

Precursor peptide-targeted mining of more than one hundred thousand genomes expands the lanthipeptide natural product family

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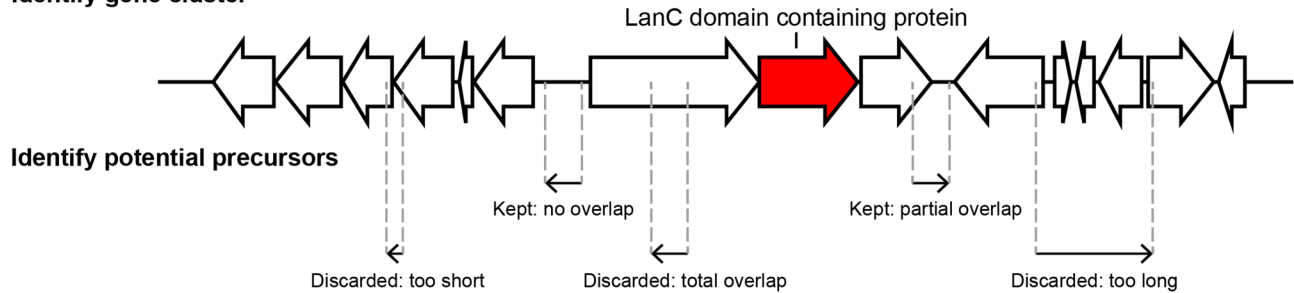
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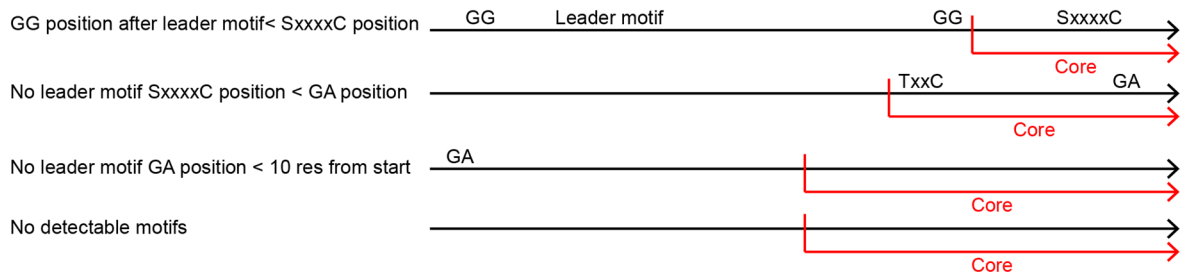
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Supplementary Figure S1. Features calculated to score precursor peptides. The DNA sequence encoding 7 genes upstream and downstream of a LanC-like domain containing protein was extracted and all potential open reading frames (ORFs) were identified. In the case that multiple potential ORFs shared a stop codon, the longest ORF within the expected range of LanA lengths was used. These ORFs were discarded if they were too long, too short, occurred entirely within an annotated gene, or did not contain a Cys residue. The remaining ORFs were then analyzed using FIMO [1] to identify conserved leader motifs. If the ORF contained a leader motif and has a GG, GA, or S/T (x)₂₋₇C motif downstream of the leader, but not within 10 residues of the end of the ORF, the approximate core was identified as starting immediately after the GG or GA motif or 1 residue before the S/T (x)₂₋₇C motif. If the ORF contained multiple GG or GA motifs, only the first one was considered, likewise with the S/T (x)₂₋₇C motif. If the ORF contained both a GG or GA motif and a S/T (x)₂₋₇C motif, the longer potential core was used. If the ORF did not contain a GG, GA, or S/T (x)₂₋₇C motif, or those motifs were within 10 residues of the end of the ORF, the C-terminal half of the peptide was identified as the potential core. If no leader peptide motif was identified in the ORF, the same analysis was performed starting from the beginning of the ORF instead of after the leader motif. Finally, if the predicted core did not contain a Cys residue, the ORF was discarded. The given features were then calculated for each potential ORF.

Identify gene cluster



Predict potential core peptides



Score potential core peptides

Molecular weight of core	Core fraction that is Cys	Second fifth of precursor fraction S+T+C
Isoelectric point of core	Core fraction that is Ser + Thr	Second fifth of precursor fraction C
Number of each amino acid in core	Core fraction that is Ser + Thr + Cys	Third fifth of precursor fraction S+T
Number of Ser + Thr in core	Core fraction that is charged residues	Third fifth of precursor fraction S+T+C
Number of Ser + Thr + Cys in core	Core fraction that is positive residues	Third fifth of precursor fraction C
Number of Charged residues in core	Core fraction that is negative residues	Fourth fifth of precursor fraction S+T
Number of positive charges in core	Core fraction that is polar residues	Fourth fifth of precursor fraction S+T+C
Number of negative charges in core	Core fraction that is aliphatic residues	Fourth fifth of precursor fraction C
Net charge of core	First fifth of precursor fraction S+T	Fifth fifth of precursor fraction S+T
Number of polar residues in core	First fifth of precursor fraction S+T+C	Fifth fifth of precursor fraction S+T+C
Number of aliphatic residues in core	First fifth of precursor fraction C	Fifth fifth of precursor fraction C
Number of aromatic residues in core	Second fifth of precursor fraction S+T	

Number of amino acid pairs

AA, AC, AD, ..., AW, AY AxA, AxC, AxD, ..., AxW, AxY ... AxxxxxA, AxxxxxC, AxxxxxD, ..., AxxxxxW, AxxxxxY
 CA, CC, CD, ..., CW, CY CxA, CxC, CxD, ..., CxW, CxY ... CxxxxxA, CxxxxxC, CxxxxxD, ..., CxxxxxW, CxxxxxY
 ⋮ ⋮ ⋮
 WA, WC, WD, ..., WW, WY WxA, WxC, WxD, ..., WxW, WxY ... WxxxxxA, WxxxxxC, WxxxxxD, ..., WxxxxxW, WxxxxxY
 YA, YC, YD, ..., YW, YY YxA, YxC, YxD, ..., YxW, YxY ... YxxxxxA, YxxxxxC, YxxxxxD, ..., YxxxxxW, YxxxxxY

Supplementary Table S1. Features and scoring for class I precursors. ORFs were identified as precursors if their score was over 10. SVM, support vector machine.

Feature	Score
SVM classification	5
Presence of Class I leader peptide MEME motif	5
Core pI less than 9	2
2 or more Cys in core	2
Leader has KLxLxK MEME motif and ends in GG sequence	1

Supplementary Table S2. Features and scoring for class II precursors. ORFs were identified as precursors if their score was over 10.

Feature	score
SVM classification	5
Presence of Class II leader peptide MEME motif	5
2 or more Cys in core	2
Hits a Pfam* hidden Markov model	5
Precursor ends with sequence KRC	4

*PF08130.1, PF04604.12, PF14867.5, PF16934.4, PF02979.15, PF07862.10

Supplementary Table S3. Features and scoring for class III precursors. ORFs were identified as precursors if their score was over 10.

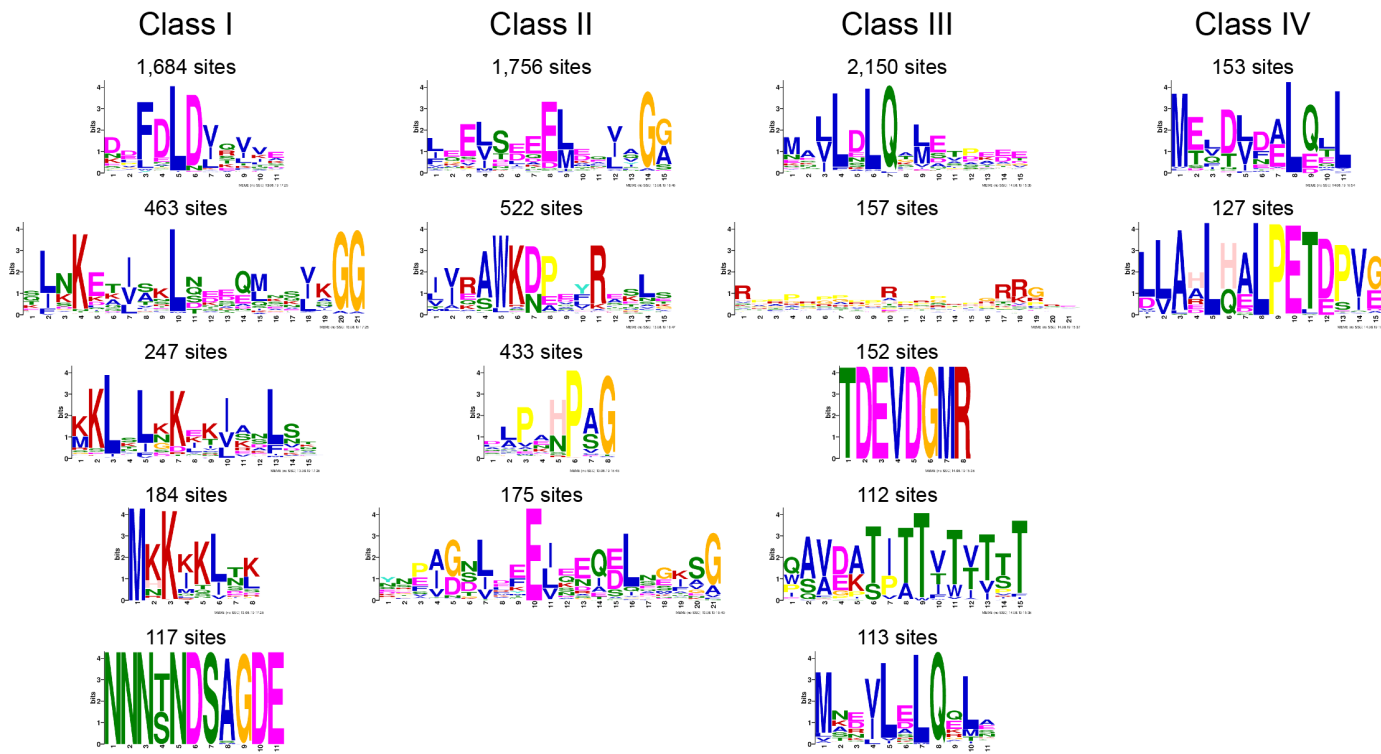
Feature	score
SVM classification	5
Presence of Class III leader peptide MEME motif	5
2 or more Cys in core	2
Has SxxSxxx motif	1
Has SxxSxxC motif	1
Core pI is between 3 and 9	1

Supplementary Table S4. Features and scoring for class IV precursors. ORFs were identified as precursors if their score was over 10.

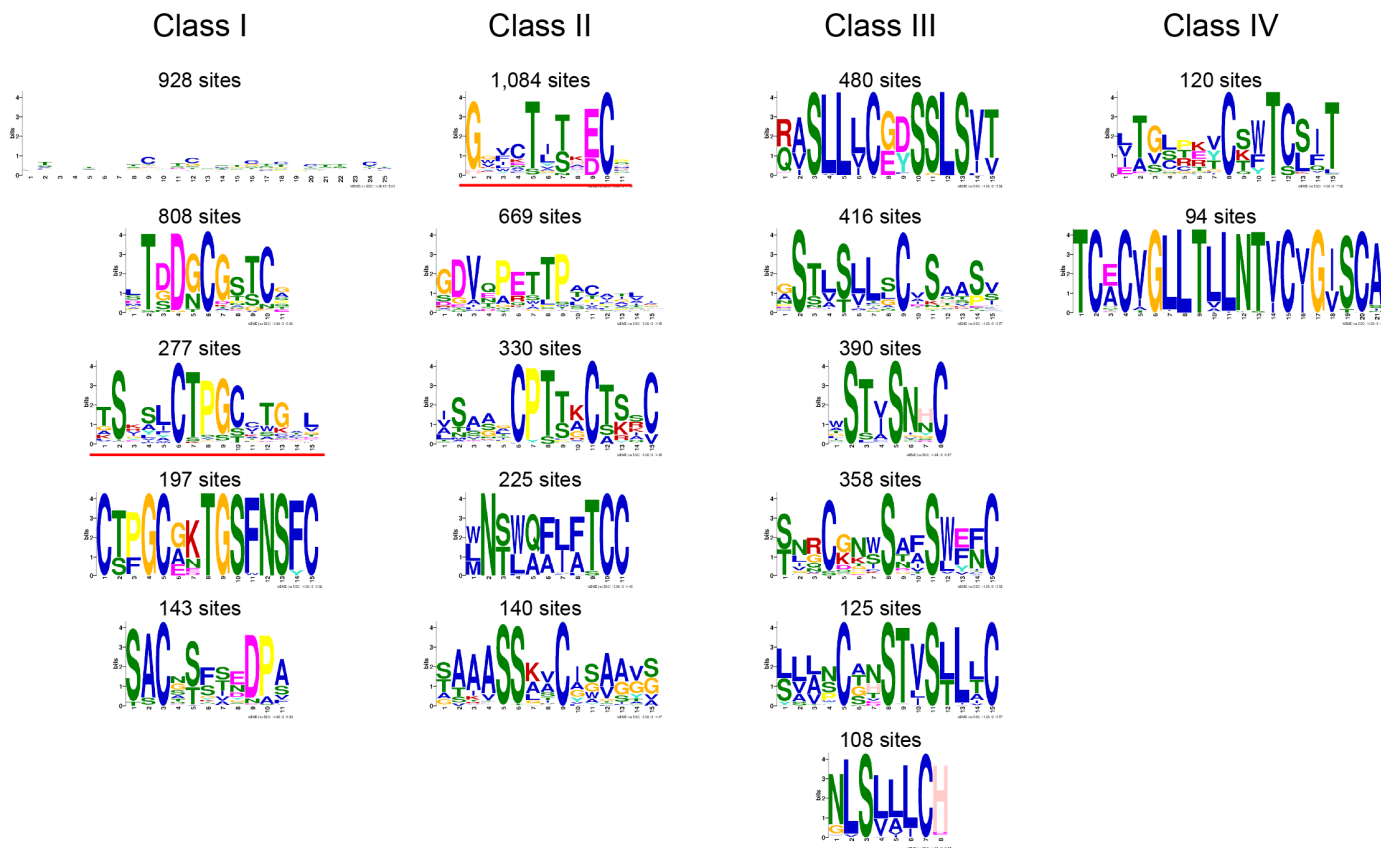
Feature	score
SVM classification	5
Presence of Class IV leader peptide MEME motif	5
2 or more Cys in core	2
Precursor ends with sequence GCD	1
Precursor ends with sequence LGS	1

Supplementary Figure S2. Sequence motifs present in more than 100 lanthipeptide precursor peptides. Some of the conserved leader peptide motifs have been shown to be important for the cognate lanthipeptide synthases (FxLD in class I [2]; E/D(-8)L/M(-9) in class II [3]; LxLQ in class III [4], and Lx₂LPE in class IV [5]). Lipid II-binding motifs, which arise for likely antibiotic lanthipeptides are underlined in red.

