: 3.29 min, *m/z* 610.1 ([M+H]⁺, C₃₅H₃₉N₅O₅⁺ Calcd 610.3).

Boc-L-4-pyridylalanine-L-homophenylalanine-AMC: *tert-butyl ((S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxo-3-(pyridin-4-yl)propan-2-yl)carbamate*. A round bottom flask charged with ³H₃N-L-homophenylalanine • TFA (146 mg, 0.325 mmol) and Boc-L-4-pyridylalanine-OH (104 mg, 0.39 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (10 mL) was added followed by the addition of N,N-diisopropylethylamine (283 μ L, 1.63 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (387 μ L, 0.65 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes and no starting material remained via TLC (1:1 hexanes:ethyl acetate v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~20 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid. The crude amide was moved directly to the next step with no purification. LC-MS Rt: 3.57 min, *m/z* 581.7 ([M+H]⁺, C₃₃H₃₆N₄O₆⁺ Calcd 581.3).

NH₂-L-4-pyridylalanine-L-homophenylalanine-AMC•TFA: *S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxo-3-(pyridin-4-yl)propan-2-aminium TFA*. A round bottom flask charged with Boc-L-4-pyridylalanine-L-homophenylalanine-AMC (172 mg, 0.294 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (10 mL) was added followed by the dropwise addition of trifluoroacetic acid (5 mL) at 0°C. The reaction was monitored via TLC (1:1 hexanes:ethyl acetate) and was complete following 1.5 hours. The solvents were removed under reduced pressure and co-evaporated with chloroform (15 mL, 4x) and ether (15 mL, 2x) to yield a tan solid. LC-MS analysis verified the formation of the desired amine. The product was directly used for the next reaction without purification.

(S8) *Cbz-L-4-pyridylalanine-L-homophenylalanine-AMC*: *Benzyl ((S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxo-3-(pyridin-4-yl)propan-2-yl)carbamate*. A round bottom flask charged with *NH₂-L-4-pyridylalanine-L-homophenylalanine-AMC-TFA* (227 mg, 0.379 mmol; excess weight is due to residual TFA salts) was purged with N₂ gas yielding positive pressure. Anhydrous ethanol (4 mL) was added followed by the addition of a solution of sodium bicarbonate (127 mg, 1.52 mmol) in water (4 mL). Subsequently, a solution of benzyl chloroformate (107 μ L, 0.76 mmol) in 1,4 dioxanes (4 mL) was added dropwise and the reaction was allowed to react overnight. Solvents were removed under reduced pressure and the reaction mixture was solubilized in dichloromethane (30 mL) and washed with H₂O (10 mL, 3x), brine (10 mL, 2x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield an orange oil. The crude amide was purified via preparative HPLC to yield (45.9 mg, 20%) as a white powder. ¹H NMR (400 MHz, MeOD) δ 1.88 – 2.27 (m, 2H), 2.44 (d, *J* = 1.3 Hz, 3H), 2.68 (dddd, *J* = 20.0, 15.3, 12.7, 7.8, 3.3 Hz, 2H), 2.96 (dd, *J* = 14.1, 9.2 Hz, 1H), 3.22 (dd, *J* = 13.9, 5.3 Hz, 1H), 4.50 (dd, *J* = 8.8, 5.3 Hz, 1H), 4.57 (dd, *J* = 9.1, 5.3 Hz, 1H), 5.05 (d, *J* = 3.5 Hz, 2H), 6.21 (d, *J* = 1.5 Hz, 1H), 7.01 – 7.37 (m, 12H), 7.47 (dd, *J* = 8.7, 2.1 Hz, 1H), 7.55 – 7.72 (m, 1H), 7.78 (d, *J* = 2.0 Hz, 1H), 8.33 (d, *J* = 5.1 Hz, 2H). LC-MS Rt: 3.79 min, *m/z* 617.5 ([M+H]⁺), C₃₆H₃₄N₄O₆⁻ Calcd 617.3.

Boc-L-phenylalanine-AMC: *tert-butyl (S)-1-(((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate*. A round bottom flask charged with *Boc-L-phenylalanine* (380 mg, 1.43 mmol) and 7-amino-4-methylcoumarin (193 mg, 1.1 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous tetrahydrofuran (30 mL) was added followed by the addition of N,N-diisopropylethylamine (958 μ L, 5.5 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (1.31 mL, 2.2 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 15 hours was monitored by TLC (3:1 hexanes:ethyl acetate) or LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~80 mL) and washed with water (30 mL, 4x) and brine (30 mL, 2x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid. The crude amide was purified via preparative HPLC (363 mg, 78%) LC-MS Rt: 5.31 min, *m/z* 423.3 ([M+H]⁺), C₂₄H₂₆N₂O₅⁺ Calcd 423.2.

H₂N-L-phenylalanine-AMC•TFA: *(S)-1-(((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-3-phenylpropan-2-aminium)•TFA*. A round bottom flask charged with *Boc-L-phenylalanine-AMC* (40 mg, 0.095 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (3 mL) was added followed by the dropwise addition of trifluoroacetic acid (1 mL) at 0°C. The reaction was monitored via TLC and was complete following 1.5 hours. The solvents were removed under reduced pressure and co-evaporated with chloroform (15 mL, 4x) and ether (15 mL, 2x) to yield a tan solid. LCMS analysis

verified the formation of the desired amine. The product was directly used for the next reaction without purification.

(S9) Cbz-L-phenylalanine-L-phenylalanine-AMC: benzyl ((S)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-3-phenylpropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate. A round bottom flask charged with ⁺H₃N-L-phenylalanine •TFA (40 mg, 0.092 mmol) and Cbz-phenylalanine-OH (33 mg, 0.11 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (5 mL) and was added followed by the addition of N,N-diisopropylethylamine (80 μL, 0.46 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (110 μL, 0.18 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes and no starting material remained via TLC (3:1 hexanes:ethyl acetate v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in dichloromethane (~20 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid. The crude amide was purified via preparative HPLC to yield (34.2 mg, 62%) as a white powder. ¹H NMR (400 MHz, DMSO) δ 2.41 (s, 3H), 2.72 (dd, *J* = 13.9, 10.5 Hz, 1H), 2.87 – 3.05 (m, 2H), 3.11 (dd, *J* = 13.8, 5.8 Hz, 1H), 4.30 (td, *J* = 9.6, 9.0, 4.2 Hz, 1H), 4.73 (q, *J* = 7.5 Hz, 1H), 4.96 (s, 2H), 6.28 (s, 1H), 7.06 – 7.39 (m, 15H), 7.39 – 7.53 (m, 2H), 7.60 – 7.79 (m, 2H), 8.37 (d, *J* = 7.8 Hz, 1H), 10.47 (s, 1H). LC-MS Rt: 5.68 min, *m/z* 603.4 ([M+H]⁺, C₃₆H₃₃N₃O₆⁺ Calcd 604.2.

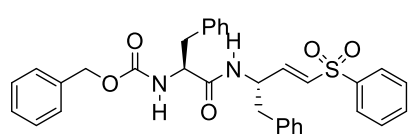
Cbz-L-arginine(Mtr)-L-homophenylalanine-AMC: Benzyl ((S)-5-(3-((4-methoxy-2,6-dimethylphenyl)sulfonyl)guanidino)-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxopentan-2-yl)carbamate. A round bottom flask charged with ⁺H₃N-L-homophenylalanine • TFA (120 mg, 0.266 mmol) and Cbz-Arginine (Mtr)-OH (180 mg, 0.346 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (10 mL) and was added followed by the addition of N,N-diisopropylethylamine (232 μL, 1.33 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (317 μL, 0.532 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 3 hours and no starting material remained via TLC (3:1 hexanes:ethyl acetate v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in DCM (~40 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid that was moved directly to the next step. LC-MS Rt: 5.73 min, *m/z* 837.9 ([M+H]⁺, C₄₃H₄₈N₆O₉S⁺ Calcd 837.3.

(S10) Cbz-L-arginine-L-homophenylalanine-AMC: benzyl ((S)-5-guanidino-1-(((S)-1-((4-methyl-2-oxo-2H-chromen-7-yl)amino)-1-oxo-4-phenylbutan-2-yl)amino)-1-oxopentan-2-yl)carbamate. A round bottom flask charged with Cbz-Arginine (Mtr)-L-homophenylalanine-AMC (130 mg, 0.158 mmol) and phenol (450 mg, 4.8 mmol, 5% wt/v) was purged with N₂ gas yielding positive pressure. Trifluoroacetic acid (6

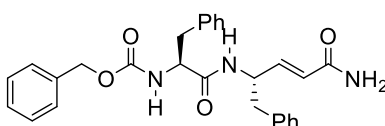
mL) and was added followed by allowing the reaction to proceed for 12 hours. The solvent was removed under a flow of N₂ gas and the crude product was subsequently purified by preparative HPLC to yield (34 mg, 34% yield) as a white powder. ¹H NMR (400 MHz, MeOD) δ 1.58 – 2.00 (m, 4H), 2.00 – 2.33 (m, 2H), 2.34 – 2.53 (m, 3H), 2.52 – 2.84 (m, 2H), 3.20 (q, *J* = 6.3, 5.8 Hz, 2H), 4.23 (t, *J* = 6.5 Hz, 1H), 4.50 (dd, *J* = 9.2, 4.9 Hz, 1H), 5.13 (s, 2H), 6.23 (t, *J* = 5.5 Hz, 1H), 6.90 – 7.41 (m, 10H), 7.47 (d, *J* = 8.7 Hz, 1H), 7.66 (dt, *J* = 18.0, 8.3 Hz, 1H), 7.84 (d, *J* = 11.3 Hz, 1H), 8.53 (s, 1H). LC-MS Rt: 3.62 min, *m/z* 625.5 ([M+H]⁺, C₃₄H₃₈N₆O₆⁻ Calcd 625.3).

(S11) *NMe-Pip-Phe-Phe-AMC*: 4-methyl-*N*-((*S*)-1-(((*S*)-1-((4-methyl-2-oxo-2*H*-chromen-7-yl)amino)-1-oxo-3-phenylpropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)piperazine-1-carboxamide. A round bottom flask charged with ³H₃N-L-phenylalanine •TFA (100 mg, 0.29 mmol) and *N*-methyl-piperazine-phenylalanine-OH (160 mg, 0.55 mmol) was purged with N₂ gas yielding positive pressure. Anhydrous dichloromethane (15 mL) and was added followed by the addition of *N,N*-diisopropylethylamine (253 μL, 1.45 mmol). Propylphosphonic anhydride in a 50% wt. solution of ethyl acetate (345 μL, 0.58 mmol) was added dropwise at 0°C resulting in a yellow color. The reaction was allowed to react for 45 minutes and no starting material remained via TLC (1:1 hexanes:ethyl acetate v/v) and LC-MS analysis. Solvents were removed and the yellow oil was solubilized in dichloromethane (~40 mL) and washed with water (10 mL, 4x) and brine (10 mL, 1x). The organic layer was dried over Na₂SO₄ and solvents removed under reduced pressure to yield a yellow solid. The crude amide was purified via preparative HPLC to yield (24.6 mg, 14%) ¹H NMR (400 MHz, CDCl₃) δ 2.38 (d, *J* = 1.2 Hz, 3H), 2.54 (s, 3H), 2.55 – 2.78 (m, 4H), 2.81 – 3.04 (m, 1H), 3.02 – 3.28 (m, 3H), 3.32 – 3.60 (m, 4H), 4.47 (dt, *J* = 8.7, 5.5 Hz, 1H), 4.87 (q, *J* = 6.9 Hz, 1H), 5.65 (d, *J* = 5.7 Hz, 1H), 6.16 (d, *J* = 1.4 Hz, 1H), 6.83 – 7.40 (m, 15H), 7.46 (d, *J* = 8.7 Hz, 1H), 7.54 – 7.72 (m, 2H), 8.30 (s, 1H), 9.45 (s, 1H). LC-MS Rt: 3.15 min, *m/z* 596.2 ([M+H]⁺, C₃₄H₃₇N₅O₅⁺ Calcd 596.3).

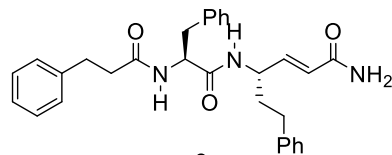
Part 5. Structures of PVH Inhibitors 1-27



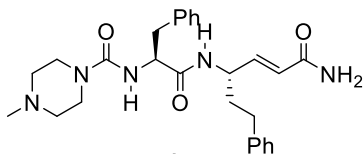
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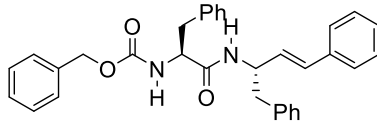
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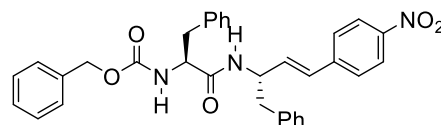
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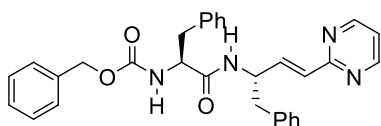
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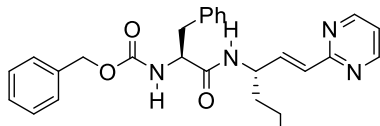
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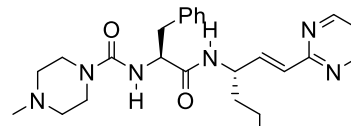
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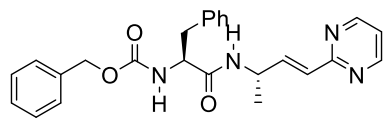
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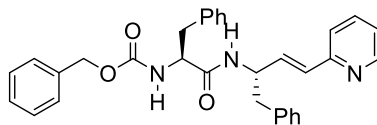
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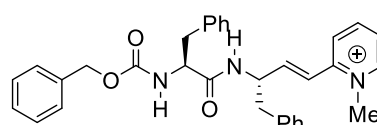
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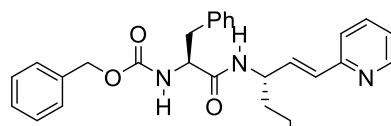
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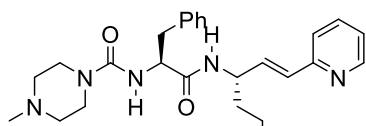
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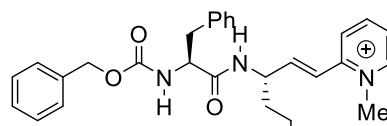
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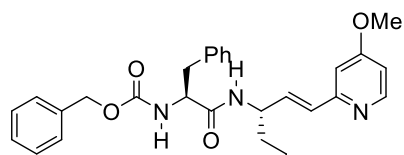
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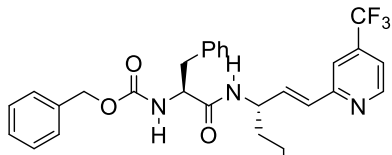
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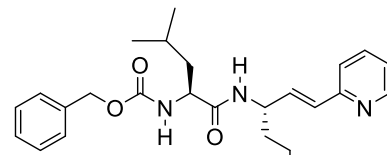
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16



17



18

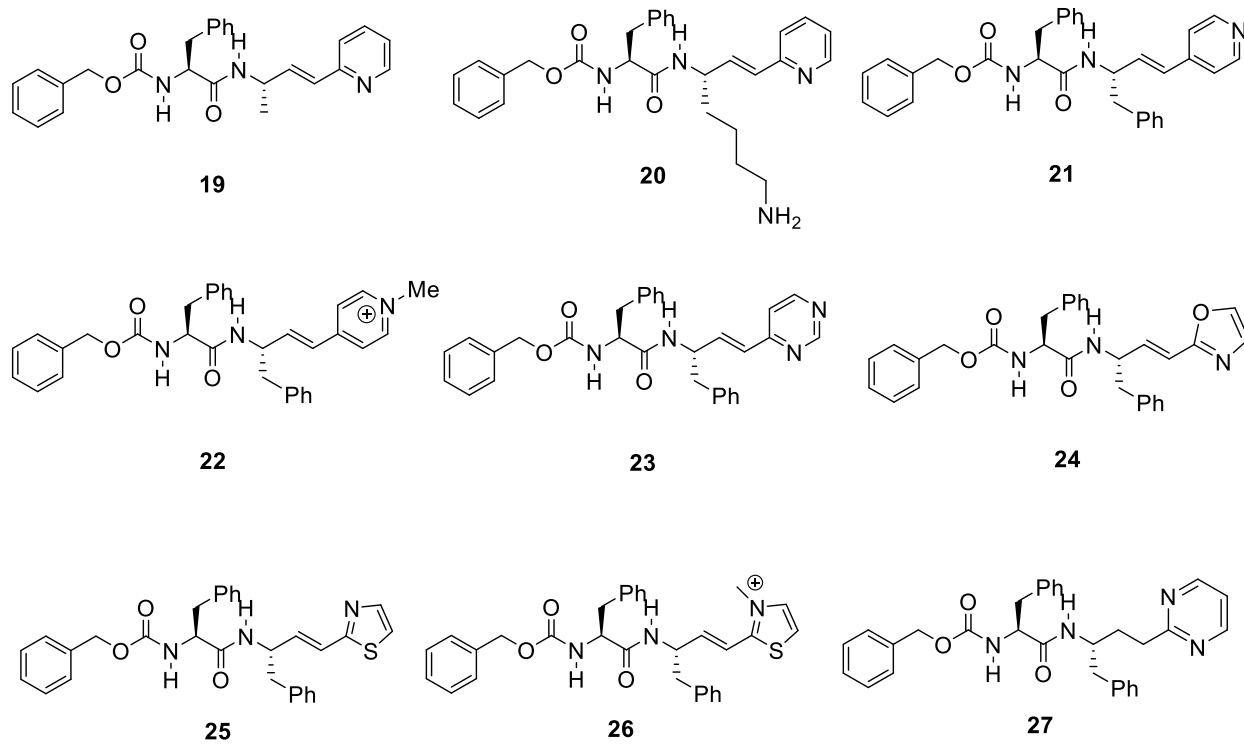
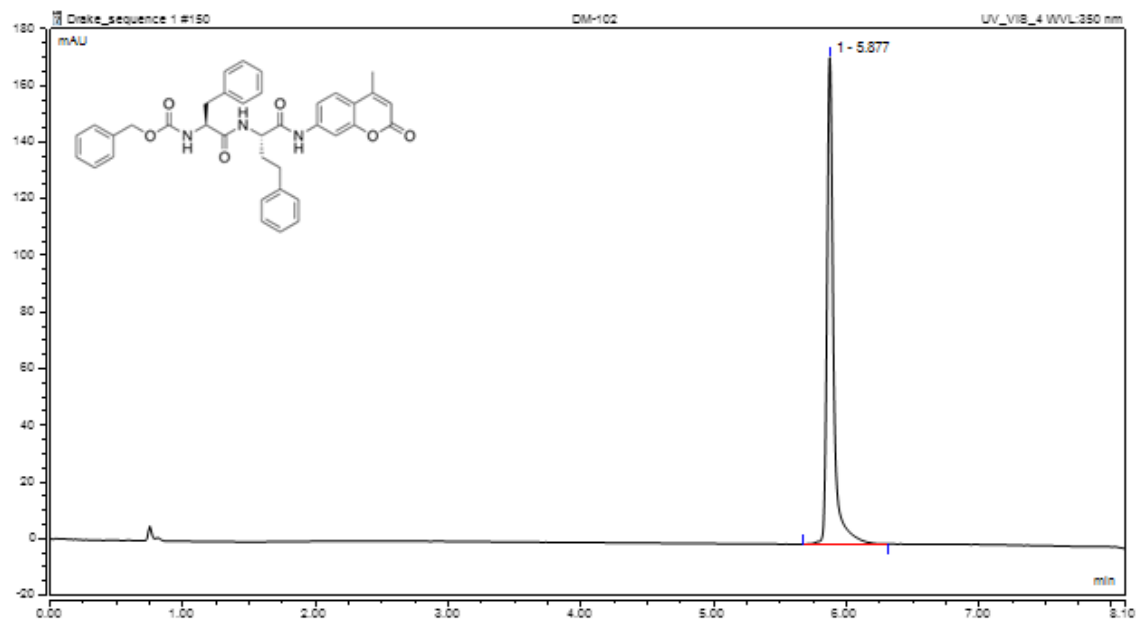


Figure S3. Structures of Inhibitors from Table 2 and Experimental Section.

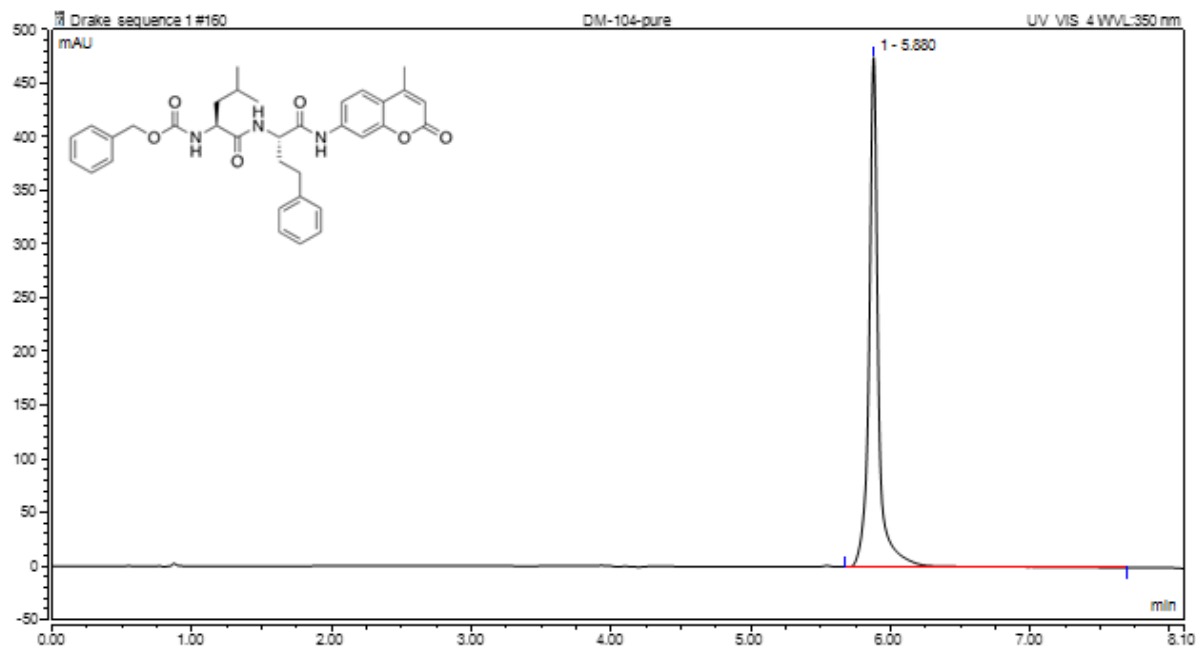
Part 6. HPLC Traces of Synthesized Substrates and Inhibitors

All cruzain substrates and inhibitors have been analyzed by LCMS, and have determined as >95% analytical purity as demonstrated in the chromatograms below.

