

SUPPLEMENTARY MATERIAL

Ceftazidime-avibactam resistance mediated by the N³⁴⁶Y substitution in various AmpC β -lactamases

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Supplementary Table S1. Primers used in this study

Primers	Nucleotide sequence (5' – 3') ^a
Cloning in pTRC99k (phenotypic analysis in <i>E. coli</i> Top10)	
PDC-5_EcoRI_F	AA <u>AGAATT</u> CATGCGCGATACCAGATTCC
PDC-5_XbaI_R	AA <u>ATCTAGA</u> TCA <u>GCCTTCAGCGGCACC</u>
DHA-1_EcoRI_F	AA <u>AGAATT</u> CATGAAAAAA <u>ATCGTTATCTG</u>
DHA-1_SalI_R	AA <u>AGTCGACTT</u> ATTCCAGTGC <u>ACTCAA</u> A
FOX-3_EcoRI_F	AA <u>AGAATT</u> CATGCAACAA <u>ACGACGTGCGT</u>
FOX-3_SalI_R	AA <u>AGTCGACT</u> CA <u>CTCGGCCAA</u> CTGACTC
AmpC _{cloacae} _EcoRI_F	AA <u>AGAATT</u> CATGATGAGAAA <u>ATCCCTT</u>
AmpC _{cloacae} _SalI_R	AA <u>AGTCGACTT</u> ACTGTAGCGCCTCGAGG
Cloning in pET-TEV (protein production in <i>E. coli</i> BL21(DE3))	
PDC-5_NdeI_F	AA <u>ACATATGGCGAGGCCCCGGCGATC</u>
PDC-5_XhoI_R	AAA <u>ACTCGAGTCAGCGCTTCAGCGGCACC</u>
DHA-1_NdeI_F	AA <u>ACATATGGATAATGTCGCGGCCGGTG</u>
DHA-1_XhoI_R	AA <u>CTCGAGTTATTCCAGTGC</u> ACTCAA
FOX-3_NdeI_F	AA <u>ACATATGCGCGGGAGGCTCCGCTGAC</u>
FOX-3_XhoI_R	AA <u>CTCGAGTCACTCGGCCAA</u> CTGACTCA
AmpC _{cloacae} _NdeI_F	AA <u>ACATATGACGCCAGTGT</u> CAGAAAAAC
AmpC _{cloacae} _NotI_R	AA <u>GCGGCCGCTT</u> ACTGTAGCGCCTCGAGG
Site-directed mutagenesis	
PDC-5_N346Y_R	AA <u>AGTCGACT</u> CA <u>GCCTCAGCGG</u> CAC <u>CTGCC</u> TGCT GCTCCAGGCCGCTGAGGA <u>TGGCGTAGGC</u> GAT <u>CTTCACC</u> CG <u>CTCGCGTAGGG</u> ATAG <u>TTGCGGTT</u>
DHA-1_N346Y_R	AA <u>AGTCGACTT</u> ATTCCAGTGC <u>ACTCAA</u> AA <u>ATGCC</u> GTG CAG <u>CTTGACT</u> CTTCGGTGTACGGGTAG <u>TTTTATT</u> C
FOX-3_N346Y_R	AA <u>AGTCGACT</u> CA <u>CTCGGCCAA</u> CTGACT <u>CAGGATGGCGT</u> GAG <u>CCGCCTTCACCCTGGC</u> CTCGTAGGGATAG <u>TTGC</u>

The restriction sites used for cloning are underlined; F, forward; R, reverse. For mutagenesis, bases introduced into the nucleotide sequence to generate the N³⁴⁶Y and I³⁴⁶Y substitutions are indicated in bold. Primers used for production of AmpC variants were identical to those used for production of native AmpC cephalosporinases.

Supplementary Table S2. Wavelength (λ) and variation in the molecular extinction coefficient ($\Delta\epsilon$) used for kinetic analyses of β -lactam hydrolysis based on spectrophotometry

β -lactam	λ (nm)	$\Delta\epsilon$ ($M^{-1} cm^{-1}$)
Amoxicillin	244	-1,200
Cephalothin	265	-8,100
Cefamandole	268	-6,000
Cefoxitin	265	-7,400
Cefotaxime	255	-8,400
Ceftazidime	256	-9,800
Aztreonam	318	-400
Cefepime	257	-8,100
Nitrocefain	486	14,600