Leader motifs



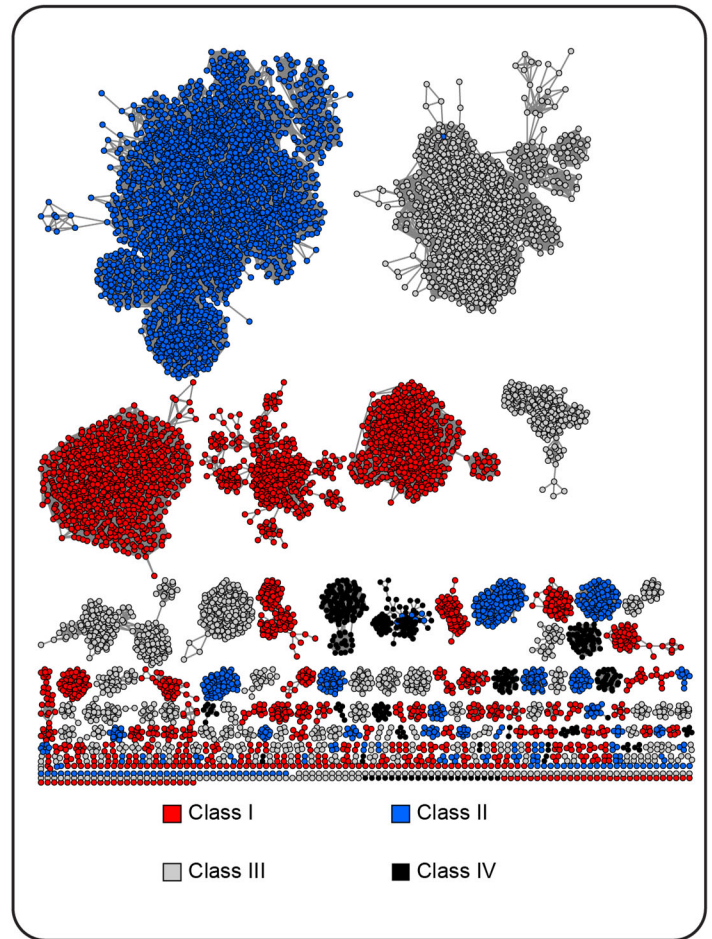
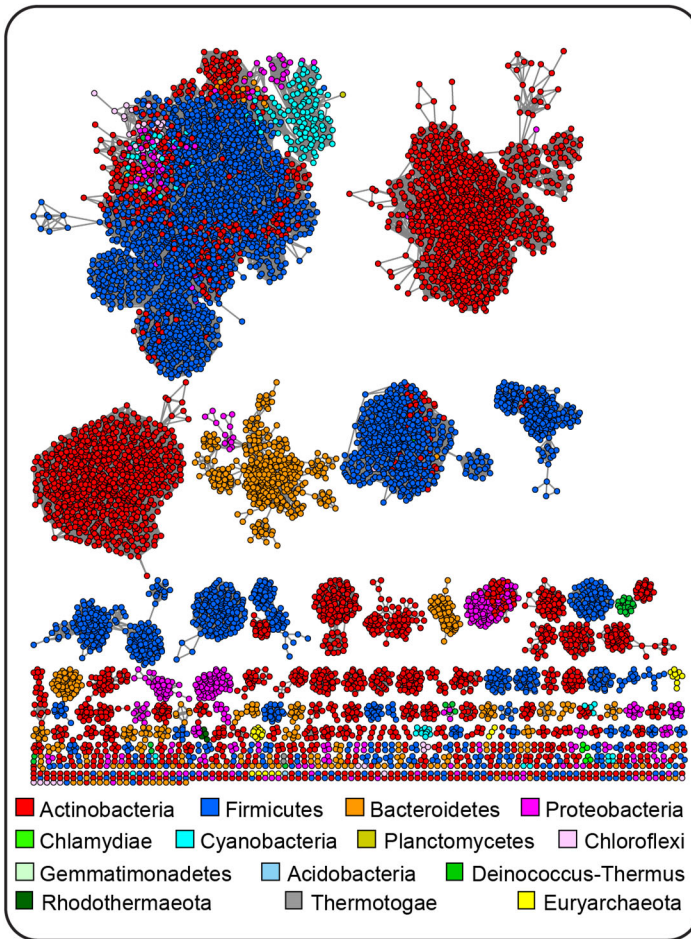
Core motifs



Supplementary Figure S3. Sequence similarity network of predicted precursor peptides generated with the Enzyme Function Initiative-Enzyme Similarity Tool [6] with permissive similarity cutoff (alignment score: 6; equivalent to an expectation value of 10^{-6}) and visualized in Cytoscape [7].

Colored by prokaryotic phylum

Colored by lanthipeptide class



Supplementary Table S5. Location of top BLAST hits of known lanthipeptides from the MIBiG [8] and BAGEL [9] databases in the sequence similarity networks presented in Figure 2. If the precursor is located in a cluster with more than 20 members, the identifier of that cluster is given. If the precursor is located in a cluster with 20 or fewer members, the label from Figure 2 is given. If the precursor occurred only once, it is annotated as a singleton.

Known Lanthipeptide	Cluster
AmfS	III 1
BacCH91	I 5
BhtA1	BhtA1
BhtA2	BhtA2
Bicereucin_beta	II 33
BLD_1648	BLD_1648
Bovicin_HJ50	Bovicin HJ50
BsaA2	I 5
Carnolysin_A1	II 16
Carnolysin_A2	II 16
Catenulipeptide	singleton
Cerecidin	II 3
Cinnamycin_B	Cinnamycin B
Cyll _L	II 46
Cyll _S	II 3
Curvopeptin	singleton
Divamide*	Cinnamycin B
Duramycin*	Cinnamycin B
Entianin	I 7
Epidermin	I 5
Ericin_A	I 7
Ericin_S	I 7
Erythreapeptin	III 4
Flavecin_A1	Flavecin A1
Flavucin	Flavucin
Gallidermin	I 5
Geobacillin_I	I 7
Geobacillin_II	Geobacillin II
Griseopeptin	III 1
Haloduracin_alpha	II 4
Haloduracin_beta	Haloduracin_beta
Informatipeptin	III 2
Lacticin 3147 α *	II 9
Lacticin 3147 β *	II 6
Lacticin 481*	II 1
Lichenicidin_VK21_A1	II 4
Lichenicidin_VK21_A2	II 7
Macedocin	II 1
Macedovicin	Bovicin HJ50
McdA1	II 1
Mersacidin	Mersacidin
Michiganin A	Michiganin A
Microbisporicin	Microbisporicin
Mutacin 1140*	I 5
Mutacin II	II 1
Mutacin K8	II 1
Nisin_A	I 7

Known Lanthipeptide	Cluster
Nisin_O	I 7
Nisin_U	I 7
Nisin_Z	I 7
Nukacin_ISK-1	II 1
Paenibacillin	Paenibacillin
Paenicidin_A	Paenicidin A
Paenicidin_B	Paenicidin A
Paenilan	I 20
Penisin	I 23
Pinensin	I 17
Plantaricin C	Plantaricin C
Pneumolancidin_PIdA2	II 11
Pneumolancidin_PIdA3	II 11
Pneumolancidin_PIdA4	II 11
Prochlorosin_1.1	Prochlorosin 1.1
Pseudomycoicidin	Geobacillin II
Roseocin α	II 29
Roseocin β	II 2
Ruminococcin_A	II 1
SAL-2242	III 1
Salivaricin_9	II 1
Salivaricin_A	II 14
Salivaricin_G32	II 1
Sap_T	I 21
SapB	III 1
Smb_A	BhtA2
Smb_B	BhtA1
SRO15-2212	III 1
SRO15-3108	II 2
Stackepeptin_A	singleton
Stackepeptin_B	singleton
Stackepeptin_C	singleton
Stackepeptin_D	singleton
Staphylococcin_C55_alpha	II 9
Staphylococcin_C55_beta	II 6
Streptin	I 14
Streptococcin_A-FF22	II 1
StreptococcinA-M49	II 1
Subtilin	I 7
Subtilomycin	Subtilomycin
Suicin_3908	Bovicin HJ50
Suicin_65	II 1
Suicin_90-1330	I 7
Thermophilin_1277	Bovicin HJ50
Thusin_A	II 4
Thusin_B	Thusin B

*top blast hit is not identical, possibly because the genome of the producer is not in the database.

I 13

n = 56

M I N E K N L F D L D V Q V T T T A G D V D P Q I T S V S A C T P G C G N T G S F N S F C C

I 14

n = 36

M N Z N T I K D F D L D L K T Z K K K D T A P V G S R Y L C T P G S C W K L V C F T T T V K

I 16

n = 33

M N K E L F D L D I N K K M E T P T E M T A Q T W T T V K V S K A V C K T G T C I C T T S C S N C K

I 17

n = 50

M E I N D N I Z N K M K L L E D L K I E S F V T S A L D S K D E L S V A K R L S G G L G N S A A G D H S H P T H T I M K T D D R L H H T P P H V C T I V I C

I 20

n = 27

M K N Q F D L D L Q V A K N E V A P K V Q P A S G L I C T P S C A T G T L N C Q V S L S F C K T C

I 21

n = 23

M P E L T E L D T L S D L P E R T S D L P S A A Y T G C S G L C T I V C T V V I C G V C

I 23

n = 23

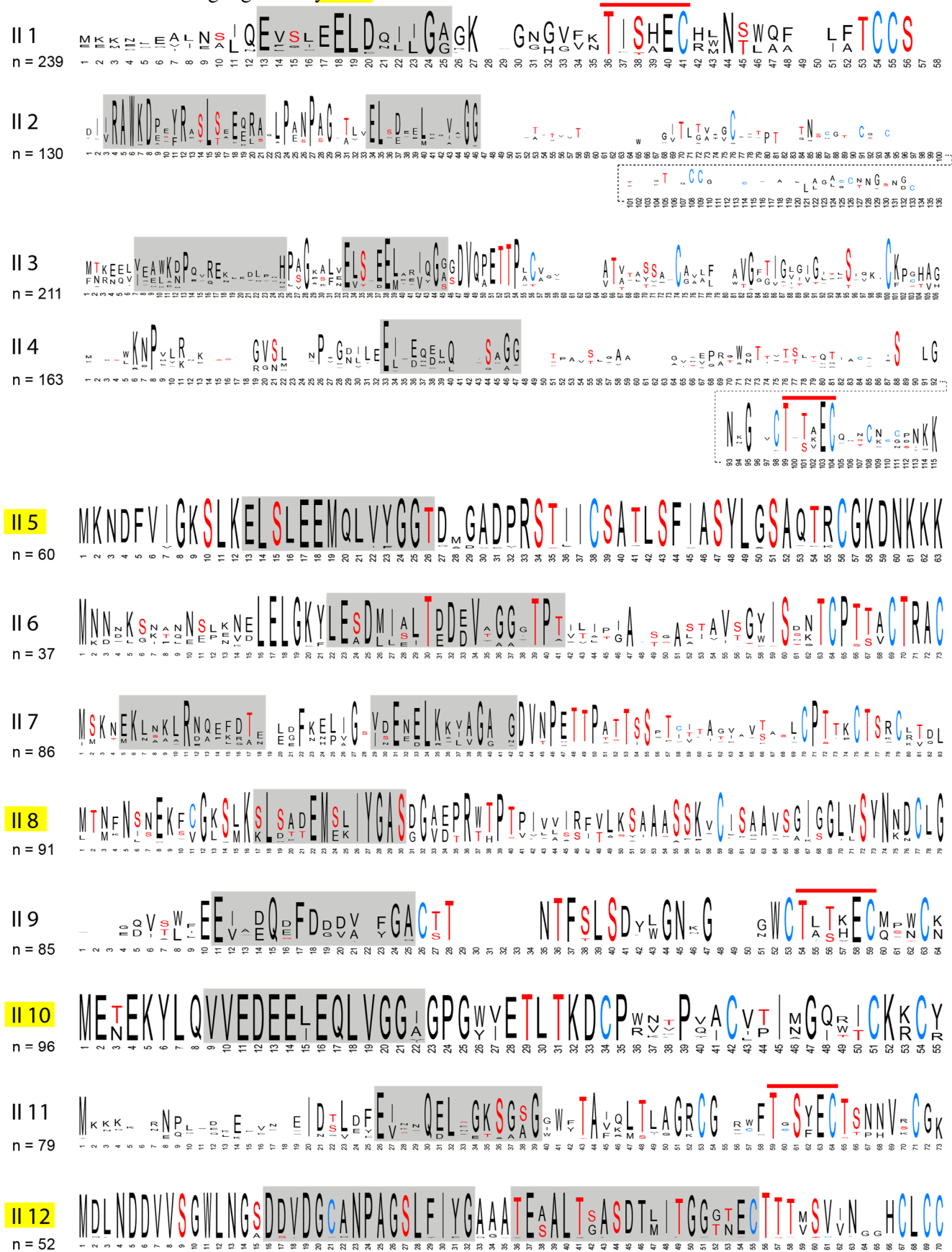
M A N N F D L D V V K S V N S V N N F G L Y T S C Y S S Q C Y S S K C Y S D S C Y S S C Y T G R H M C G Y T H G Y S C

I 31

n = 21

M E N T E F S L E L D V T E V A T E Q D Y V S S G V T S T G C C K N

Supplementary Figure S5. Sequence logos generated from alignments of class II precursor peptides in clusters with 20 or more members (in Figure 2) using WebLogo [10]. Conserved leader motifs that are shared among multiple clusters (from Figure S2) are boxed in gray and potential lipid II-binding motifs are overlined in red. Families with no previously characterized members are highlighted in yellow.



II 13
 n = 33
 1 MAE 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33
 S D Y L G E P K K F A R L I A A S D E F R A R Y E E P R A V I A E Y G V P G L P T P L P A R D E G E F D L E L E A G A A G T G T v S S T C T T
 101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 122 123

II 14
 n = 65
 1 M S F M K N S K D I L T N A I E E V S E K E L M E V A G G K K G S G W F A T I T D D C P N S V F V C C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51

II 16
 n = 36
 1 M E F M Z K K V V G C F E D M S I A E N T V Q G S G D V E T P T T P A C A I A A A A S S V K T A K G A A I S A I A V S G A V I S A V K C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36

II 17
 n = 27
 1 T R E L E A L V K A W D E F K Q E L L S N P A K A V A A E L G E P D E V V L E E T A Z K V Y L V L P P A A G A I E L S E E L E A V A G G
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27
 101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 122 123

II 18
 n = 34
 1 M E K M Y F A G D L E E L E E I S L E I S G G G A E Q R G I S Q G N D G K L C T L T W E C G L C P T H T C W C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34

II 19
 n = 27
 1 M S D Y L G G Y D E A E L V E L S E A D V Y G G T T P W S C A T V T L V A T L V A S A A C P T T K C T S C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27

II 21
 n = 48
 1 M K K F A L E M T E E L K E L A G G S E F A T P M T V T P T T I T I P I S L A G C P T T K C A S I V S P C N D
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48

II 23
 n = 34
 1 M S E K L N M S G D F Y L E F R A Y Q A P A N N G L L A T T M E C N T F G T C T N L E C S T L G C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34

II 24
 n = 26
 1 M E N L N A T N P V G S V L S E L I D T E M P T V A G G A Q A L G F T D G N C L T I T A D C T P W L G C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26

II 25
 n = 32
 1 M D D P P I L T E D E L R S A V R Q V F K R A Q T D W E F R Q L C L S D P A A A I R Q V S G K S L P S G F A L Q F T D T R E S V G
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32

II 26
 n = 39
 1 M L N K E L L Q N A L Q Q Q Y K E A N L A E A I K L I T A G A Q K G V F T G E Y A Q L M S L E E L S E D L L V A G G
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39
 101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 122 123

II 27
 n = 30
 1 M A V G L L M K A G N V S E E L A V L N N E H S L N A S L D T I C G T M G S L G C G S F G C G S L S S C C
 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30

II 28 MQAV¹EEK²VQ³SG⁴TDRNADEVFDLV¹⁰GDLEQE¹⁵PPPLASPTNSGSGTACGTCAGvYCC⁵⁵
n = 34

II 29 VDI¹VRS²WKD³ADYR⁴LS⁵LGSE⁶EAP⁷HP⁸SGE⁹GL¹⁰LTA¹¹EL¹²DEEL¹³TE¹⁴INGAAG¹⁵SG¹⁶v¹⁷LG¹⁸LGCC¹⁹s²⁰CLPW²¹YS²²G²³ P²⁴ WT²⁵ VCG²⁶LA²⁷C²⁸NP²⁹GK³⁰PC³¹KN³²
n = 31

II 30 MNNKFTGKINELELEQLVGDNQVVGGIPTIPATISFVGVTFVTVLNAGAPTSGCTKSCNK
n = 34

II 31 VDI¹VRS²WKD³ADYR⁴LS⁵LGSE⁶EAP⁷HP⁸SGE⁹GL¹⁰LTA¹¹EL¹²DEEL¹³TE¹⁴INGAAG¹⁵SG¹⁶v¹⁷LG¹⁸LGCC¹⁹s²⁰CLPW²¹YS²²G²³ P²⁴ WT²⁵ VCG²⁶LA²⁷C²⁸NP²⁹GK³⁰PC³¹KN³²
n = 31

II 32 MAQKDFPQKINSQLEEISDN¹⁵SAVGAG²⁰WAQLS²⁵FISEALGNKGAVCTGTIECQNNCR
n = 33

II 33 M¹ I² D³ N⁴ M⁵ N⁶ K⁷ Z⁸ F⁹ H¹⁰ N¹¹ I¹² G¹³ S¹⁴ F¹⁵ E¹⁶ L¹⁷ Q¹⁸ E¹⁹ M²⁰ D²¹ F²² S²³ G²⁴ A²⁵ E²⁶ G²⁷ T²⁸ V²⁹ E³⁰ P³¹ Q³² A³³ T³⁴ P³⁵ T³⁶ I³⁷ A³⁸ S³⁹ P⁴⁰ S⁴¹ T⁴² P⁴³ C⁴⁴ A⁴⁵ R⁴⁶ V⁴⁷ A⁴⁸ S⁴⁹ I⁵⁰ V⁵¹ S⁵² G⁵³ A⁵⁴ V⁵⁵ S⁵⁶ A⁵⁷ V⁵⁸ V⁵⁹ S⁶⁰ G⁶¹ L⁶² A⁶³ S⁶⁴ A⁶⁵ T⁶⁶ K⁶⁷ D⁶⁸ L⁶⁹ D⁷⁰
n = 26

II 36 MPPVASPDIDISSAEEINRWLSDTSLDGS DSPAGPLFTGGRYVVQEITATWGGCNSWTVYCSCTGSPICCC
n = 28

II 46 MENL²S³VVP⁴SFEEL⁵SVEEMEA¹⁰IQGS¹⁵GDVQAE²⁰TTPVCAVAATAAASSAACG³⁵WVGGG⁴⁰IFTGVTVVVSLKHC⁶⁸
n = 23

Supplementary Figure S6. Sequence logos generated from alignments of class III precursor peptides in clusters with 20 or more members (in Figure 2) using WebLogo [10]. Conserved leader motifs that are shared among multiple clusters (from Figure S2) are boxed in gray. Families with no characterized members are highlighted in yellow.