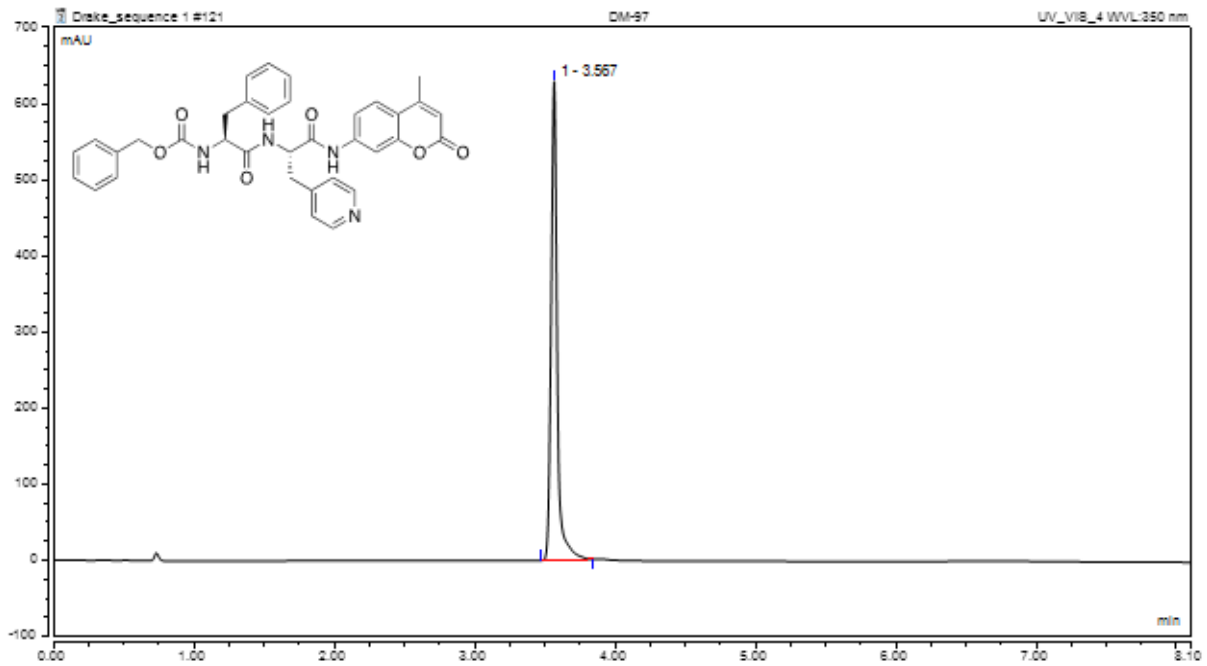
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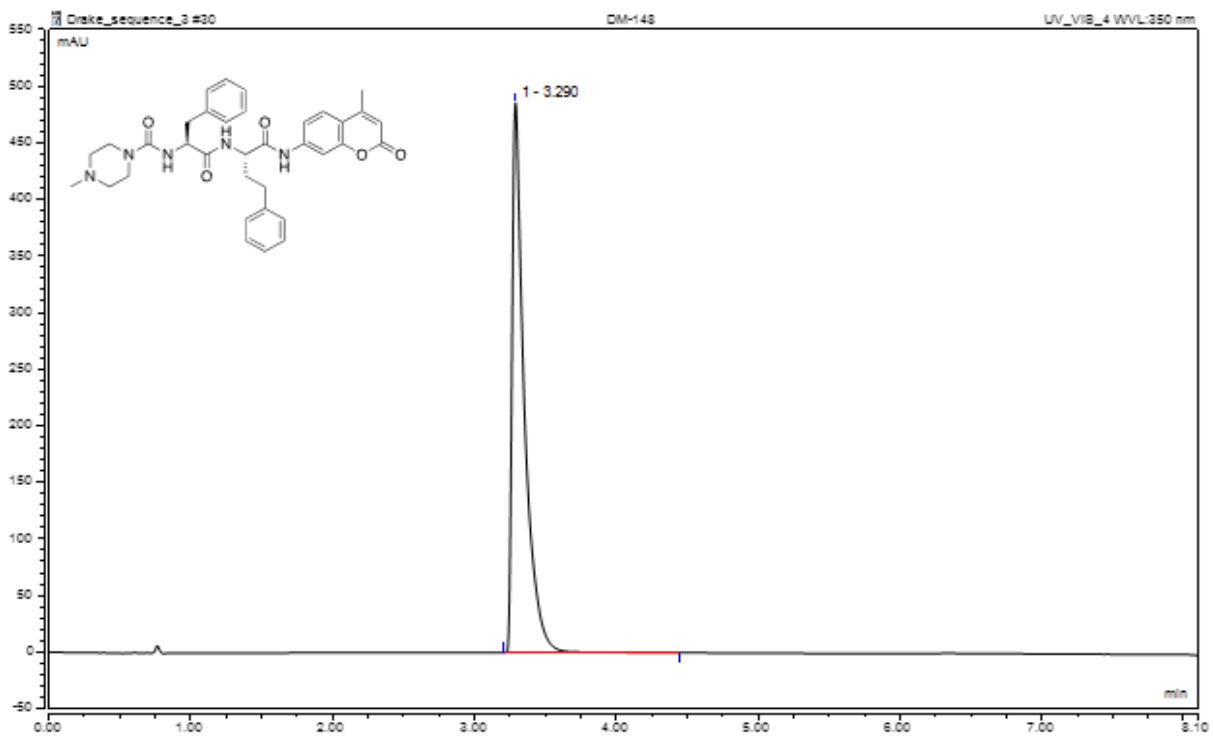
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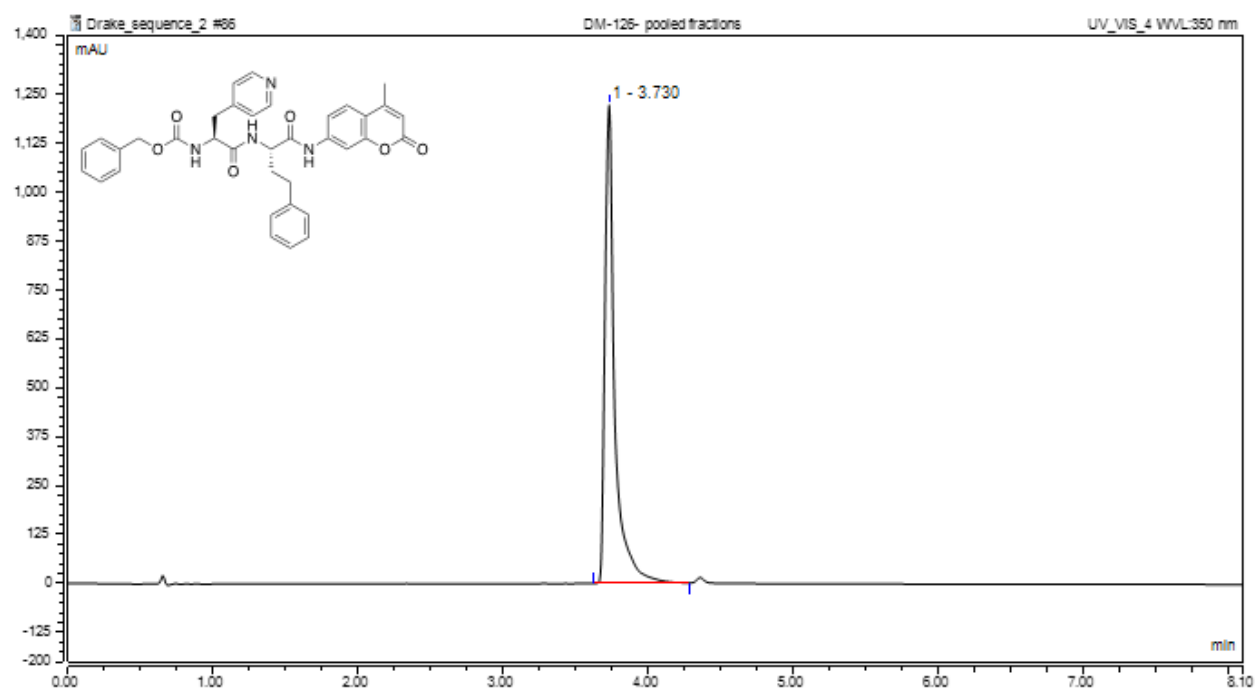
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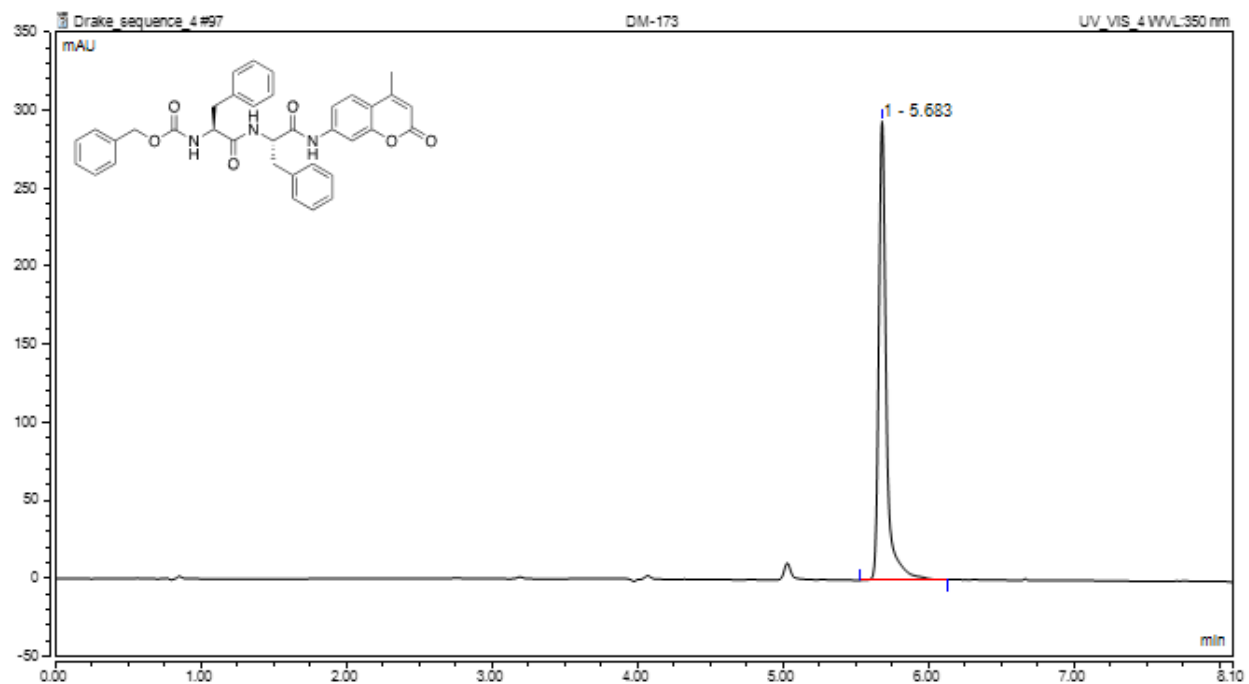
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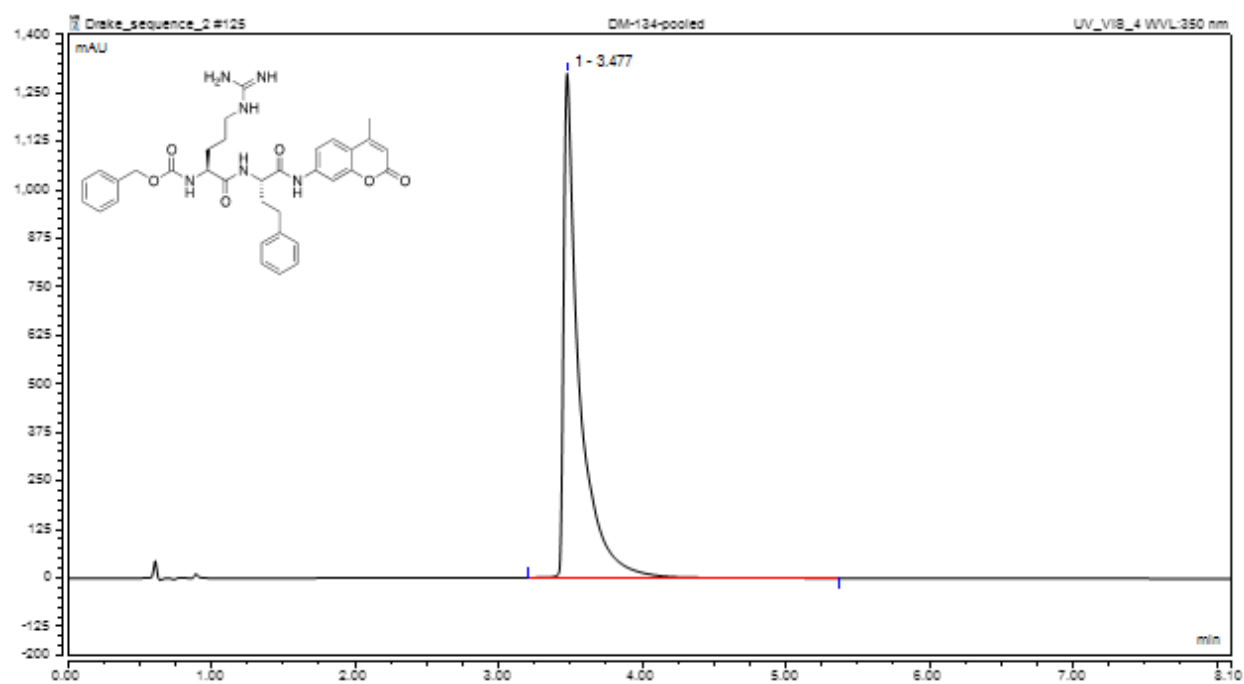
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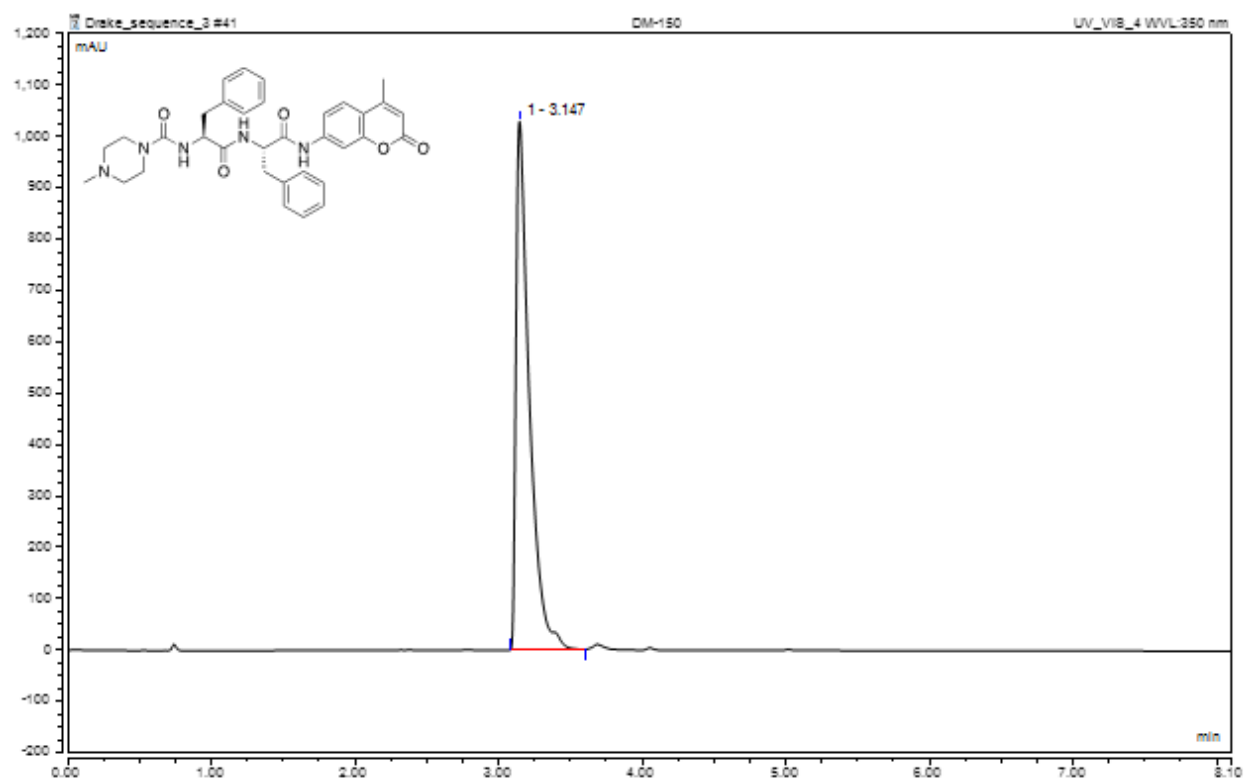
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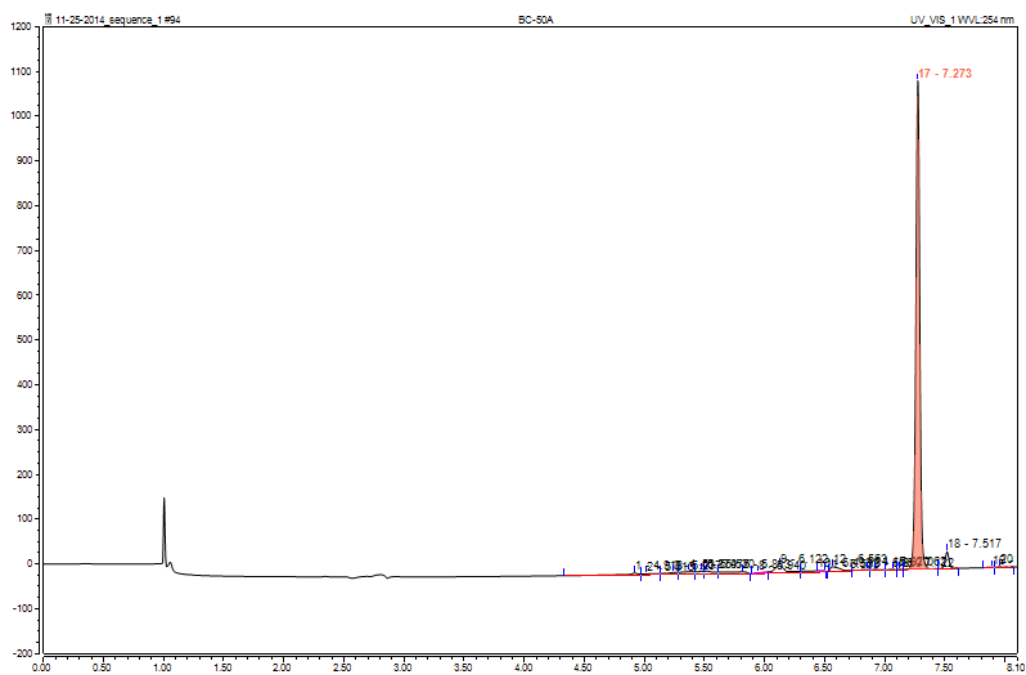
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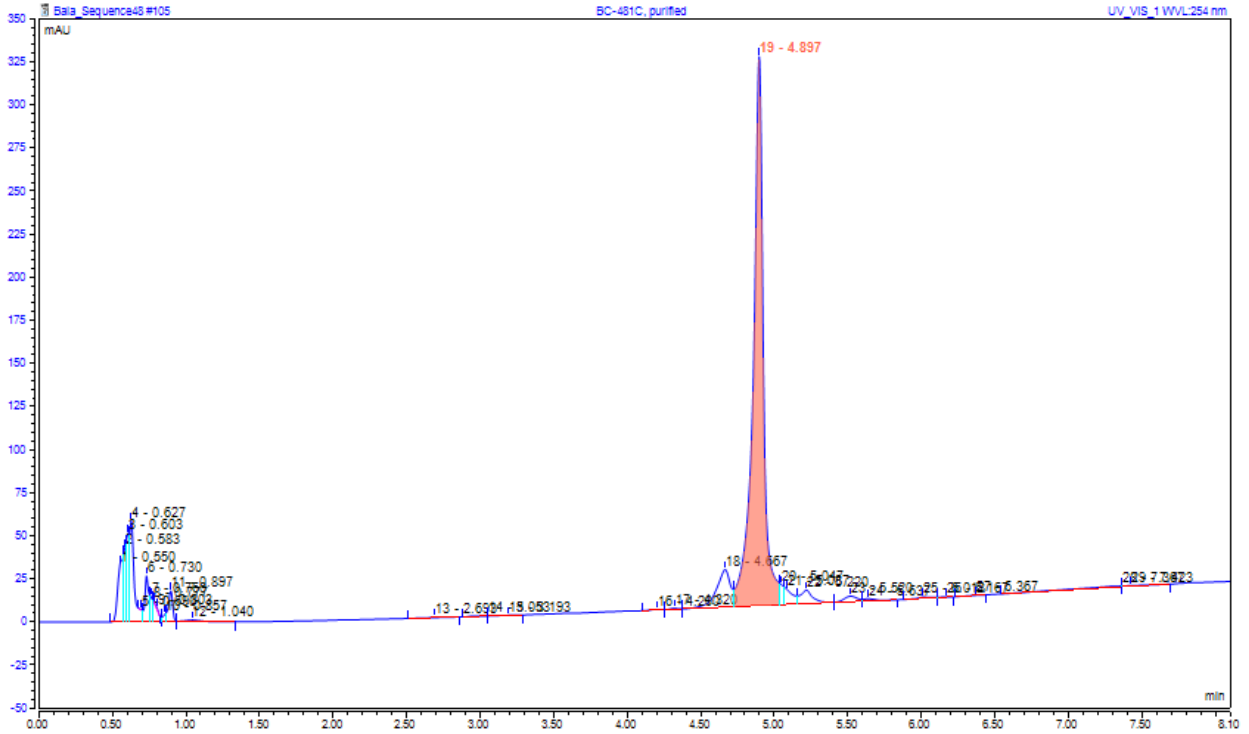
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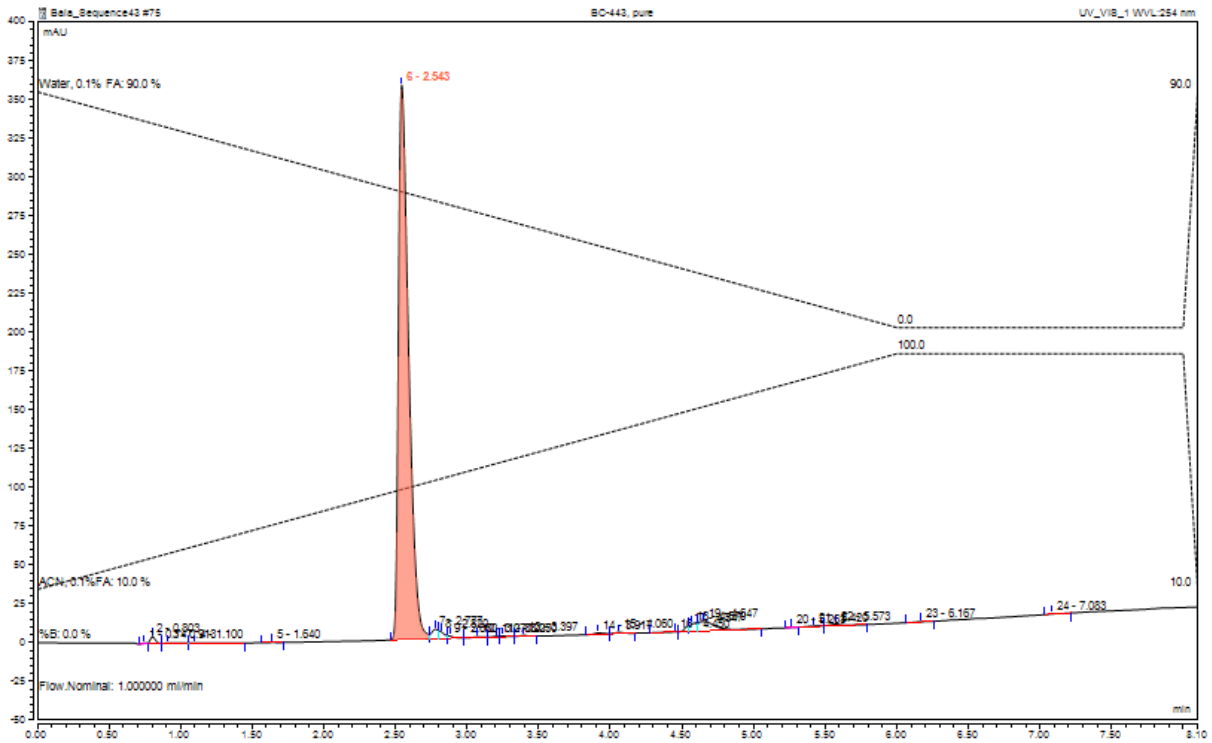
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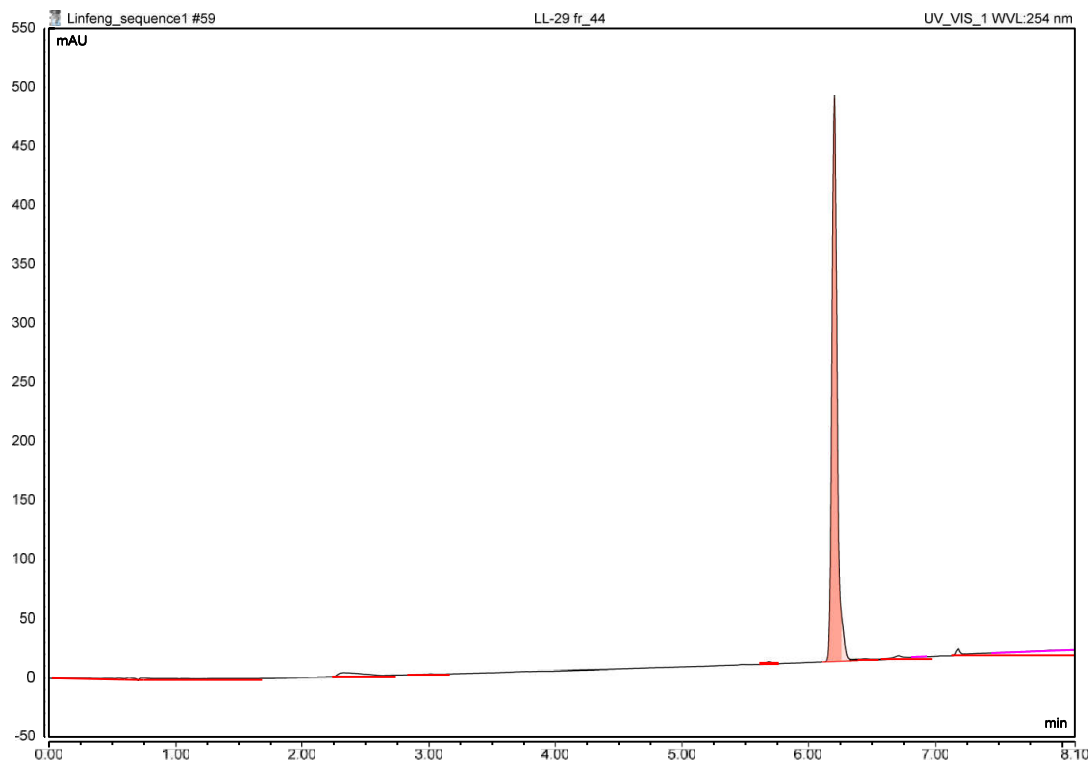
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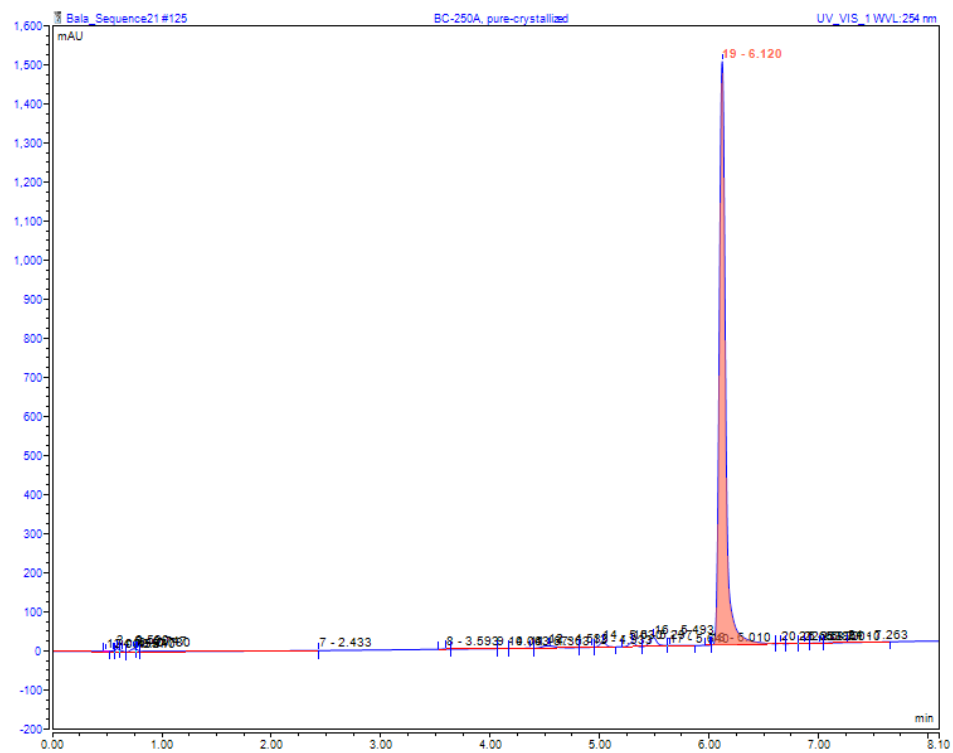
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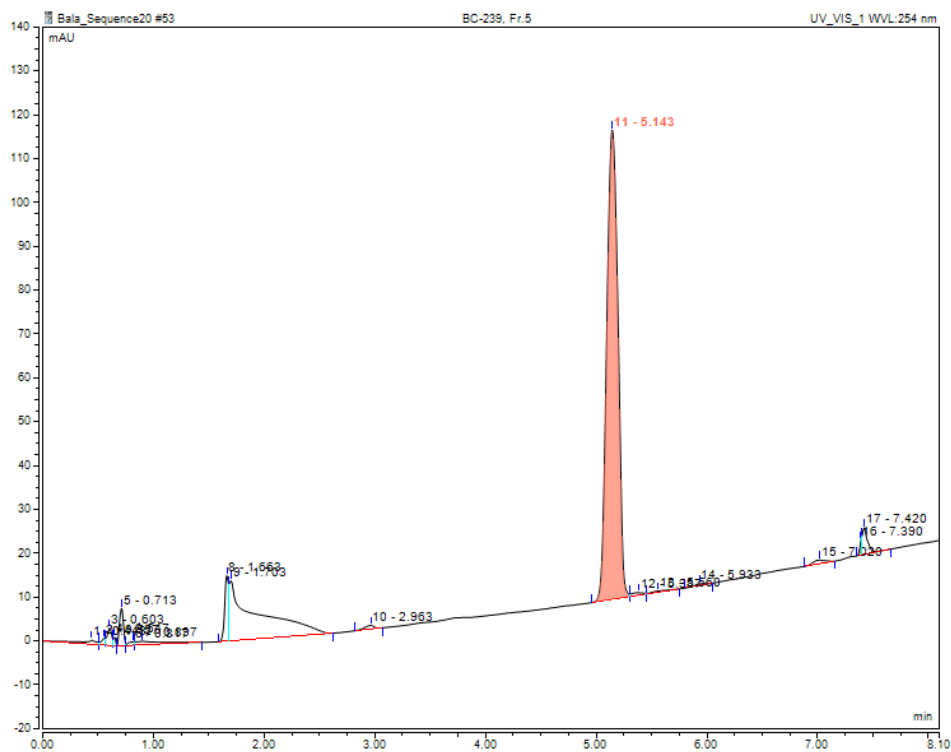
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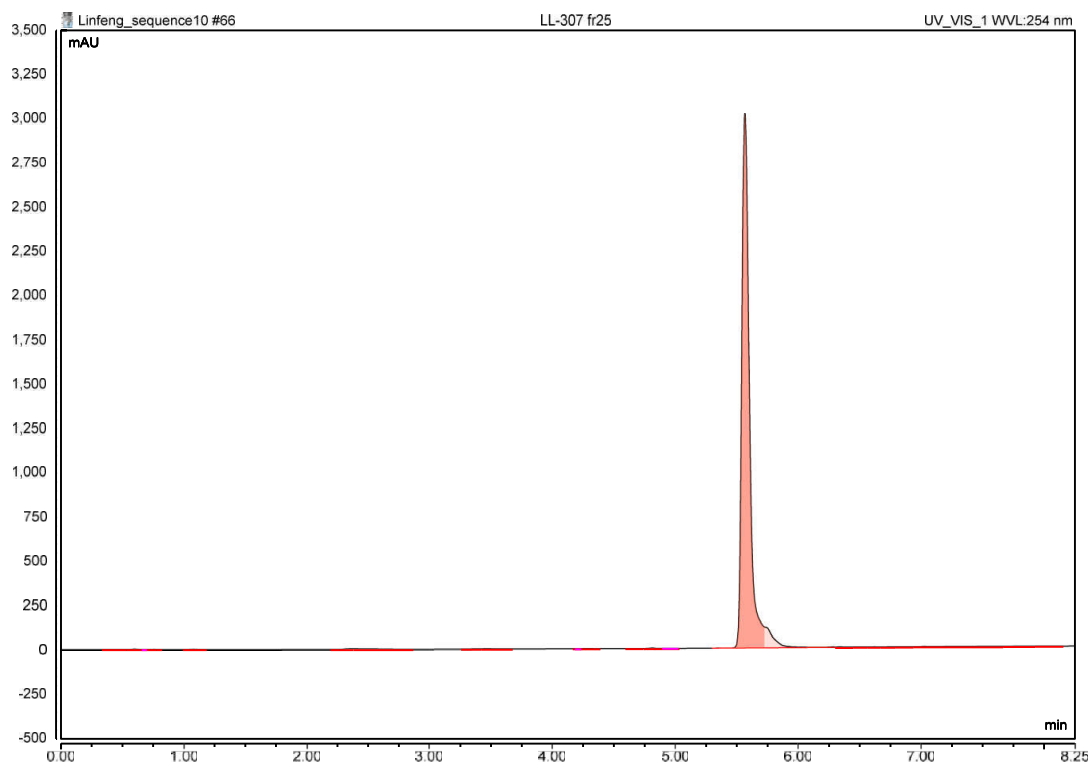
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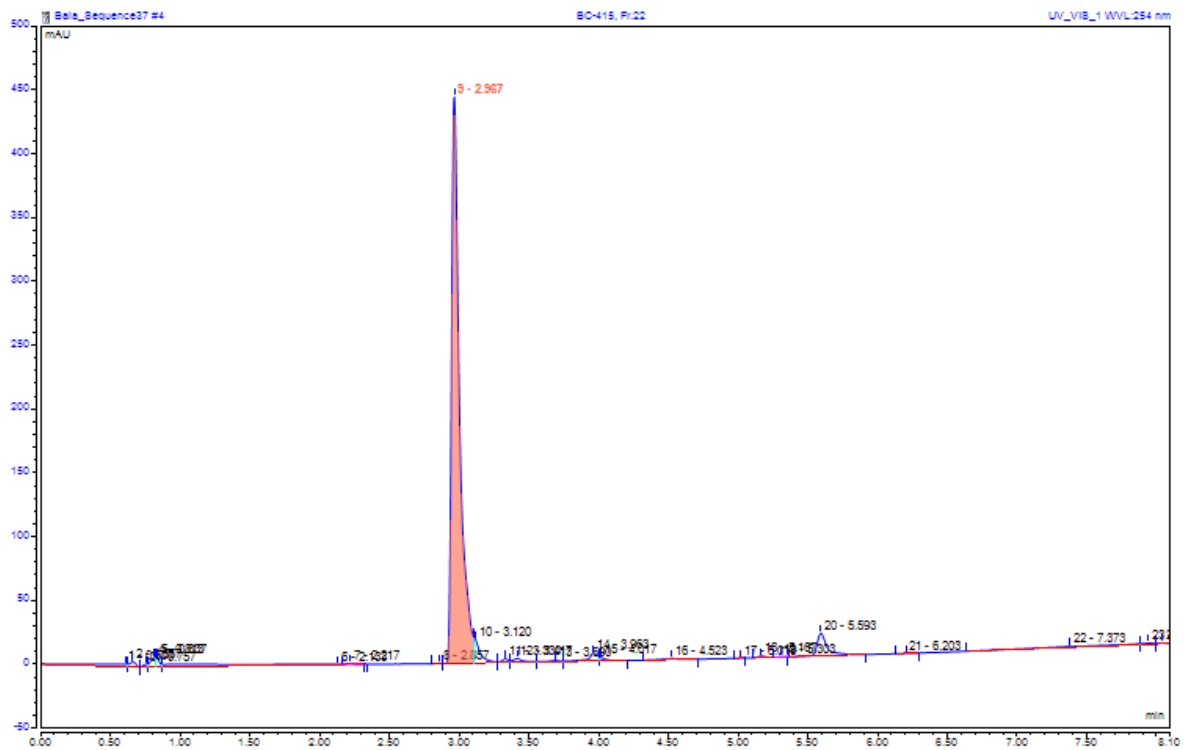
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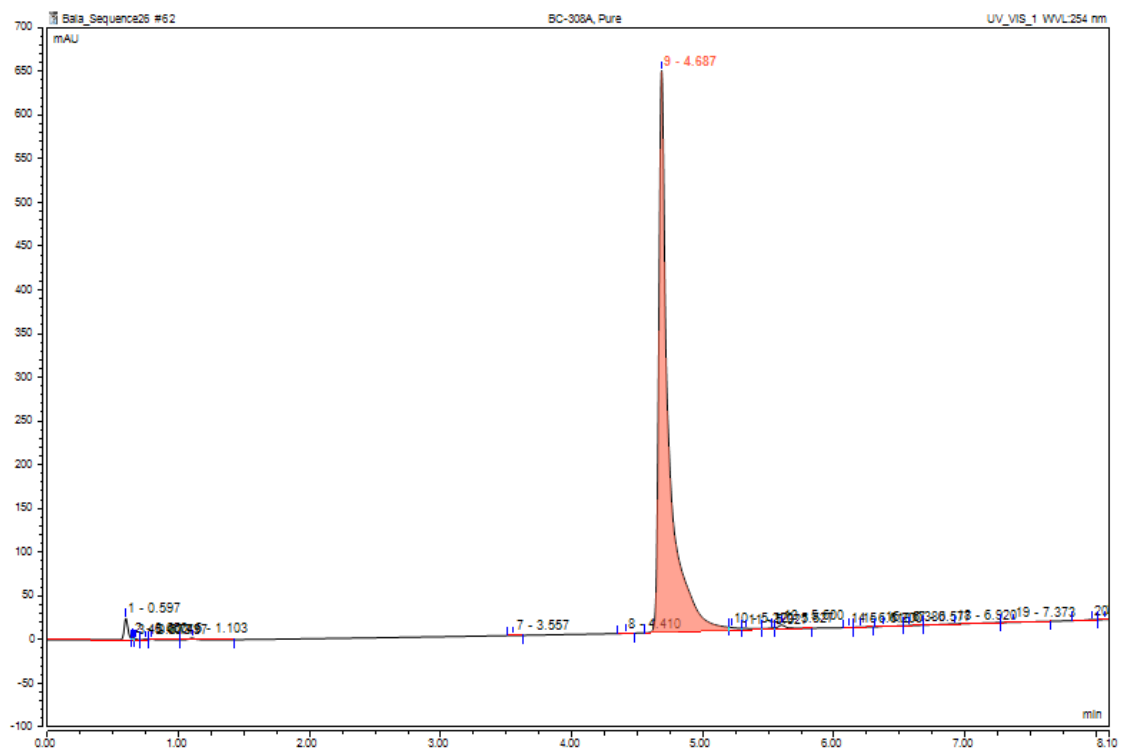