	Ambler positions 64-67										SXXX	
	1	10	20	30	40	50	60	70	80	90	100	
<i>E. cloacae</i> AmpC	---	MMRKS--	----LCCAL	LLGISCSALA	TPVSEKQLAE	VVANTITPLM	KAQSVPGMAV	AVIYQGKPHY	YTFGKADIAA	NKPVTQPTLF	EL
<i>P. aeruginosa</i> AmpC	---	MRDTRF	PCLCGIAAST	LLFATTTPAIA	GEAPADRDKA	LVDAAVQPVM	KANDIPGLAV	AISLKGEPEHY	FSYGLASKED	GRQVTPETLF	EI	SVSKTFT
<i>E. coli</i> AmpC	---	MFKT--	----TLCTL	LITASCSTFA	AP---QQIND	IVHRTITPLI	EQQKIPGMAV	AVIYQGKPHY	FTWGYADI	KQPVTTQPTLF	EL	SVSKTFT
<i>C. freundii</i> AmpC	---	MMKKS--	----ICCAL	LLTASFSTFA	AAKTEQQIAD	IVNRTITPLM	QEQAIPGMAV	AIIYEGKPHY	FTWGKADIAN	NHPVTQPTLF	EL	SVSKTFN
<i>H. alvei</i> AmpC	MRKKMQNT--	-LKLLSVITC	LAATAQGALA	ANIDESKIKD	TVDDLIQPLM	QKNNIIPGMSV	AVTVNGKNYI	YNYGLAAKQP	QQPVTTENTLF	EV	SLSKTFA	
<i>M. morganii</i> AmpC	---	MMKKS--	----LSATL	ISALLAFSAP	GFSAADNVAA	VVDSTIKPLM	AQQDIPGMAV	AVSVKGKPHY	FNYGFADVQA	KQPVTTENTLF	EL	SVSKTFT
<i>P. stuartii</i> AmpC	---	MDNS--	-MKNIFRQGR	LFIASLAMT	SISAFALTQQ	EVDDIIQPLM	KQEQAIPGMVS	AISVNGKQAI	YHYGVQSKQT	QIPVSDRTLY	EI	SLSKTFT
<i>S. marcescens</i> AmpC	-----	-----	MTKMNR	LAALALIAALI	IPTAHAAQQQ	DIDAVIQPLM	KKYVGPGMAI	AVSVDGKQOI	YPYGVASKQT	GKPITEQPTLF	EV	SLSKTFT
CMY-1	---	MQQRQS	ILWG--AVAT	LMWAGLAHAG	EASPVDPLR	VVDASIOPLL	KEHRIPGMAV	AVLKDGAHY	FNYGVANRES	GAGVSEQPTLF	EI	SVSKTLT
DHA-1	---	MMKKS--	----LSATL	ISALLAFSAP	GFSAADNVAA	VVDSTIKPLM	AQQDIPGMAV	AVSVKGKPHY	FNYGFADIQA	KQPVTTENTLF	EL	SVSKTFT
ACC-1	---	MQNT--	-LKLLSVITC	LAATVQGALA	ANIDESKIKD	TVDDLIQPLM	QKNNIIPGMSV	AVTVNGKNYI	YNYGLAAKQP	QQPVTTENTLF	EV	SLSKTFA
ACT-1	---	MMMTKS--	----LCCAL	LLSTSCSVLA	TPMSEKQLAE	VVERTVTPLM	KAQAIPGMAV	AVIYEGQPHY	FTFGKADVA	NKPVTQPTLF	EL	SISKTFT
MIR-1	---	MMMTKS--	----LSCAL	LLSVASSAFA	APMSEKQLAE	VVERTVTPLM	NAQAIPGMAV	AVIYEGQPHY	FTFGKADVA	NKPVTQPTLF	EL	SISKTFT
MOX-1	---	MQQRQS	ILWG--AVAT	LMWAGLAHAG	EASPVDPLR	VVDASIOPLL	KEHRIPGMAV	AVLKDGAHY	FNYGVANRES	GASVSEQPTLF	EI	SVSKTLT
BIL-1	---	MMKKS--	----LCCAL	LLTASFSTFA	AAKTEQQIAD	IVNRTITPLM	QEQAIPGMAV	AVIYEGQPHY	FTWGKADIAN	NHPVTQPTLF	EL	SVSKTFN
LAT-1	---	MMKKS--	----LCSAL	LLTASFSTFA	AAKTEQQIAD	IVNRTITPLM	QEQAIPGMAV	AVIYEGQPHY	FTWGKADIAN	NHPVTQPTLF	EL	SVSKTFN
FOX-3	---	MQORRA	FALL--TLGS	LLLAPCTYA-	--SGEAPLTA	TVDGIIQPM	KEYRIPGI	AVLKDGAHY	FNYGVANRES	GQRVSEQPTLF	EI	SVSKTLT
CFE-1	---	MMKKS--	----ICCAL	LLTASFSTFA	AAKTEQQIAD	IVNRTITPLM	QEQAIPGMAV	AIIYEGQPHY	FTWGKADIAN	NRPVTQPTLF	EL	SVSKTFN
CMY-2	---	MMKKS--	----LCCAL	LLTASFSTFA	AAKTEQQIAD	IVNRTITPLM	QEQAIPGMAV	AVIYEGQPHY	FTWGKADIAN	NHPVTQPTLF	EL	SVSKTFN