Supplementary Figure S7. Sequence logos generated from alignments of class IV precursor peptides in clusters with 20 or more members (in Figure 2) using WebLogo [10]. Conserved leader motifs that are shared among multiple clusters (from Figure S2) are boxed in gray. Families with no previously characterized members are highlighted in yellow.



Supplementary Table S6. Twenty most abundant proteins in class I BGCs that belong to at least one Pfam family. If a protein has multiple domains from different Pfam families, those families are separated by a slash. Potential secondary modification enzymes are highlighted in **yellow**. Split LanBs, with the glutamylation and elimination domains on separate polypeptides, are denoted sLanB.

Pfam families	Description	Count
LANC_like (PF05147)	LanC	2,204
Lant_dehydr_N (PF04738)/Lant_dehydr_C (PF14028)	LanB	1,745
ABC_tran (PF00005)	LanT	657
Lant_dehydr_C (PF14028)	LanB elimination domain	562 (193 sLanBs)
PCMT (PF01135)	Protein-L-isoaspartate(D-aspartate) O-methyltransferase	571
Gallidermin (PF02052)	Gallidermin- and nisin-like precursor peptides	426
ABC2_membrane_4 (PF12730)	LanT	363
HTH_31 (PF13560)	transcriptional regulator	307
Lant_dehydr_N (PF04738)	PEARL	304 (193 sLanBs)
ABC_membrane (PF00664)/ABC_tran (PF00005)	LanT	283
Lant_dehydr_C (PF14028)/PCMT (PF01135)	LanB elimination domain and Protein-L-isoaspartate(D-aspartate) O-methyltransferase fusion protein	261
Peptidase_C39 (PF03412)/ABC_membrane (PF12730)/ABC_tran (PF00005)	LanT _p	258
HATPase_c_2 (PF13581)	Histidine kinase-like ATPase	197
Response_reg (PF00072)/Trans_reg_C (PF00486)	transcriptional regulator	180
Peptidase_S8 (PF00082)	LanP _A	172
Flavoprotein (PF02441)	LanD	146
Acetyltransf_1 (PF00583)	N-acetyltransferase	142
DUF397 (PF04149)	Domain of Unknown Function	140
MFS_1 (PF07690)	Major Facilitator Superfamily protein	128
Leukocidin (PF07968)	Leukocidin/Hemolysin toxin family protein	127

Supplementary Table S7. Twenty most abundant proteins in class II BGCs that belong to at least one Pfam family. If a protein has multiple domains from different Pfam families, those families are separated by a slash. Potential tailoring enzymes are highlighted in **yellow**.

Pfam families	Description	Count
DUF4135 (PF13575)/LANC_like (PF05147)	LanM	2,163
Peptidase_C39 (PF03412)/ABC_membrane (PF12730)/ABC_tran (PF00005)	LanT _p	1,270
ABC_tran (PF00005)	LanT	886
ABC2_membrane_4 (PF12730)	LanT	761
Mersacidin (PF16934)	Mersacidin-like precursor peptide	705
L_biotic_typeA (PF04604)	Type A lanthipeptide precursor peptide	532
HTH_3 (PF01381)	transcriptional regulator	348
Lantibiotic_a (PF14867)	Alpha precursor peptide	339
Response_reg (PF00072)/GerE (PF00196)	transcriptional regulator	308
Peptidase_S8 (PF00082)	LanP _A	291
ABC_membrane (PF00664)/ABC_tran (PF00005)	LanT	222
ABC_tran (PF00005)/DUF4162 (PF13732)	LanT	213
FMN_red (PF03358)	Flavin mononucleotide reductase	186
HisKA (PF00512)/HATPase_c (PF02518)	2-component response regulator	180
Response_reg (PF00072)/Trans_reg_C (PF00486)	transcriptional regulator	164
GerE (PF00196)	transcriptional regulator	140
Nhase_alpha (PF02979)	precursor peptide with nitrile hydratase family leader peptide	119
Nif11 (PF07862)	precursor peptide with Nif11 family leader peptide	109
FtsX (PF02687)	FtsX-like permease family	104
LANC_like (PF05147)	LanC (split LanM possibly from sequencing errors)	95

Supplementary Table S8. Twenty most abundant proteins in class III BGCs that belong to at least one Pfam family. If a protein has multiple domains from different Pfam families, those families are separated by a back slash. Potential tailoring enzymes are highlighted in yellow. LanP_P is a Pro oligopeptidase that is distinct from the LanP_A subtilin-like S8 peptidases.

Pfam families	Description	Count
ABC_membrane (PF00664)/ABC_tran (PF00005)	LanT	1,514
Pkinase (PF00069)/LANC_like (PF05147)	LanKC	1,511
ABC_tran (PF00005)	LanT	781
Response_reg (PF00072)/GerE (PF00196)	transcriptional regulator	715
GerE (PF00196)	transcriptional regulator	263
MFS_1 (PF07690)	Major facilitator superfamily protein	243
adh_short (PF00106)	short chain dehydrogenase	156
GAF_2 (PF13185)/PAS_3 (PF08447)/GAF_2 (PF13185)/SpoIIIE (PF07228)/HATPase_c_2 (PF13581)	Unknown	135
trypsin (PF00089)	Protease	133
Pkinase (PF00069)	protein kinase	133
HTH_20 (PF12840)	transcriptional regulator	123
Acetyltransf_1 (PF00583)	N-acetyltransferase	122
BPD_transp_1 (PF00528)	Binding-protein-dependent transport system, inner membrane component	121
FtsX (PF02687)/FtsX (PF02687)	FtsX-like permease	119
GAF_2 (PF13185)/PAS_4 (PF08448)/GAF_2 (PF13185)/SpoIIIE (PF07228)	Unknown	114
Acetyltransf_3 (PF13302)	N-acetyltransferase	113
Peptidase_S9 (PF00326)	LanP _P	112
FecCD (PF01032)	FecCD transport family	102
Mac (PF12464) /Hexapep (PF00132)	Acetyltransferase	97
DUF4265 (PF14085)	Domain of unknown function	93

Supplementary Table S9. Twenty most abundant proteins in class IV BGCs that belong to at least one Pfam family. If a protein has multiple domains from different Pfam families, those families are separated by a back slash. Known class IV BGCs comprise only 4 genes, so these entries may include proteins encoded by genes that are not part of the gene cluster. Potential tailoring enzymes are highlighted in yellow.

Pfam families	Description	Count
Pkinase (PF00069)/LANC_like (PF05147)	LanL	340
ABC_membrane (PF00664)/ABC_tran (PF00005)	LanT	164
Peptidase_S9 (PF00326)	LanP _P	112
MFS_1 (PF07690)	Major facilitator Superfamily protein	101
ABC2_membrane (PF01061)	LanT	83
ABC_tran (PF00005)/DUF4162 (PF13732)	LanT	82
HATPase_c_2 (PF13581)	Histidine kinase-like ATPase domain	48
ABC_tran (PF00005)	LanT	48
Nif3 (PF01784)	NGG1p interacting factor 3	46
NAD_binding_10 (PF13460)	NAD(P)H-binding protein	35
STAS_2 (PF13466)	STAS domain containing protein	31
DAO (PF01266)	FAD dependent oxidoreductase	30
BPD_transp_1 (PF00528)	Binding-protein-dependent transport system, inner membrane component	30
TrmK (PF04816)	N-methyltransferase	29
Glycos_transf_2 (PF00535)	Glycosyl transferase	28
SBP_bac_3 (PF00497)	Bacterial extracellular solute-binding protein	26
Methyltransf_19 (PF04672)	Methyltransferase	26
GDP_Man_Dehyd (PF16363)	GDP-mannose 4,6-dehydratase	26
SNF2_assoc (PF08455)/SNF2_N (PF00176)/Helicase_C (PF00271)	Helicase	24
Response_reg (PF00072)/Sigma70_r4_2 (PF08281)	Response regulator	25

Supplementary Table S10. Distribution among the lanthipeptide classes of Pfams that are in the 20 most abundant protein families in a single class. Values are presented on the basis of domains, so Pfam families that occur in multidomain proteins and single domain proteins are counted together. The Pfam are listed in order of decreasing frequency.

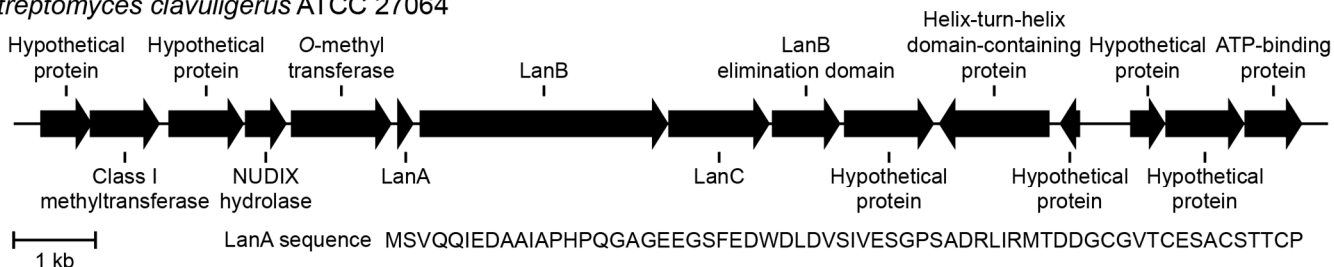
Pfam protein family	Top 20 most abundant in class	Class I	Class II	Class III	Class IV
PCMT (PF01135)	I	835	1	1	0
Flavoprotein (PF02441)	I	146	26	24	4
Acetyltransf_1 (PF00583)	I, III	149	47	142	12
FMN_red (PF03358)	II	34	186	7	4
adh_short (PF00106)	III	57	30	157	19
Acetyltransf_3 (PF13302)	III	57	12	113	9
Mac (PF12464)	III	0	0	97	0
TrmK (PF04816)	IV	0	0	0	29
DAO (PF01266)	IV	6	1	9	30
NAD_binding_10 (PF13460)	IV	21	10	19	38
GDP_Man_Dehyd (PF16363)	IV	2	0	65	26
Glycos_transf_2 (PF00535)	IV	36	8	39	29
Methyltransf_19 (PF04672)	IV	64	12	9	26

Supplementary Figure S8. Example putative biosynthetic gene clusters encoding the enzymes in Table S10. LanA genes that were not annotated in the genome are indicated in red. Because BGC boundaries are not known, the genes encoding the noted enzymes may or may not be part of the lanthipeptide BGC for all panels of Supplementary Figure S8.

O-methyltransferase containing clusters

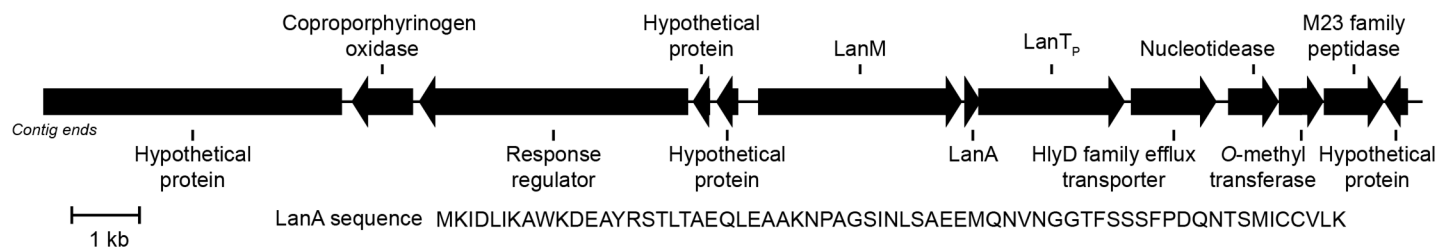
Class I

Streptomyces clavuligerus ATCC 27064



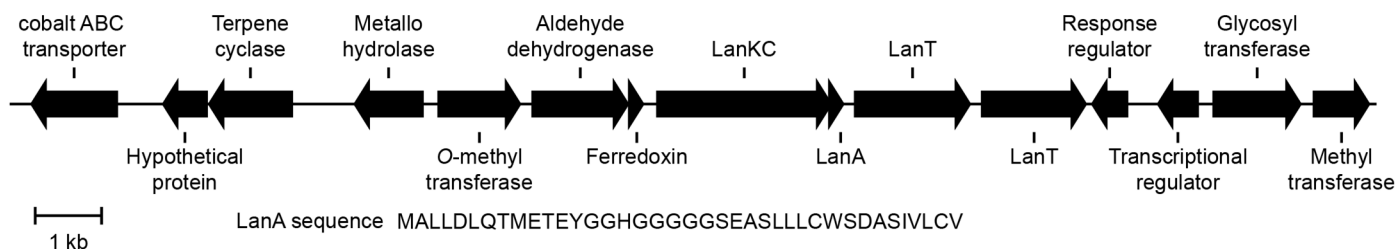
Class II

Marinicella sediminis strain F2



Class III

Streptomyces varsoviensis strain NRRL ISP-5346

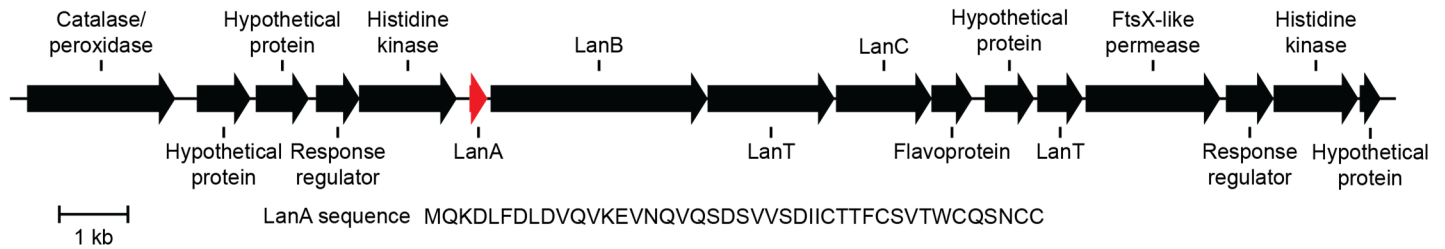


Supplementary Figure S8 continued.