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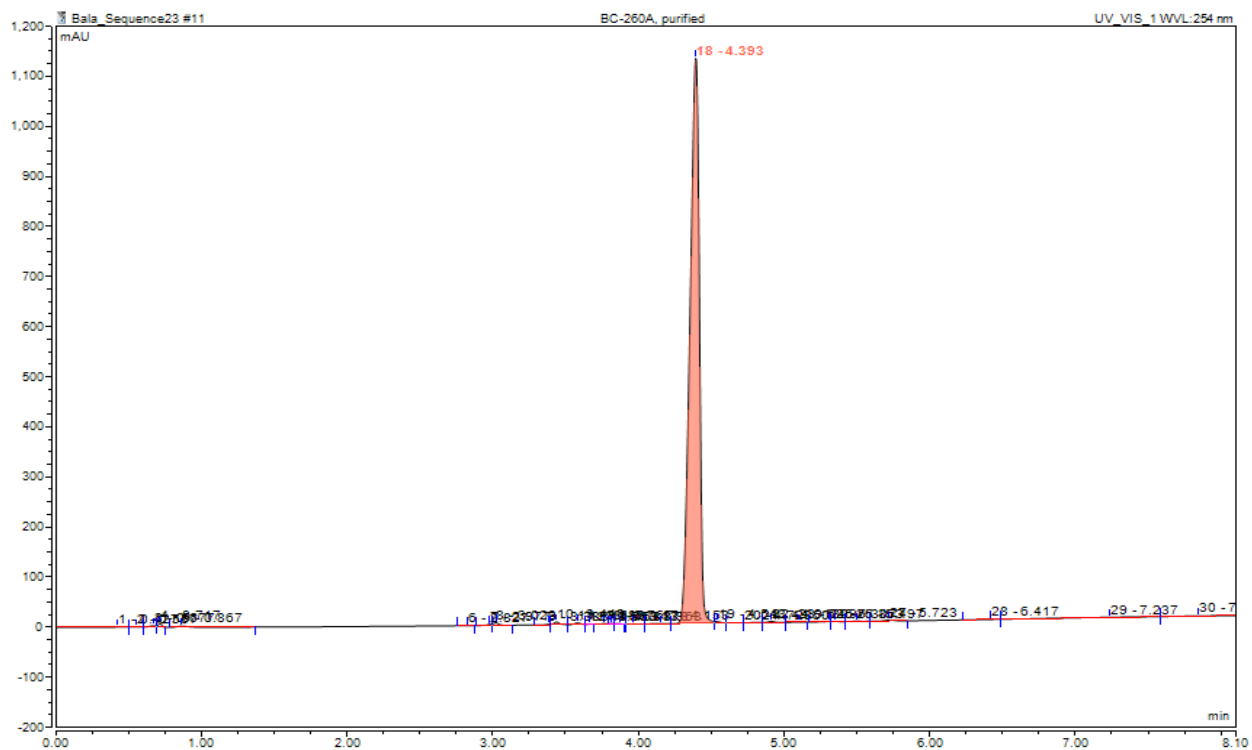
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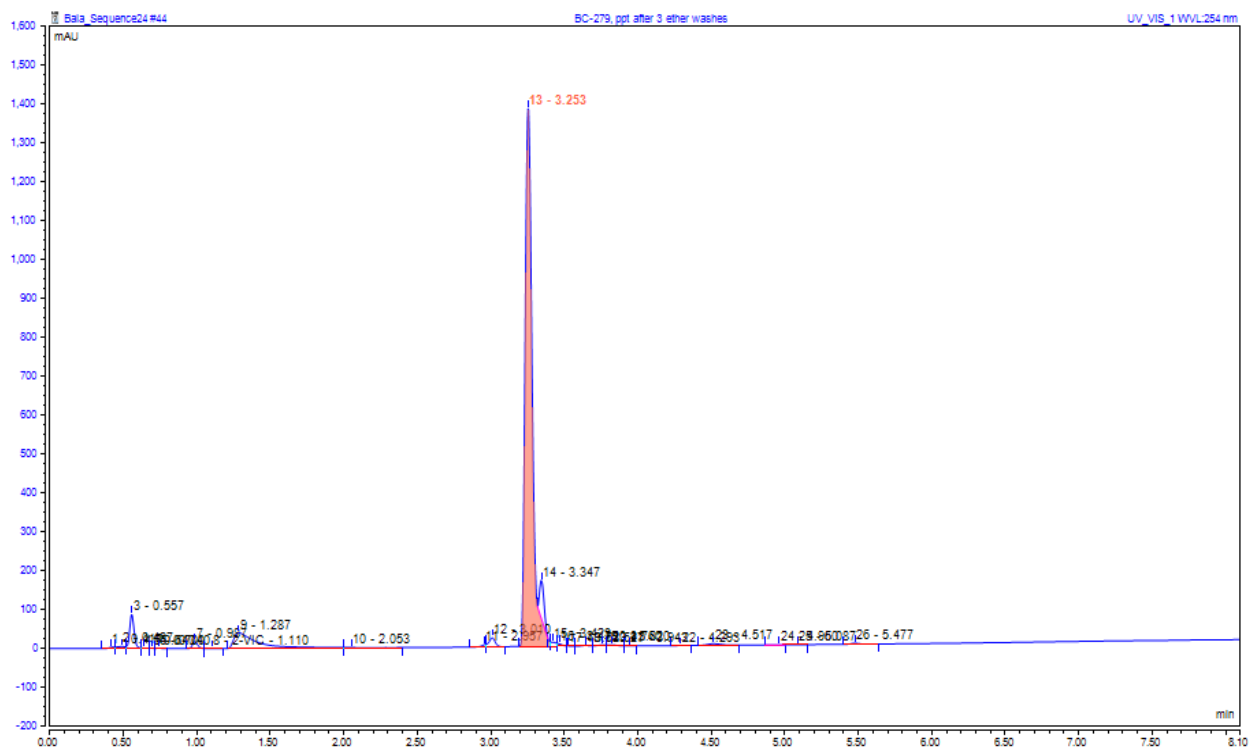
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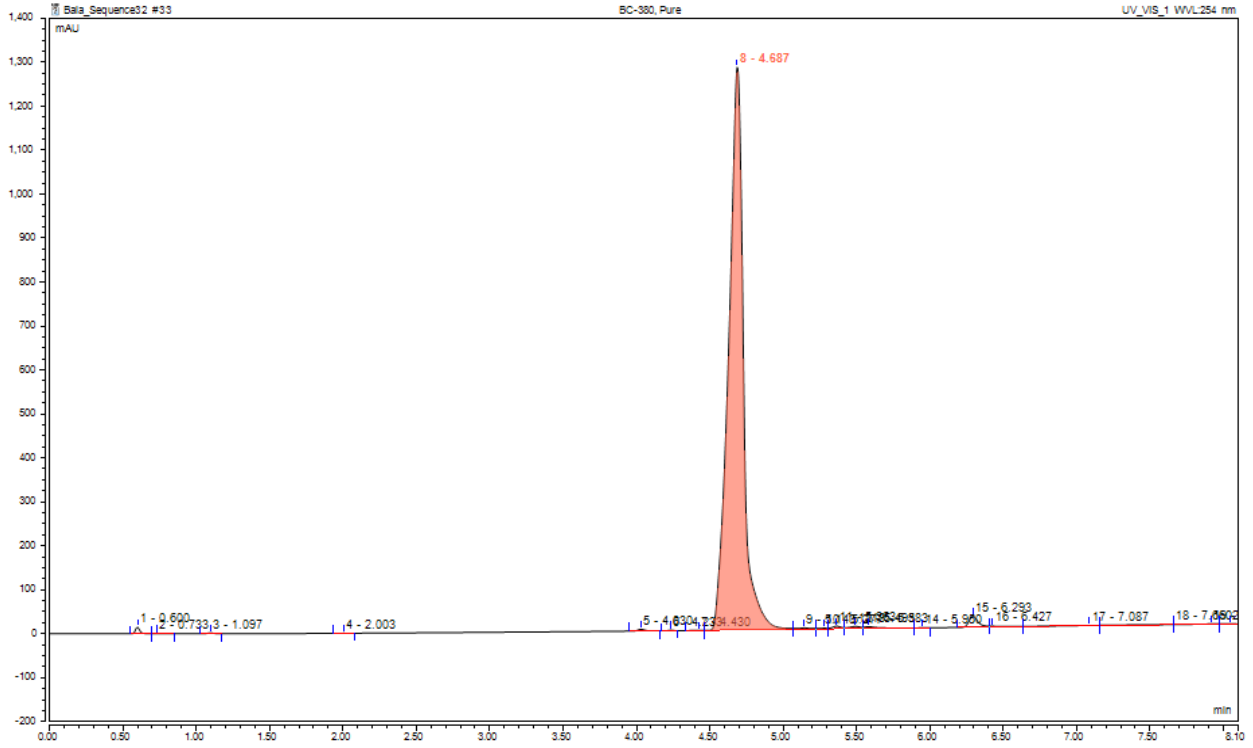
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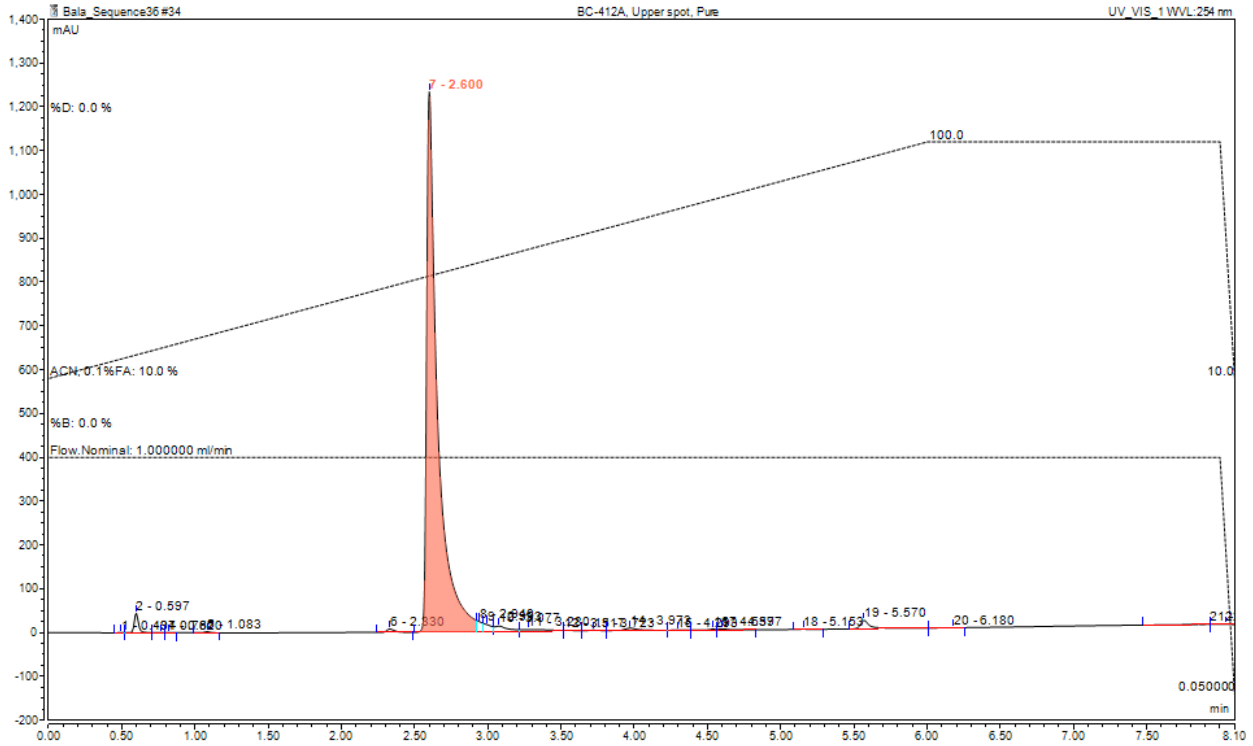
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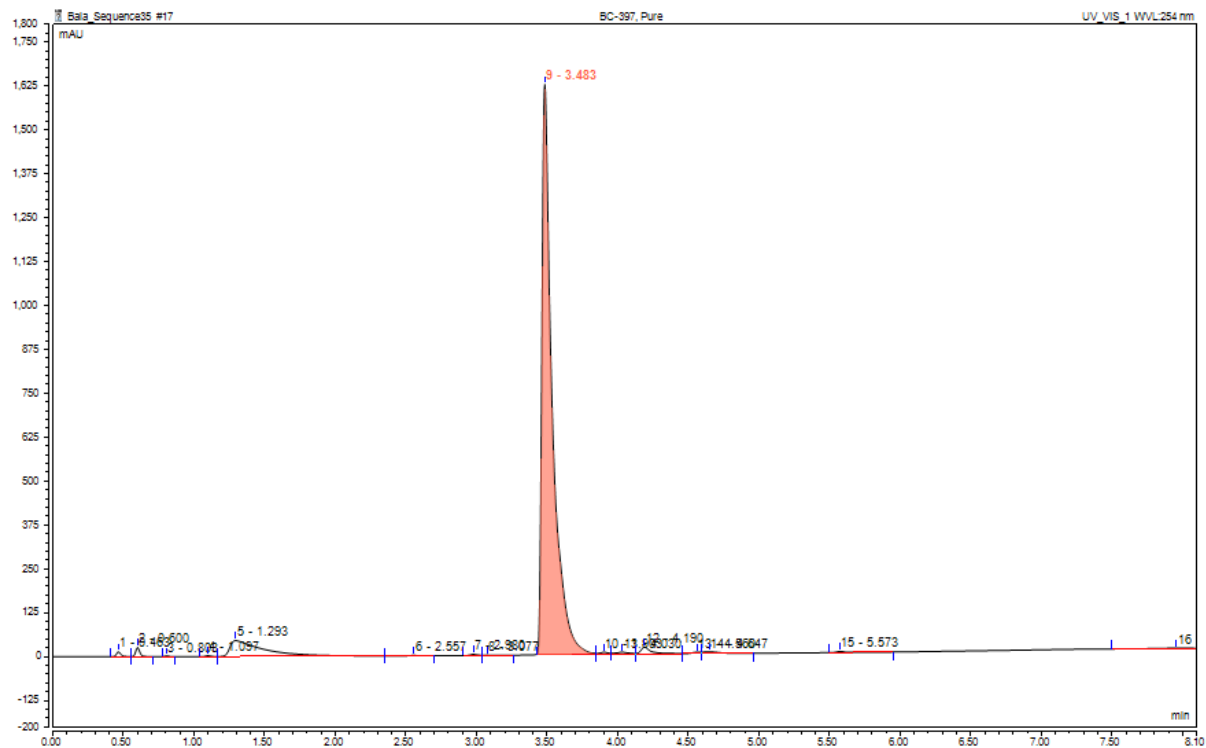
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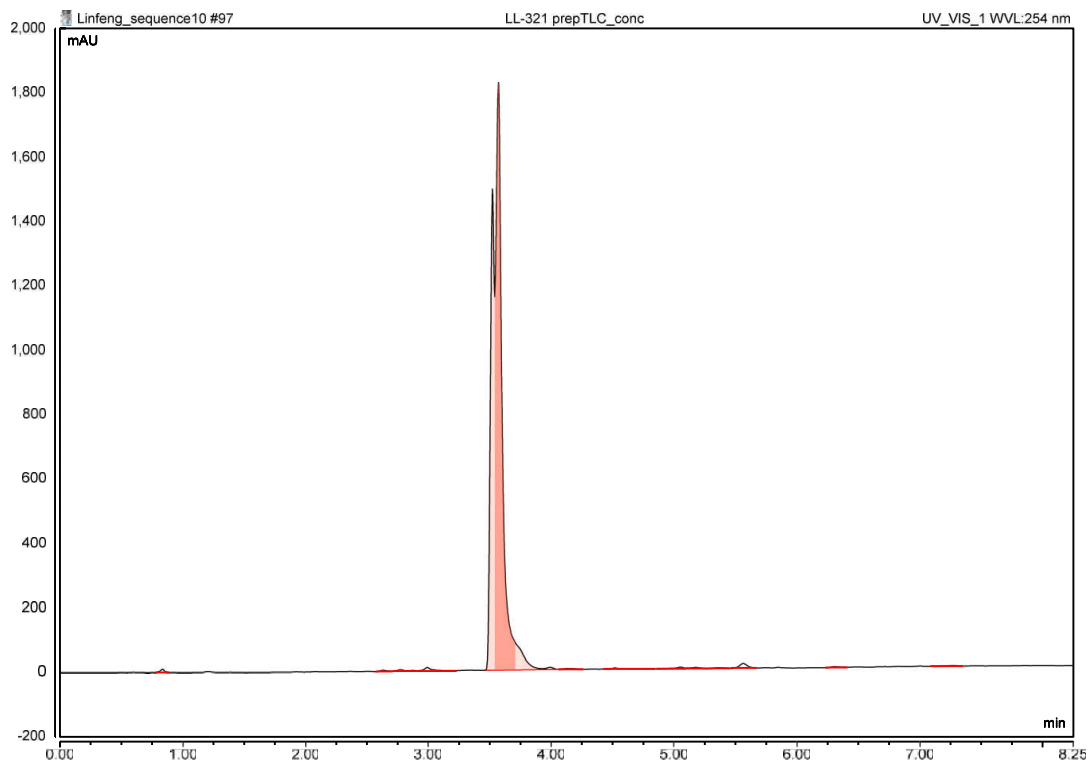
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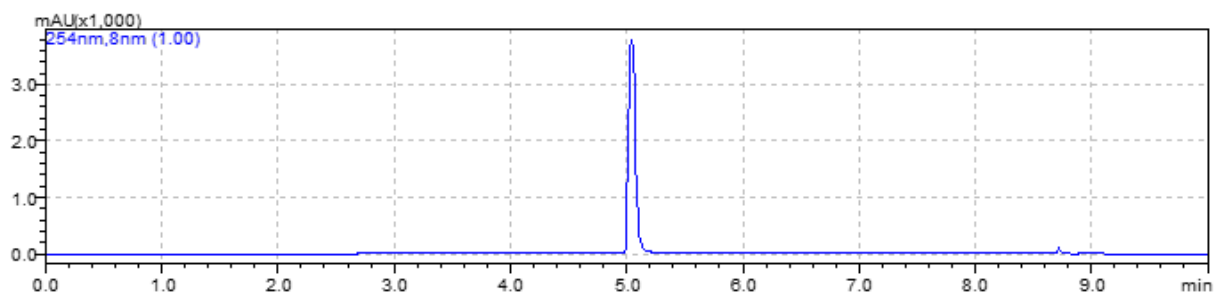
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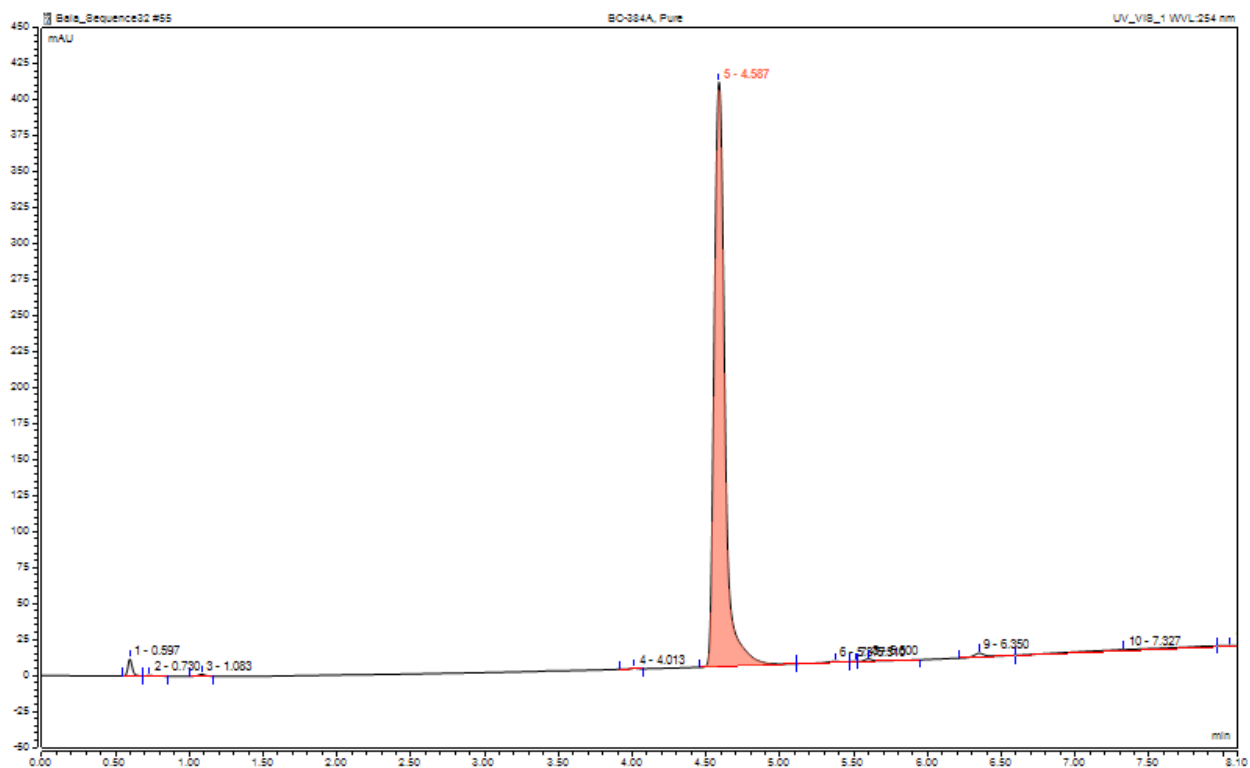
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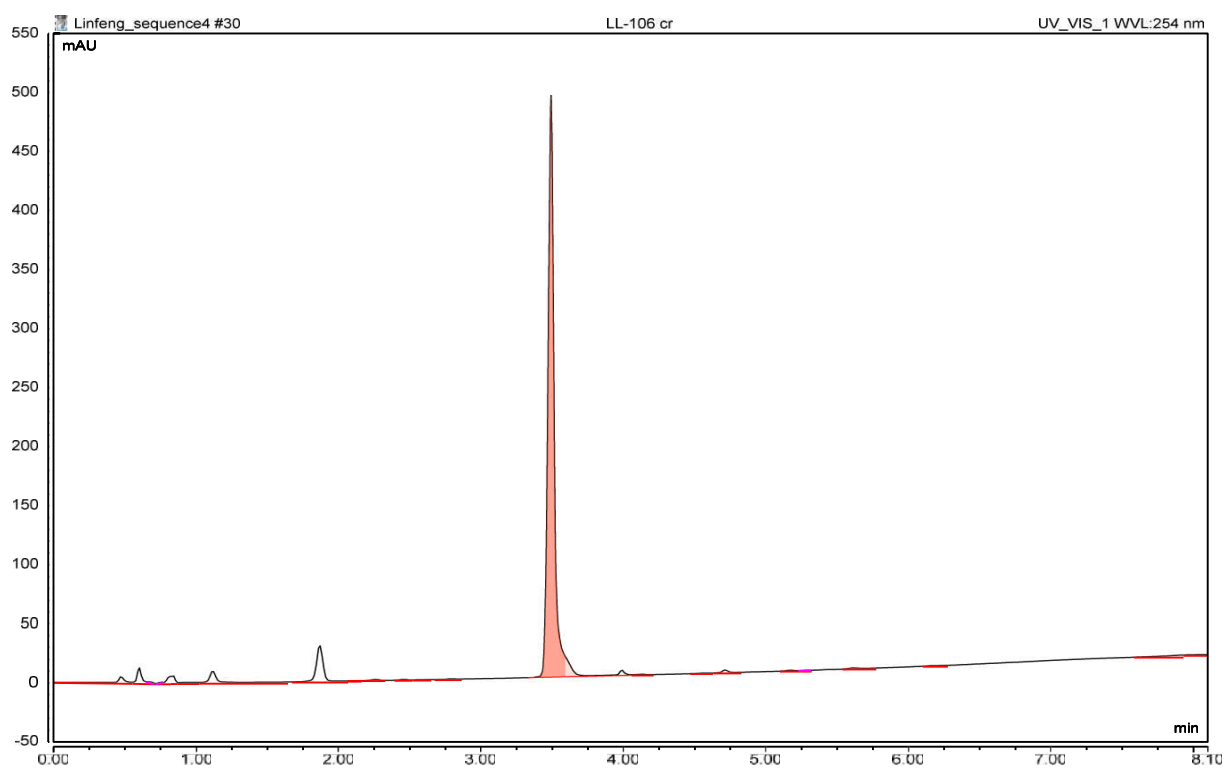
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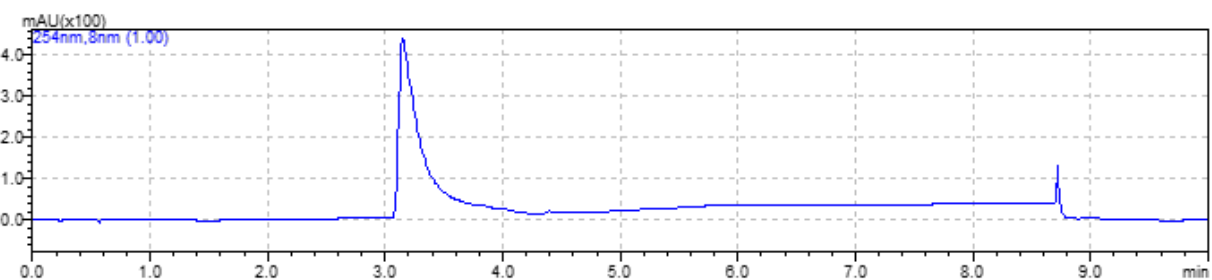
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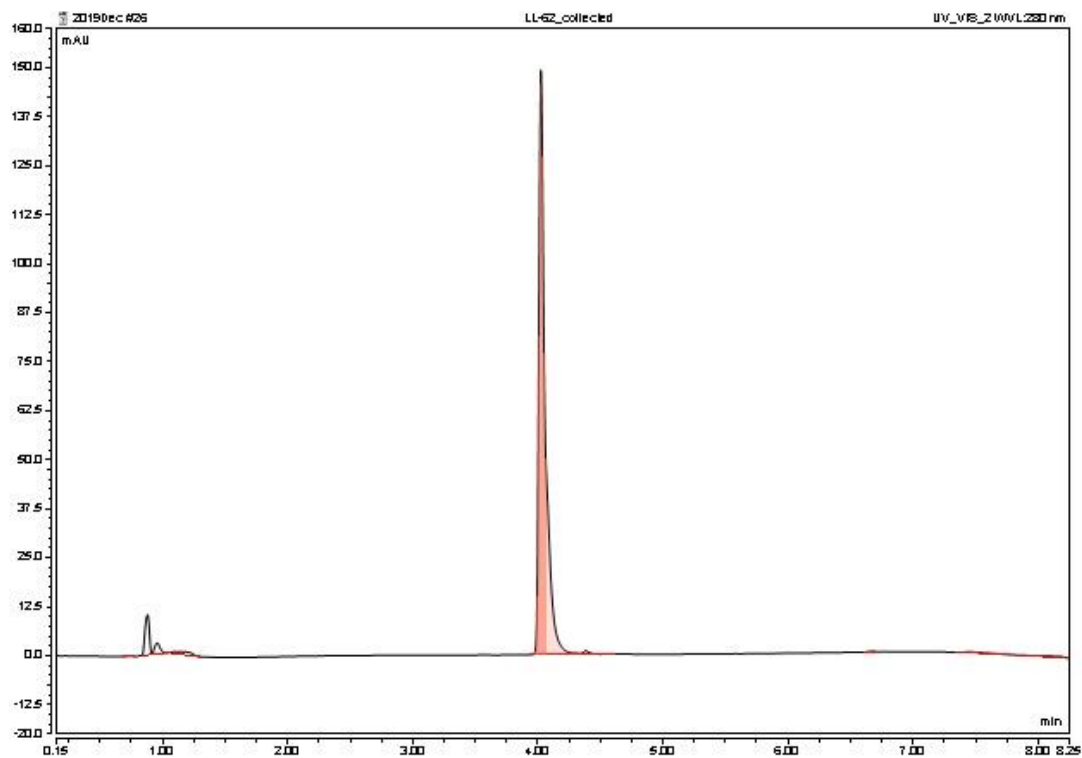
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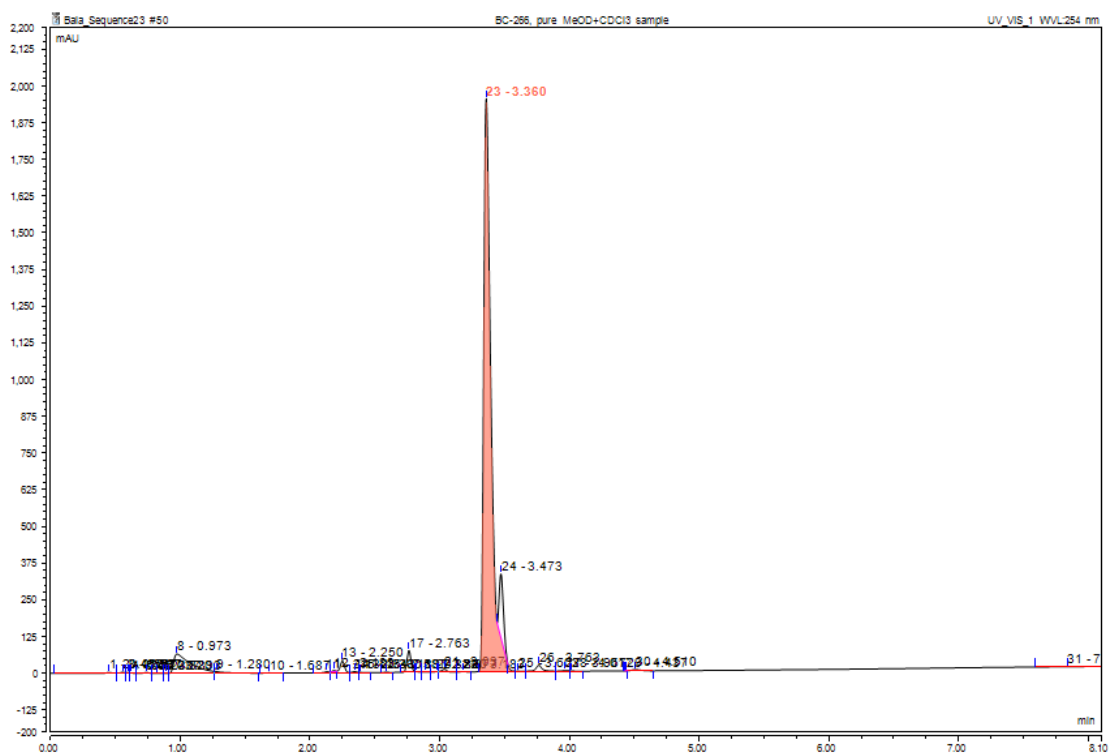
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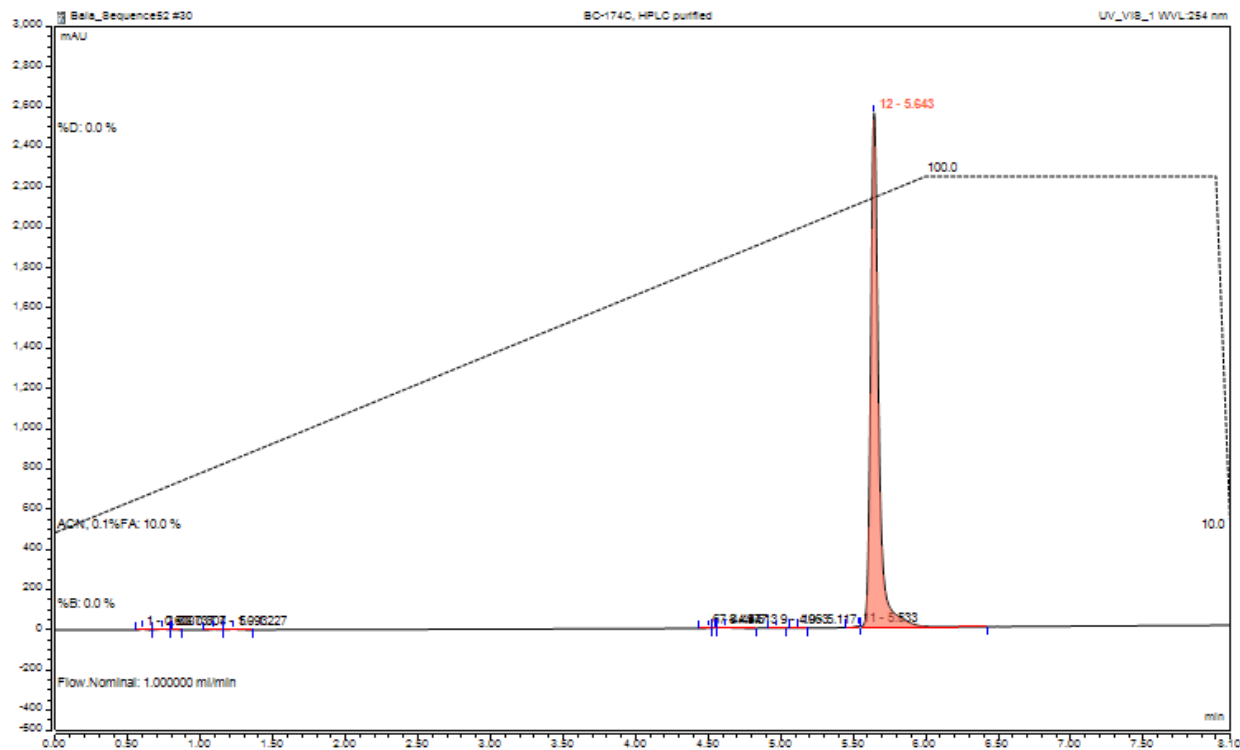
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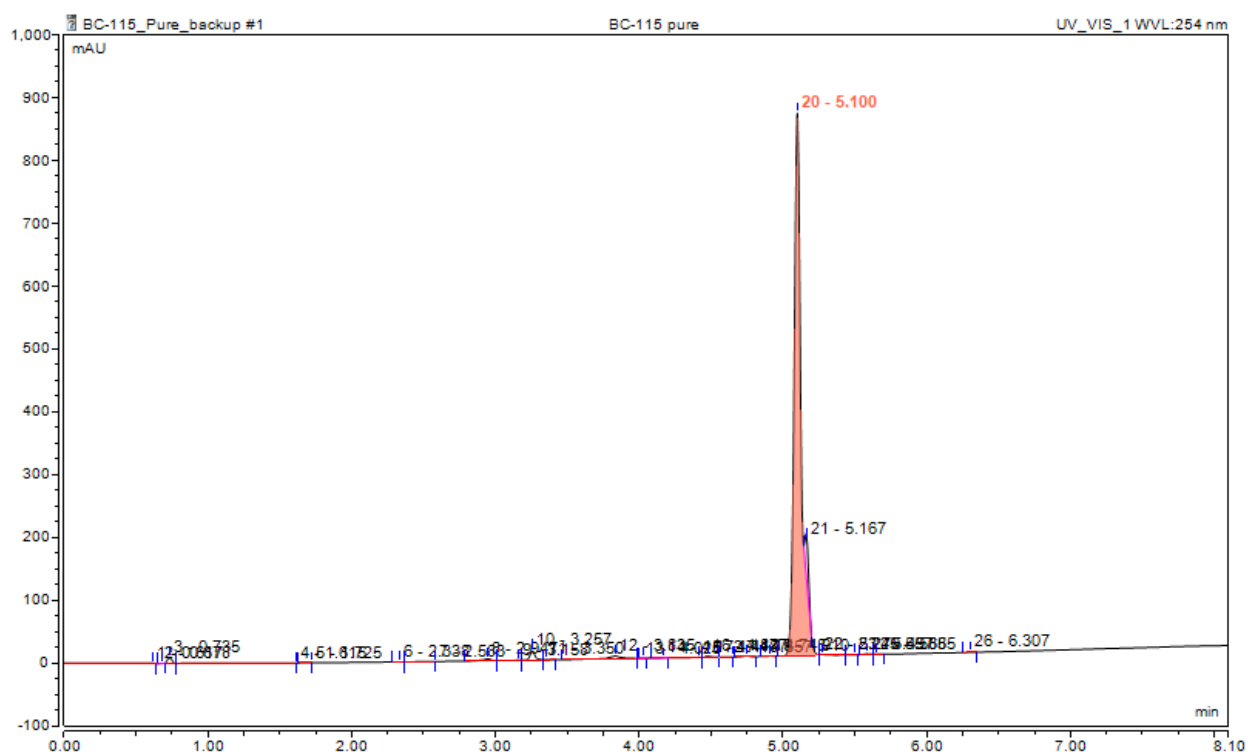
Compound 22:



Compound 25:

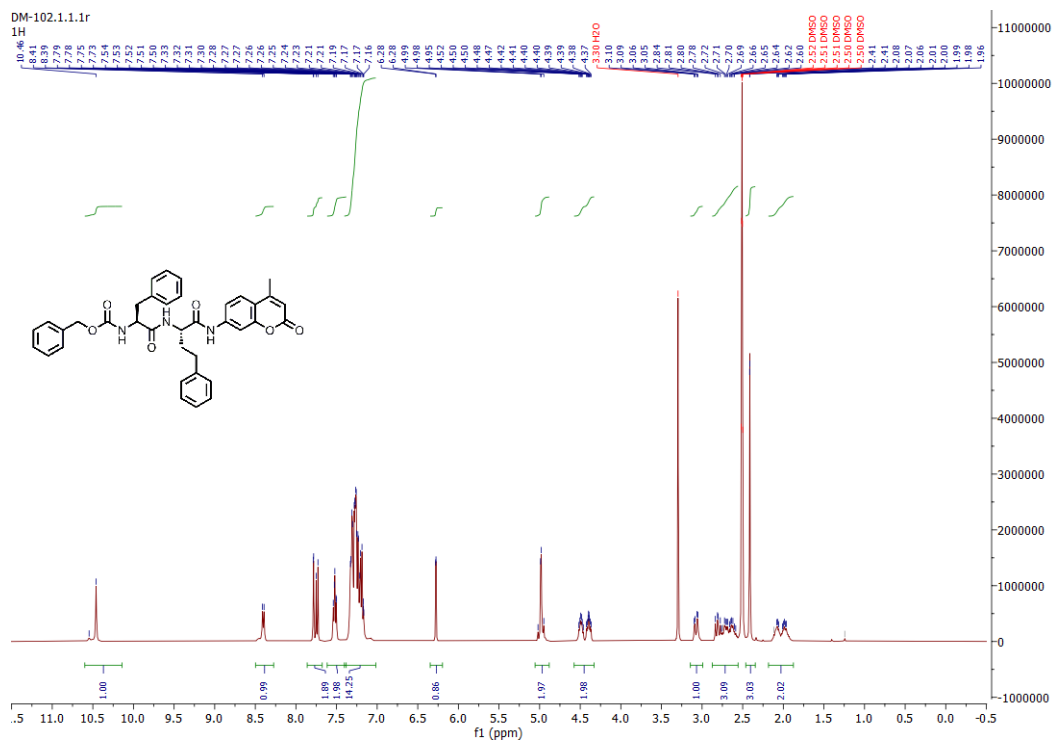


Compound 27:

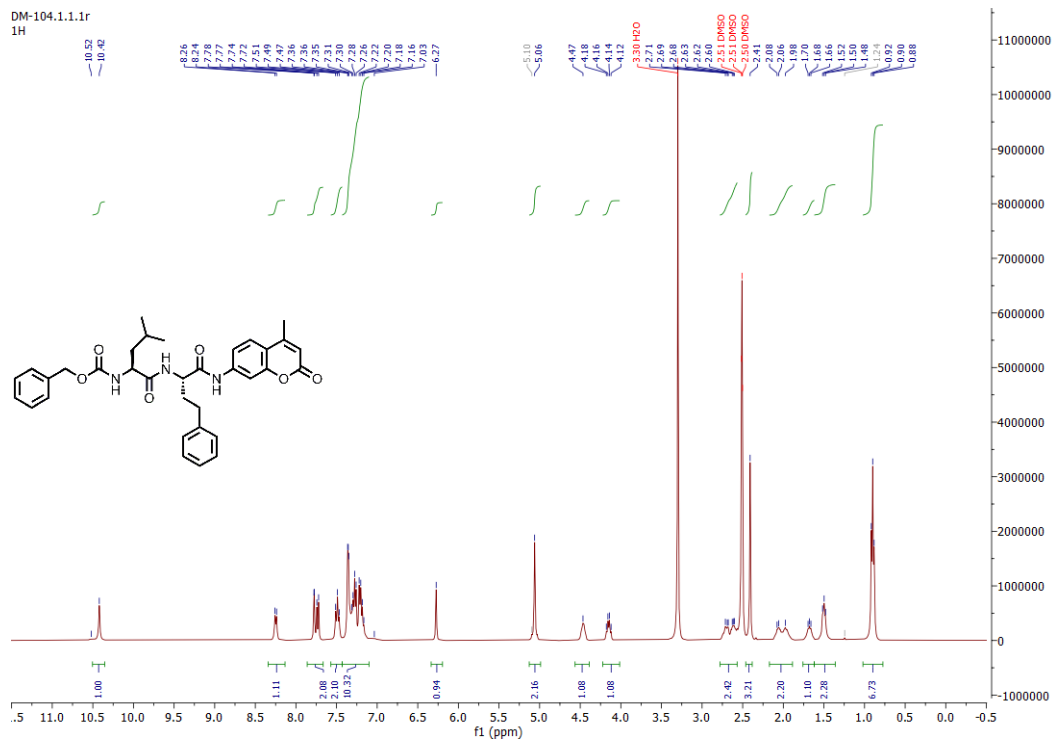


Part 7. NMR Spectra of Synthesized Substrates and Inhibitors

S2:

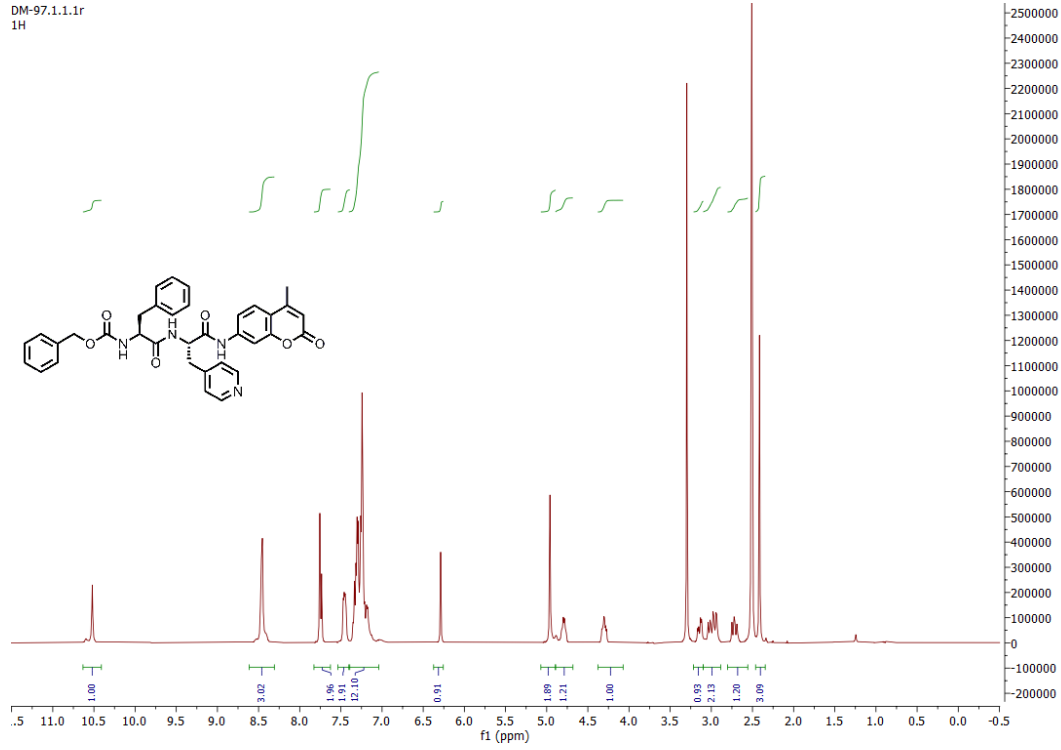


S3:



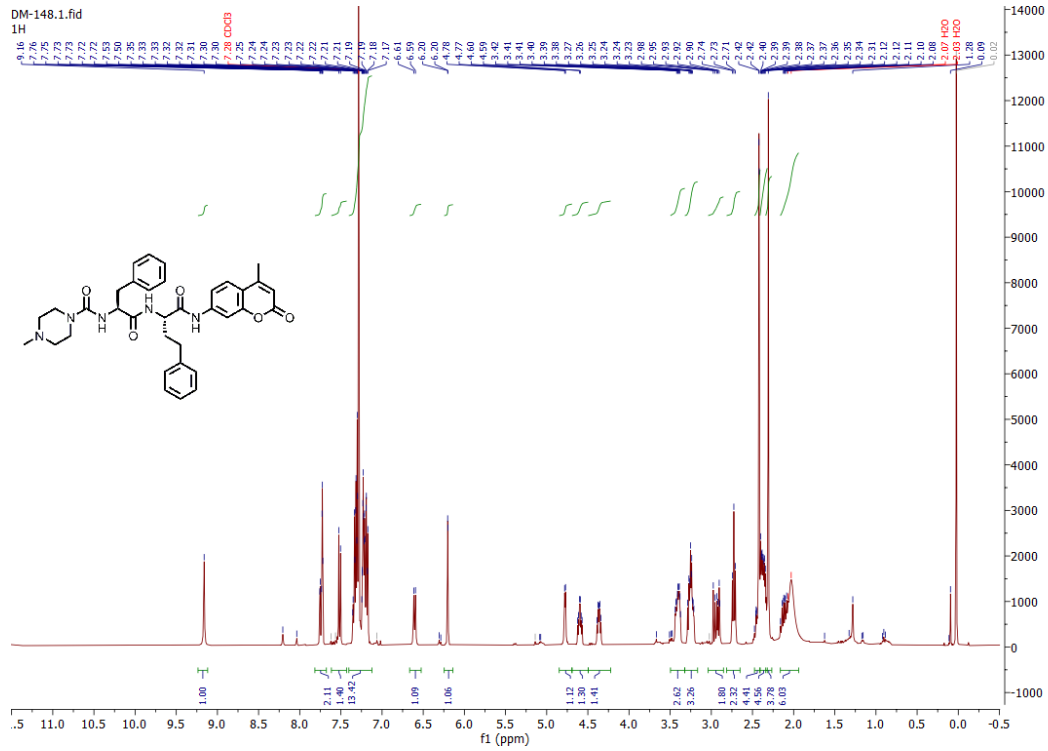
S5:

DM-97.1.1.1r
1H



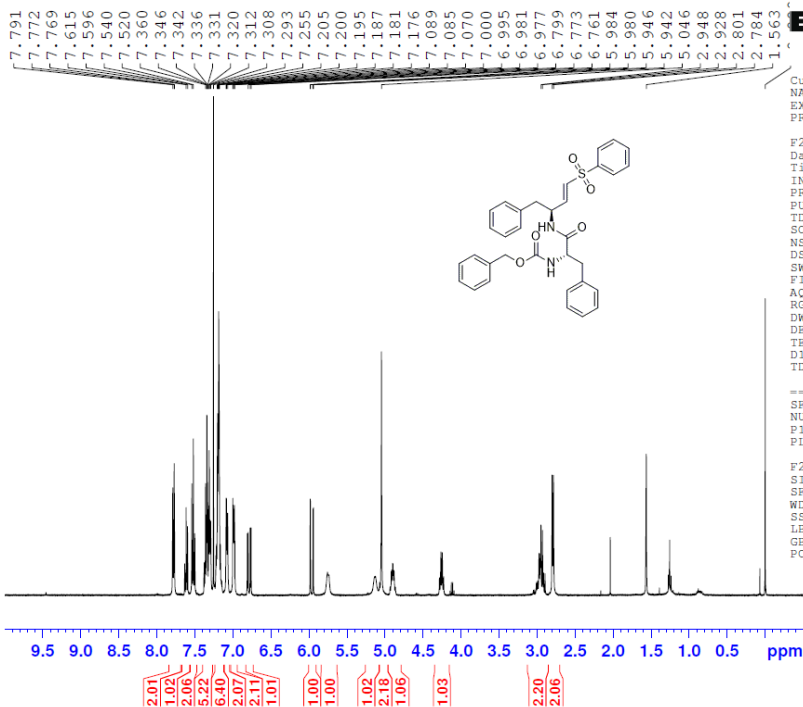
S7:

DM-148.1.fid
1H



¹H NMR spectrum of compound 1

BC-50A 1HNMR, CDCl3



Current Data Parameters
 NAME BC-50A
 EXPNO 1
 PROCNO 1

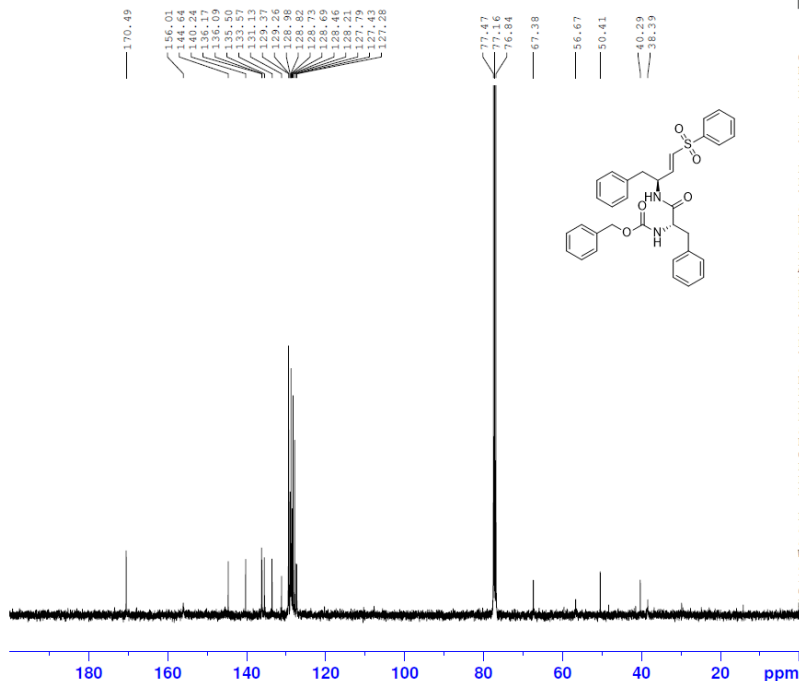
F2 - Acquisition Parameters
 Date_ 20150202
 Time_ 12.14
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 192.64
 DW 62.400 usec
 DE 6.50 usec
 TE 305.0 K
 D1 1.0000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 26.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300114 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

¹³C NMR spectrum of compound 1

BC-50A, C13 NMR, Pure, CDCl3
 CARBON_TAMU CDCl3 /data bala 35



Current Data Parameters
 NAME BC-50A
 EXPNO 4
 PROCNO 1

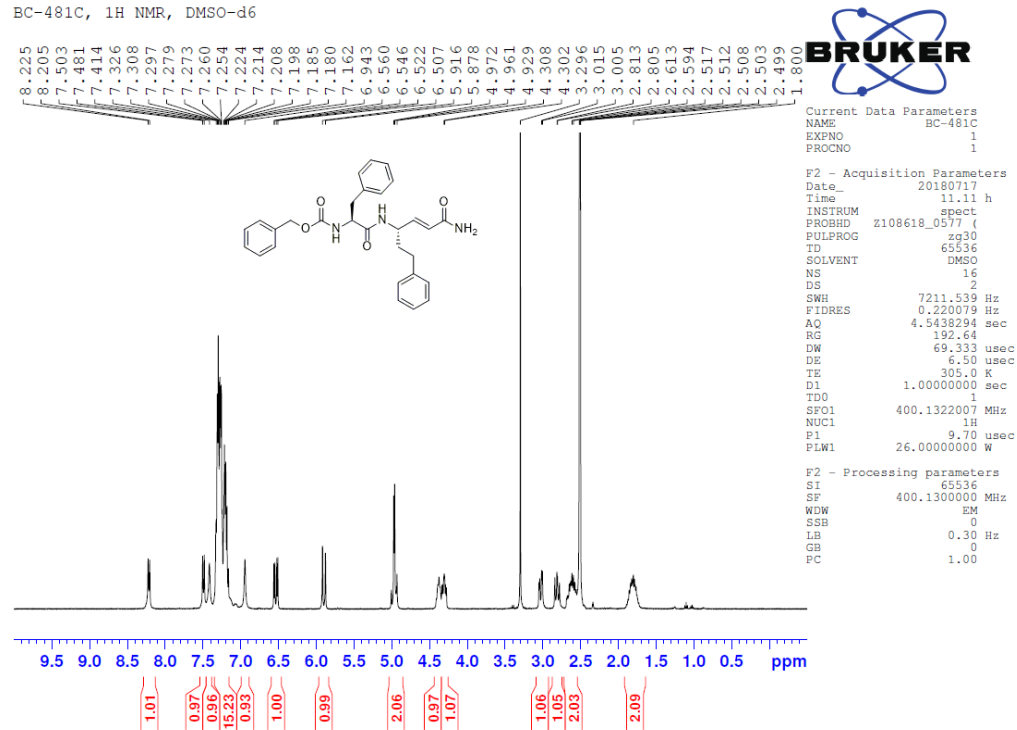
F2 - Acquisition Parameters
 Date_ 20200128
 Time_ 22.18 h
 INSTRUM spect
 PROBHD Z108618_0577 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1008
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.133596 Hz
 AQ 1.3631488 sec
 RG 192.64
 DW 20.800 usec
 DE 6.50 usec
 TE 305.0 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

SFO1 100.6228298 MHz
 NUC1 13C
 P0 3.33 usec
 P1 10.00 usec
 PLW1 50.0000000 W
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG2 waltz65
 PCPD2 90.00 usec
 PLW2 26.0000000 W
 PLW12 0.38839999 W

F2 - Processing parameters
 SI 32768
 SF 100.6127564 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

