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# **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see Authors & Referees and the Editorial Policy Checklist.

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For	all statistical analyse	es, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.				
n/a	a Confirmed					
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement					
	A statement o	🔀 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly				
	The statistical test(s) used AND whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.					
$\boxtimes$	A description of all covariates tested					
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons					
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficien AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)					
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give P values as exact values whenever suitable.					
$\boxtimes$	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings					
$\times$	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes					
	$\square$ Estimates of effect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated					
		Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.				
So	ftware and c	ode				
Poli	cy information abou	at <u>availability of computer code</u>				
Da	ata collection	Epifluorescence microscopy: Olympus IX71 microscope equipped with a CellSens Dimension 2.3 Software Nikon Eclipse microscope Nikon A1R MP (2 photon)				

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

#### Data

Data analysis

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets

Western Blot imaging: LiCor Sytem Odyssey Clx

GraphPad Prism 5, 2012 GraphPad Software Inc.;

Matlab Version 2014b, 2007, Mathworks Inc; Sigma Plot 11 (Systat Software, San Jose, CA, USA); ImageJ, Fiji, version: 2.0.0-rc-69/1.52p; Build 269a0ad53f

ÄKTA chromatography: UNICORN™ 7.0.2 software (GE Healthcare Life Sciences)

Phyton Seaborn package, python.org; version 0.9; 10.5281/zenodo.1313201

- A list of figures that have associated raw data
- A description of any restrictions on data availability

#### Data Availability

The data that supports the finding of this study is readily available within this paper, its Supplementary file and in the Source Data file. The data set for Figure 1e, 2b,

		nentary Figures 1a, 1b, 1c, 3a, 3b, 5b and 6 are provided in the Source Defrom the corresponding author on reasonable request.	ata file. All data sets generated during and/or analyzed		
Field-spe	ecific re	eporting			
Please select the o	ne below that	is the best fit for your research. If you are not sure, read the app	propriate sections before making your selection.		
X Life sciences		Behavioural & social sciences Ecological, evolutionary &	environmental sciences		
or a reference copy of	the document with	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>			
:£:					
Lite scier	ices st	udy design			
All studies must dis	sclose on these	points even when the disclosure is negative.			
Sample size	provided a suf	Sample size calculation were not performed a priori. In quantitative data, the aim was to collect as much data as possible, the sample size provided a sufficient accuracy to distinguish statistically significant differences between conditions. In descriptive experiments we chose a sample size that was large enough to show difference between the conditions.			
Data exclusions	No data were	were excluded from the analysis, except raw images which were out of focus.			
Replication		Replicates and independent experiments were carefully kept that way. Most of our Experiments were replicated independently at least 3 times. Specific n can be found in the each Figure legend.			
Randomization	Randomization	was not performed for this study.			
Blinding	Blinding was n	ot performed in this study.			
We require informati	ion from authors	pecific materials, systems and about some types of materials, experimental systems and methods use by your study. If you are not sure if a list item applies to your research, res	ed in many studies. Here, indicate whether each material,		
Materials & ex	perimental :	systems Methods			
n/a Involved in th	•	n/a Involved in the study	_		
Antibodies		ChIP-seq			
Eukaryotic cell lines    Flow cytometry   MRI-based neuroimaging					
	nd other organis	— [ —			
Human research participants					
Clinical dat	ta				
Antibodies					
Antibodies used	g c c n r	abbit polyclonal anti- $\alpha/\beta$ -Synuclein antibody; dilution used 1:500; Cat. Nullinea pig polyclonal anti-synaptophysin antibody; dilution 1:100; Cat. Nullinea pig labeled with ATTO 647N; dilution 1:500; Cat. No. I onkey anti-rabbit; dilution 1:500; Cat. No. 711-175-152, Dianova GmbH nouse monoclonal (clone 3E6) anti EGFP; dilution 1:500; Cat. No. A11120 abbit polyclonal (affinity purified) anti beta-Actin directly labeled with CymbH	o. 101004; Synaptic System GmbH N0602-At647N-S; Synaptic System GmbH 0, Thermo-Scientific		

Validation

anti- $\alpha/\beta$ -Synuclein: antiserum, Synthetic peptide corresponding to AA 2 to 25 from human  $\alpha$ -Synuclein; Reacts with: rat, mouse, zebrafish and human  $\alpha$  synuclein and  $\beta$  synuclein. Validated for IF, IHC.

anti-synaptophysin antibody:Synthetic peptide corresponding to AA 301 to 313 from human Synaptophysin1,

Reacts with: human, rat, mouse, hamster, cow, chicken, frog. Validated fro IF and IHC.

Specific for synaptophysin 1, no cross-reactivity to other synaptophysins.

Mouse monoclonal anti EGFP (clone 3E6) validated in IF, IHC, WB and more applications source Thermo-fisher.

Rabit polyclonal anti Beta actin, antigen is the AA2 to AA16 of mouse Beta Actin, Externally validated for WB stated in suppliers webpage: Synaptic System GmbH

## Eukaryotic cell lines

Policy information about cell lines

Cell line source(s)

Wildtype HEK293 were purchased from the German Collection for Microorganisms and Cell Cultures (DSMZ), Braunschweig, Germany. Stably transfected lines like the Report cell line and the aSyn cell line were produced by Sirion Biotech GmbH (Martinsried, Germany). A full report on their generation was provided using DZMZ HEK-203 cell lines.

Authentication

We relied on the authentication from DSMZ and Sirion Biotechnology GmbH

Mycoplasma contamination

Cells were tested negative for mycoplasma.

Commonly misidentified lines (See ICLAC register)

HEK-293 were not reported by the ICLAC to be misidentified.

## Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

Xenopus laevis tadpoles (albinos)

Wild animals

Provide details on animals observed in or captured in the field; report species, sex and age where possible. Describe how animals were caught and transported and what happened to captive animals after the study (if killed, explain why and describe method; if released, say where and when) OR state that the study did not involve wild animals.

Field-collected samples

For laboratory work with field-collected samples, describe all relevant parameters such as housing, maintenance, temperature, photoperiod and end-of-experiment protocol OR state that the study did not involve samples collected from the field.

Ethics oversight

All procedures for animal handling were approved by the governmental animal care and use office (Niedersächsisches Landesamt für Verbraucherschutz und Lebensmittelsicherheit, Oldenburg, Germany, Az.12/0779) and were in accordance with the German Animal Welfare Act as well as with the guidelines of the Göttingen University Committee for Ethics in Animal Experimentation

Note that full information on the approval of the study protocol must also be provided in the manuscript.

## Human research participants

Policy information about  $\underline{\text{studies involving human research participants}}$ 

Population characteristics

Study participants consisted of individuals who were in treatment at the Paracelsus Elena Klinik, Kassel, Germany and had been diagnosed with a variety of neurological disorders non-related to  $\alpha$ Syn aggregation disorders.

Recruitment

CSF samples from all individuals were collected after the informed consent of the participant at the Paracelsus Elena Klinik (Kassel, Germany) in accordance with the principles of Declaration of Helsinki and following identical standard operating procedures.

Ethics oversight

The use of the CSF samples in this study was approved by the ethical committee of the Medical Center Göttingen with the approval numbers 36/7/02 and 9/7/04

Note that full information on the approval of the study protocol must also be provided in the manuscript.