

Supplementary information

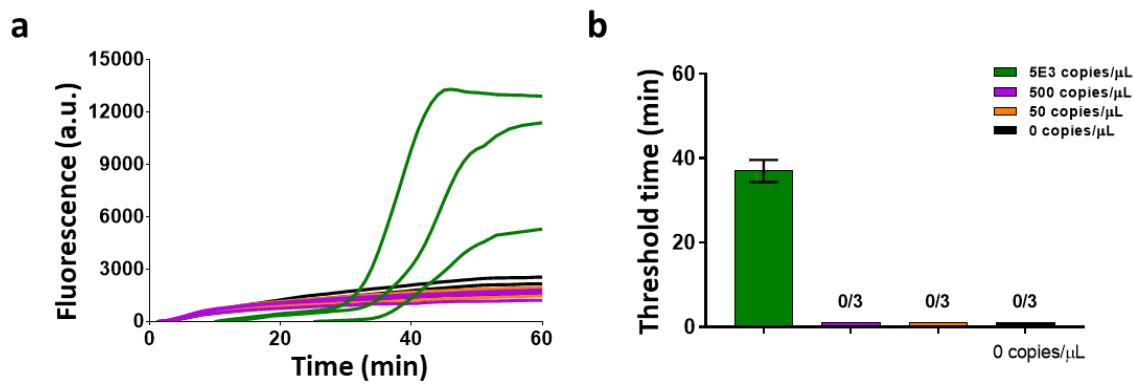


Fig. S1. Characterization of SARS-CoV-2 virus in nasal fluid in a 96 μL reaction. (a-b) Raw fluorescence data and amplification threshold times (for 3 replicates of data) for viral detection in a 96 μL reaction with 50% nasal fluid per reaction. Thermal lysis at 95°C was conducted for 1 min. of the virus in nasal fluid sample before addition of RT-LAMP reagents for the final reaction. The bar graphs show mean and standard deviation. Fraction indicates number of replicates amplified.