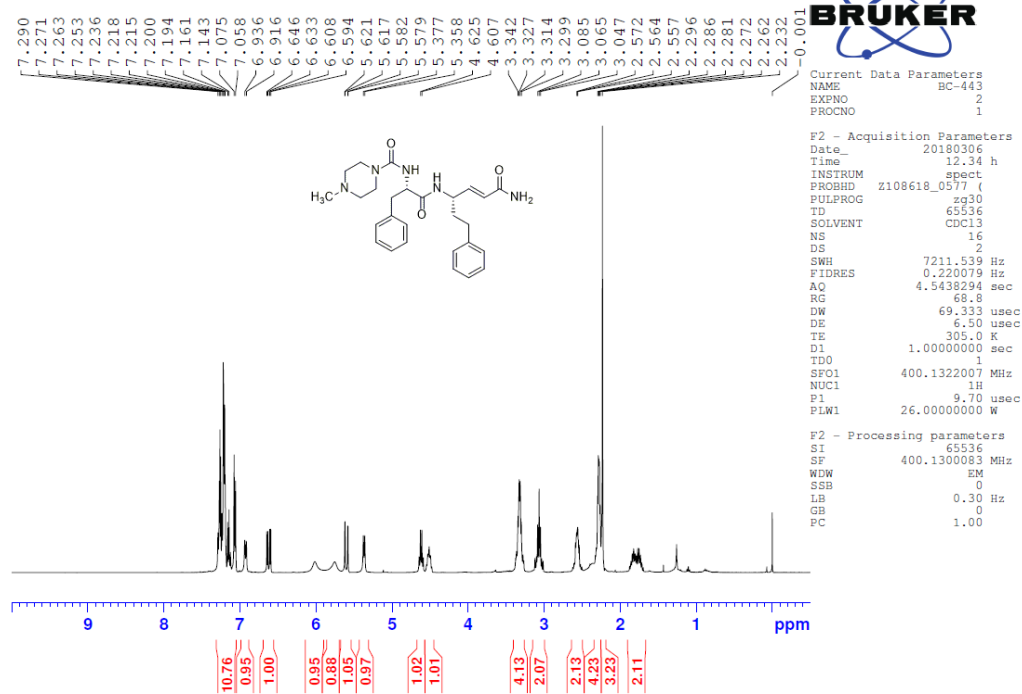
¹H NMR spectrum of compound 3

BC-481C, ¹H NMR, DMSO-d₆



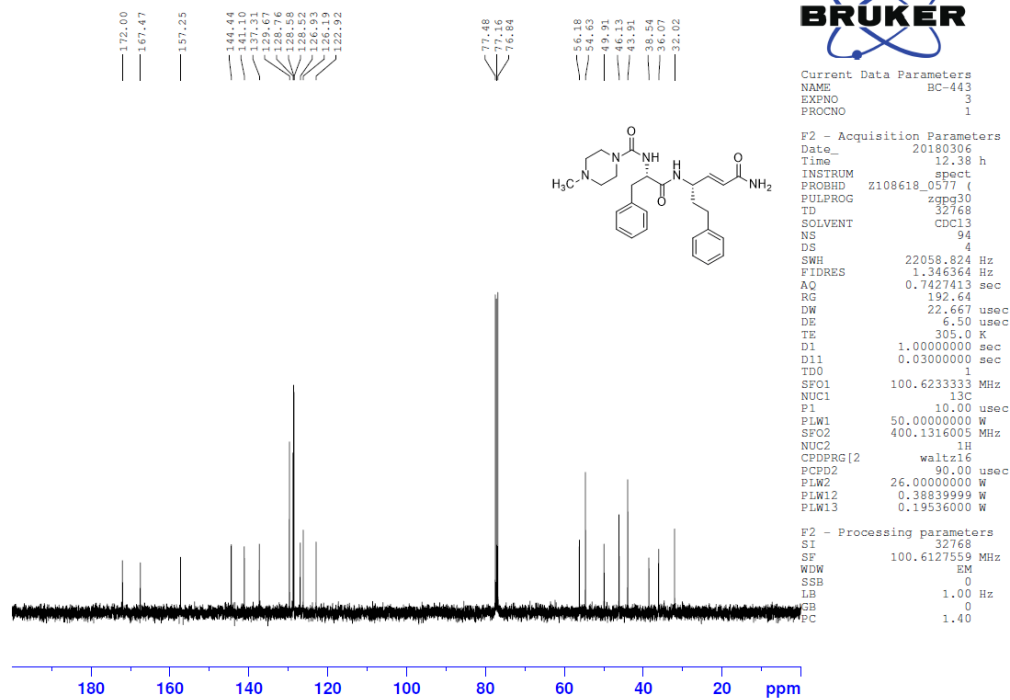
¹H NMR spectrum of compound 4

BC-443, purified ¹H NMR, CDCl₃

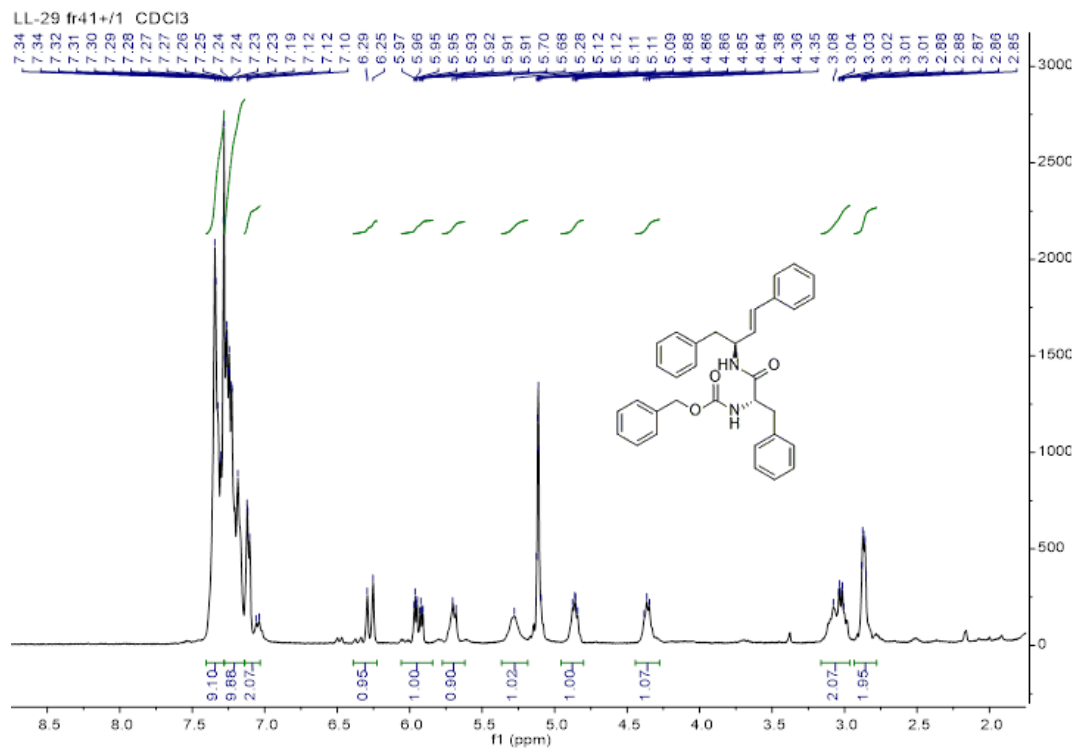


¹³C NMR spectrum of compound 4

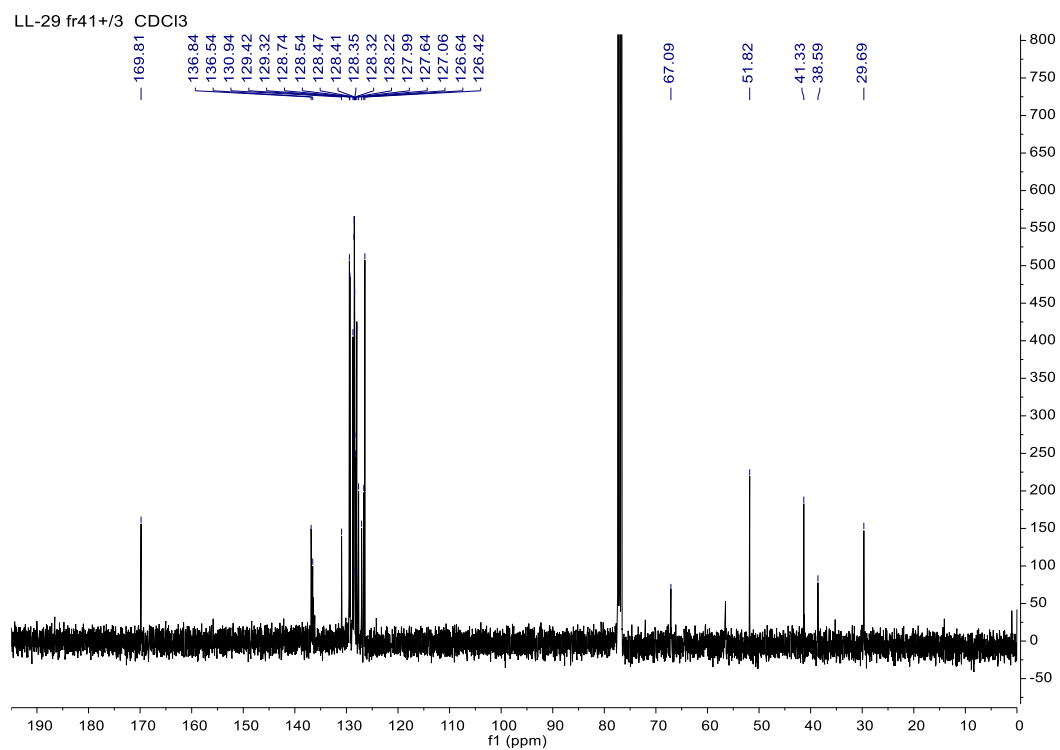
BC-443, purified ¹³C NMR, CDCl₃



¹H NMR spectrum of compound **5**

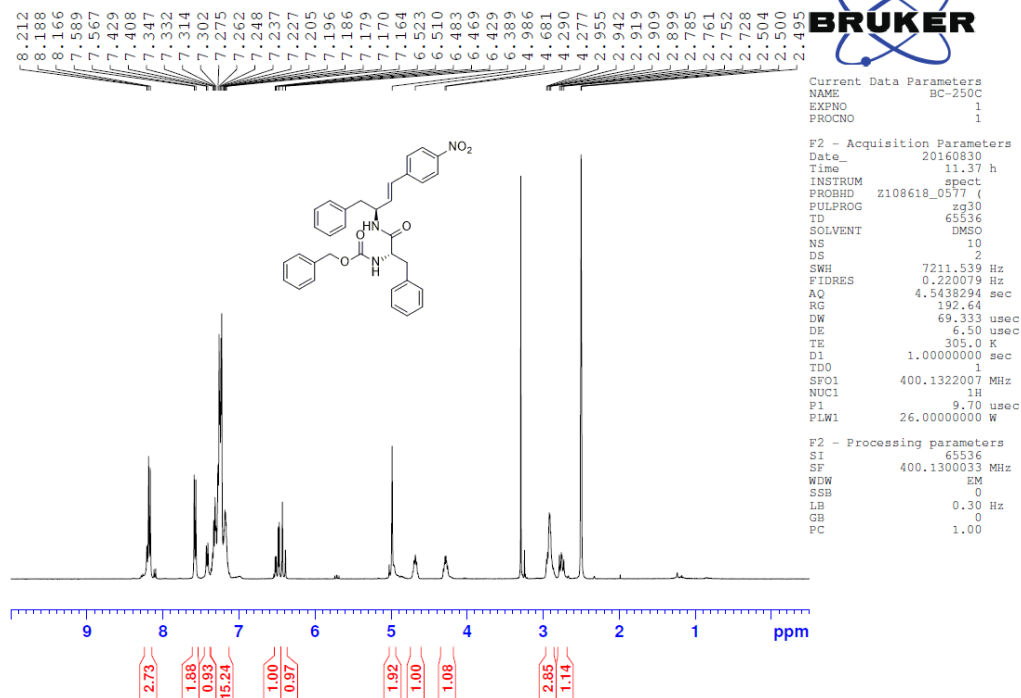


¹³C NMR spectrum of compound **5**



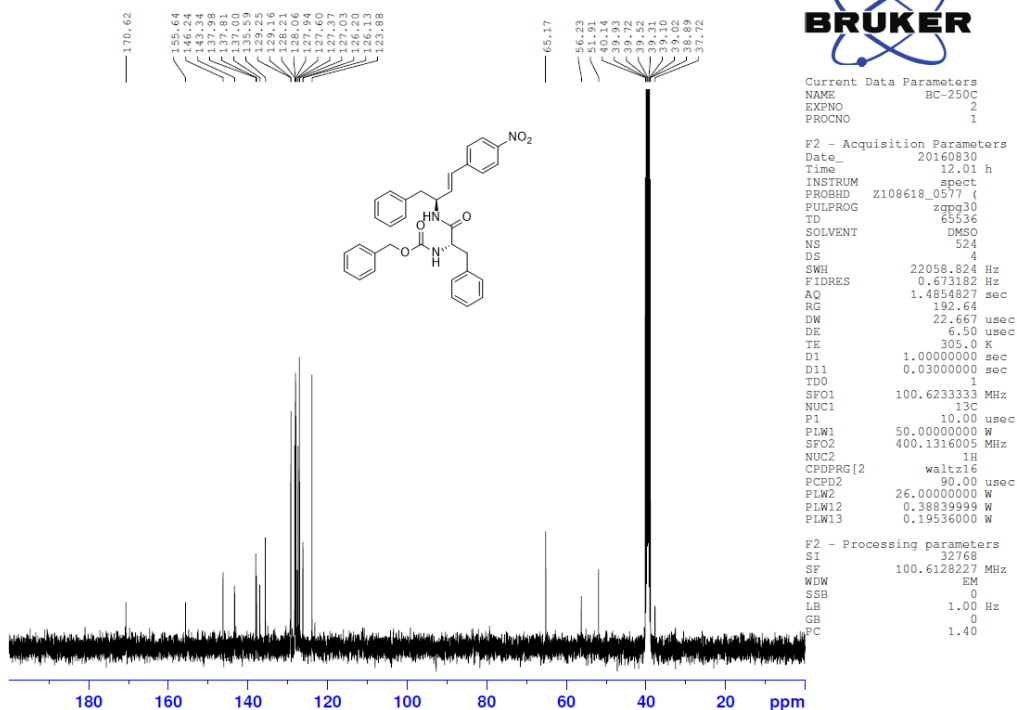
¹H NMR spectrum of compound 6

BC-250C, ¹H NMR, purified-twice, DMSO-d₆



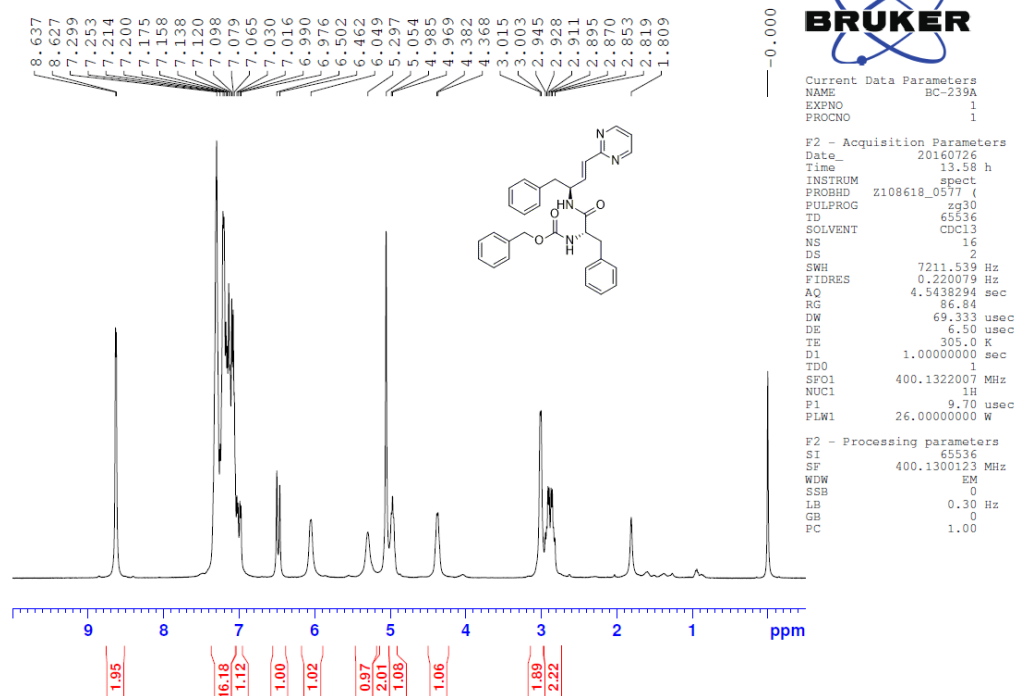
¹³C NMR spectrum of compound 6

BC-250C, ¹³C NMR, purified-twice, DMSO-d₆



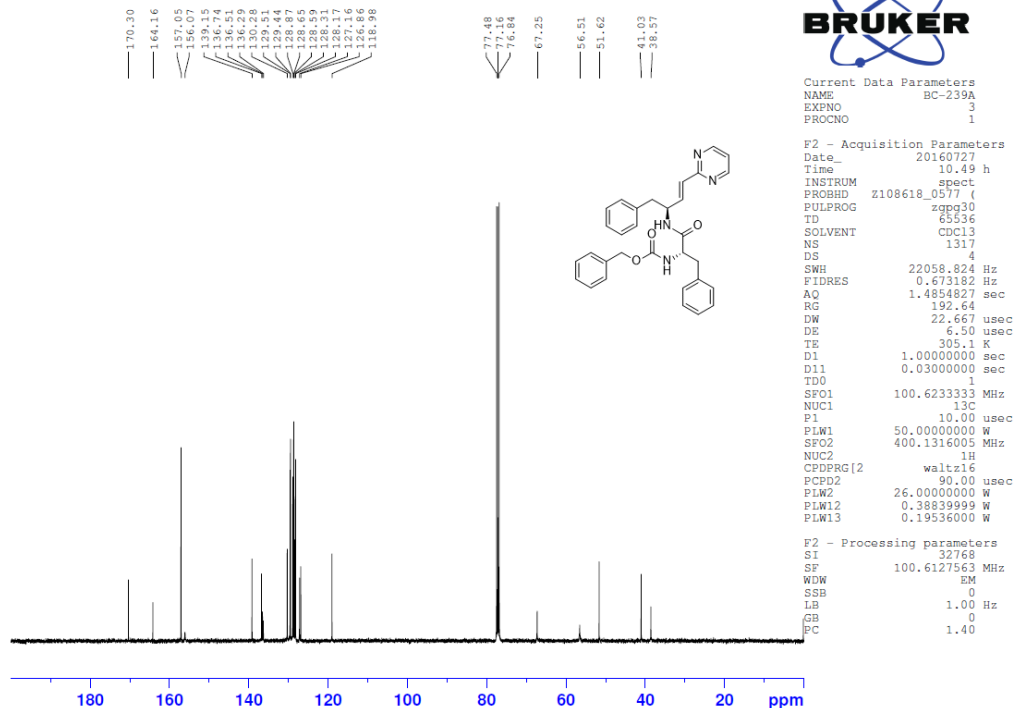
¹H NMR spectrum of compound 7

BC-239A ¹H NMR Purified-crystallized CDCl₃



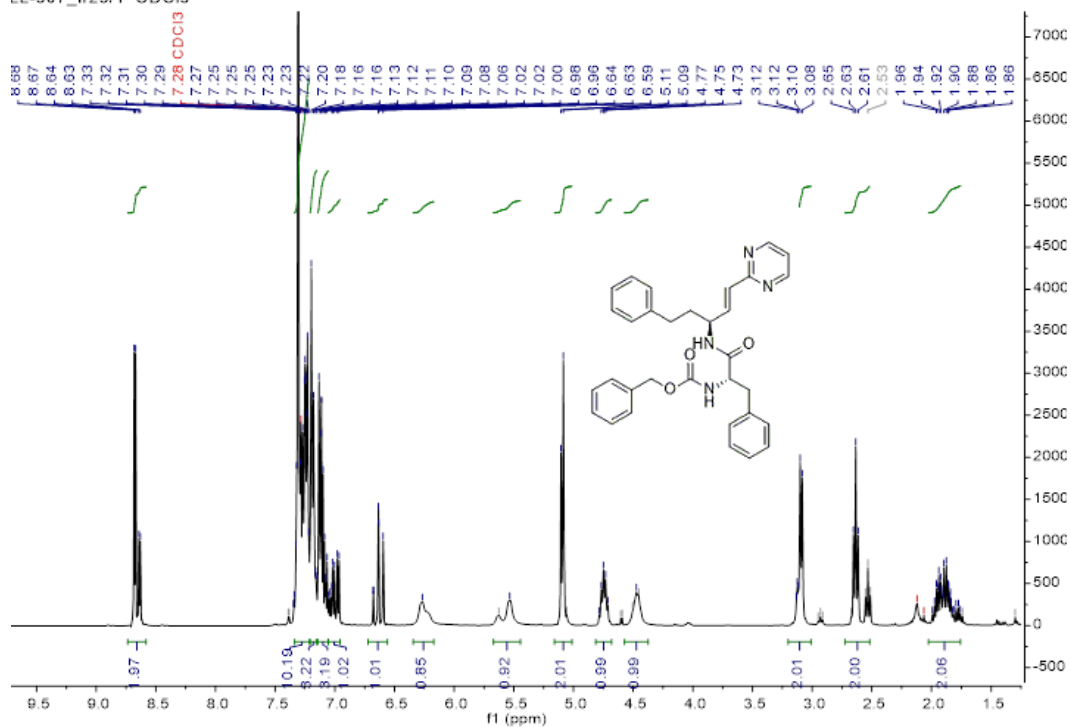
¹³C NMR spectrum of compound 7

BC-239A ¹³C NMR Purified-crystallized CDCl₃



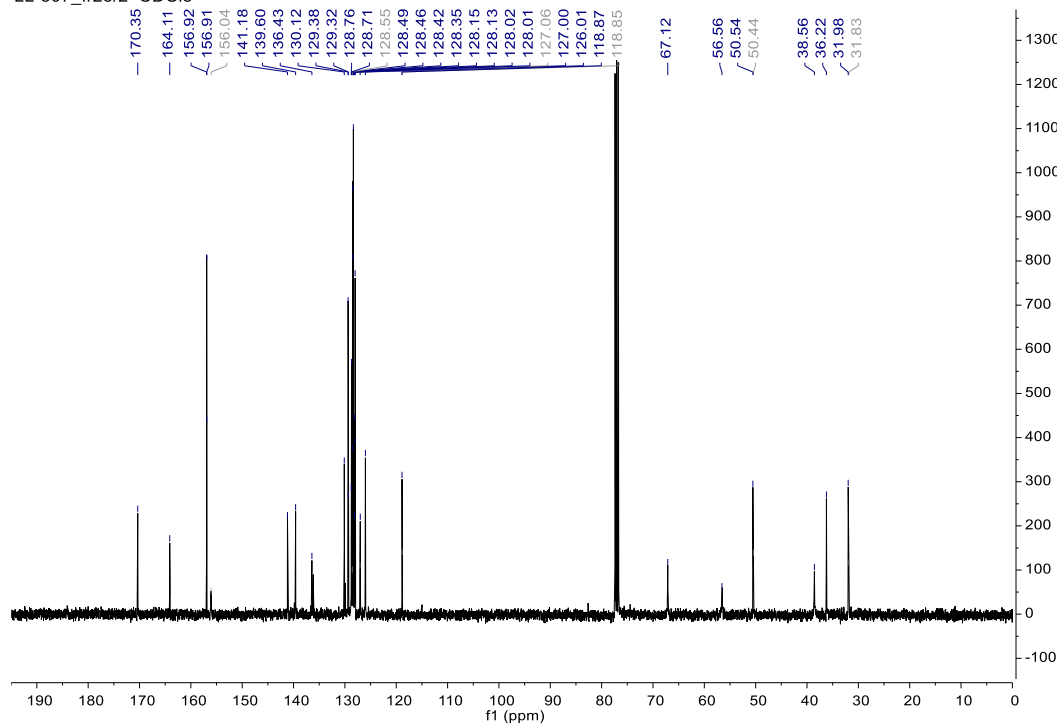
¹H NMR spectrum of compound 8

LL-307_fr25/1 CDCl₃



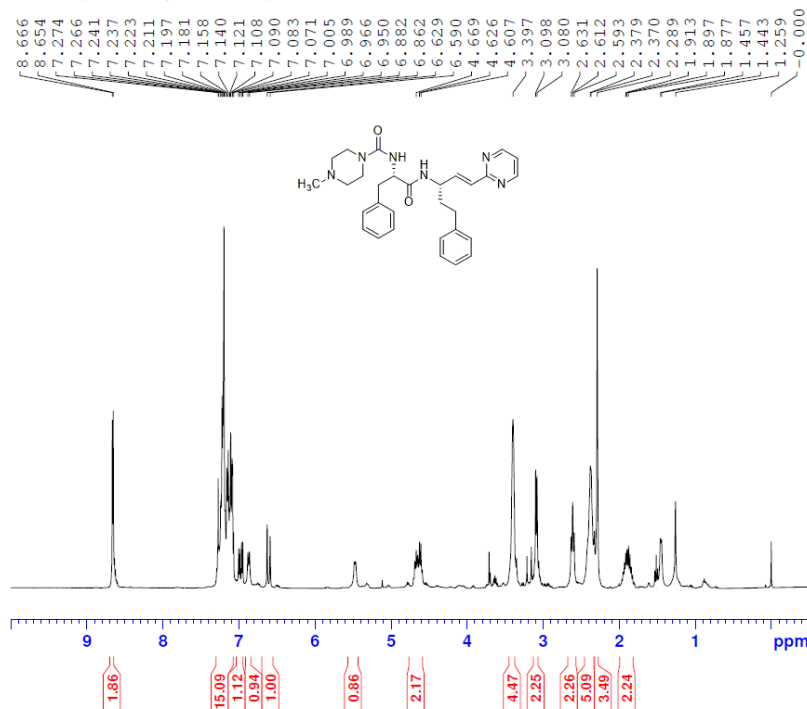
¹³C NMR spectrum of compound 8

LL-307_fr25/2 CDCl₃



¹H NMR spectrum of compound 9

BC-415A, ¹H NMR, Purified, CDCl₃



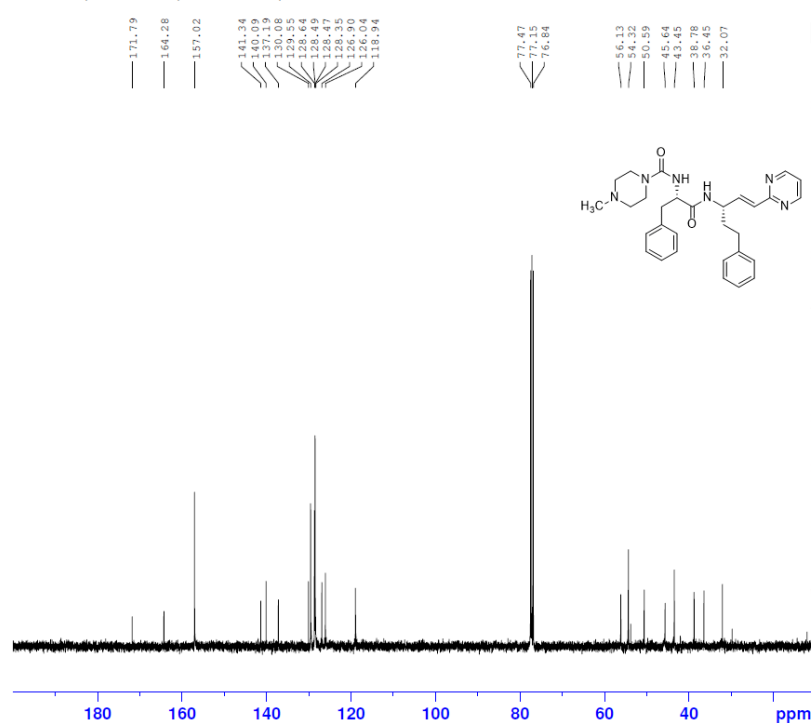
Current Data Parameters
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 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171113
 Time 10.41 h
 INSTRUM spect
 PROBHD Z108618_0577 (
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4807.692 Hz
 FIDRES 0.144719 Hz
 AQ 6.8157439 sec
 RG 55.4
 DW 104.000 usec
 DE 6.50 usec
 TE 305.0 K
 D1 1.00000000 sec
 TDO 1
 SFO1 400.1322007 MHz
 NUC1 1H
 P1 9.70 usec
 PLW1 26.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300041 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR spectrum of compound 9

BC-415A, ¹³C NMR, Purified, CDCl₃



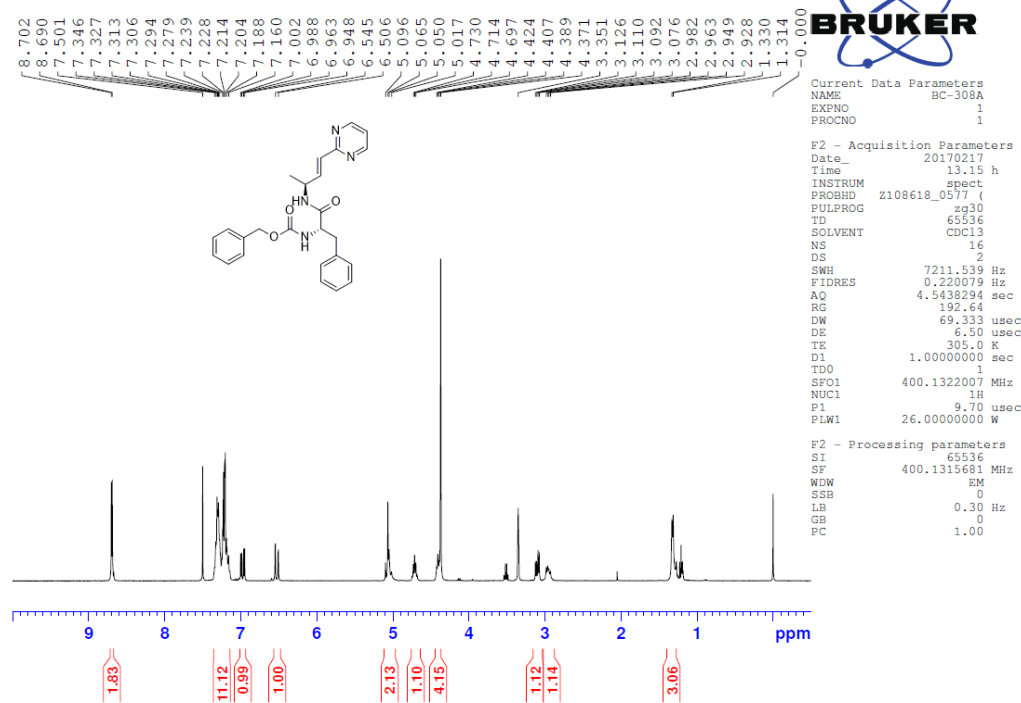
Current Data Parameters
 NAME BC-415A
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171113
 Time 10.48 h
 INSTRUM spect
 PROBHD Z108618_0577 (
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 127
 DS 4
 SWH 22050.024 Hz
 FIDRES 0.673182 Hz
 AQ 1.4854827 sec
 RG 192.64
 DW 22.667 usec
 DE 6.50 usec
 TE 305.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1
 SFO1 100.6233333 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 50.00000000 W
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 90.00 usec
 PLW2 26.00000000 W
 PLW12 0.38839999 W
 PLW13 0.19536000 W

F2 - Processing parameters
 SI 32768
 SF 100.6127600 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

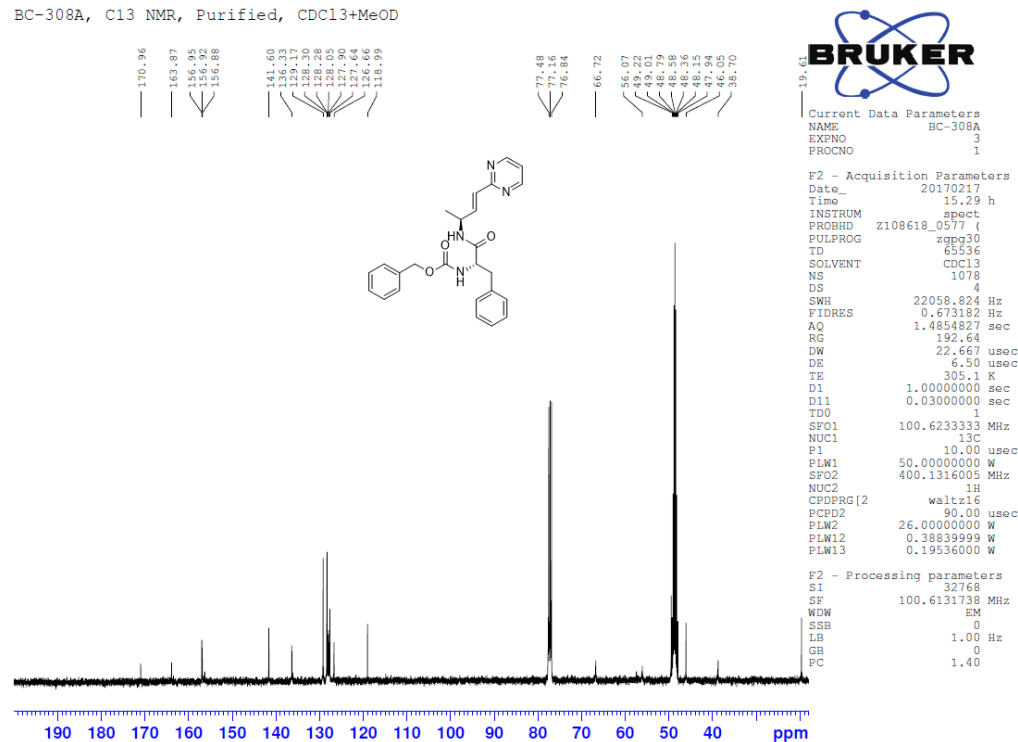
¹H NMR spectrum of compound 10

BC-308A, ¹H NMR, Purified, CDCl₃+MeOD



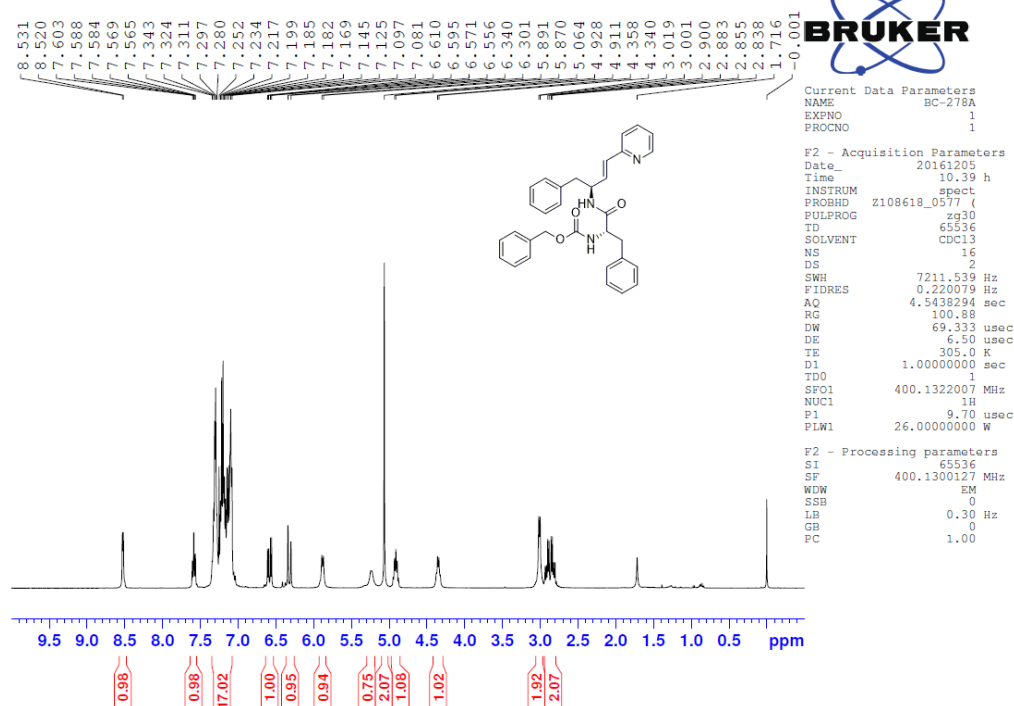
¹³C NMR spectrum of compound 10

BC-308A, ¹³C NMR, Purified, CDCl₃+MeOD



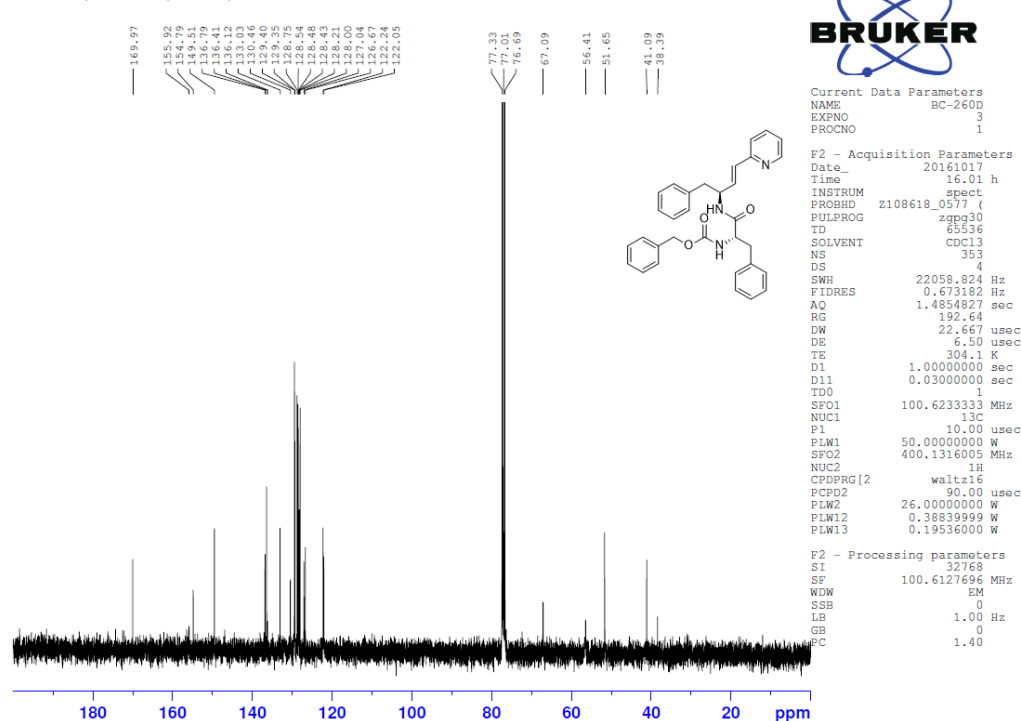
¹H NMR spectrum of compound 11

BC-260D (BC-278A), ¹H NMR, Purified, CDCl₃



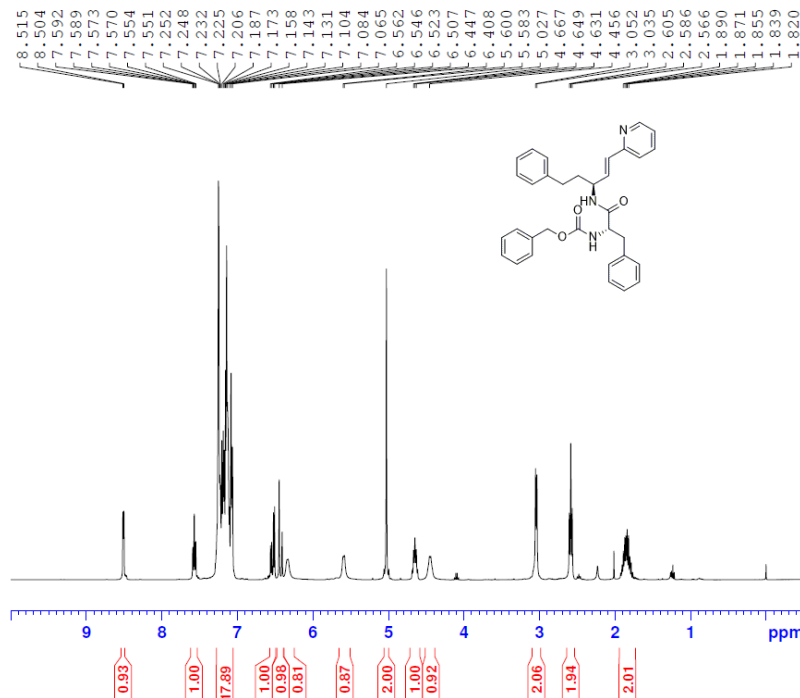
¹³C NMR spectrum of compound 11

BC-260D, ¹³C NMR, Pure, CDCl₃



¹H NMR spectrum of compound 13

BC-380, ¹H NMR, CDCl₃



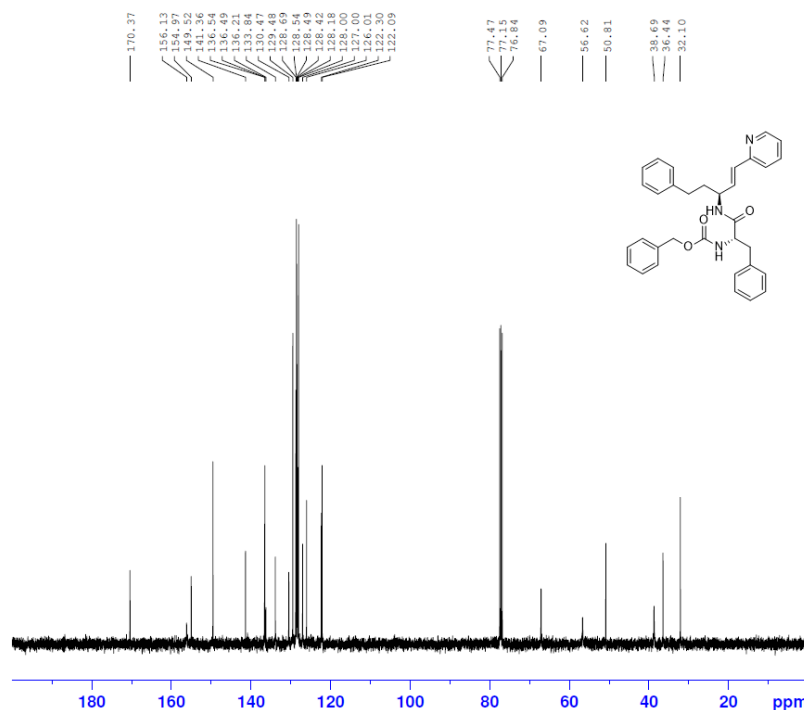
Current Data Parameters
 NAME BC-380
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20170710
 Time 12.14 h
 INSTRUM spect
 PROBHD Z108618_0577 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4807.692 Hz
 FIDRES 0.146719 Hz
 AQ 6.8157439 sec
 RG 24.52
 DW 104.000 usec
 DE 6.50 usec
 TE 305.0 K
 D1 1.00000000 sec
 TDO 1
 SFO1 400.1322007 MHz
 NUC1 1H
 P1 9.70 usec
 PLW1 26.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300207 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR spectrum of compound 13

BC-380, ¹³C NMR, CDCl₃



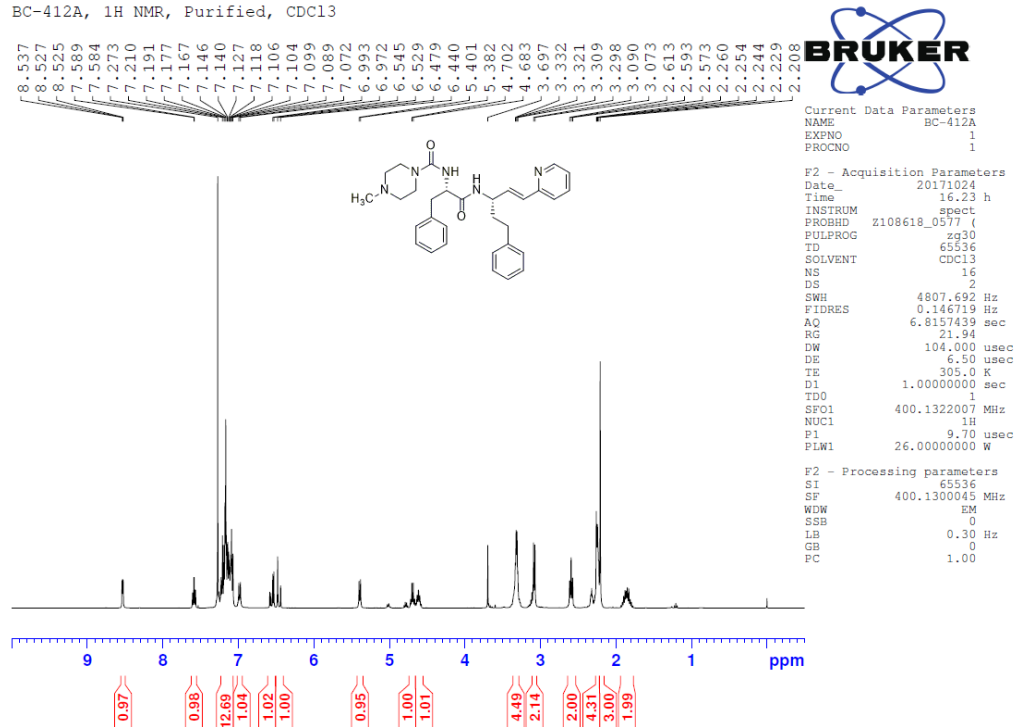
Current Data Parameters
 NAME BC-380
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20170710
 Time 12.18 h
 INSTRUM spect
 PROBHD Z108618_0577 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 41
 DS 4
 SWH 22058.824 Hz
 FIDRES 0.673182 Hz
 AQ 1.4854827 sec
 RG 192.64
 DW 22.667 usec
 DE 6.50 usec
 TE 305.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1
 SFO1 100.6233333 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 50.00000000 W
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG2 waltz16
 ECPD2 90.00 usec
 PLW2 26.00000000 W
 PLW12 0.38839999 W
 PLW13 0.19536000 W

F2 - Processing parameters
 SI 32768
 SF 100.6127666 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

