SUPPORTING INFORMATION

Characterization of the structure and interactions of P450 BM3 using hybrid mass spectrometry approaches

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Figure S1. Native MS of full-length P450 BM3 and its component domains in ligand-free and ligand-bound states. Panel A: The native MS spectrum of full-length P450 BM3 shows multiple charge states as well as the presence of dimeric (major component) and monomeric (minor component) protein. **Panel B**: The native MS spectrum of the heme domain protein shows only three prevalent charge states. **Panel C**: The MS spectrum of the heme domain does not change significantly upon association with the tight binding BM3 substrate NPG. **Panel D**: The MS spectrum of the CPR domain shows a number of different charge states. **Panel E**: The MS spectrum of the CPR. Multiple species are observed, corresponding to 1, 2, 3 and 4 molecules of NADP⁺ binding to the CPR domain.

Table S1. Experimentally and theoretically derived collision cross-section (CCS) values for full-length P450 BM3 and its domains. CCS values were determined for the full-length, dimeric P450 BM3 protein and its component heme and reductase (CPR) domains using IM-MS with helium as the carrier gas. In addition, CCS values were determined for ligand-bound domains. Theoretical CCS values were calculated using crystallographic structures (1BU7, 4DQK and 1BVY) using the IMoS suite with helium as the carrier gas. These calculations gave CCS values of 39.1 nm² for the heme domain (13% difference to the experimental data) and 46.1 nm² for the reductase domain model (23% difference to the experimental data).

Experimental			Theoretical	Expected mass	Measured mass
Protein	CCS (nm ²)	STDEV	ImoS (nm ²)	Da	Da
Full-length protein	99.66	1.21	-	239889.20	244915.91 ± 76.08
Heme domain	34.56	0.17	39.1	56147.08	56637.51 ± 0.71
Heme domain + NPG	34.71	0.24	-	56406.58	56952.95 ± 1.56
CPR domain	37.53	1.08	46.1	64145.98	64536.33 ± 30.81
CPR domain + 1 NADP ⁺	37.05	0.38	-	64889.38	65229.05 ± 11.67
CPR domain + 2 NADP ⁺	37.39	0.15	-	65632.78	66054.80 ± 2.79
CPR domain + 3 NADP ⁺	38.03	0.48	-	66376.18	66809.45 ± 14.95
CPR domain + 4 NADP ⁺	38.31	0.43	-	67119.58	67563.77 ± 20.17



Figure S2. Relative deuterium uptake plots for ligand-free and ligand-bound full-length P450 BM3 dimeric protein. Using a LEAP Technologies dual-armed robot, accurate deuterium incubations could be accomplished for HDX-MS analysis. Observing the deuterium uptake from 1 minute to 180 minutes for each peptide fragment allows determination of the solvent accessibility of the protein. Four states are compared above; ligand-free (red), NADP⁺-bound (blue), NPG-bound (green) and NADP⁺/NPG-bound (purple). Plots were generated using Waters DynamX software. In Panel A a small segment of the protein exhibits very poor deuterium uptake over time. In Panel B a longer segment of the protein exhibits poor deuterium uptake over time. However, this segment begins with higher deuterium levels than Panel A. In Panels C and D deuterium uptake increases over time. However, in Panel C there is an observable difference between different ligand-bound states.



Figure S3. The coverage maps for the ligand-free P450 BM3 heme and CPR domains. Panel A: The heme domain HDX-MS data show 91.5% coverage with 3.18 redundancy. **Panel B:** The CPR domain data show 89.8% coverage with 3.75 redundancy.



Figure S4. The coverage maps for the ligand-bound, full-length P450 BM3 dimeric protein. All four states of the protein (ligand-free, NADP⁺-bound, NPG-bound and NADP⁺/NPG-bound)

show 99.1% coverage and 8.51 redundancy.