Flavoprotein containing clusters

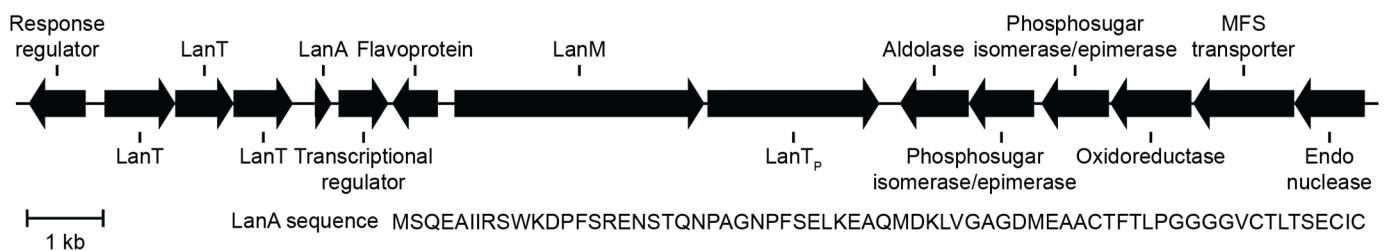
Class I

Brevibacillus sp. BC25



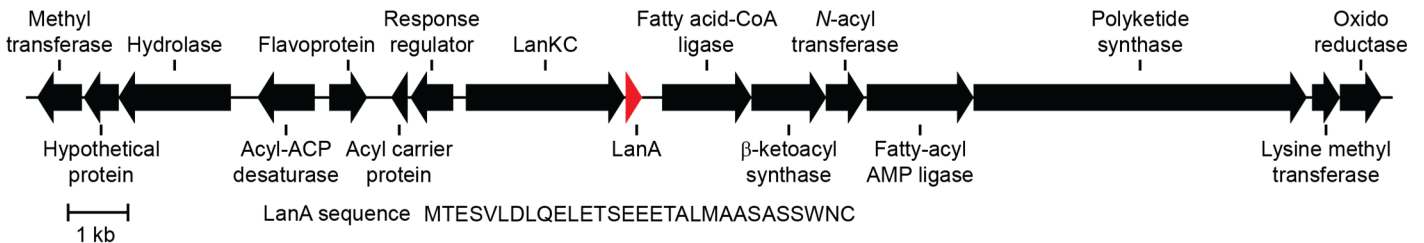
Class II

Bacillus velezensis strain GH1-13



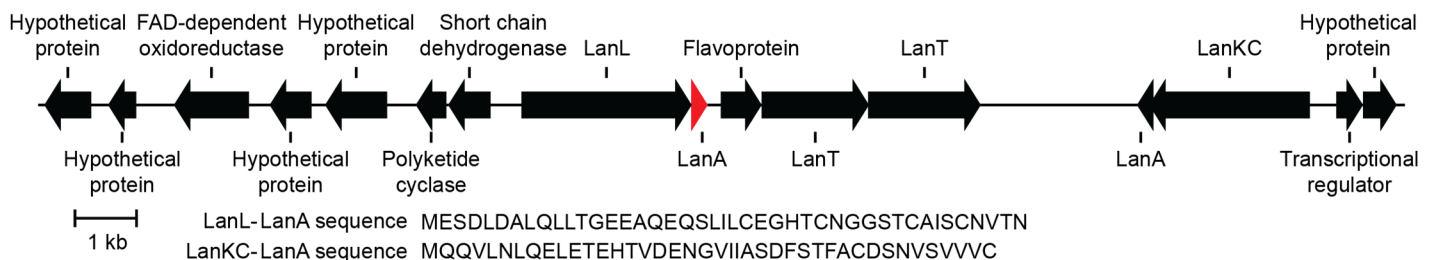
Class III

Streptomyces bicolor strain NRRL B-5348



Class IV

Streptomyces sp. Ach 505

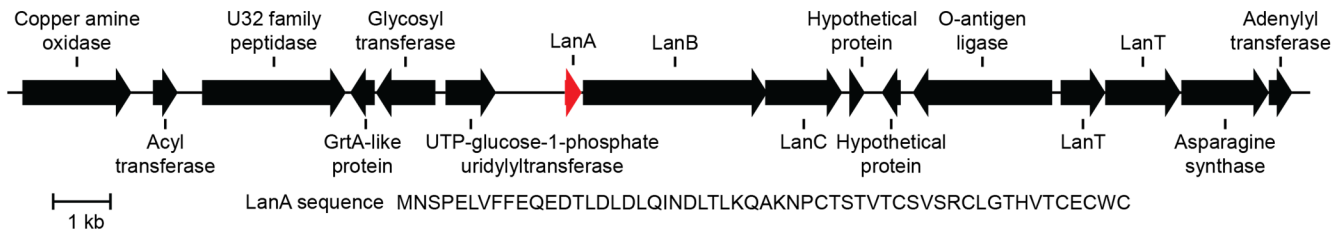


Supplementary Figure S8 continued.

Acyltransferase (Acyltransf_1) containing clusters

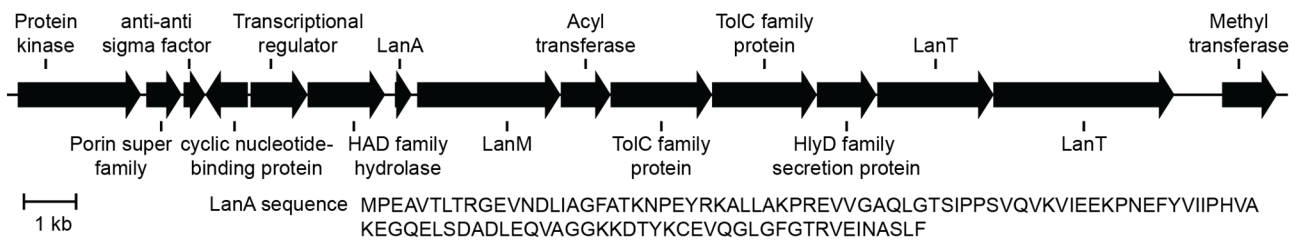
Class I

Paenibacillus polymyxa strain NCTC10343



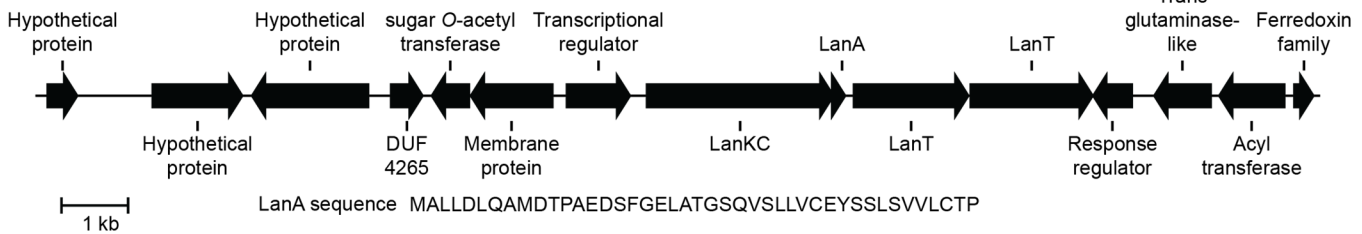
Class II

Thermoanaerobaculum aquaticum strain MP-01



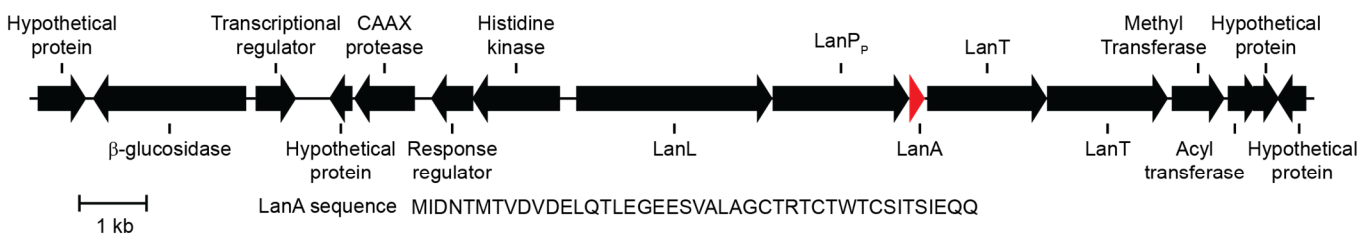
Class III

Streptomyces sp. CNS654



Class IV

Streptomyces baarnensis strain NRRL B-2842

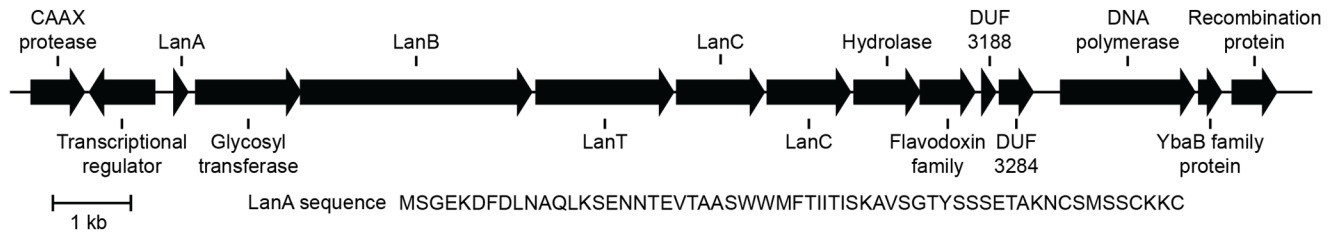


Supplementary Figure S8 continued.

FMN reductase (FMN_red) containing clusters

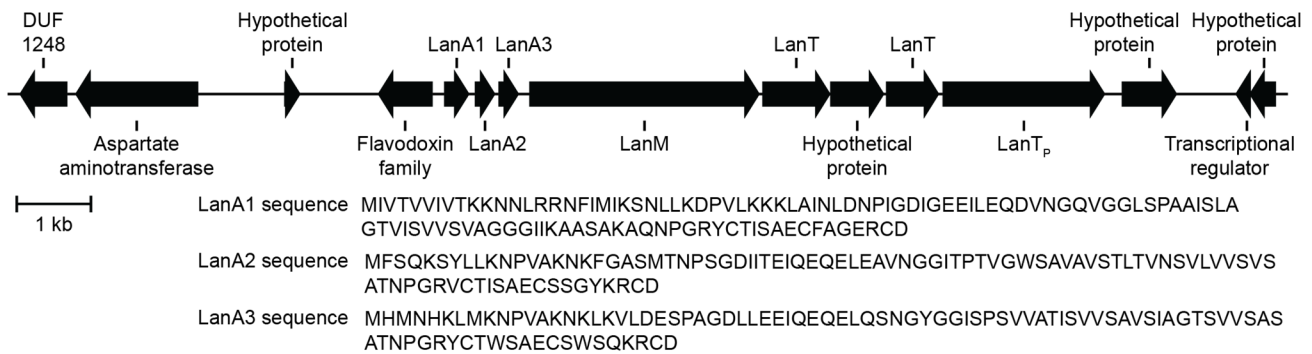
Class I

Listeria monocytogenes strain AL-OM-1-WH2



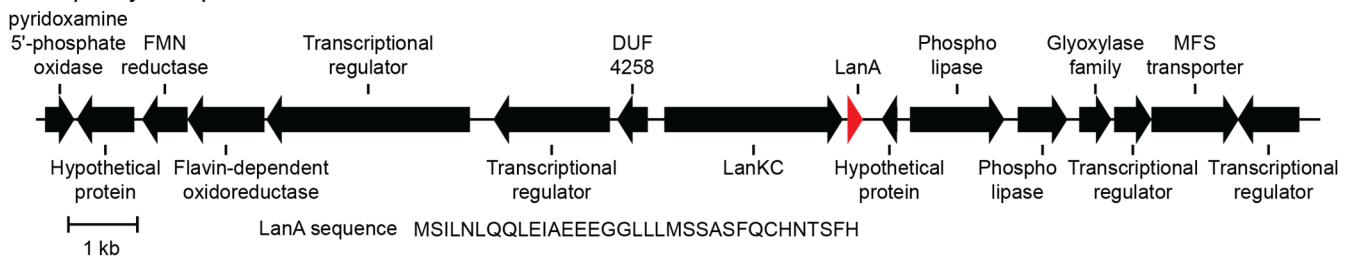
Class II

Bacillus cereus VD166



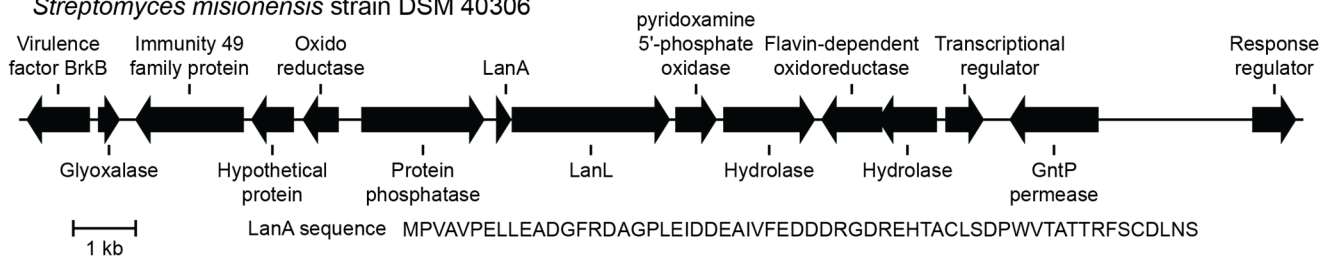
Class III

Streptomyces sp. CNS654



Class IV

Streptomyces misionensis strain DSM 40306

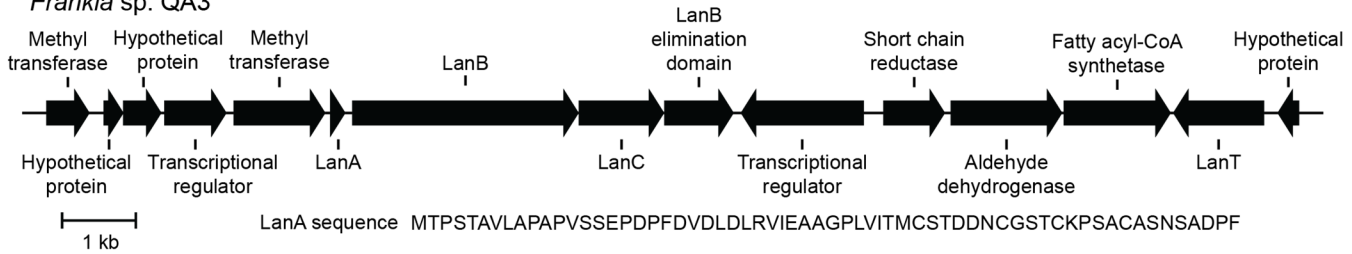


Supplementary Figure S8 continued.

Short chain dehydrogenase (adh_short) containing clusters

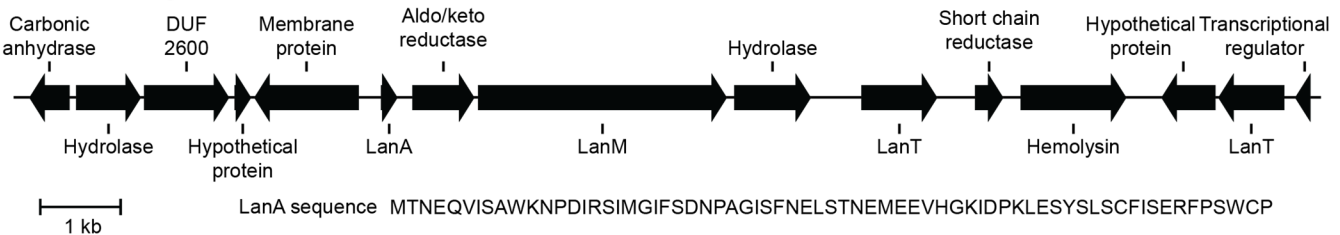
Class I

Frankia sp. QA3



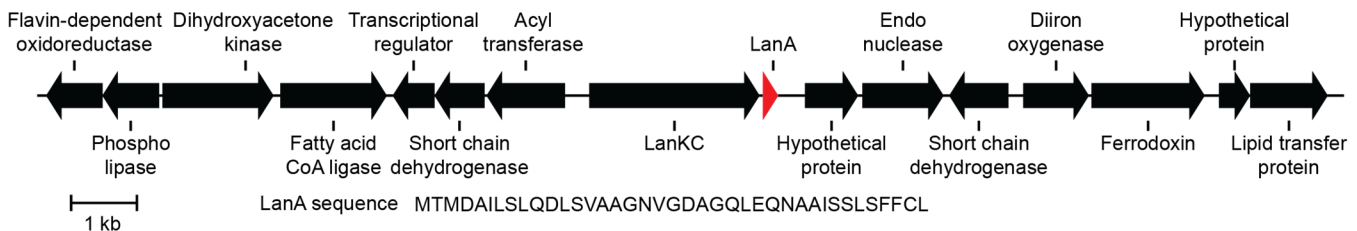
Class II

Bacillus thuringiensis MC28



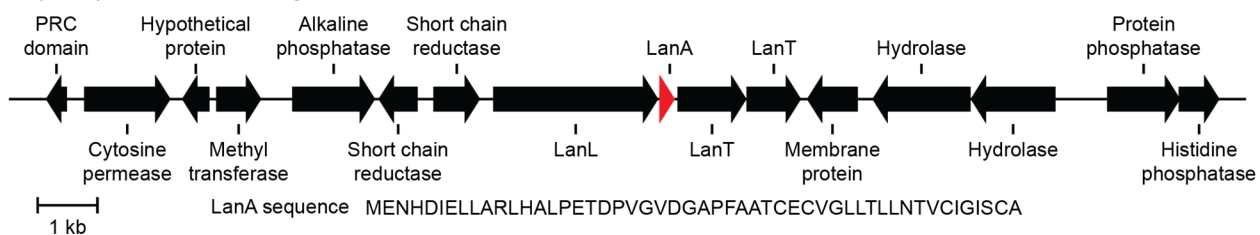
Class III

Rhodococcus erythropolis SK121



Class IV

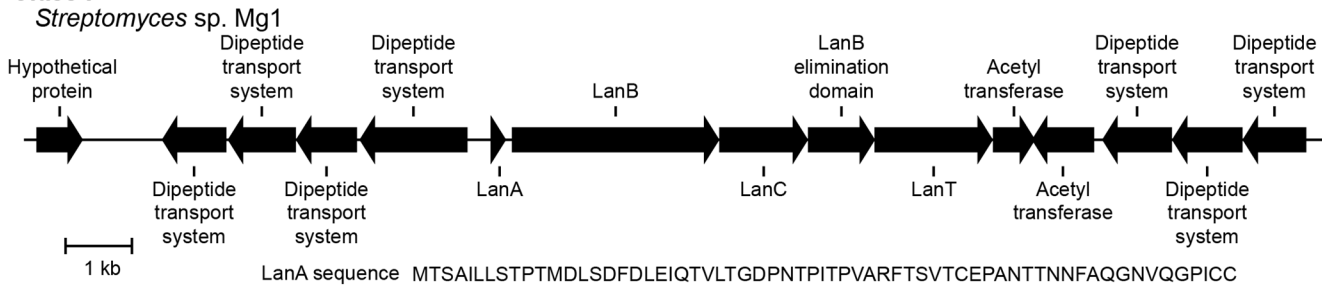
Streptomyces flavochromogenes strain NRRL B-2684