	Ambler position 120										Ambler positions 150-152 YXN	
	101	110	120	130	140	150	160	170	180	190	200	
<i>E. cloacae</i> AmpC	GVLGGDAIAR	GEISLDDAVT	RYWPQLTGKQ	WQGIRMLDLA	TYTAGGLPLQ	VPDEVTDN-A	SLLRFYQNWQ	PWPKPGTTRL	YANASIGLFG	ALAVKPSGMP	
<i>P. aeruginosa</i> AmpC	ATLAGYALAQ	DKMLRDDRAS	QHWPALQGSR	FDGISLDDLA	TYTAGGLPLQ	FPDSVQKDQA	QIRDYYRQWQ	PTYAPGSQLR	YSNPSIGLFG	YLAARSLGQP		
<i>E. coli</i> AmpC	GVLGGDAIAR	GEIKLSDPTT	KYWPELTAKQ	WNGITLLHLA	TYTAGGLPLQ	VPDEVKSS-S	DLLRFYQNWQ	PAWAPGTQRL	YANSSIGLFG	ALAVKPSGLS		
<i>C. freundii</i> AmpC	GVLGGDRIAR	GEIKLSDPVT	KYWPELTGKQ	WRGISLHLA	TYTAGGLPLQ	IPGDVTDK-A	ELLRFYQNWQ	PQWTPTGAKRL	YANSSIGLFG	ALAVKSSGMS		
<i>H. alvei</i> AmpC	ATLASYAQVS	GKLSDLQSVS	HYVPELRGSS	FDHVSVLNVG	THTSG-LQLF	MPEDIKNT-T	QIMAYLKA	PADAAGTHR	YSNIGTGLLG	MIAAKSLGVS		
<i>M. morganii</i> AmpC	GVLGAVSVAK	KEMTLDNDAE	KYQPELALPQ	WKGITLLDLA	TYTAGGLPLQ	VPDAVKSR-A	DLLHFYQQWQ	PSRKPGDMRL	YANSSIGLFG	ALTANAAGMP		
<i>P. stuartii</i> AmpC	ATLATYAQIQ	GKLDQSOSV	HYVPELRGSS	FDNVSMVNLA	THTSG-LSLF	VPSDIKTN-D	QIMAYYQKWL	PDNEVGQYRS	YSNLGVGLLG	IVTAKQLNMP		
<i>S. marcescens</i> AmpC	ATLAVYAAQQQ	GKLSKDPAS	RYVPELRGSA	FDGVSLLNLA	THTSG-LPLF	VPDDVTDN-A	QIMAYYQKWL	PKHPAGSYRV	YSNLGIGMLG	MIAAKSLDQP		
CMY-1	ATLGAYAVVK	GAMQLDDKAS	RHAPWLKGSA	FDSITMGELA	TYTAGGLPLQ	FPEEVDS	E KMRAYYRQWA	PVYSPGSHRQ	YSNPSIGLFG	HIAASSLKQP		
DHA-1	GVLGAVSVAK	KEMALNDPAA	KYQPELALPQ	WKGITLLDLA	TYTAGGLPLQ	VPDAVKSR-A	DLLNFYQQWQ	PSRKPGDMRL	YANSSIGLFG	ALTANAAGMP		
ACC-1	ATLASYAQVS	GKLSDLQSVS	HYVPELRGSS	FDHVSVLNVG	THTSG-LQLF	MPEDIKNT-T	QIMAYLKA	PADAAGTHR	YSNIGTGLLG	MIAAKSLGVS		
ACT-1	GVLGGDAIAR	GEISLGDPVT	KYWPELTGKQ	WQGIRMLDLA	TYTAGGLPLQ	VPDEVKDN-A	SLLRFYQNWQ	PQWKPGTTRL	YANASIGLFG	ALAVKPSGMS		
MIR-1	GVLGGDAIAR	GEIALGDPVA	KYWPELTGKQ	WQGIRMLDLA	TYTAGGLPLQ	VPDEVTDT-A	SLLRFYQNWQ	PQWKPGTTRL	YANASIGLFG	ALAVKPSGMS		
MOX-1	ATLGAYAVVK	GAMQLDDKAS	RHAPWLKGSA	FDSITMGELA	TYTAGGLPLQ	FPEEVDS	E KMRAYYRQWA	PVYSPGSHRQ	YSNPSIGLFG	HIAASSLKQP		
BIL-1	GVLGRDAIAR	GEIKLSDPVT	KYWPELTGKQ	WQGIRLLHLA	TYTAGGLPLQ	IPDDVRDK-A	ALLHFYQNWQ	PQWTPTGAKRL	YANSSIGLFG	ALAVKPSGMS		
LAT-1	GVLGGDCIAR	GEIKLSDPVT	KYWPELTGKK	WQGIRLLHLA	TYTAGGLPLQ	IPDDVRDK-A	ALLHFYQNWQ	PQWTPTGAKRL	YANSSIGLFG	ALAVKPSGMS		
FOX-3	ATLGAYAAVK	GGFVLDDKVS	QHAPWLKGSA	LDGVTMAELA	TYTAGGLPLQ	FFDKVDSN-D	KMOTYYRSWS	PVYPAGTHR	YSNPSIGLFG	HIAANSLGQP		
CFE-1	GVLGGDAIAR	GEIKLSDPVT	QYWPELTGKQ	WQGISLHLA	TYTAGGLPLQ	VPDDVTDK-A	ALLRFYQNWQ	PQWAPGAKRL	YANSSIGLFG	ALAVKPSGMS		