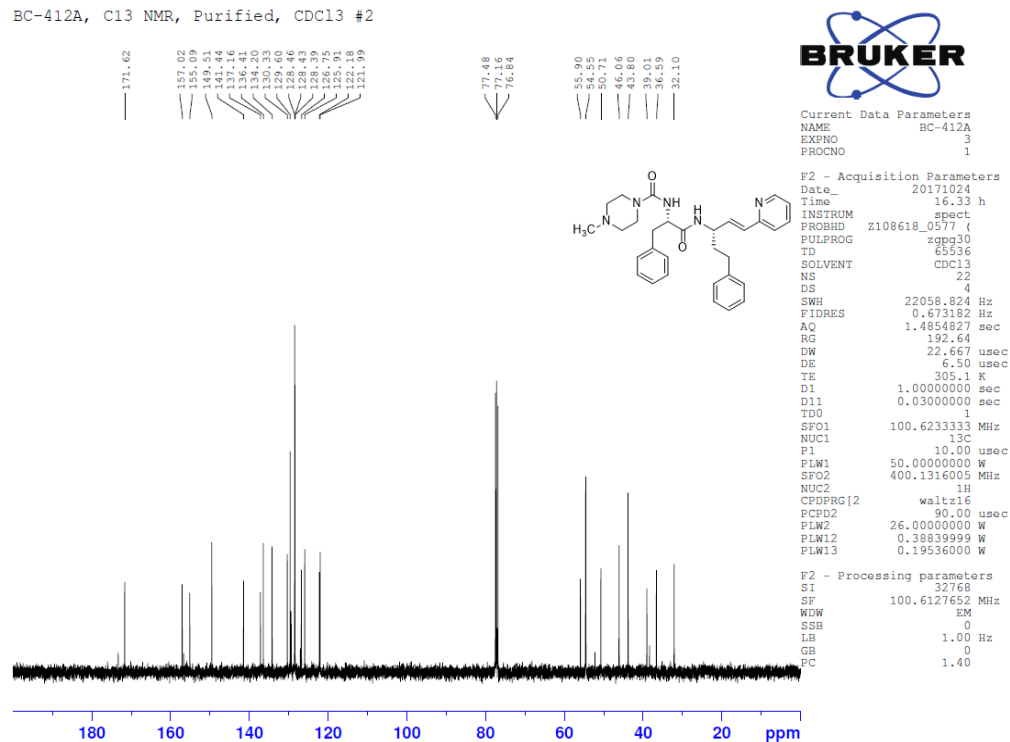
¹H NMR spectrum of compound 14

BC-412A, ¹H NMR, Purified, CDCl₃



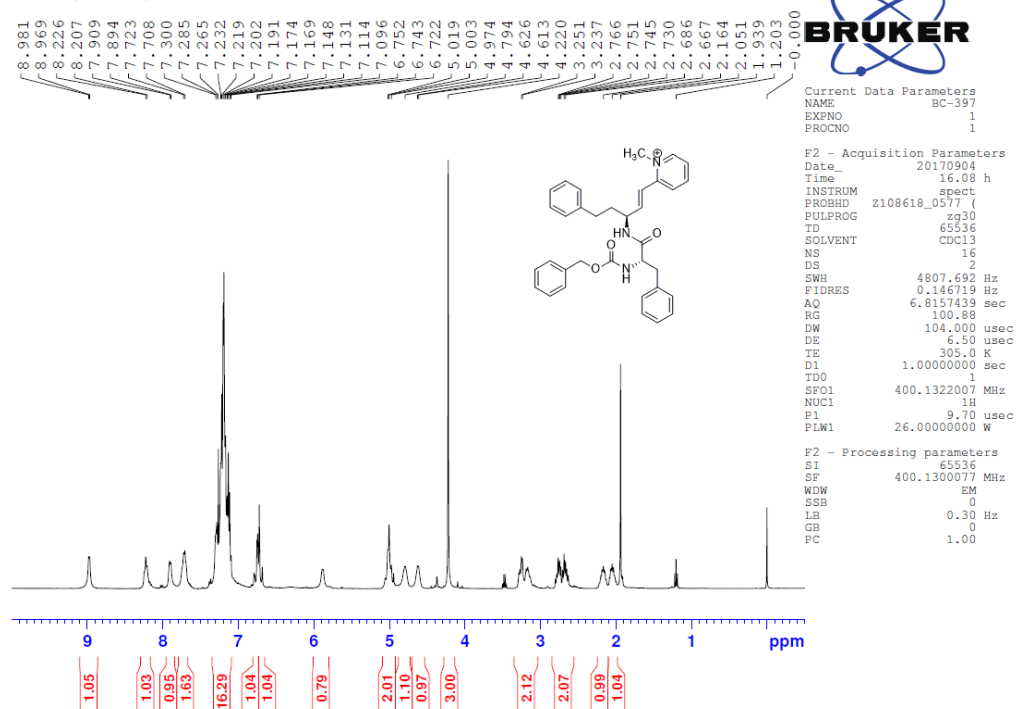
¹³C NMR spectrum of compound 14

BC-412A, ¹³C NMR, Purified, CDCl₃ #2

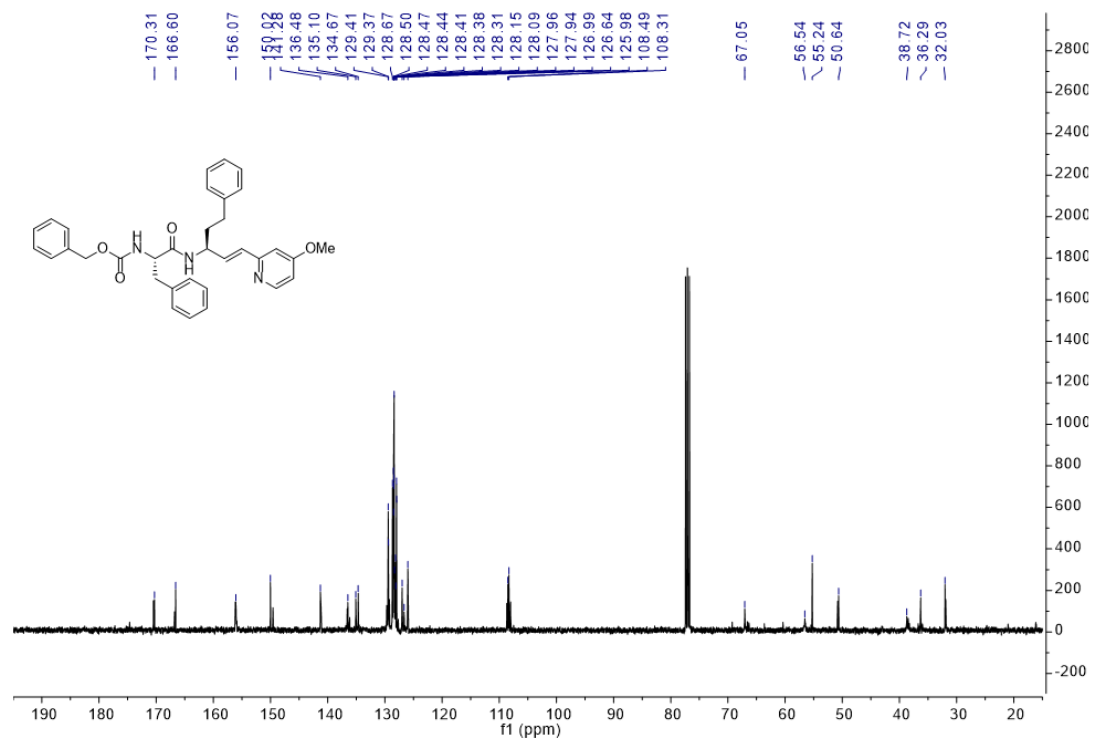
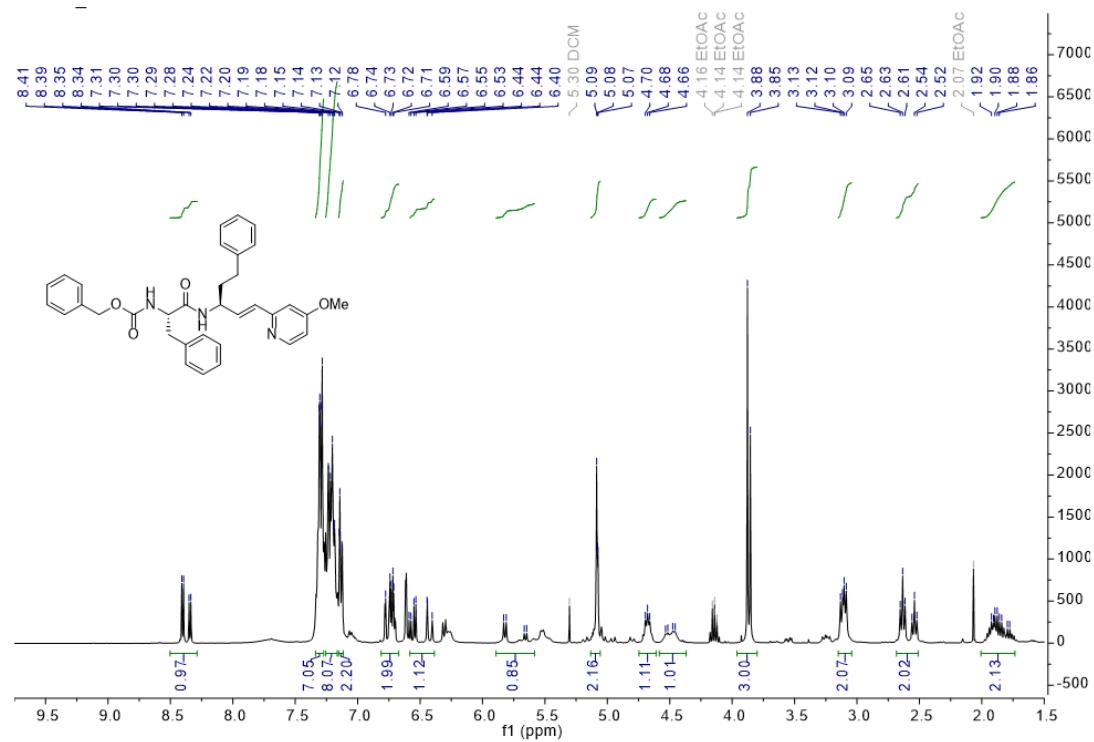


¹H NMR spectrum of compound 15

BC-397, ¹H NMR, CDCl₃

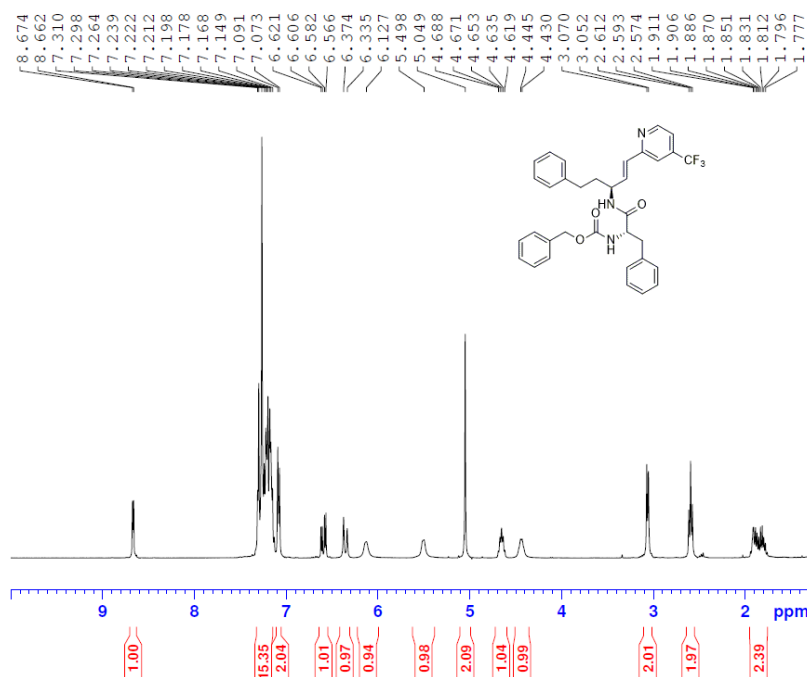


¹H NMR spectrum of compound **16**



¹H NMR spectrum of compound 17

BC-582A, ¹H NMR, Purified, CDCl₃
PROTON_TAMU CDCl₃ /data bala 48



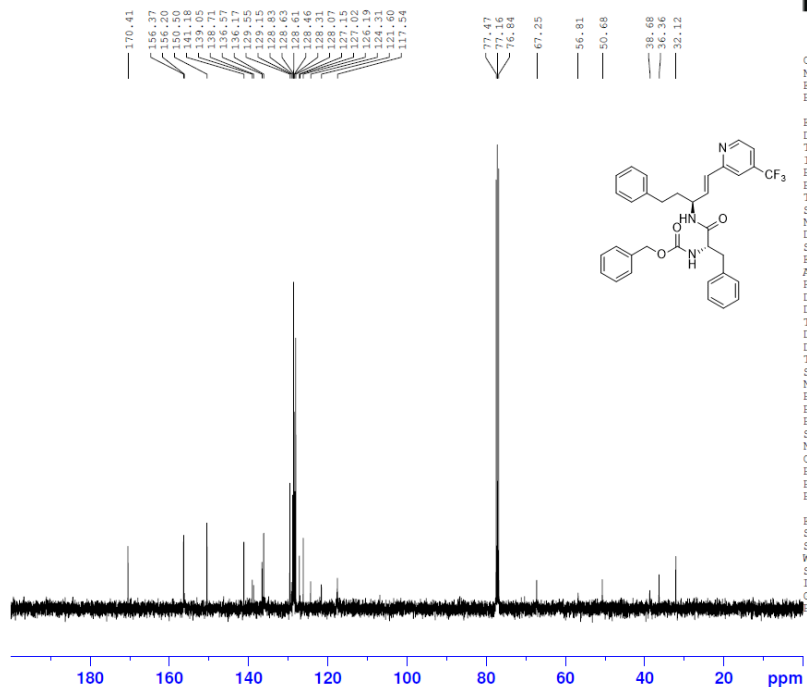
Current Data Parameters
NAME BC-582A
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190924
Time 15.38 h
INSTRUM spect
PROBHD Z108618_0577 ()
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 2
DS 2
SWH 8012.820 Hz
FIDRES 0.244532 Hz
AQ 4.0894465 sec
RG 31.82
DW 62.400 usec
DE 6.50 usec
TE 305.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1324708 MHz
NUC1 1H
P0 3.23 usec
P1 9.70 usec
PLW1 26.00000000 W

F2 - Processing parameters
SI 65536
SF 400.1300177 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR spectrum of compound 17

BC-582A, ¹³C NMR, Purified, CDCl₃
CARBON_TAMU CDCl₃ /data bala 48



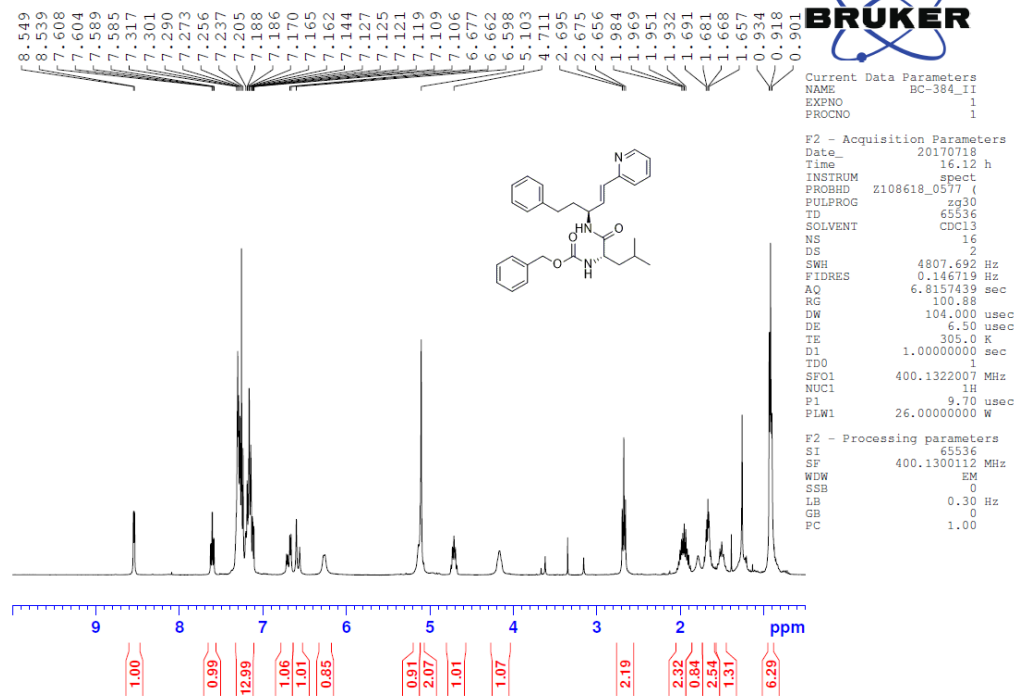
Current Data Parameters
NAME BC-582A
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190924
Time 15.43 h
INSTRUM spect
PROBHD Z108618_0577 ()
PULPROG zg1g30
TD 65536
SOLVENT CDCl₃
NS 54
DS 4
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 1.3631488 sec
RG 192.64
DW 20.800 usec
DE 6.50 usec
TE 305.0 K
D1 3.00000000 sec
D11 0.03000000 sec
TD0 1
SF01 100.6228298 MHz
NUC1 13C
P0 3.33 usec
P1 10.00 usec
PLW1 50.00000000 W
SF02 400.1316005 MHz
NUC2 1H
CPDPRG2 waltz65
PCPD2 90.00 usec
PLW2 26.00000000 W
PLW12 0.38839999 W

F2 - Processing parameters
SI 32768
SF 100.6127593 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

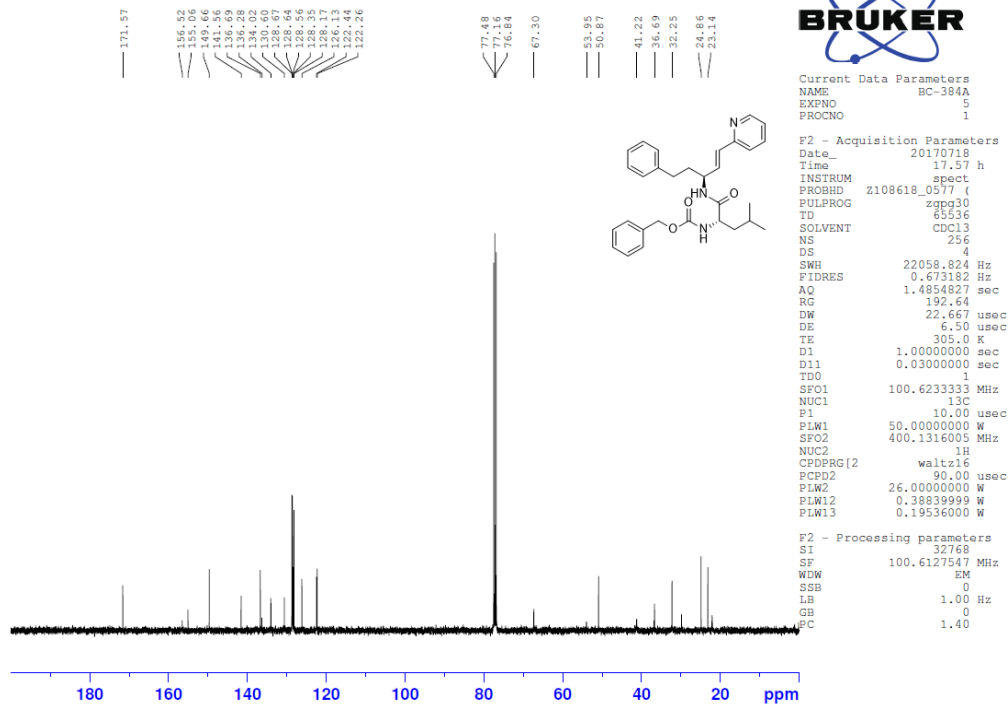
¹H NMR spectrum of compound 18

BC-384_II, purified, ¹H NMR, CDCl₃

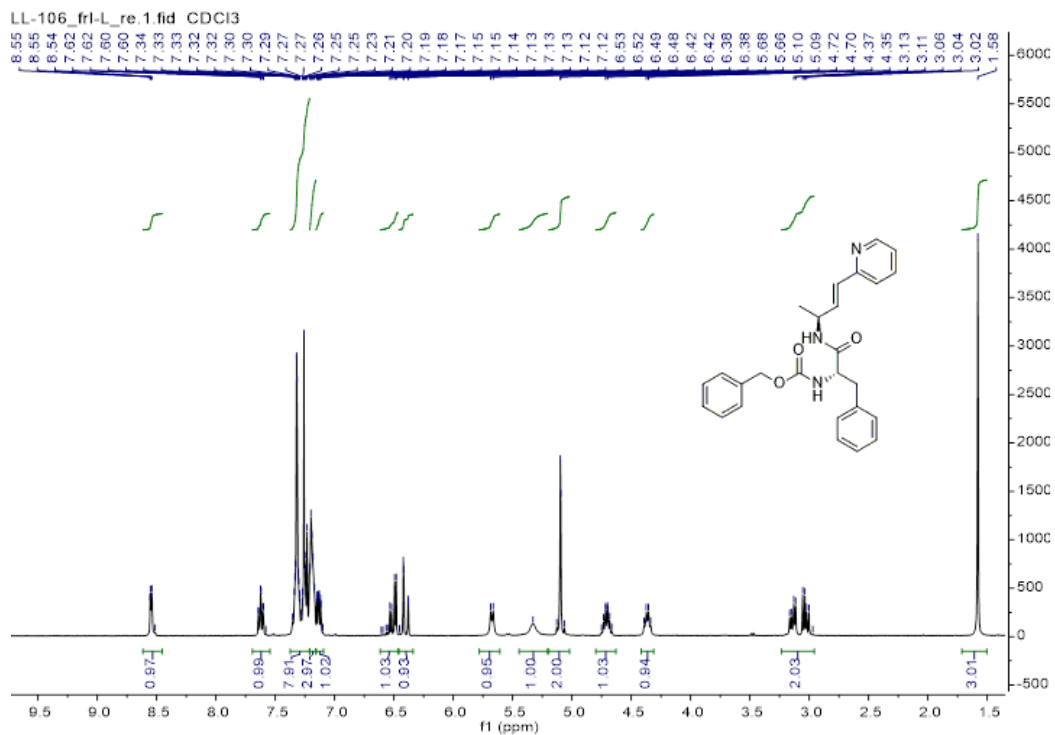


¹³C NMR spectrum of compound 18

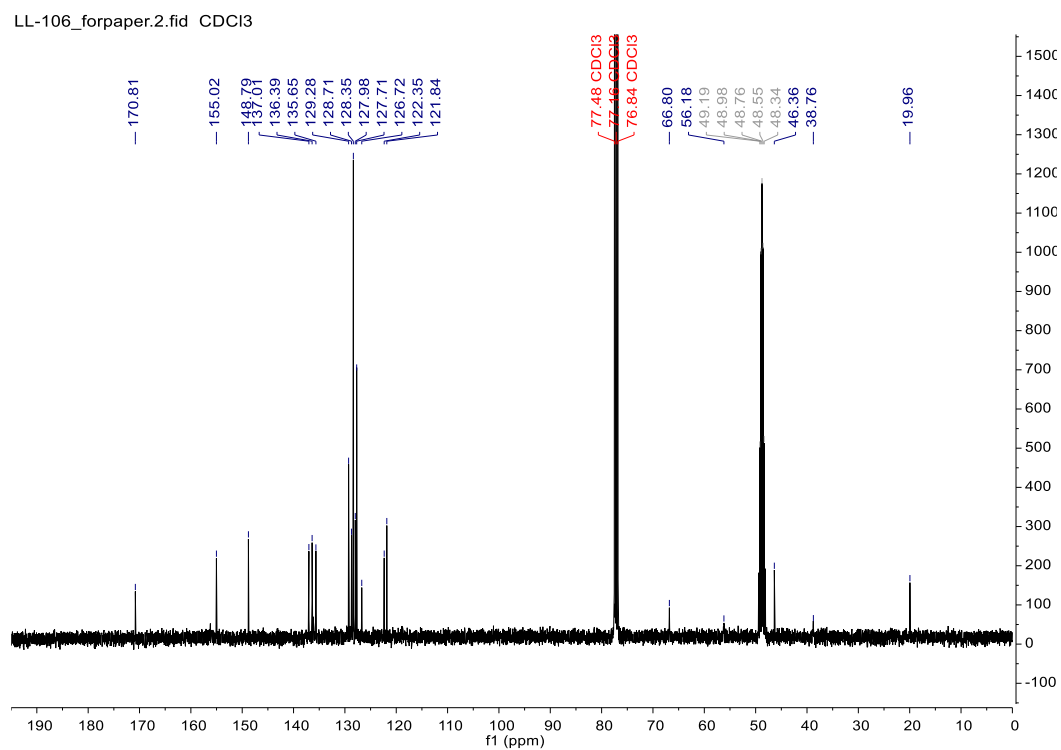
BC-384A, #3, purified, ¹³C NMR, CDCl₃



¹H NMR spectrum of compound **19**

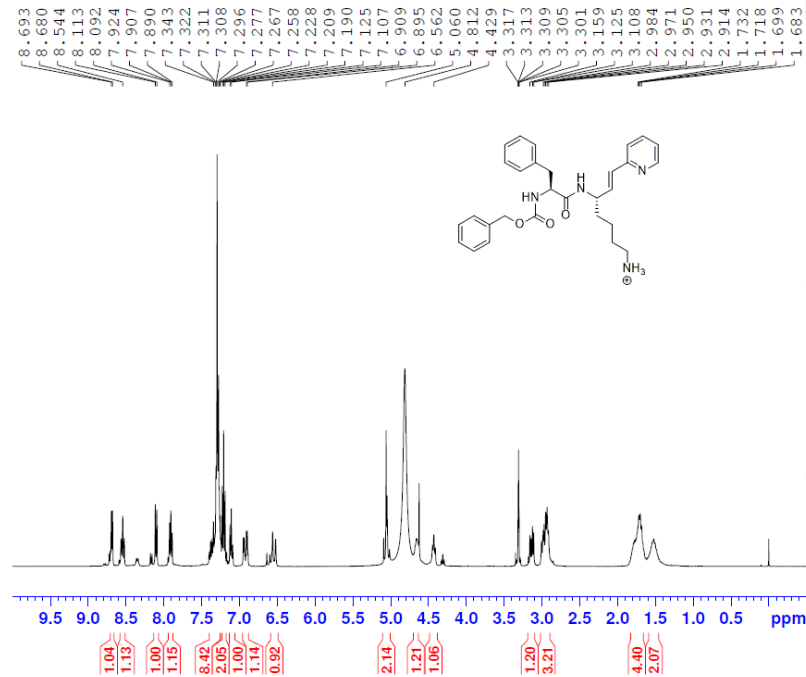


¹³C NMR spectrum of compound **19**



¹H NMR spectrum of compound 20

BC-566, HPLC purified, MeOD, ¹H NMR
 PROTON_TAMU MeOD /data bala 4



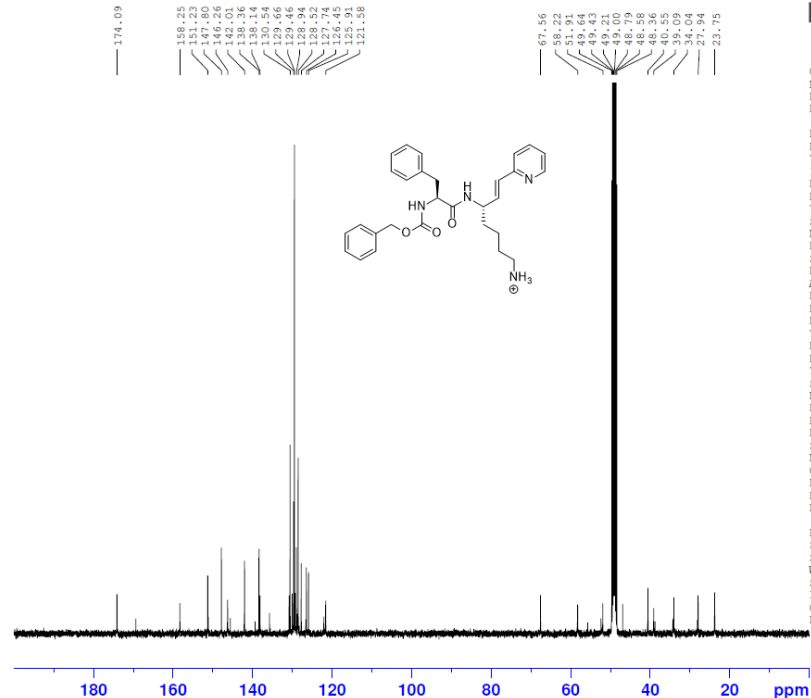
Current Data Parameters
 NAME BC-566
 EXPO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190805
 Time_ 12.35 h
 INSTRUM spect
 PROBHD Z108618_0577 ()
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.244532 Hz
 AQ 4.0894465 sec
 RG 55.4
 DW 62.400 usec
 DE 6.50 usec
 TE 305.0 K
 D1 1.00000000 sec
 TDO 1
 SFO1 400.1324708 MHz
 NUC1 1H
 P0 3.23 usec
 P1 9.70 usec
 PLW1 26.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300079 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR spectrum of compound 20

BC-566, HPLC purified, MeOD, ¹³C NMR
 CARBON_TAMU MeOD /data bala 4

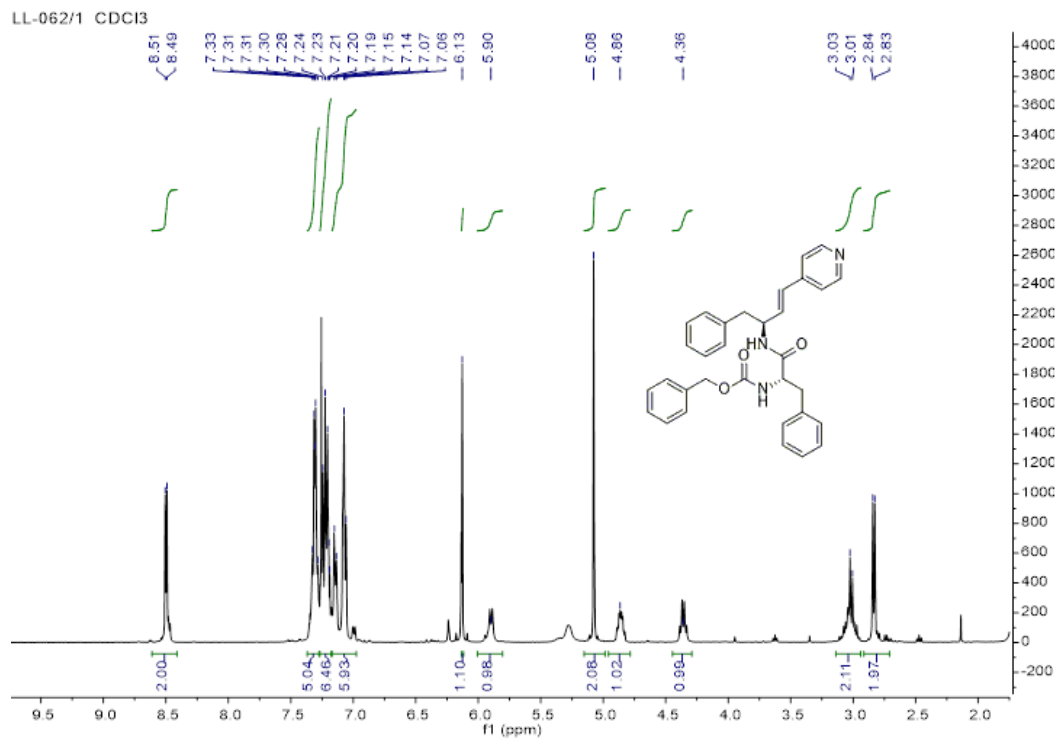


Current Data Parameters
 NAME BC-566
 EXPO 3
 PROCNO 1

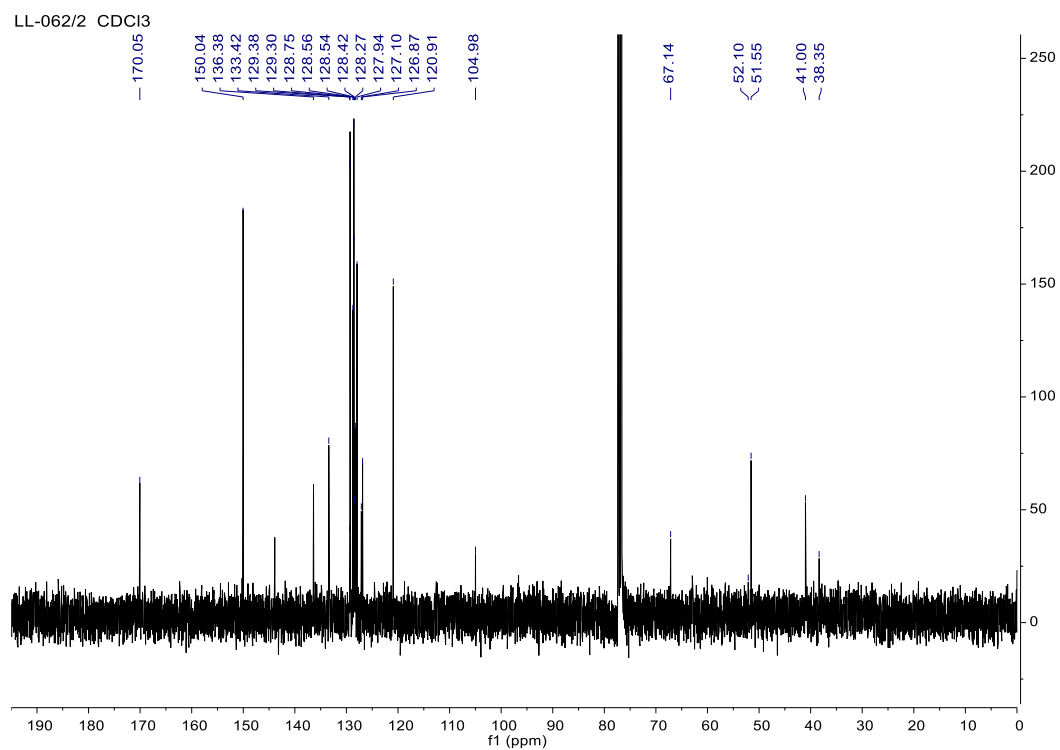
F2 - Acquisition Parameters
 Date_ 20190806
 Time_ 0.04 h
 INSTRUM spect
 PROBHD Z108618_0577 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOD
 NS 2444
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.733596 Hz
 AQ 1.3631488 sec
 RG 192.64
 DW 20.800 usec
 DE 6.50 usec
 TE 305.0 K
 D1 3.00000000 sec
 D11 0.03000000 sec
 TDO 1
 SFO1 100.6228298 MHz
 NUC1 13C
 P0 3.33 usec
 P1 10.00 usec
 PLW1 50.00000000 W
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG2 waltz165
 PCPD2 90.00 usec
 PLN2 26.00000000 W
 PLW12 0.38839999 W

