#### **Supplementary Figures to:**

# Partner independent fusion gene detection by multiplexed CRISPR/Cas9 enrichment and long-read Nanopore sequencing

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# A 3' unknown sequence

# 

crRNA sequence: 5'TGCTAGCTAGCGGCTAGGCT3' Read direction: Downstream

B <u>5' unknown sequence</u>

crRNA sequence: 5'AGCCTAGCCGCTAGCTAGCA3' Read direction: Upstream

### Supplementary Figure 1:

Examples of crRNA design for directional sequencing are shown. (A) crRNAs (grey) are designed containing the sequence of the forward strand of the target gene 20 bp prior to a PAM-sequence (NGG, red underlined) if the unknown gene partner is downstream (3') of the known target sequence. (B) crRNAs (grey) are designed containing the sequence of the reverse strand of the target gene 20 bp prior to a PAM-sequence (NGG, red underlined) if the unknown gene partner is upstream (5') of the known target sequence.





(A-E) Coverage plots showing deconvoluted on-target coverage across multiple genomic loci for two different cut positions (red = PP1 and black = PP2).





Supplementary Figure 3:

(A) Fusion gene configurations for the identified *EWSR1-FLI1*, *AGAP3-BRAF* and *TRIM24-BRAF* fusion genes. (+) or (-) indicate genomic location on the forward or reverse strand of the gene, respectively. Dashed lines (red) indicate break-position for each sample. (B) Breakpoint PCR (n=1) for the *SS18-SSX1* fusion gene in the HS-SYII cell line.



Supplementary Figure 4:

Coverage plots for the (A) ES1 tumor sample for the two fusion partners *EWSR1* (targeted) and *FL11*, (B) BL tumor sample the *MYC* gene (targeted) and IGH locus, and (C) for the ALL1 sample for the two fusion partners *BCR* (targeted) and *ABL1*. Dotted lines (green) indicate cut position, dashed lines (red) indicate breakpoint positions and arrows indicate the desired read direction.



Supplementary Figure 5:

(A) Breakpoint PCR (n=1) for the reciprocal *PAX3-FOXO1* and *FOXO1-PAX3* fusion genes in the RH tumor sample. (B) Breakpoint PCR (n=1) for the *BCR-ABL1* and *DRICH1-BCR* fusions in the CML tumor sample. (C) Sanger sequencing trace for the *DRICH1-BCR* fusion gene identified in the CML sample. (D) Graph depicting the number of fusion genes called by NanoFG depending on the set threshold of fusion-supporting reads (n=1-5) for the samples CHP-100, ES1, C1-WGA and C2-WGA.
(E) Breakpoint PCR (n=1) for the *AGAP3-BRAF* and *TRIM24-BRAF* fusion genes in the non-amplified tumor material of C1 and C2.



Supplementary Figure 6:

(A) Coverage plots for the *KMT2A* fusion partners *MLLT1*, *MLLT2*, *MLLT3* and *MLLT4*. Dashed lines (red) indicate breakpoint positions. (B) IGV screenshot showing reads identifying the *KMT2A-MLLT2* and *KMT2A-MLLT3* fusion genes (red) and their reciprocal translocations (blue).



Supplementary Figure 7:

(A) Breakpoint PCR (n=1) with the fusion-specific breakpoint primers for the cell lines KOPN8 (*MLLT1*), ALLPO (*MLLT2*), Monomac-1 (*MLLT3*), ML2 (*MLLT4*) and the tumor samples C1 (*AGAP3-BRAF*) and C2 (*TRIM24-BRAF*). \* indicates the bands at the correct height in the correct sample. For the complex *KMT2A-MLLT4* fusion, primers that span the inversion (KMT2A-MLLT4-i), translocation (KMT2A-MLLT4-t) and the complete rearrangement (KMT2A-MLLT4-c) were tested. (**B**) Breakpoint PCR (n=1) for the KOPN8 and freshly isolated Monomac-1 cell lines for the MLLT1 and MLLT3 fusion genes. Asterisk indicates the bands at the correct height in the correct sample. (C) Schematic of rearranged *KMT2A-MLLT4* fusion gene. Sanger-traces showing the exact breakpoints (vertical lines) and breakpoint-positions. The discordant nucleotides between breakpoints ("G" between *KMT2A/KMT2A* inversion; "CG" between *KMT2A* inversion/*MLLT4*) were most likely introduced during non-homologous end joining.



Supplementary Figure 8:

Plot depicting the relationship between coverage at the target-locus, distance between the cut and the breakpoint, and sequencing throughput. Samples are highlighted according to <0.1 Gbs (red) or >0.1 Gbs (grey) throughput. Samples with one fusion supporting read are circled in brown.