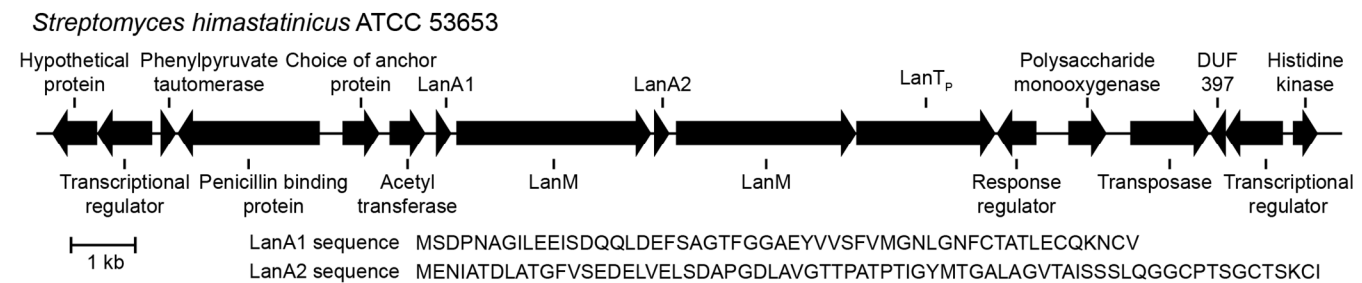
Supplementary Figure S8 continued.

Acyltransferase (Acetyltransf_3) containing clusters

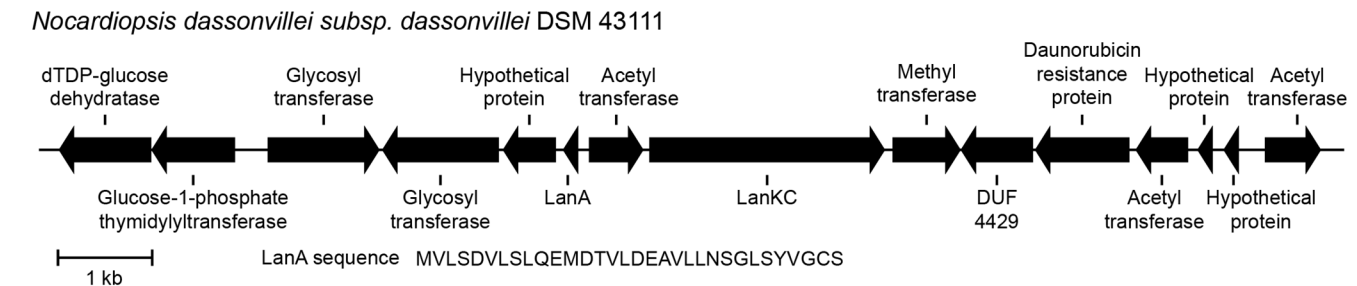
Class I



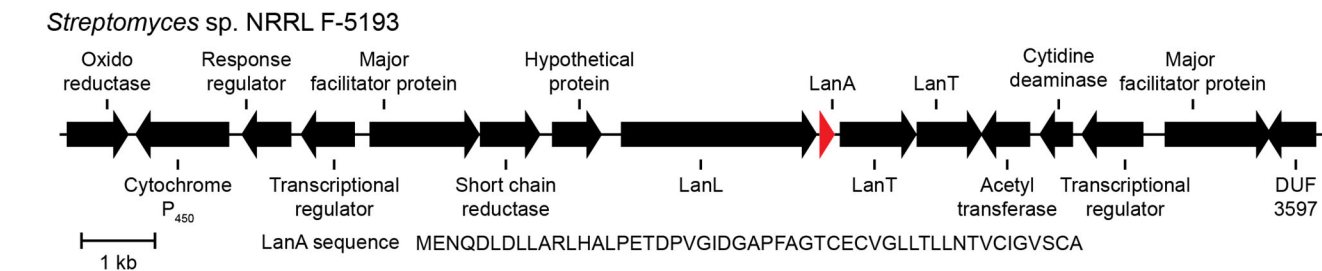
Class II



Class III



Class IV

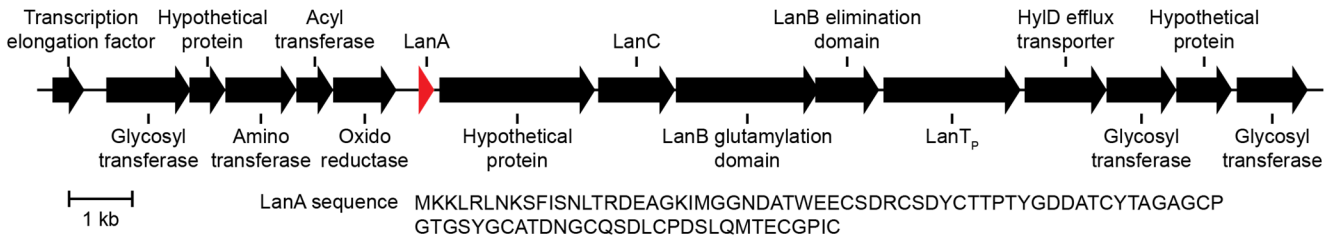


Supplementary Figure S8 continued.

Glycosyltransferase (Glycos_transf_2) containing clusters

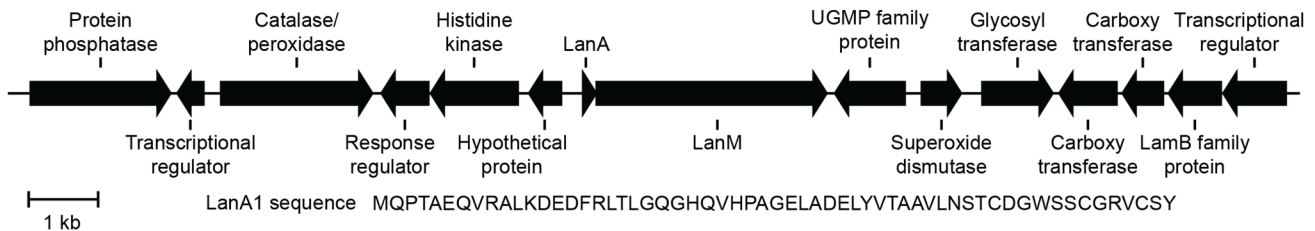
Class I

Pedobacter heparinus DSM 2366



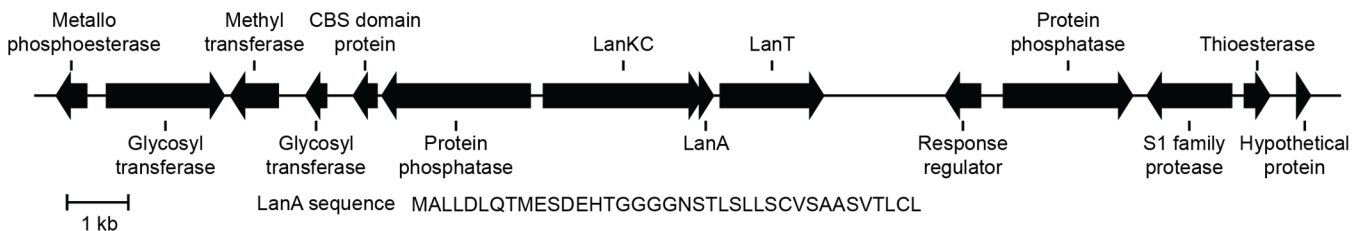
Class II

Kitasatospora aureofaciens strain NRRL B-1286



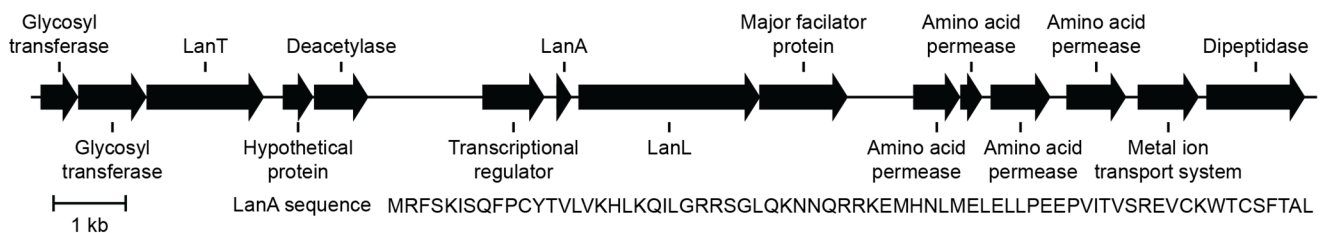
Class III

Streptomyces pseudovenezuelae strain DSM 40212



Class IV

Streptomyces sp. NRRL F-5193

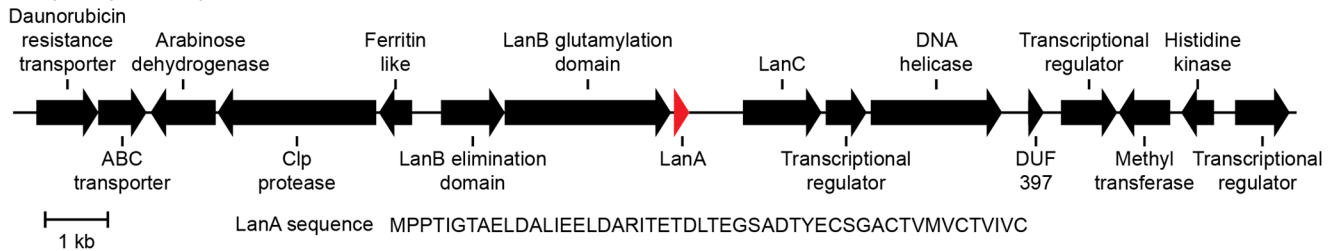


Supplementary Figure S8 continued.

Methyltransferase (Methyltransf_19) containing clusters

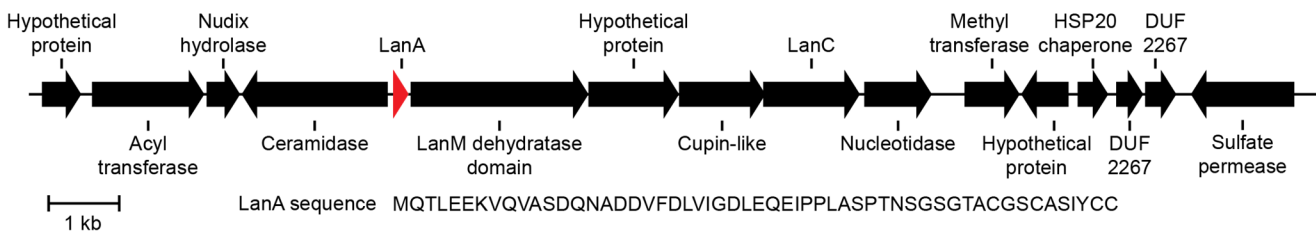
Class I

Streptomyces sulphureus DSM 40104



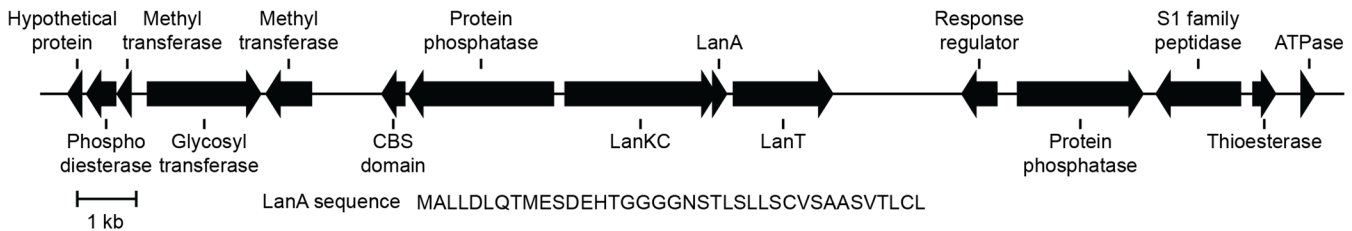
Class II

Streptomyces sp. WZ.A104



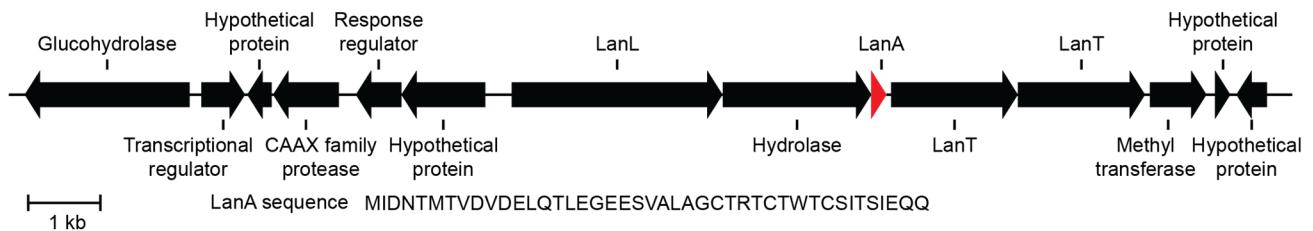
Class III

Streptomyces canus strain DSM 40017



Class IV

Streptomyces globisporus C-1027



Supplementary Table S11. Distribution of select Pfam protein families from BGCs. The enzymes belonging to these families potentially carry-out secondary post-translational modifications. Values are presented on the basis of domains, so Pfam protein families that occur in multidomain proteins or single domain proteins are counted together. NRPS: non-ribosomal peptide synthetase, PKS: polyketide synthase, FAS: fatty acid synthase.

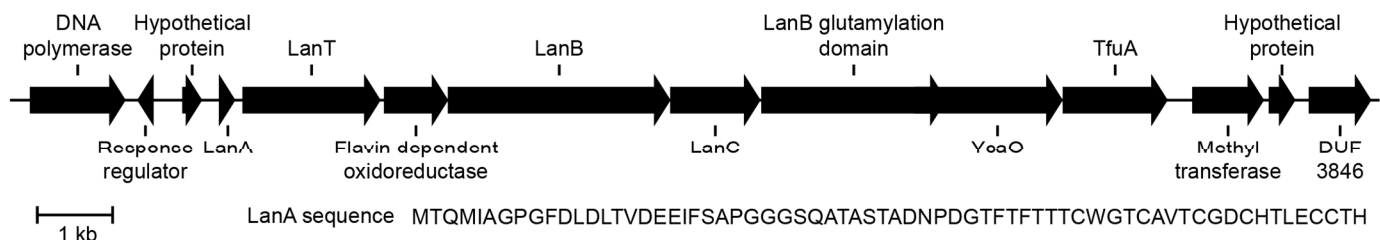
Pfam protein families	Class I	Class II	Class III	Class IV
YcaO (PF02624)	4	42	2	0
Radical_SAM (PF04055)	39	49	42	43
p450 (PF00067)	45	44	50	23
Condensation (PF00668) (NRPS)	45	31	10	2
Ketoacyl-synt_C (PF02801) (PKS/FAS)	34	5	92	8
ADH_N (PF08240) (zinc-dependent dehydrogenase)	56	63	38	27
2OG-Fell_Oxy_3 (PF13640)	0	5	4	0

Supplementary Figure S9. Example biosynthetic gene clusters encoding the enzymes in Table S11. LanA genes that were not annotated in the genome are indicated in red. Because BGC boundaries are not known, the noted enzymes may or may not be part of the lanthipeptide BGC for all panels of Supplementary Figure S9.