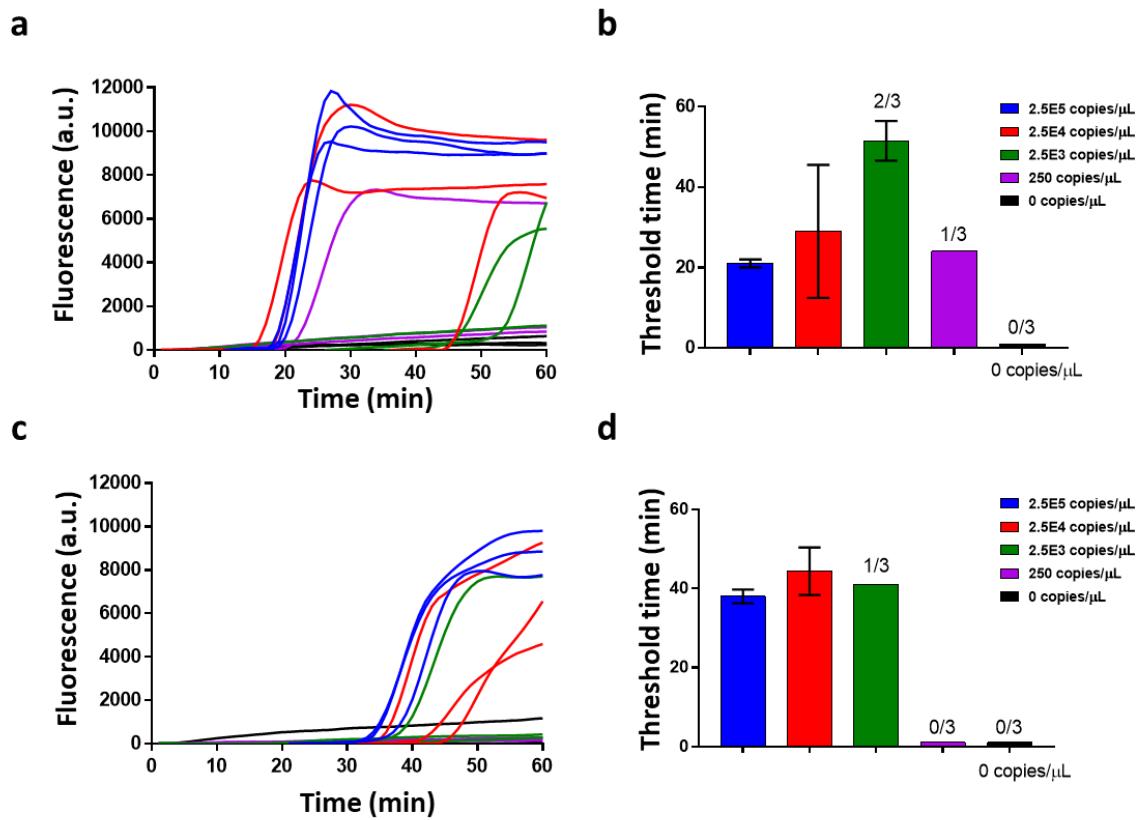
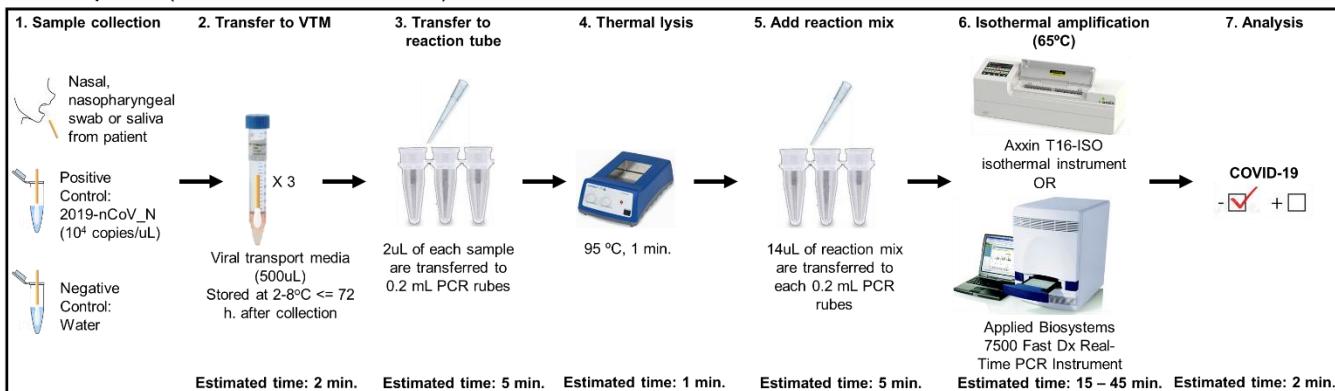


Fig. S2. Characterization of SARS-CoV-2 virus in mock swab samples transported in 100 μ L Viral Transport Media. (a-b) Raw fluorescence data and amplification threshold times (for 3 replicates of data) for viral detection in a 16 μ L reaction with 12.5% VTM per reaction from a 100 μ L VTM sample. **(c-d)** Raw fluorescence data and amplification threshold times (for 3 replicates of data) for viral detection in a 16 μ L reaction with 12.5% VTM per reaction from a 100 μ L VTM sample. The bar graphs show mean and standard deviation.

RT-LAMP protocol (estimated total time: 30 - 60 min.)



RT-PCR protocol (estimated total time: 119 min.)