CMY-2

GVLGGDAIAR GEIKLSDPVT KYWPELTGKQ WQGIRLLHLA TYTAGGLPLQ IPDDVRDK-A ALLHFYQNWQ PQWTGAKRL YANSSIGLFG ALAVKPSGMS

<i>E. cloacae</i>	AmpC
<i>P. aeruginosa</i>	AmpC
<i>E. coli</i>	AmpC
<i>C. freundii</i>	AmpC
<i>H. alvei</i>	AmpC
<i>M. morganii</i>	AmpC
<i>P. stuartii</i>	AmpC
<i>S. marcescens</i>	AmpC
CMY-1	
DHA-1	
ACC-1	
ACT-1	
MIR-1	
MOX-1	
BIL-1	
LAT-1	
FOX-3	
CFE-1	
CMY-2	

201 210 220 230 240 250 260 270 280 290 300

E. cloacae AmpC
P. aeruginosa AmpC
E. coli AmpC
C. freundii AmpC
H. alvei AmpC
M. morganii AmpC
P. stuartii AmpC
S. marcescens AmpC
CMY-1
DHA-1
ACC-1
ACT-1
MIR-1
MOX-1
BIL-1
LAT-1
FOX-3
CFE-1
CMY-2

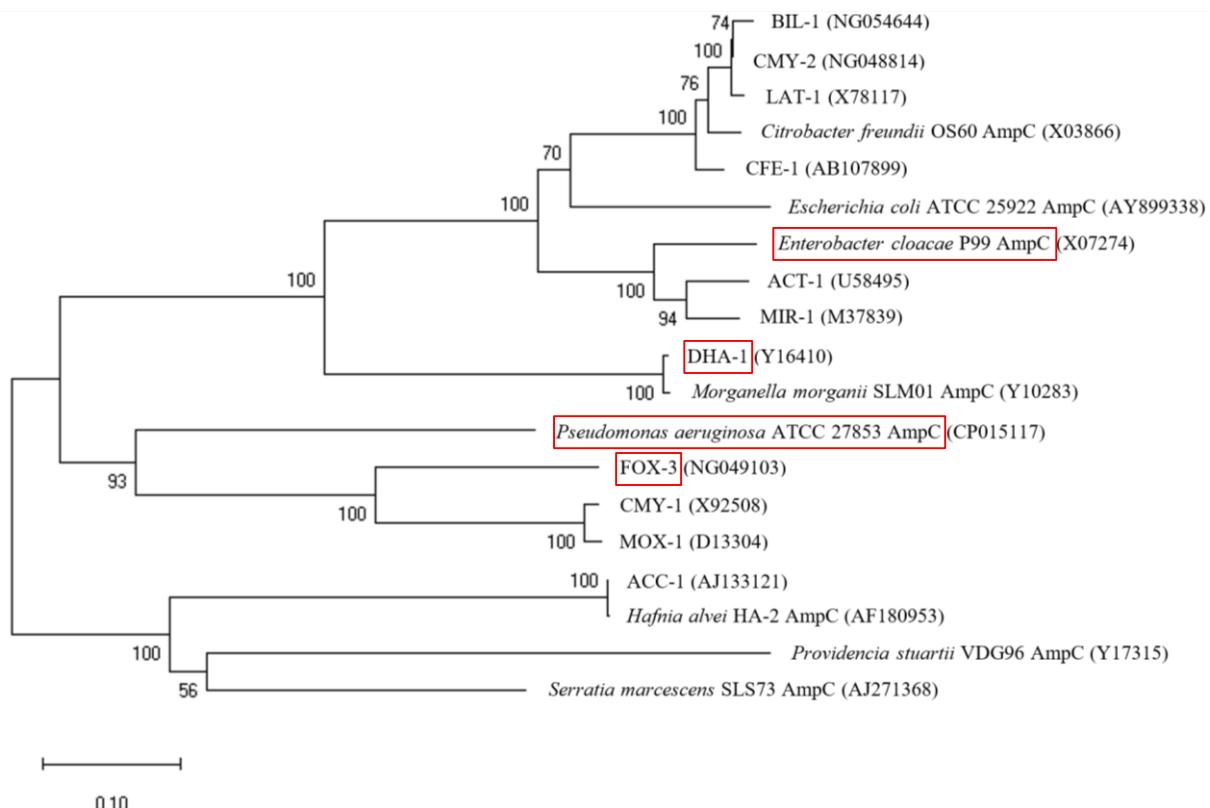
Ambler positions 315-317 **KTG** Ambler positions 346

P. aeruginosa AmpC

401404

Supplementary Figure S1. Alignment of the amino acid sequence of representative chromosomal and plasmid-borne AmpC β -lactamases.

Conserved motifs are highlighted in red. Additional key residues in interaction with avibactam are shown in green-colored font. The asparagine residue studied here, Asn346 in Ambler numbering scheme, is highlighted in yellow. This residue is substituted by an isoleucine in *H. alvei* AmpC, *P. stuartii* AmpC, ACC-1, MOX-1, and FOX-3. Residues of the Ω loop are indicated in orange. The deletion of RVGPGPL in the cAmpC for *P. aeruginosa* (PDC-5), which emerged under the selective