YcaO containing clusters

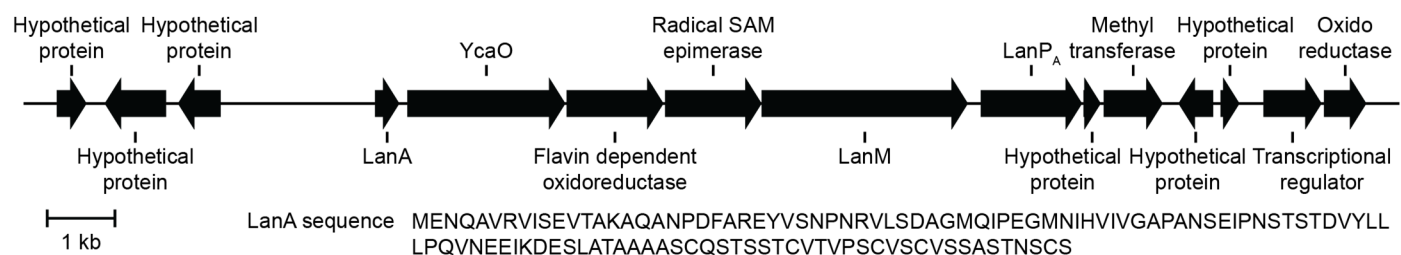
Class I

Actinokineospora enzanensis DSM 44649



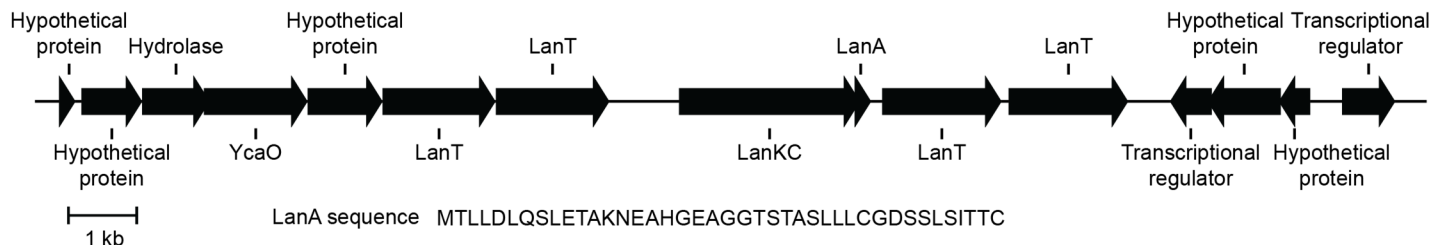
Class II

Chryseobacterium vrystaatense strain LMG 22846



Class III

Nocardiopsis valliformis DSM 45023

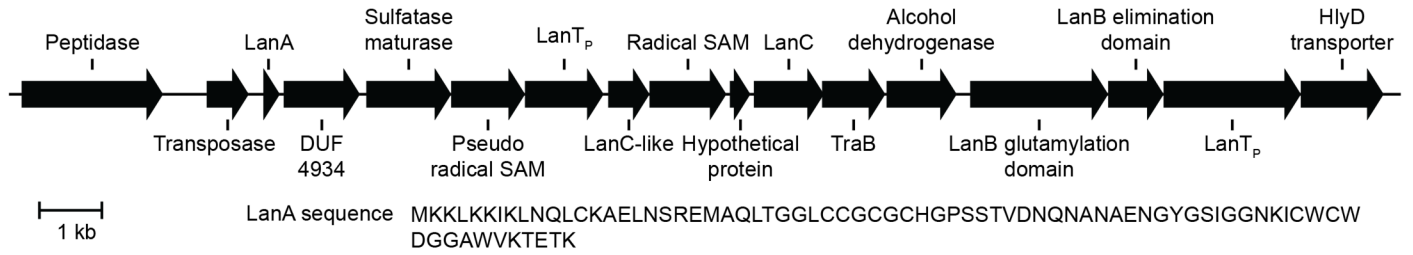


Supplementary Figure S9 continued.

Radical SAM containing clusters

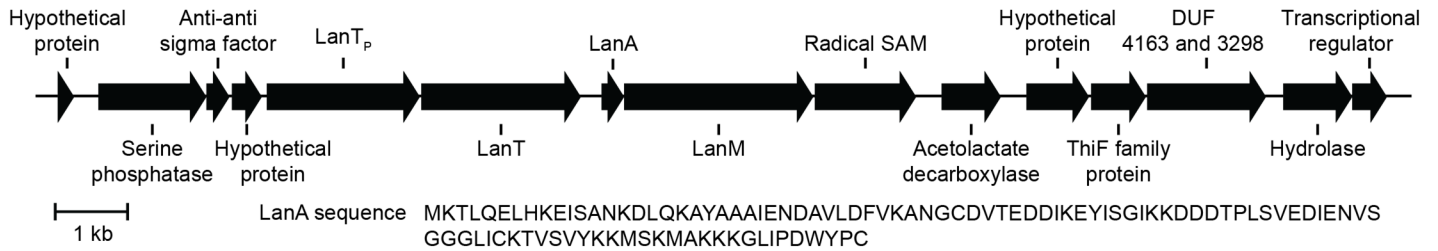
Class I

Tannerella forsythia 92A2



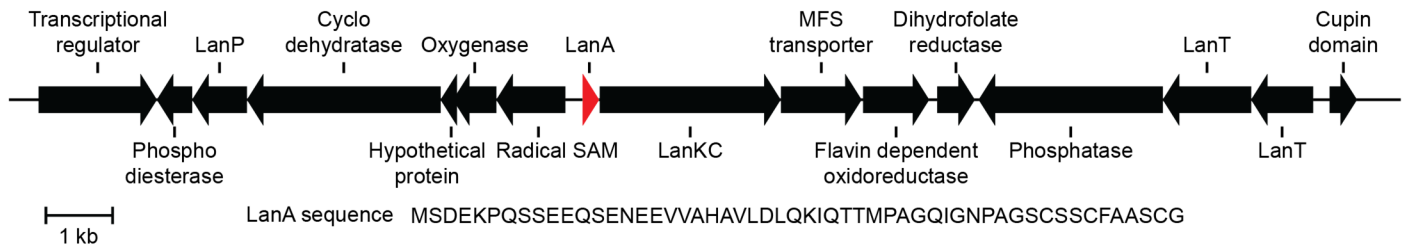
Class II

Butyrivibrio proteoclasticus strain P18



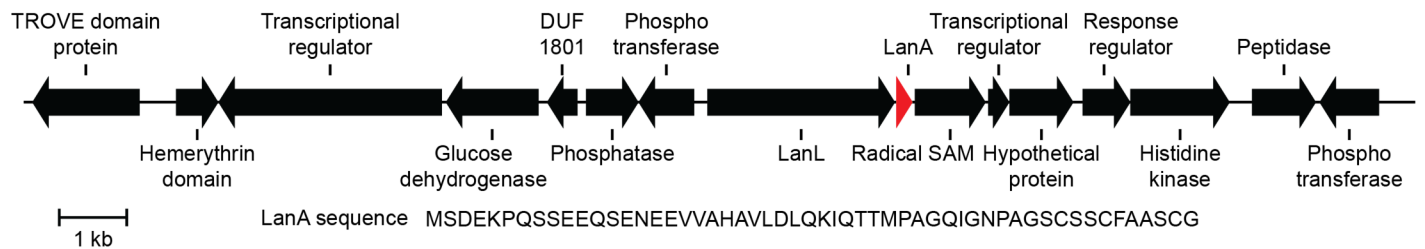
Class III

Kibdelosporangium aridum strain A82846



Class IV

Kibdelosporangium aridum strain A82846

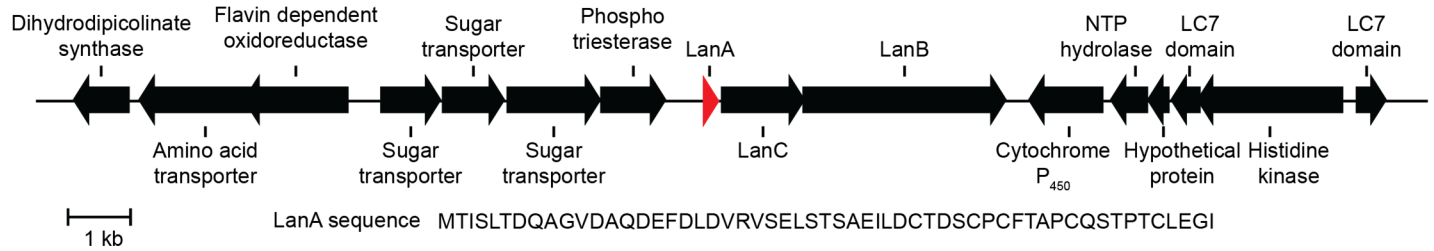


Supplementary Figure S9 continued.

Cytochrome P₄₅₀ containing clusters

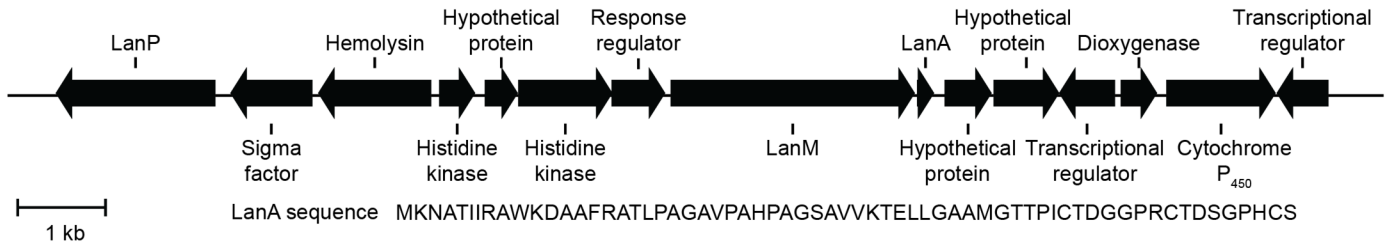
Class I

Actinomadura madurae strain DSM 43067



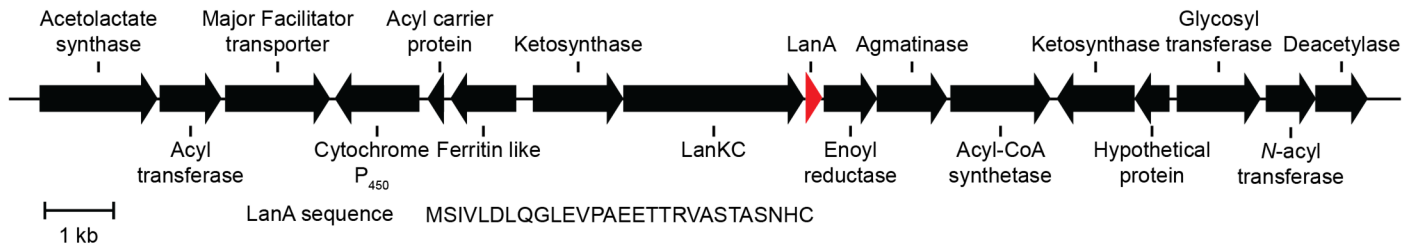
Class II

Lentzea terrae strain NEAU-LZS 42



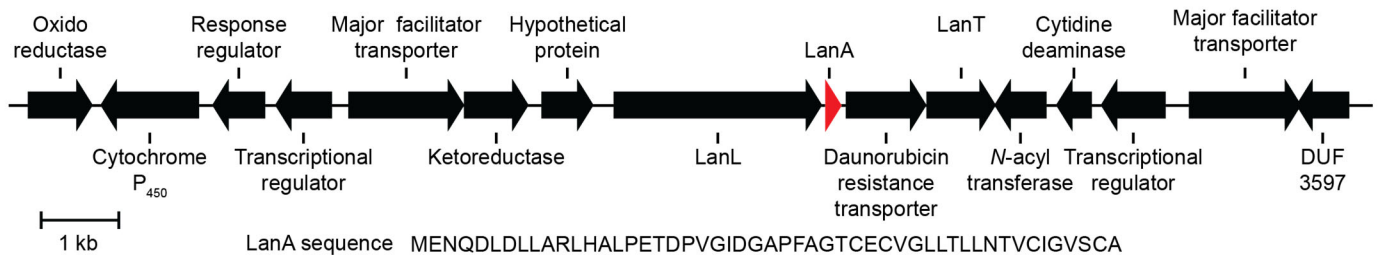
Class III

Streptomyces sp. NRRL F-4428



Class IV

Streptomyces sp. NRRL F-5193

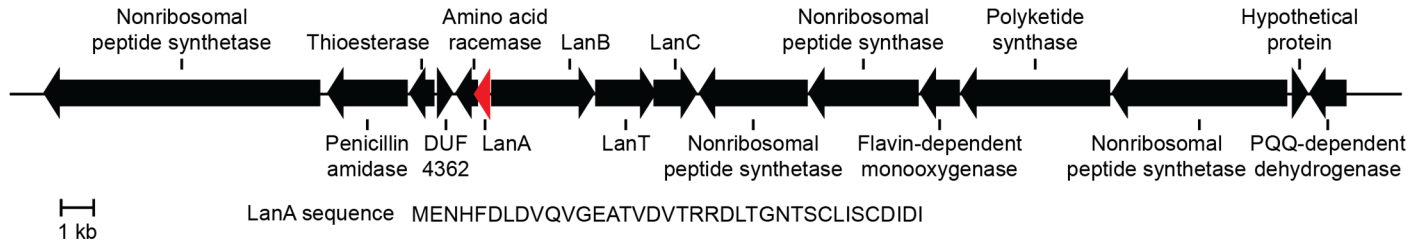


Supplementary Figure S9 continued.

Nonribosomal peptide synthetase containing clusters

Class I

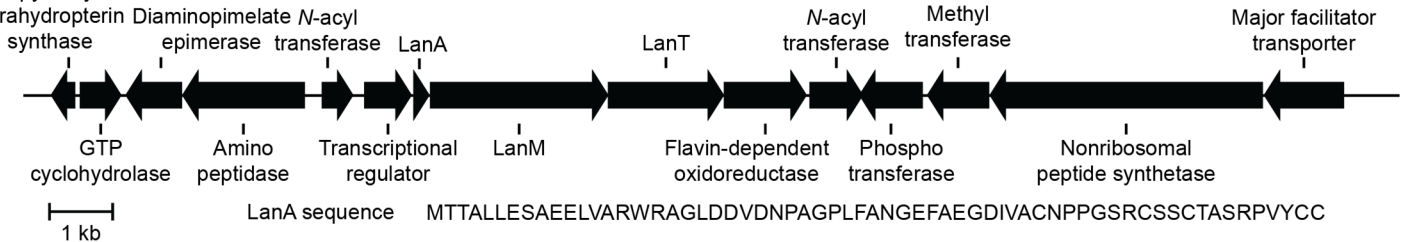
Tumebacillus avium strain AR23208



Class II

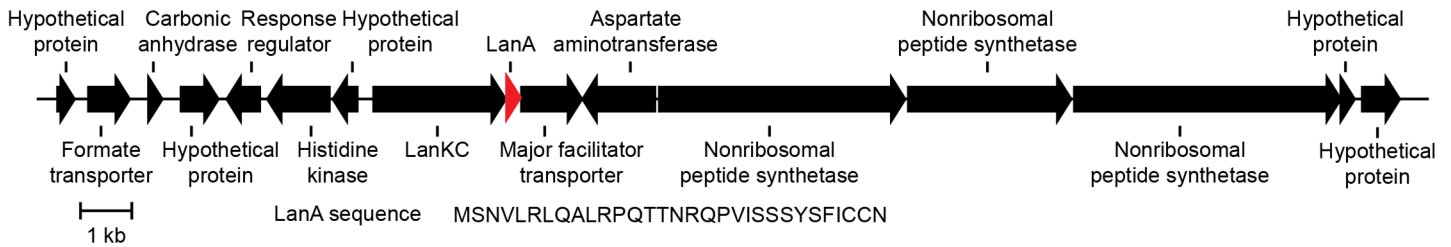
Saccharothrix variispora strain DSM 43911

6-pyruvoyl tetrahydropterin synthase



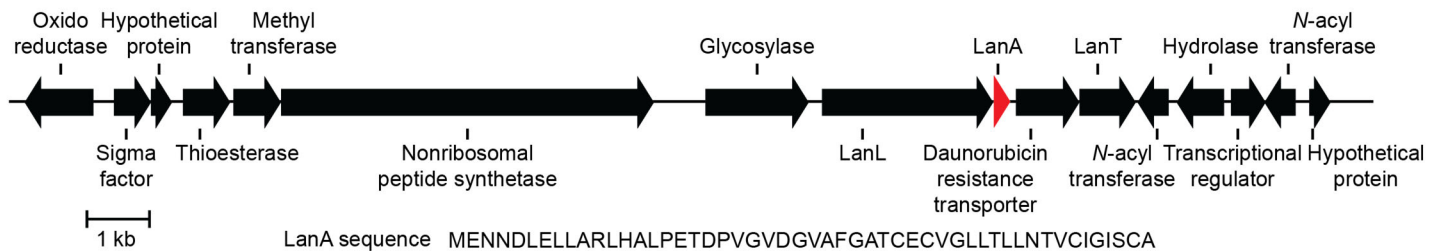
Class III

Streptomyces sp. NRRL F-4428



Class IV

Streptomyces venezuelae

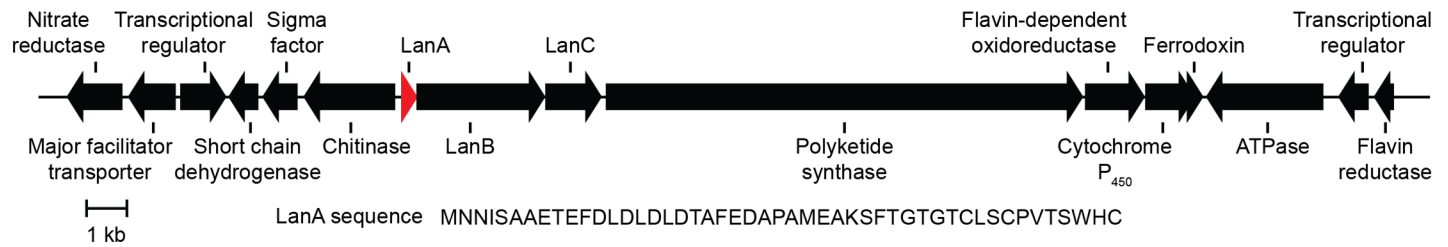


Supplementary Figure S9 continued.