F2 - Processing parameters
 SI 32768
 SF 100.6126322 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR spectrum of compound 21

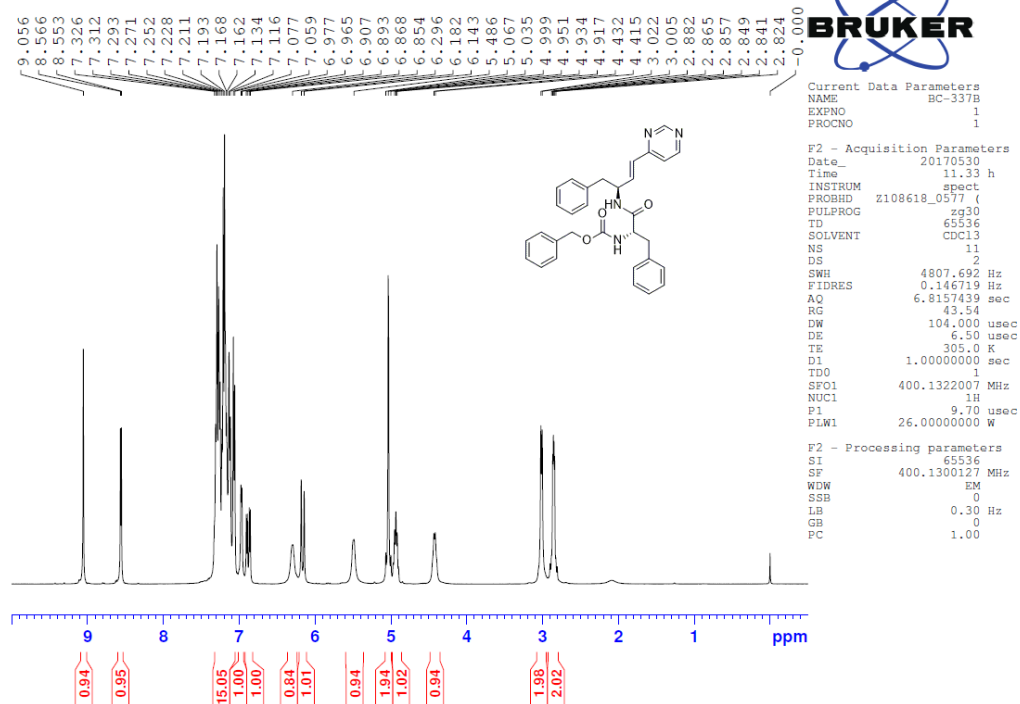


¹³C NMR spectrum of compound 21



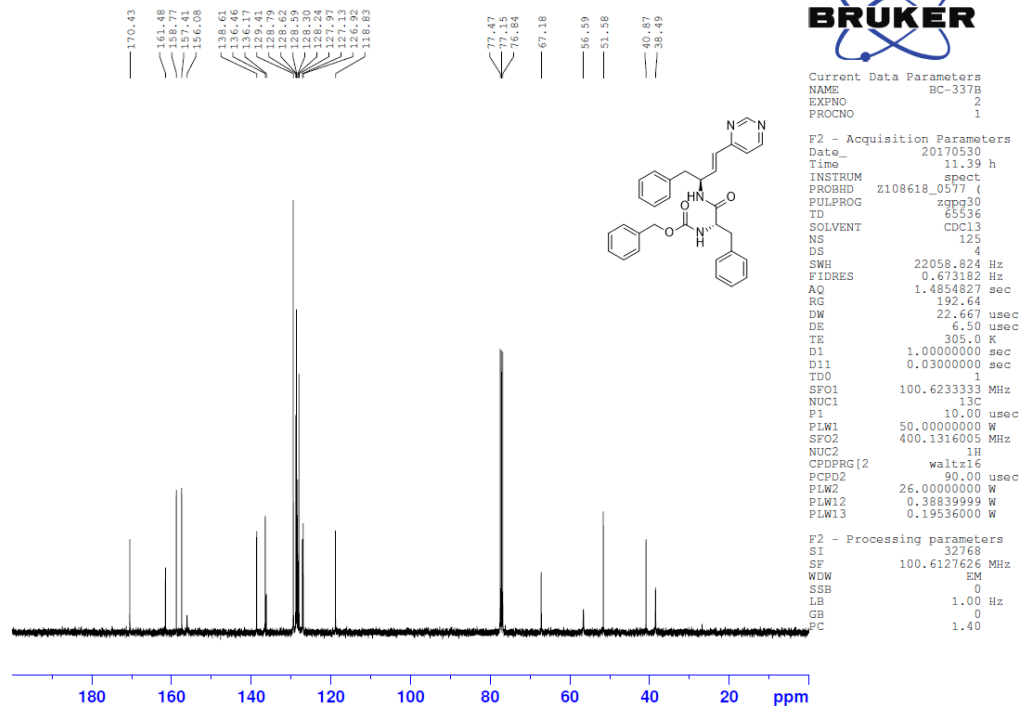
¹H NMR spectrum of compound 23

BC-337B, ¹H NMR, Purified, CDCl₃



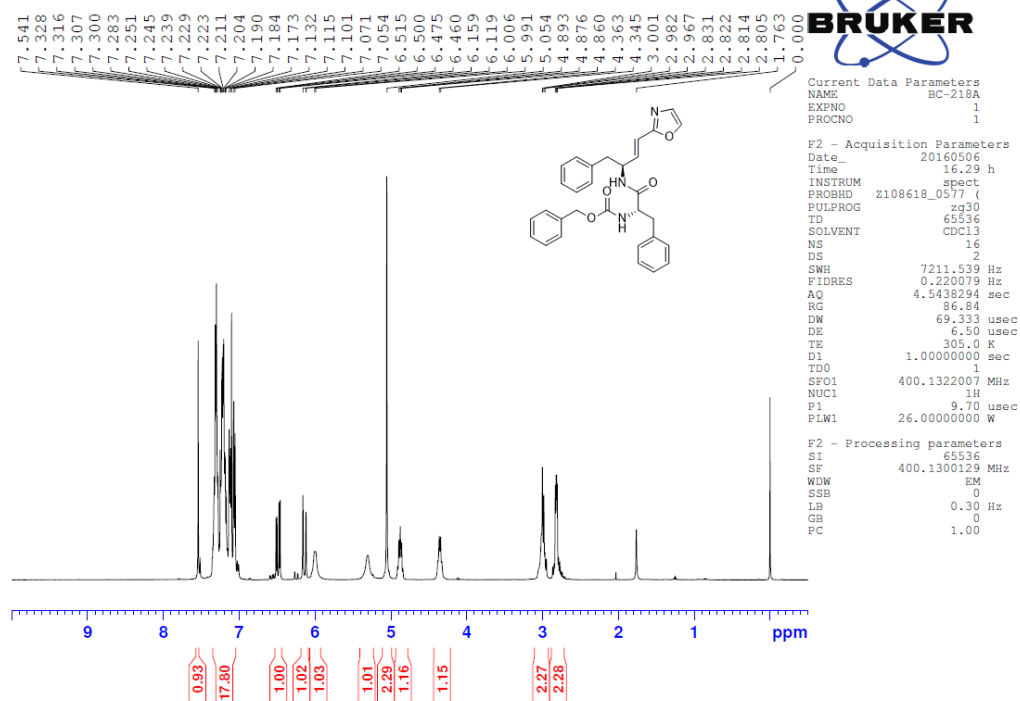
¹³C NMR spectrum of compound 23

BC-337B, ¹³C NMR, Purified, CDCl₃



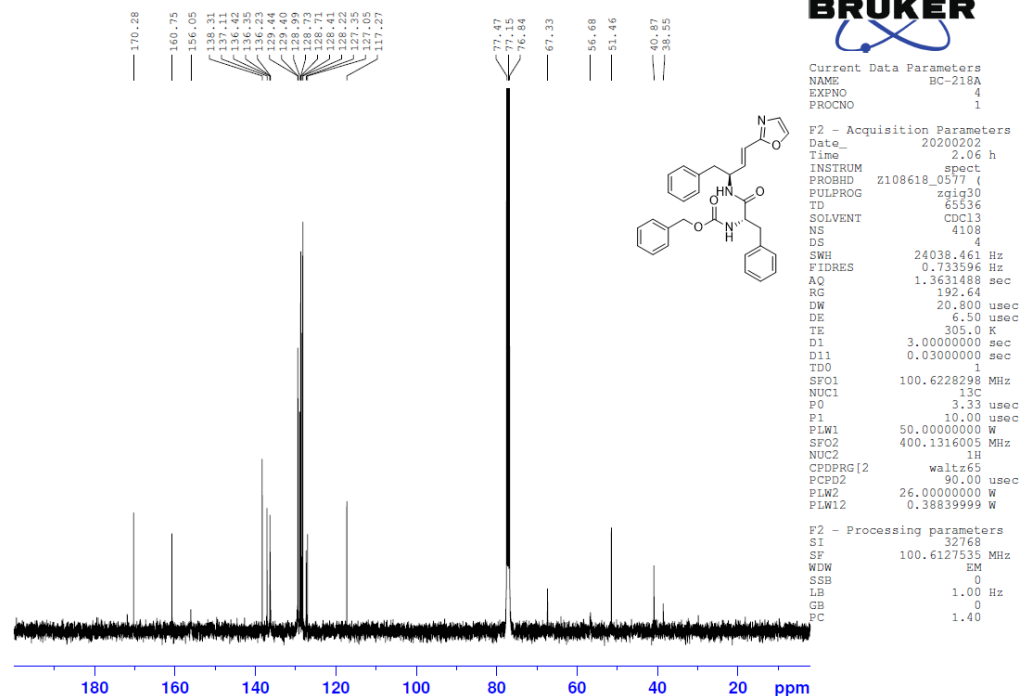
¹H NMR spectrum of compound 24

BC-218A Purified-crystallized 1HNMR, CDCl3



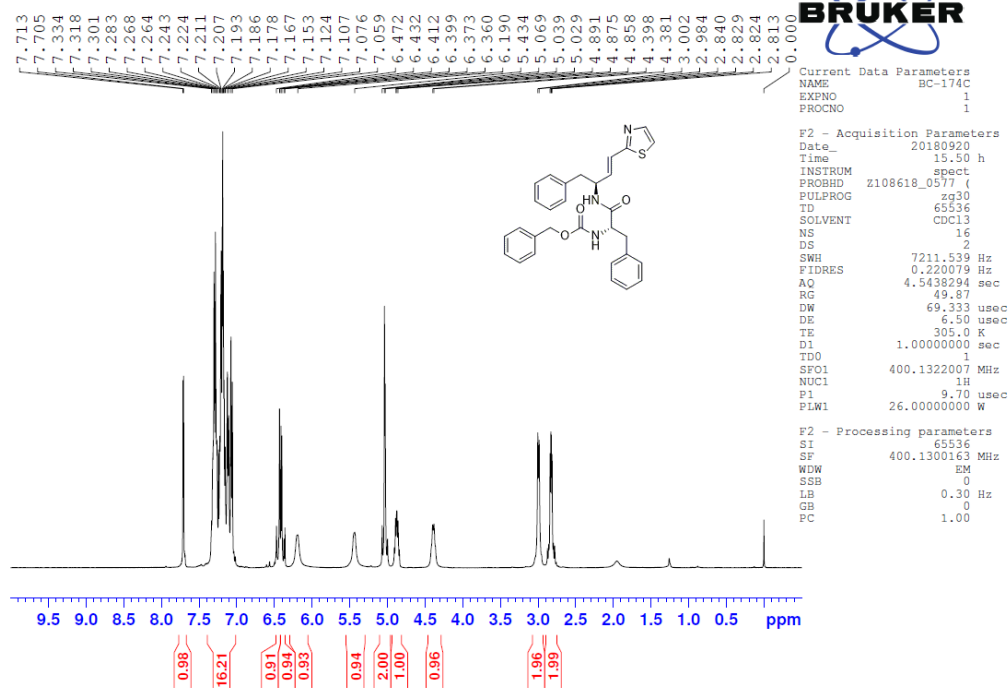
¹³C NMR spectrum of compound 24

BC-218A, C13 NMR, CDCl3
 CARBON_TAMU CDCl3 /data bala 14



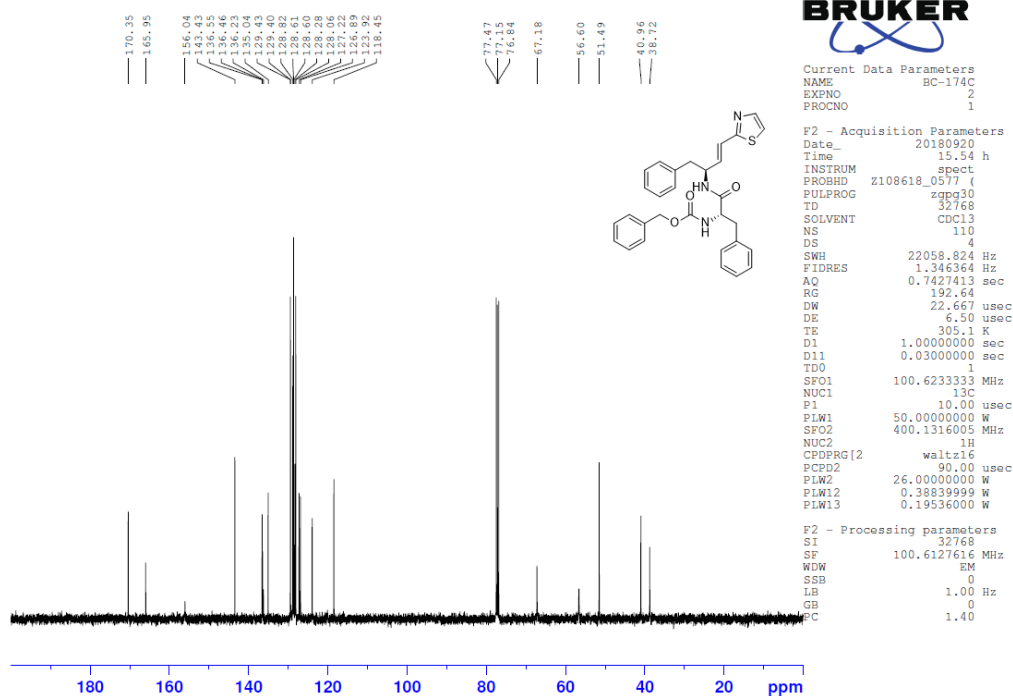
¹H NMR spectrum of compound 25

BC-174C, ¹H NMR, HPLC Purified, CDCl₃



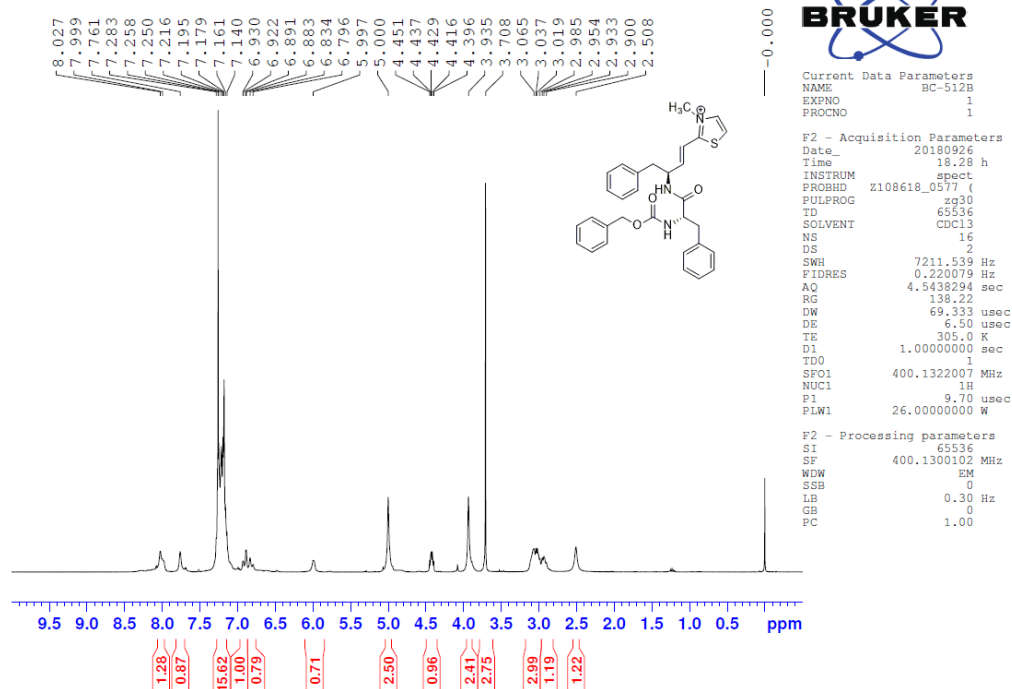
¹³C NMR spectrum of compound 25

BC-174C, ¹³C NMR, HPLC Purified, CDCl₃



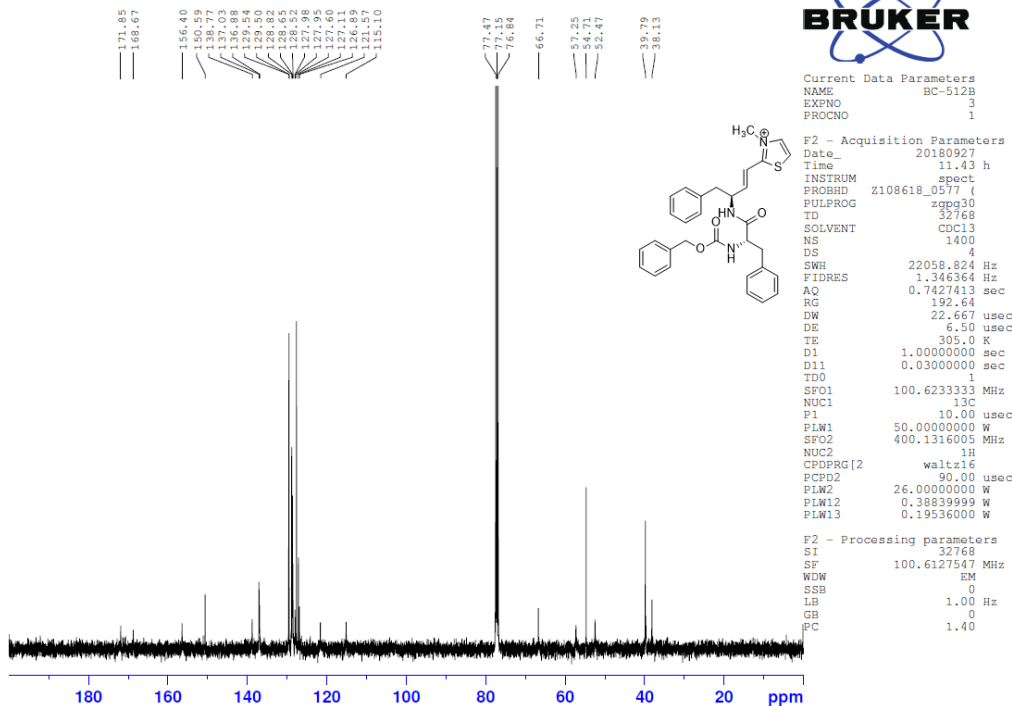
¹H NMR spectrum of compound 26

BC-512B, ¹H NMR, solid after 3 ether washes, CDCl₃



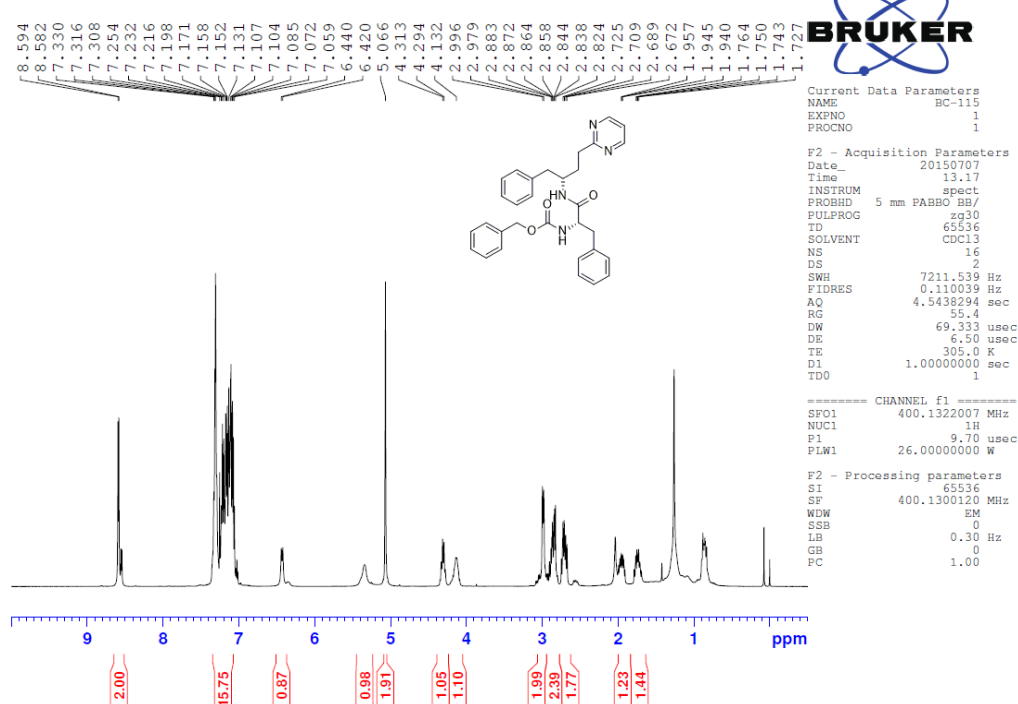
¹³C NMR spectrum of compound 26

BC-512B, ¹³C NMR, solid after 3 ether washes, CDCl₃



¹H NMR spectrum of compound 27

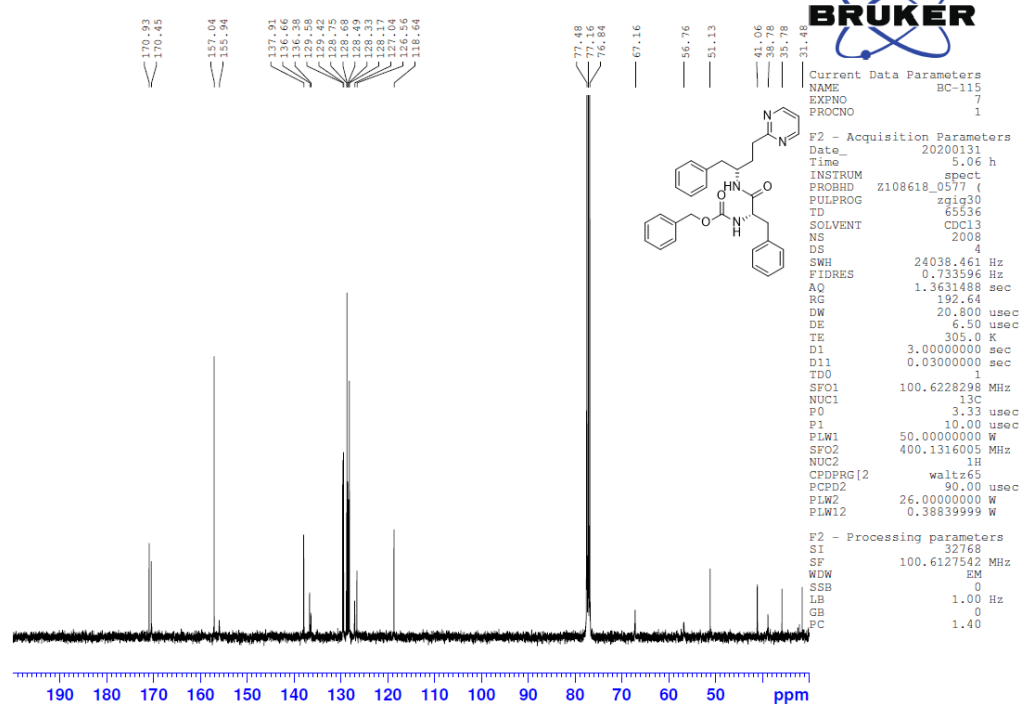
BC-115 1HNMR Purified CDCl₃



¹³C NMR spectrum of compound 27

BC-115, C13 NMR, CDCl₃

CARBON_TAMU CDCl₃ /data bala 24



Part 8. Unpublished Data

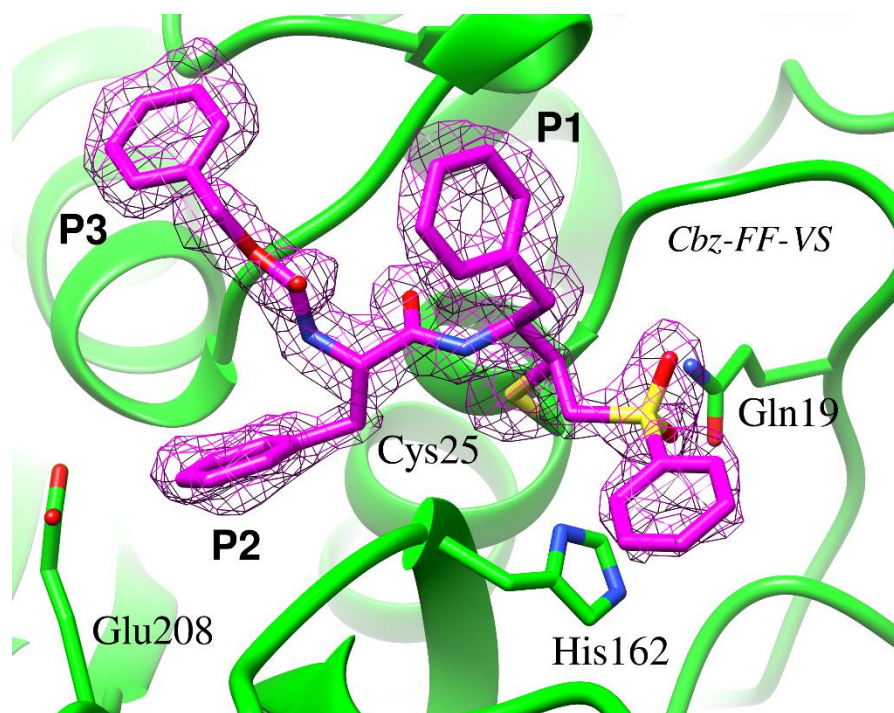


Figure S4. Crystal structure of cruzain bound to compound **1**. A covalent bond is evidently formed between the sulfur of Cys₂₅ and the β -carbon of the former olefin bond (Tang, *et al.*).