pressure of ceftazidime combined to avibactam, is highlighted in blue. Residue E, replaced by K in another mutant is also highlighted in blue. Genbank accession numbers of AmpC β -lactamases are as follows: X07274 (*E. cloacae* P99), CP015117 (*P. aeruginosa* ATCC 27853), AY899338 (*E. coli* ATCC 25922), X03866 (*C. freundii* OS60), AF180953 (*H. alvei* HA-2), Y10283 (*M. morganii* SLM01), Y17315 (*P. stuartii* VDG96), AJ271368 (*S. marcescens* SLS73), X92508 (CMY-1), Y16410 (DHA-1), AJ133121 (ACC-1), U58495 (ACT-1), M37839 (MIR-1), D13304 (MOX-1), NG054644 (BIL-1), X78117 (LAT-1), NG049103 (FOX-3), AB107899 (CFE-1), NG048814 (CMY-2).



Supplementary Figure S2. Unrooted evolutionary tree constructed with a set of 19 sequences representative of chromosomal and plasmid-borne AmpC β -lactamases. Amino acid sequences are designated by their GenBank accession number. Evolutionary analyses were conducted in MEGA X using the neighbor-joining method (1). The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) are shown next to the branches. The evolutionary distances are in the units of the number of amino acid substitutions per site. Red boxes indicate AmpC cephalosporinases used in our study.

SUPPLEMENTARY REFERENCES

- 1 Kumar S., Stecher G., Li M., Knyaz C., and Tamura K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution* 35:1547-1549.