Polyketide synthase containing clusters

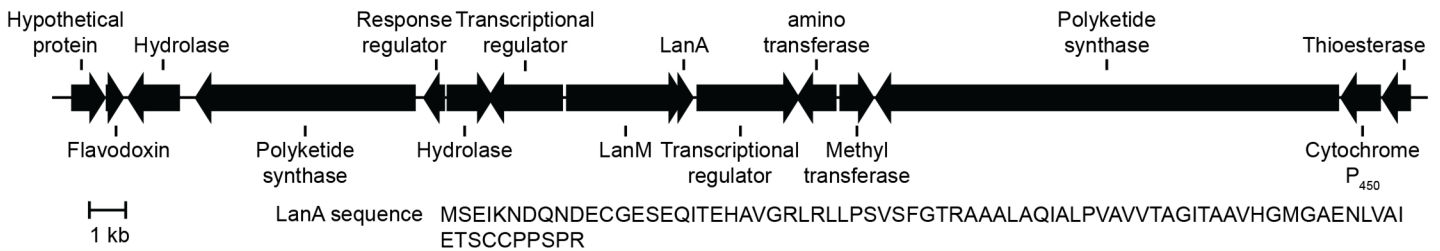
Class I

Streptomyces formicae strain KY5



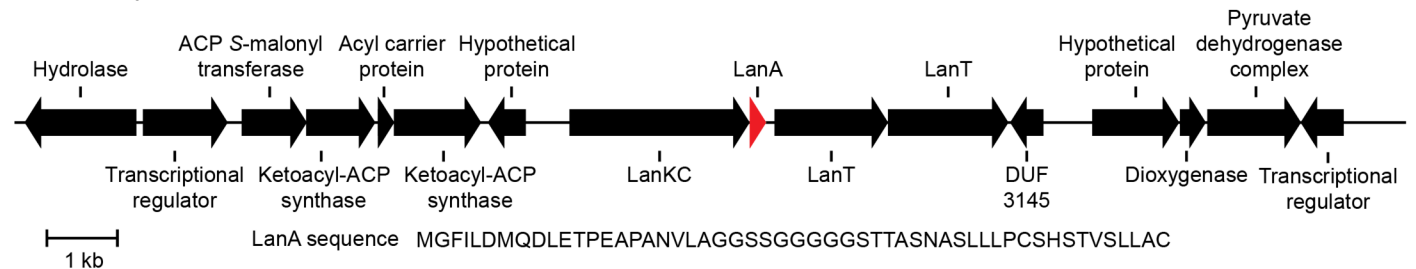
Class II

Actinoplanes brasiliensis strain DSM 43805



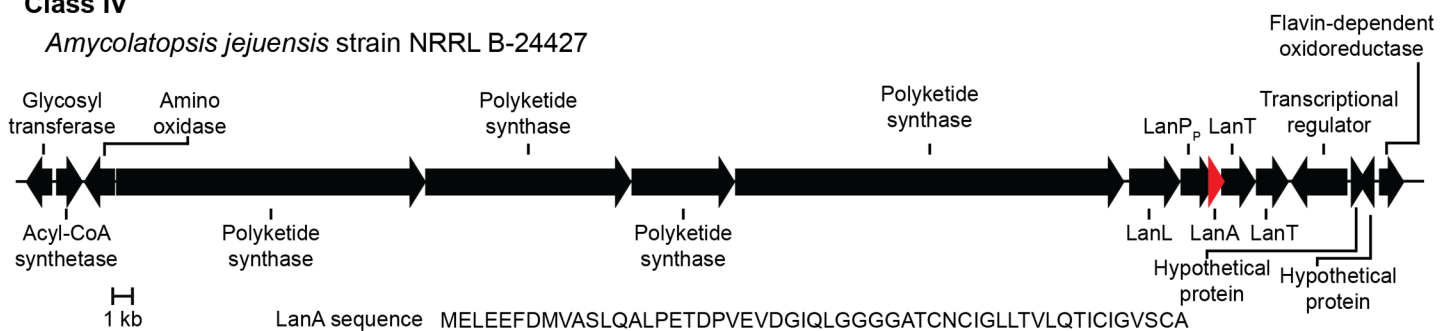
Class III

Nocardia sp. NRRL S-836



Class IV

Amycolatopsis jejuensis strain NRRL B-24427

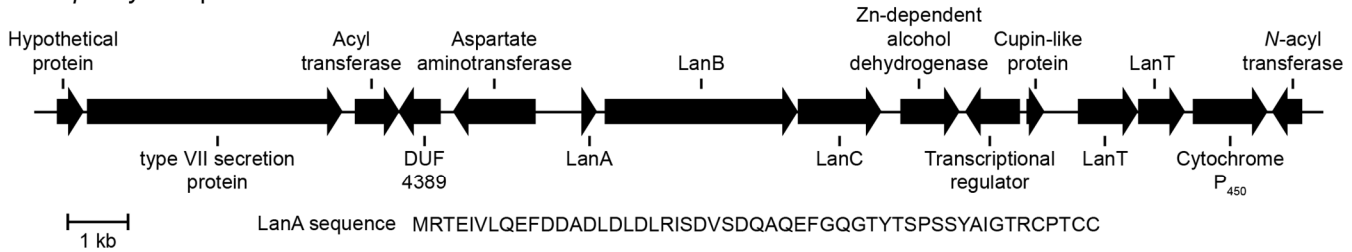


Supplementary Figure S9 continued.

