Supplemental figures:

Characterization of the small molecule ARC39, a direct and specific inhibitor of acid sphingomyelinase *in vitro*

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Supplemental Figure S1. *Left:* Fold change of *SMPD1* mRNA relative to *HPRT1*, ASM protein level (representative blot) after treatment with 20 μ M ARC39 as indicated in HepG2 cells. *Right:* ASM enzyme activity. Enzyme activity is normalized to the protein concentration and control values are normalized to their mean. L-ASM activity here was determined with the conventional ASM assay (without ZnCl₂ in the assay buffer). Data represented as mean \pm SD, n=3 experiments. Two-way ANOVA was used. *P*<0.05 for time as a source of variation in ASM activity.



Supplemental Figure S2. ARC39 leads to lysosomal accumulation of exogenous sphingomyelin. HepG2 cells were incubated for 24h with 1 μ M BODIPY FL sphingomyelin (SM) together with PBS as control, 10 μ M ARC39 or 10 μ M amitriptyline (Ami) as a positive control. Then, they were incubated 30 min at 37°C with 25 nM LysoTracker Red DND-99. Fresh medium was added then the cells were analyzed by confocal microscopy, magnification 100×.



Supplemental Figure S3. Sphingolipidomic analysis after treatment with ARC39 in HepG2 cells. Cells were treated with 20 μ M ARC39 and at the indicated time points the endogenous levels of the following sphingolipids were determined by mass spectrometry in whole -cell lysates. (A) Sphingomyelins (SMs). (B) Ceramides (Cers). Numbers indicate the chain length and saturation of the fatty acyl chain. (C) Dihydrosphingosine (dhSph), Sphingosine (Sph) and sphingosine-1-phosphate (S1P), respectively. Data represented as mean \pm SD, n=3 experiments. Two -way ANOVA was used followed by Bonferroni correction. *P<0.05 **P<0.01 ***P<0.001 ***P<0.0001.



Supplemental Figure S4. Effect of ARC39 on lysosomes. L929 cells were treated 2 h (up) or 24 h (down) with PBS, 40 μ M ARC39 for 2 h, 20 μ M ARC39 for 24 h, 10 μ M amitriptyline (2h), 10 μ M desipramine (2h) or 10 mM NH₄Cl as a positive control. Then, LysoTracker Red (LTR) was added to a final concentration of 25 nM for 30 min at 37°C. Fresh medium was added and the cells were analyzed by confocal microscopy. Magnification 100× (up) and 40× (down).



Supplemental Figure S5. Toxicity of ARC39 *in vivo* in a single-dose regimen. The compound was applied i.p. for one time at the indicated doses. After 24 h in (A) and after 10 d in the same individuals in (B), serum was collected and the following serological markers were assessed: Blood urea nitrogen (BUN), glutamate-oxaloacetate transaminase (GOT) / aspartate transaminase, glutamate-pyruvate transaminase (GPT) / alanine transaminase, lactate dehydrogenase (LDH), creatine phosphokinase (CPK) and amylase. On day 10, only the three groups that did not exhibit morbidity or lethality between 24 h-10 d are shown. Data represented as mean \pm SD, n=3 mice per group. One-way ANOVA was used in (A) and two-way ANOVA was in (B), both followed by Bonferroni correction. **P*<0.05 ***P*<0.01 #*P*<0.0001.



Supplemental Figure S6. Toxicity of ARC39 *in vivo* in a multiple-dose regimen. The compound was applied i.p. every 12 h for 96 h. After 12 h in (A) and after 10 d in the same individuals in (B), serum was collected and the following serological markers were assessed: Blood urea nitrogen (BUN), glutamate-oxaloacetate transaminase (GOT) / aspartate transaminase, glutamate-pyruvate transaminase (GPT) / alanine transaminase, lactate dehydrogenase (LDH), creatine phosphokinase (CPK) and amylase. On day 10, only the two groups that did not exhibit morbidity or lethality between 12 h-10 d are shown. Data represented as mean \pm SD, n=3 mice per group. One-way ANOVA was used in (A) and two-way ANOVA was in (B), both followed by Bonferroni correction. **P*<0.05 ***P*<0.01.