MGSSHHHHHH	SSGLVPRGSH	MTIKEMPQPK	TFGELKNLPL	LN <mark>TDKPVQAL</mark>	M <mark>KI</mark> ADELGEI
FKFEAPGRVT	RYLSSQRLIK	EACD <mark>ESRF</mark> DK	NLSQA <mark>LKF</mark> V <mark>R</mark>	DFAGDGLFTS	WTHEKNWKKA
HNILLPSFSQ	QAMKGYHAMM	VDIAVQLVQK	WERL <mark>NADEHI</mark>	<mark>EVPEDM</mark> TRLT	LD <mark>TIGLCGF</mark> N
YRFNSFY <mark>RDQ</mark>	PHPFITSMVR	<mark>aldeam</mark> nklq	RANPDDPAYD	ENKRQFQEDI	KVMNDLVDKI
<mark>IAD</mark> RKASGEQ	SDDLLTHMLN	GKDPETGEPL	DDEN <mark>IRYQII</mark>	<mark>TF</mark> LIAGHETT	SGLLS <mark>FAL</mark> Y <mark>F</mark>
LVKNPHVLQK	AAEEAARVLV	DPVPSYKQVK	<mark>QLKYVG</mark> MVLN	EALRLWPTAP	AFSLYAKEDT
VLGGEYPLEK	GDELMVLIP <mark>Q</mark>	LHRDKTIWGD	<mark>D</mark> VEEFRPERF	ENPSAIPQHA	FKPFGN <mark>GQRA</mark>
<mark>CIG</mark> QQFALHE	ATLVLGMMLK	HFDF <mark>E</mark> DHTNY	EL <mark>DIKETLTL</mark>	<mark>KPEGFV</mark> VKAK	SKKIPLGGIP
SPSTEQSAKK	VRKKAENAHN	TPLLV <mark>LYGSN</mark>	<mark>MGTAEGTA</mark> RD	LADIAMS <mark>KGF</mark>	<mark>apqvatl</mark> dsh
AGNLPREGAV	LI <mark>VTASYNG</mark> H	<mark>ppdnakq</mark> fvd	WLDQASADE <mark>V</mark>	KGVRYSVF <mark>GC</mark>	<mark>GDKNW</mark> ATTY <mark>C</mark>
KVPAFIDETL	AA <mark>K</mark> GAENIA <mark>D</mark>	<mark>RGEADASD</mark> DF	EGTY <mark>EEWREH</mark>	<mark>MWSDVAA</mark> YFN	LDIENSEDNK
STLSLQFVD <mark>S</mark>	<mark>aad</mark> mplakmh	GAFSTNVVAS	KELQQPGSAR	STRHLEIELP	KEASYQEGDH
LGVIPRNY <mark>EG</mark>	IVNRVTARFG	LDASQQIRL <mark>E</mark>	AE <mark>EEKLAHLP</mark>	<mark>laktvsv</mark> ee <mark>l</mark>	<mark>lqyv</mark> elqdpv
T <mark>RTQLRA</mark> MAA	KT <mark>VCPPHKVE</mark>	<mark>l</mark> eal <mark>l</mark> ekqay	KE <mark>QVLAKRLT</mark>	<mark>MLELL</mark> EKYPA	CEM <mark>KFSEFIA</mark>
<mark>llpsirpry</mark> y	SISSSPRVDE	KQ <mark>ASITVSVV</mark>	SGEAWSGYGE	YKGIASNYLA	ELQEGDTITC
FISTPQSEFT	LPKDPETPLI	MVGPGTGVAP	FRGFVQARKQ	LKEQGQSLGE	AHLY <mark>FG</mark> CRSP
HEDYLYQEEL	E <mark>NAQSEGIIT</mark>	<mark>L</mark> HTAFS <mark>RMPN</mark>	<mark>QPKT<mark>YVQH</mark>VM</mark>	EQDGKKLIEL	LDQGAHFYIC
GDGSQMAPAV	EATLMKSYAD	VHQVSEADAR	LWLQQLEEKG	RYAKDVWAG*	

Figure S5. The P450 BM3 residues showing the highest degree of change when comparing full-length P450 BM3 with its isolated domains. The residues are coloured red or cyan to indicate areas of the greatest deshielding or shielding respectively (\pm 5% deuterium uptake), when subtracting the isolated domains from the full-length P450 BM3 protein. These residues correspond to conformational changes causing alterations in solvent accessibility. In particular, the residues highlighted in cyan correspond to the sites hypothesized to be involved with dimerization. A His-tag was used to facilitate protein purification and is shown in green. Residues with no coverage are not coloured.