Zinc-dependent alcohol dehydrogenase containing clusters

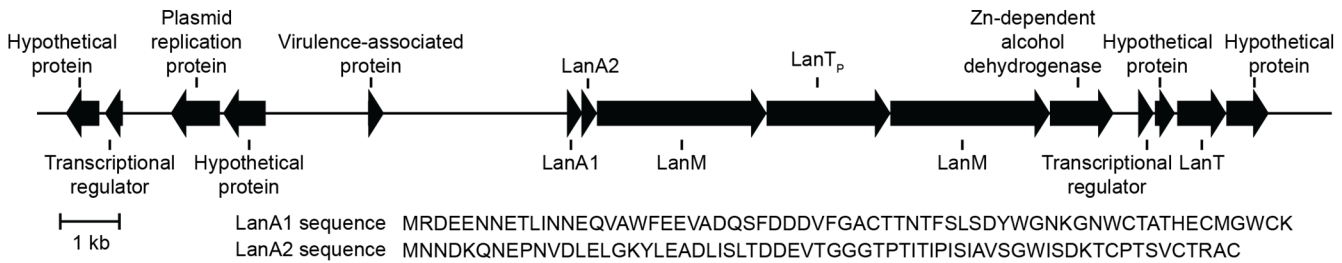
Class I

Streptomyces sp. CB02959



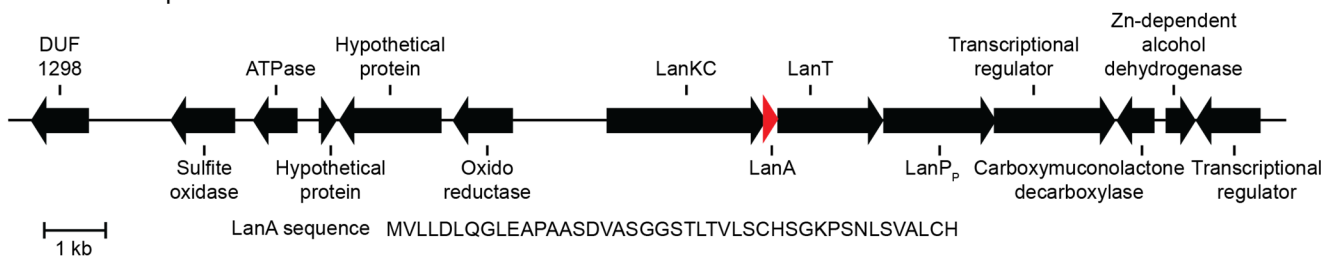
Class II

Enterococcus faecalis strain 4928STDY7071440



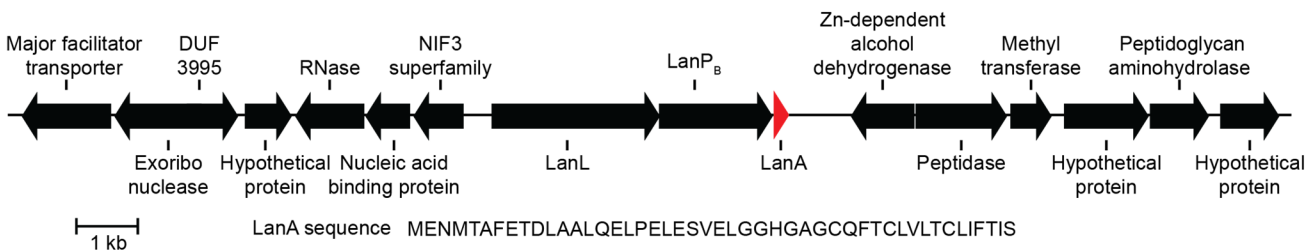
Class III

Nonomuraea sp. ATCC 55076

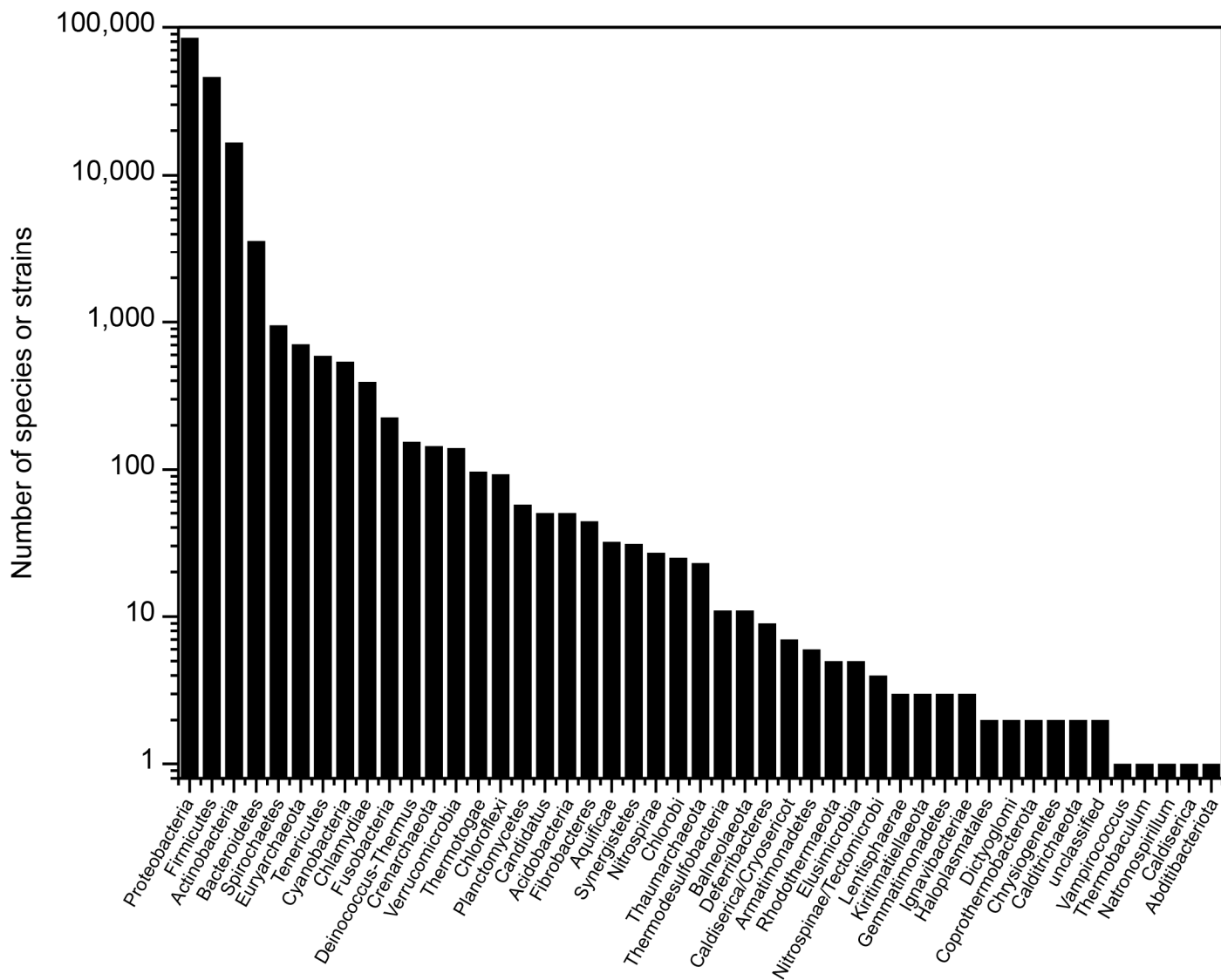


Class IV

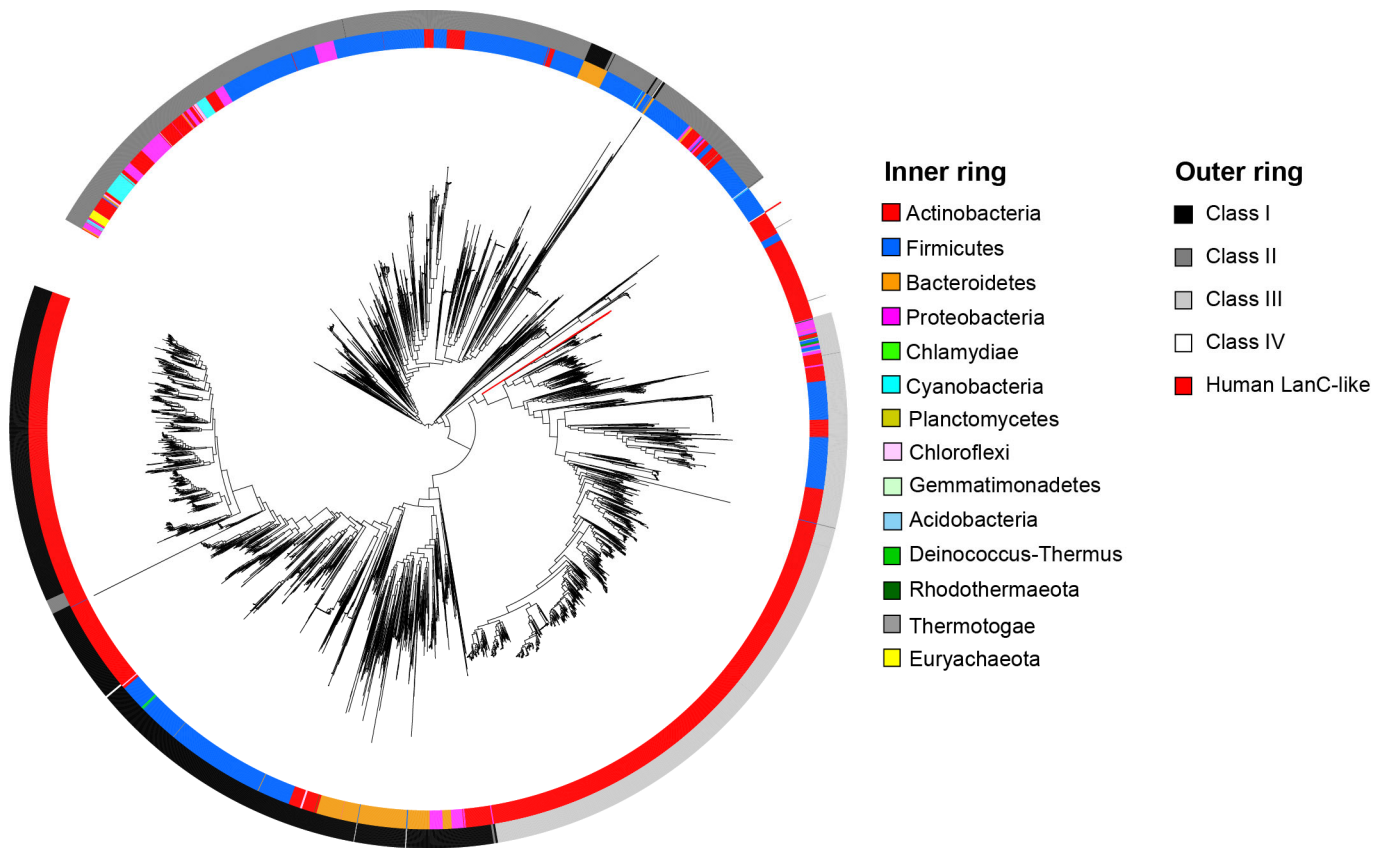
Kitatospora sp. CB01950



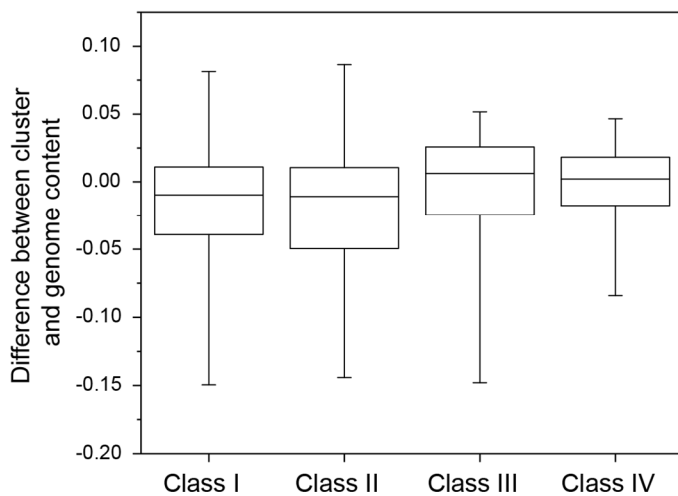
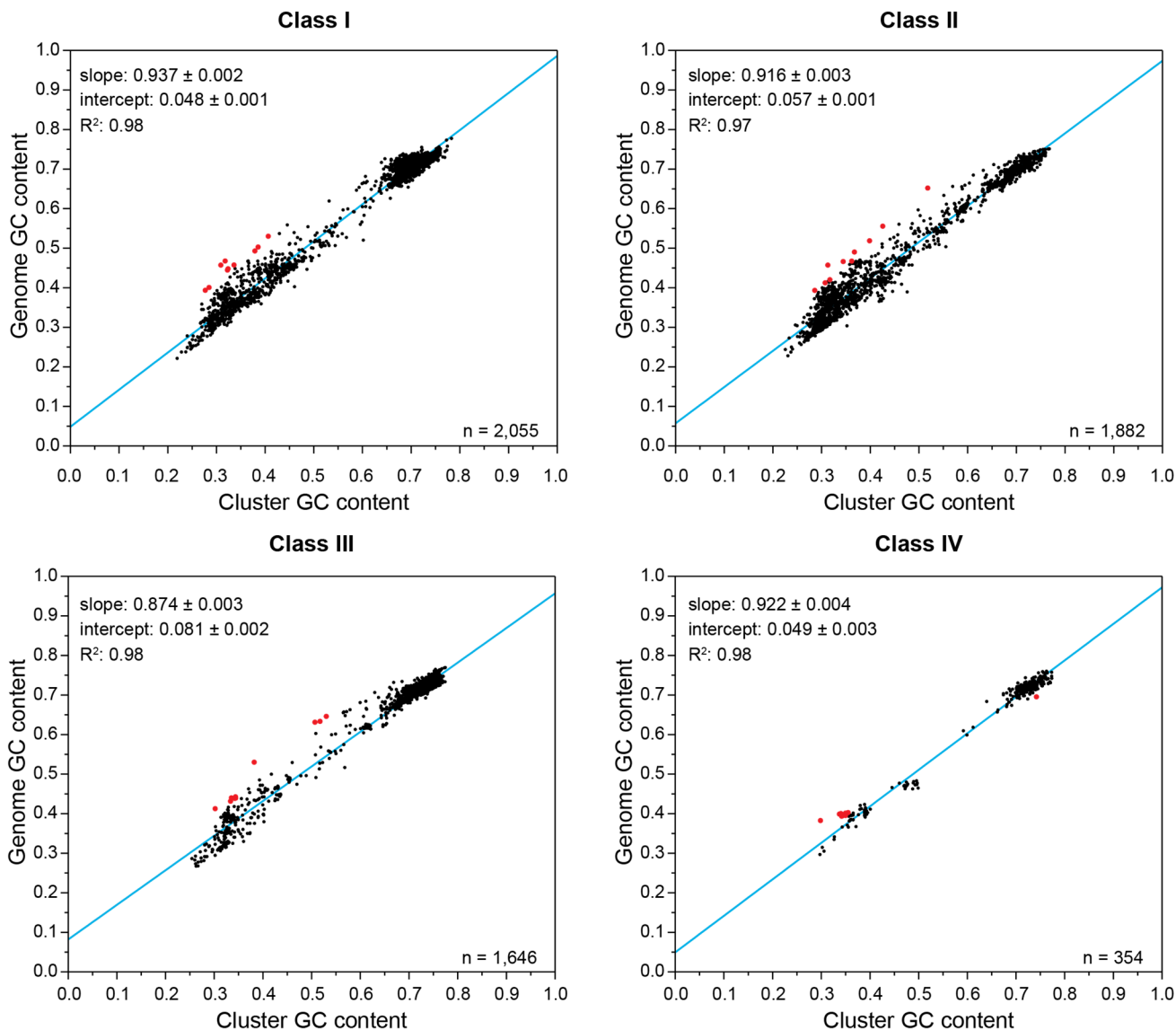
Supplementary Figure S10. Phylogenetic distribution of genomes in the dataset used for this study.



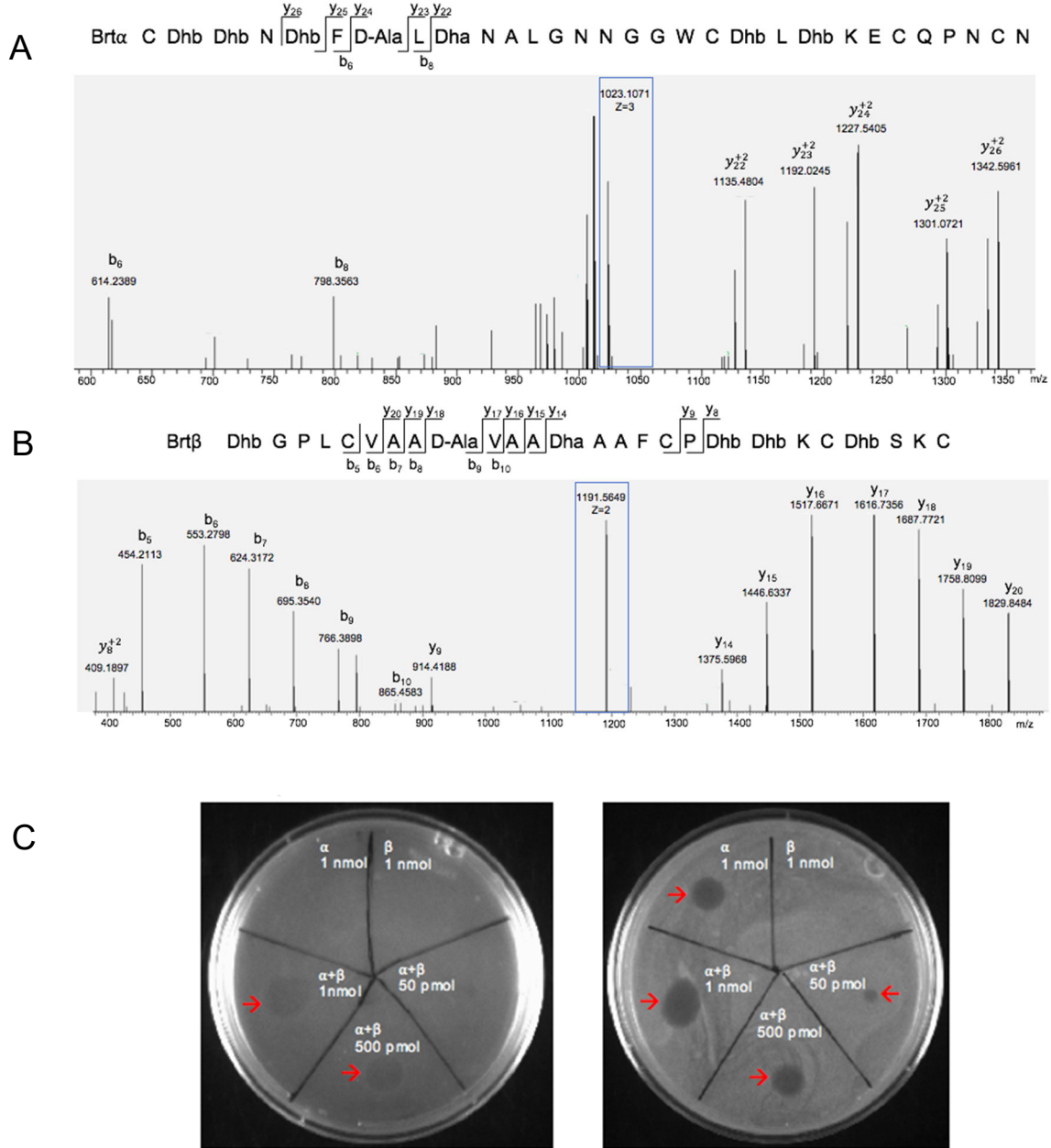
Supplementary Figure S11. An approximate maximum likelihood, midpoint rooted phylogenetic tree of LanC and LanC-like domains including human LanC-like proteins. The branches for the human LanC-like proteins are colored in red.



Supplementary Figure S12. GC content of clusters versus the cognate genome. The blue diagonal line is the linear regression with slope and intercept provided. Red points are the 10 greatest regression outliers. The box plot represents the differences between the cluster GC content and the genome GC content. The boxes indicate 1 standard deviation of the mean with the whiskers representing the maximum and minimum values.



Supplementary Figure S13. ESI MS/MS and bioactivity of birimositide α and β . A+B) Collision induced dissociation fragmentation spectrum of each peptide. The multiply-charged monoisotopic mass is boxed in blue. C) Peptides were screened against *Lactococcus lactis* sp. *cremoris* (left) and *Micrococcus luteus* ATCC 4698 (right). Red arrows indicate zones of growth inhibition.



Supplementary Table S12. Expected and observed monoisotopic masses for Brt α and Brt β using orbitrap ESI MS/MS.

Ion	Predicted (m/z)	Observed (m/z)	Error (ppm)
(Brt α + 3H) ³⁺	1023.1071	1023.1071	0.0000
(Brt α + 3H) ³⁺ y ₂₆ ⁺²	1342.5938	1342.5961	-1.7131
(Brt α + 3H) ³⁺ y ₂₅ ⁺²	1301.0752	1301.0721	2.3826
(Brt α + 3H) ³⁺ y ₂₄ ⁺²	1227.5410	1227.5405	0.4073
(Brt α + 3H) ³⁺ y ₂₃ ⁺²	1192.0225	1192.0245	-1.6778
(Brt α + 3H) ³⁺ y ₂₂ ⁺²	1135.4804	1135.4804	0.0000
(Brt α + 3H) ³⁺ b ₈	798.3603	798.3563	5.0103
(Brt α + 3H) ³⁺ b ₆	614.2391	614.2389	0.3256
(Brt β + 2H) ²⁺	1191.5649	1191.5649	0.0000
(Brt β + 2H) ²⁺ y ₂₀	1829.8495	1829.8484	0.6011
(Brt β + 2H) ²⁺ y ₁₉	1758.8124	1758.8099	1.4214
(Brt β + 2H) ²⁺ y ₁₈	1687.7753	1687.7721	1.8960
(Brt β + 2H) ²⁺ y ₁₇	1616.7382	1616.7356	1.6082
(Brt β + 2H) ²⁺ y ₁₆	1517.6698	1517.6671	1.7790
(Brt β + 2H) ²⁺ y ₁₅	1446.6327	1446.6337	-0.6913
(Brt β + 2H) ²⁺ y ₁₄	1375.5956	1375.5968	-0.8723
(Brt β + 2H) ²⁺ y ₉	914.4223	914.4188	3.8276
(Brt β + 2H) ²⁺ b ₁₀	865.4600	865.4583	1.9643
(Brt β + 2H) ²⁺ b ₉	766.3916	766.3898	2.3487
(Brt β + 2H) ²⁺ b ₈	695.3545	695.3540	0.7191
(Brt β + 2H) ²⁺ b ₇	624.3174	624.3172	0.3203
(Brt β + 2H) ²⁺ b ₆	553.2803	553.2798	0.9037
(Brt β + 2H) ²⁺ b ₅	454.2119	454.2113	1.3210
(Brt β + 2H) ²⁺ y ₈ ⁺²	409.1884	409.1897	-3.1770

References

1. Grant CE, Bailey TL, Noble WS: **FIMO: Scanning for occurrences of a given motif.** *Bioinformatics* 2011, **27**(7):1017-1018.
2. van der Meer JR, Rollema HS, Siezen RJ, Beerthuyzen MM, Kuipers OP, de Vos WM: **Influence of amino acid substitutions in the nisin leader peptide on biosynthesis and secretion of nisin by *Lactococcus lactis*.** *J Biol Chem* 1994, **269**(5):3555-3562.
3. Patton GC, Paul M, Cooper LE, Chatterjee C, van der Donk WA: **The importance of the leader sequence for directing lanthionine formation in lactacin 481.** *Biochemistry* 2008, **47**(28):7342-7351.
4. Müller WM, Enslé P, Krawczyk B, Süßmuth RD: **Leader peptide-directed processing of labyrinthopeptin A2 precursor peptide by the modifying enzyme LabKC.** *Biochemistry* 2011, **50**(39):8362-8373.
5. Hegemann JD, van der Donk WA: **Investigation of substrate recognition and biosynthesis in class IV lanthipeptide systems.** *J Am Chem Soc* 2018, **140**(17):5743-5754.
6. Zallot R, Oberg NO, Gerlt JA: **'Democratized' genomic enzymology web tools for functional assignment.** *Curr Opin Chem Biol* 2018, **47**:77-85.
7. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T: **Cytoscape: a software environment for integrated models of biomolecular interaction networks.** *Genome Res* 2003, **13**(11):2498-2504.
8. Medema MH, Kottmann R, Yilmaz P, Cummings M, Biggins JB, Blin K, de Bruijn I, Chooi YH, Claesen J, Coates RC *et al*: **Minimum Information about a Biosynthetic Gene cluster.** *Nat Chem Biol* 2015, **11**(9):625-631.
9. van Heel AJ, de Jong A, Song C, Viel JH, Kok J, Kuipers OP: **BAGEL4: a user-friendly web server to thoroughly mine RiPPs and bacteriocins.** *Nucleic Acids Res* 2018, **46**(W1):W278-w281.
10. Crooks GE, Hon G, Chandonia JM, Brenner SE: **WebLogo: a sequence logo generator.** *Genome Res* 2004, **14**(6):1188-1190.