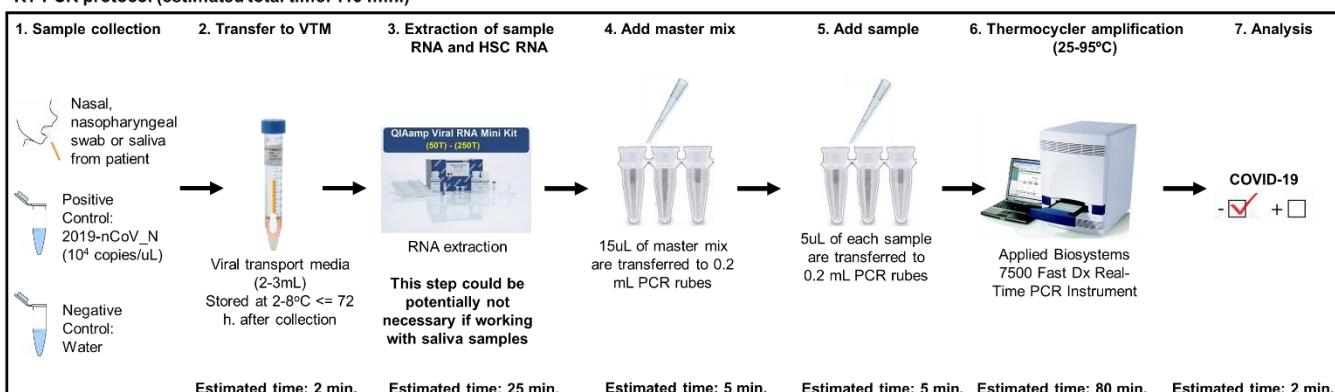


Fig. S3. RT-LAMP assay process flow and comparison side by side with the RT-PCR assay process flow.

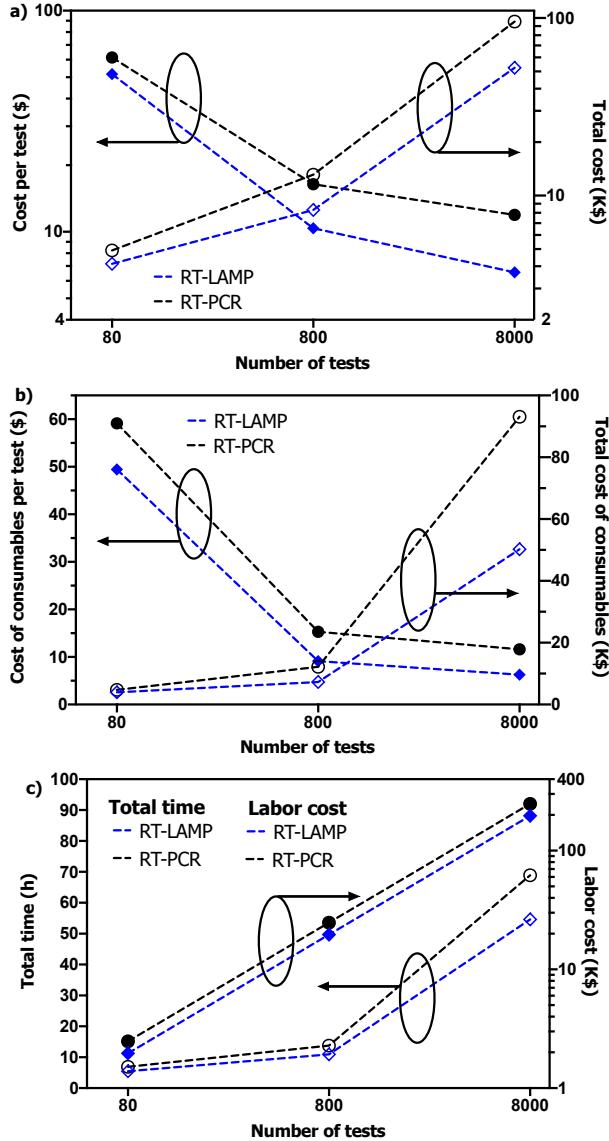


Fig. S4: Resources modeling results. a) Cost per test and total cost (excluding labor) for the three scenarios considered; b) Total cost of consumables and Cost of consumables per test for the 3 models designed; c) Labor cost and Total time for the 3 models designed. The labor cost assumes a gross salary rate = \$1/min.

Table S1. RT-LAMP primer sequences for 12 primer sets (3 primer sets each for 4 target genes)