A

MGSSHHHHHH	SSGLVPRGSH	MTIKEMPQPK	TFGELKNLPL	LNTDKPVQAL	MKIADELGEI
FKFEAPGRVT	RYLSSQRLIK	EACDESRFDK	NLSQALKFVR	DFAGDGLFTS	WTHEKNWKKA
HNILLPSFSQ	QAMKGYHAMM	VDIAVQLVQK	WERLNADEHI	EVPEDMTRL <mark>T</mark>	LDTIGLCGFN
YRFNSFYRDQ	PHPFITSMVR	ALDEAMNKLQ	RANPDDPAYD	ENKRQFQEDI	KVMNDLVDKI
IADRKASGEQ	SDDLLTHMLN	GKDPETGEPL	DDENIRYQII	TFLIAGHETT	SGLLSFALYF
LVKNPHVLQK	AAEEAARVLV	DPVPSYKQVK	QLKYVGMVLN	EALRLWPTAP	AFSLYAKEDT
VLGGEYPLEK	GDELMVLIPQ	LHRDKTIWGD	DVEEFRPERF	ENPSAIPQHA	FKPFGNGQRA
CIGQQFALHE	ATLVLGMMLK	HFDFEDHTNY	ELDIKETLTL	KPEGFVVKAK	SKKIPLGGIP
SPSTEQSAKK	VRKKAENAHN	TPLLVLYGSN	MGTAEGTARD	LADIAMSKGF	APQVATLDSH
AGNLPREGAV	LIVTASYNGH	PPDNAKQFVD	WLDQASADEV	KGVRYSVFGC	GDKNWATTYQ
KVPAFIDETL	AAKGAENIAD	RGEADASDDF	EGTYEEWREH	MWSDVAAYFN	LDIENSEDNK
STLSLQFVDS	aadm <mark>p</mark> lakmh	GAFSTNVVAS	KELQQPGSAR	STRHLEIELP	KEASYQEGDH
LGVIPRNYEG	IVNRVTARFG	LDASQQ <mark>IRL</mark> E	<mark>AEE</mark> EKLAHLP	LAKTVSVEEL	LQYVELQDPV
TRTQLRAMAA	KTVCPPHKVE	LEALLEKQAY	KEQVLAKRLT	MLELLEKYPA	CEMKFSEFIA
LLPSIRPRYY	SISSSPRVDE	KQASITVSVV	SGEAWSGYGE	YKGIASNYLA	ELQEGDTITC
FISTPQSEFT	LPKDPETPLI	MVGPGTGVAP	FRGFVQARKQ	LKEQGQSLGE	AHLYFGCRSP
HEDYLYQEEL	ENAQSEGIIT	LHTAFSRMPN	QPKTYVQHVM	EQDGKKLIEL	LDQGAHFYIC
GDGSQM <mark>APAV</mark>	<mark>E</mark> ATLMKSYAD	VHQVSEADAR	LWLQQLEEKG	RYAKDVWAG*	

B

MGSSHHHHHH	SSGLVPRGSH	MTIK <mark>EMPQPK</mark>	TFGEL <mark>KNLPL</mark>	LNTDKPVQAL	MKIADELGEI
FKFEAPGRVT	RYLSS <mark>Q</mark> RLIK	EACDESRFDK	NLSQALKFV <mark>R</mark>	DFAGDGLF <mark>T</mark> S	W <mark>THEKNWKKA</mark>
HNILL _{PSFSQ}	QAMKGYHAMM	VDIAVQLV <mark>QK</mark>	WERLNADE <mark>H</mark> I	EVPEDMTRLT	LDTIGLCGFN
YRFNSFYRDQ	PHPFITSMVR	ALDEAMN <mark>KLQ</mark>	RANPDDPAYD	ENKRQFQEDI	KVMNDLVDKI
IADRKASGEQ	SDDLLTHMLN	GKDPETGEPL	DDENIRYQII	TFLIAGHETT	SGLLSFALY
LVKNPHVLQK	AAEEAARVLV	DPVPSYKQVK	<mark>QLKYVGM</mark> VLN	EAL <mark>RL</mark> WPTAP	AFS <mark>L</mark> YAKEDI
VLGGEYPLE <mark>K</mark>	GDELMVLIPQ	<mark>l</mark> hrdktiwGd	DVEEFRPERF	ENPSAIPQHA	FKPFGNGQRA
CIGQQFALHE	ATLVLGMMLK	HFDFEDHTNY	ELDIKETLTL	KPEGFVVKAK	SKKIPLGGIE
SPSTE <mark>Q</mark> SAKK	VRKKAENAHN	TPLLVLYGSN	MGTAEGTARD	LADIA <mark>M</mark> SKGF	<mark>apqvatl</mark> dsh
AGNLPREGAV	L <mark>IVTA</mark> SYNGH	PPDNAKQF <mark>VD</mark>	<mark>WLDQA</mark> SADE <mark>V</mark>	KGVRYSVFGC	<mark>gdkn</mark> watty <mark>ç</mark>
KVPAFIDETL	<mark>aakgaen</mark> iad	RGEADASDDF	EGTYEE <mark>w</mark> reh	MWSD <mark>VAAYF</mark> N	LDIENSEDNK
STL <mark>SLQFVDS</mark>	<mark>aa</mark> dmplakmh	GAFS <mark>TNVVAS</mark>	<mark>ke</mark> lqqpgsar	STRHLEIELP	<mark>keasy</mark> Qegdh
L <mark>G</mark> VIPRNYEG	IVNRVTARFG	LDASQQ <mark>IRL</mark> E	AEEEKLAHLP	LAKTVSVEEL	L <mark>qy</mark> v <mark>el</mark> qdpv
TRTQL <mark>RAMAA</mark>	KTVCP <mark>PHKVE</mark>	<mark>l</mark> ealle <mark>kqay</mark>	<mark>ke</mark> qvlakrlt	MLELLEKYPA	CE <mark>MKFSEFIA</mark>
LLPSIRPRYY	SISSSPRVDE	KQASI <mark>TVSVV</mark>	SGEAWSGYGE	YK <mark>GIASNYL</mark> A	ELQEGDTITC
FISTPQSEF <mark>T</mark>	LP <mark>KDPETPL</mark> I	MVGPGTGVA <mark>P</mark>	<mark>FRG</mark> FVQARKQ	LKEQGQSLGE	AHL <mark>YFGCRSE</mark>
HEDYLY <mark>QEEL</mark>	ENAQSEGIIT	LHTAFSRMPN	<mark>QPKT</mark> YVQHVM	EQDGKKLIEL	LDQGAHFYIC
GDGSQMAPAV	EATLM <mark>KSYAD</mark>	VHQVSEA <mark>DAR</mark>	LWLQQLEEKG	RYAKD <mark>VWAG</mark> *	

С

MGSSHHHHHH	SSGLVPRGSH	MTIKEMPQPK	TFGEL <mark>KNLPL</mark>	<mark>l</mark> ntdkpvqal	MKIADELGEI
FKFEAPGRVT	RYLSSQRLIK	EACDESRFDK	NLSQALKFVR	DFAGDGLFTS	WTHEKNWKKA
HNILLPSFSQ	QA <mark>MKGYHAMM</mark>	VDIAVQLVQK	WERLNADE <mark>H</mark> I	EVPEDMTRLT	LDTIGL <mark>CGFN</mark>
YRFNSFYRDQ	PHPFITSMVR	ALDEAMN <mark>KLQ</mark>	RANPDDPAYD	ENKRQFQEDI	KVMNDLVDKI
IADRKASGEQ	SDDL <mark>L</mark> THMLN	GKDPETGEPL	DDENIRYQII	TFLIAGHETT	SGLLSFALYE
LVKNPHVLQK	AAEE <mark>AARVLV</mark>	<mark>dpvpsy</mark> kQvk	QLKYVGMVLN	EALRLWPTAP	AFS <mark>L</mark> YAKEDI
VLGGEYPLEK	GDELM <mark>VLIPQ</mark>	LHRDKTIWGD	DVEEFRPERF	ENPSAIPQHA	F <mark>KPFGNGQR</mark> A
CIGQQ FALHE	ATLVLGMMLK	HFDFEDHTNY	ELDIKETLTL	KPEGFVVKAK	SKKIPLGGI
SPSTEQSAKK	VRKKAENAHN	TPLLVLYGSN	MGTAEGTARD	LADIA <mark>MSKGF</mark>	<mark>APQVATL</mark> DSH
AGNLPREGAV	LIVTA <mark>SYNGH</mark>	PPDNAKQ <mark>F</mark> VD	WLDQASADEV	KGVRYSVFGC	<mark>gdkn</mark> watty <mark>(</mark>
KVPAFIDETL	<mark>aakgaeni</mark> ad	RGEADASDDF	EGTYEE <mark>w</mark> REH	MWSD <mark>VAAYF</mark> N	LDIENSEDNA
STLSL <mark>Q</mark> FVDS	AADMPLAKMH	GAFS <mark>TNVVAS</mark>	<mark>ke</mark> lqqpgsar	STRHLEIELP	KEASYQEGDH
L <mark>G</mark> VIPRNYEG	IVNRVTARFG	LDASQQ <mark>IRL</mark> E	AEEEKLAHLP	LAKTVSVEEL	LQY <mark>VEL</mark> QDP\
TRTQL <mark>RA</mark> MA <mark>A</mark>	KTVCPPHKVE	LEALLE <mark>KQAY</mark>	<mark>ke</mark> qvlakrlt	MLELLEKYPA	CEMKFSEF <mark>I<i>F</i></mark>



Figure S6. The P450 BM3 residues showing the highest degree of change when comparing ligand-free and ligand-bound full-length P450 BM3. The residues are coloured red or cyan to indicate areas of the greatest deshielding or shielding respectively (\pm 5% deuterium uptake), when subtracting ligand-free values from NADP⁺-bound (A), NPG-bound (B) and NADP⁺/NPG-bound protein (C). These residues correspond to conformational changes causing alterations in solvent accessibility. In particular, the residues highlighted in cyan correspond to the sites hypothesized to be involved with dimerization. The His-tag used to facilitate protein purification is shown in green. Residues with no coverage are not coloured.