Gene Orf1a	P1	F3	ATCTAGTTGAATGGCCTACA
		B3	ACAAGCACAGGTTGAGAT
		FIP	TGAGTTTTCATAAACAGTGCCAAA-CAGGTGGTGTGTTCACT
		BIP	AACCCGTCCTTGATTGGCTT-AATTAACAATTCCCAACCGTC
		Loop F	TGTTAGTTAGCCACTGCGAAGT
	P2	F3	AAACCCGTCCTTGATTGG
		B3	CTTTAAGTTTAGCTCCACCAAT
		FIP	TCACAAGCACAGGTTGAGATAAATT-GAAGGTGTAGAGTTCTTAGAGAC
		BIP	GTCACCTGTGCAAAGGAAATTAAG-AGAGTCAGCACACAAAGC
	P3	F3	CGGTGGACAAATTGTCAC
		B3	CTTCTCTGGATTTAACACACTT
		FIP	TCAGCACACAAAGCCAAAAATTAT-CTGTGCAAAGGAAATTAAGGAG
		BIP	TATTGGTGGAGCTAAACTAAAGCC-CTGTACAATCCCTTGAGTG
Gene S	P1	F3	TCTTCACACGTGGTGT
		B3	CAGTGGAAAGCAAAATAAACAC
		FIP	GAAAGGTAAGAACAAAGTCCTGAGT-TTACCCCTGACAAAGTTTCAG
		BIP	TTCCAATGTTACTGGTCCATGC-GACAGGGTTATCAAACCTCT
		Loop B	TATACATGTCCTGGGACCAATGG
	P2	F3	GGTGTATTACCCCTGACAAAG
		B3	GTACCAAAATCCAGCCTC
		FIP	TGGAACCAAGTAACATTGGAAAAGA-TTTCAGATCCTCAGTTTACATTC
		BIP	CTCTGGGACCAATGGTACTAAGAG-GACTTCTCAGTGGAAAGCA
		Loop B	AACCCTGTCCTACCATTAAATGATG
	P3	F3	CCTGACAAAGTTTCAGATCC
		B3	GTACCAAAATCCAGCCTC
		FIP	GCATGGAACCAAGTAACATTGGAAA-TCAGTTTACATTCAACTCAGGA
		BIP	CTCTGGGACCAATGGTACTAAGAG-GACTTCTCAGTGGAAAGCA
		Loop B	AACCCTGTCCTACCATTAAATGATG
Gene Orf 8	P1	F3	ACGCCTAACGAACATGAA
		B3	AGAACCGACCTCATCCAG

		FIP	GGTTGATGTTGAGTACATGACTGTA- CTTGTTCCTTAGGAATCATCACA
		BIP	ATATGTAGTTGATGACCCGTTGTC- TAAAGGTGCTGATTTCTAGCT
		Loop F	CTACATTCTGGTGAAATGCAGCTA
	P2	F3	CTCAACATCAACCATAATGTAATG
		B3	CAATTAGGTTCTGGCAATT
		FIP	GGTGCTGATTTCTAGCTCCACTC- GACCCGTGCTTATTAC
		BIP	TGCTGGATGAGGCTGGTCTA- TGTAAAAGGTAACAGGAAACTG
		Loop B	ATCACCCATTCACTGACATCGATATC
	P3	F3	AGCTGCATTTCACCAAGAA
		B3	CGATATCGATGTACTGAATGG
		FIP	TGAATAGGACACGGGTATCA- GTAGTTACAGTCATGTACTCAA
		BIP	GAGTAGGAGCTAGAAAATCAGCAC- TGATTTAGAACAGCCTCATC
Gene N	P1	F3	GTTCCTCATCACGTAGTCG
		B3	GTGGCCCTTGTGTTGTT
		FIP	GCCAGCCATTCTAGCAGGAG- CAACAGTTAAGAAATTCAACTCC
		BIP	GATGCTGCTCTTGCTTGCT- ACCAGACATTTGCTCTCAA
		Loop B	GCTGCTTGACAGATTGAACCAG
	P2	F3	AGACGAATTCTCGTGGTGGT
		B3	TTGTTAGCAGGATTGCGG
		FIP	TGGCCCAGTTCTAGGTAGT- GACGGTAAAATGAAAGATCTCAG
		BIP	CTTCCCTATGGTGCTAACAAAGAC- TGGTGTATTCAAGGCTCC
		Loop B	GGCATCATATGGGTGCAACTGAG
	P3	F3	GTCATTTGCTGAATAAGCATAT
		B3	GAGTCAGCACTGCTCATG
		FIP	TAAGGCTTGAGTTCTCATCAGCCTT- ACGCATACAAACATTCCCC
		BIP	CAGAGACAGAAGAACAGCAAAC- GATTGTTGCAATTGTTGGAG
		Loop B	GTGACTCTCTTGCTGCAGATT

Table S2. Supply chain: RT-LAMP and RT-PCR assay

See excel file.

Video S1. Uniform filling of the amplification chambers.

See video

Video S2. Time stamped videos of amplification on the cartridge for 5000 copies/ μ L of virus in VTM.

See video

Video S3. Time stamped videos of amplification on the cartridge for negative control (VTM only).

See video