

## **The Role of Host Genetic Factors in Coronavirus Susceptibility: Review of Animal and Systematic Review of Human Literature**

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## **Abstract**

**Background:** The recent SARS-CoV-2 pandemic raises many scientific and clinical questions. One set of questions involves host genetic factors that may affect disease susceptibility and pathogenesis. New work is emerging related to SARS-CoV-2; previous work on other coronaviruses in humans or other host species may be relevant.

**Objectives:** To review existing literature on host genetic factors and their association with infection and disease with coronaviruses in humans and in other host species.

**Methods:** We conducted a systematic review of literature on host genetic factors in humans associated with coronavirus outcomes. We also reviewed studies of host genetic factors associated with coronavirus outcomes in non-human species. We categorized articles, summarized themes related to animal studies, and extracted data from human studies for analyses.

**Results:** We identified 1,187 articles of potential relevance. Forty-five studies examined human host genetic factors related to coronavirus, of which 35 involved analysis of specific genes or loci; aside from one meta-analysis on respiratory infections, all were candidate-driven studies, typically investigating small numbers of research subjects and loci. Multiple significant loci were identified, including 16 related to susceptibility to coronavirus (of which 7 identified protective alleles), and 16 related to outcomes or clinical variables (of which 3 identified protective alleles). The types of cases and controls used varied considerably; four studies used traditional replication/validation cohorts. Of the other studies, 28 involved both human and non-human host genetic factors related to coronavirus, and 174 involved study of non-human (animal) host genetic factors related to coronavirus.

**Key findings:** We have outlined key genes and loci from animal and human host genetic studies that may bear investigation in the nascent host genetic factor studies of COVID-19. Previous human studies

have been limited by relatively low numbers of eligible participants and limited availability of advanced genomic methods. These limitations may be less important to studies of SARS-CoV-2.

**Key words:** Coronavirus; COVID-19; Host genetic factors; SARS-CoV-2

## Introduction

The ongoing severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pandemic raises many scientific and clinical questions. One unknown is the extent to which individuals vary in susceptibility to infection and disease (COVID-19). Various hypotheses have been suggested to explain observed differences, including sex, age, comorbidities, and genetic factors.<sup>1</sup> As with many complex diseases, the explanations likely involve a combination of genetic and non-genetic factors. In this context, genetic factors involve an interplay between virus and host genetics.<sup>2</sup>

Large, international studies and collaborations have formed to investigate host genetic factors related to COVID-19, including disease severity and susceptibility. These investigations include analyses of existing public and private datasets, as well as the establishment of new cohorts (e.g., <https://blog.23andme.com/23andme-research/genetics-and-covid-19-severity/>).<sup>3</sup>

While SARS-CoV-2 has seized recent attention, there are many other coronaviruses and a significant related body of literature exists about host genetic factors and their association with infection and outcomes in both humans and non-human host species. The *Coronavirinae* subfamily of the *Coronaviridae* family consists of four genera. The alphacoronaviruses include two major human coronaviruses, HCoV-229E (of which multiple HCoV-229E-like strains have been identified) and HCoV-NL63. Alphacoronaviruses that affect other species include mouse hepatitis virus (MHV), feline coronavirus (FCoV), which includes feline infectious peritonitis virus (FIPV) and feline enteric coronavirus (FECV), canine coronavirus (CCoV), and transmissible gastroenteritis coronavirus (TGEV) and porcine transmissible gastroenteritis coronavirus (TGEV) in pigs. The betacoronaviruses consist of four lineages: lineage A (HCoV-OC43 and HCoV-HKU1), lineage B (SARS-CoV-1 and SARS-CoV-2), lineage C (Middle East Respiratory Syndrome (MERS) and many bat coronaviruses), and lineage D (coronaviruses only identified in bats to date). HCoV-OC43, HCoV-229E, HCoV-HKU1, and HCoV-NL63 can result in a variety of

presentations, including “common cold” and severe but rarely fatal disease; they are also frequently detected as co-infections with other viruses.<sup>4</sup> There are other rare coronaviruses observed in humans as well as in other species.<sup>5,6</sup> Relative to other coronaviruses, SARS-CoV-2 has unique biological properties and related clinical impact, but data regarding other coronaviruses may be relevant.

In various species, much work has focused on the genes encoding the relevant coronavirus receptor, including effects of viral and host genetic changes. Among other cell surface determinants,<sup>7</sup> these receptor genes include *ACE2* for HCoV-NL63,<sup>8</sup> SARS-CoV-1,<sup>9,10</sup> and SARS-CoV-2,<sup>11</sup> *ANPEP* for HCoV-229,<sup>12,13</sup> FIPV,<sup>14</sup> CCoV,<sup>15</sup> and TGEV,<sup>16</sup> *DPP4* for MERS,<sup>17-19</sup> and *Ceacam1* for MHV.<sup>20</sup> Host genetic studies have - to varying degrees and in different ways - analyzed these genes, as well as other genes identified through targeted and agnostic methods. Studies to date have been disparate in terms of the virus and species studied, as well as the aims of the particular study. This has resulted in a rich body of literature that is difficult to efficiently leverage for SARS-CoV-2-related work.

We aimed to perform a review of the literature to outline previous studies of host genetic factors related to coronaviruses, with the objective of performing a systematic review to encapsulate genes and loci interrogated through these efforts. We do not attempt to fully describe the findings nor recapitulate what is known about the underlying host biology related to coronavirus infection and disease. As the majority of studies are candidate-driven, we did not attempt to conduct a meta-analysis. However, one goal is that the data presented here can help populate lists of genes that - along with data from related work<sup>21-23</sup> - may bear scrutiny in the developing and important large-scale host genetic studies related to SARS-CoV-2.<sup>24,25</sup> We present an overview of themes and interrogated genes/loci from animal studies, and perform a systematic review on human studies.

## Methods

We conducted an initial search of the PubMed database (last queried May 4, 2020) using each of the following phrases: “host genetics”; “genetic resistance”; “genetic susceptibility”; “genetic factors”; “genetics”; “GWAS” along with each of the following terms: “coronavirus”; “SARS”; “MERS”; “COVID-19”; “COVID19”. We also identified additional articles by searching for specific coronaviruses or coronavirus-associated conditions (e.g., “canine coronavirus”; “middle east respiratory syndrome”) along with the term “genetics”. Articles were included in the search regardless of publication date. Articles included electronic, ahead-of-print publications available in the PubMed database. We also identified and categorized relevant articles from the references of initially selected articles. We did not include articles only available on non-peer reviewed preprint servers, though recognize that a substantial number of these manuscripts will be on PubMed soon.

Each abstract was reviewed by a single reviewer. Full articles were reviewed when insufficient data were available in the abstract, or when no abstract was available. Publications were classified into the following categories: 1) Study of human host genetic factors related to coronavirus; 2) Study of non-human (animal) host genetic factors related to coronavirus; 3) Study of non-genetic (including non-DNA-based analyses - see further explanation below) host factors related to coronavirus, including involving immunopathogenesis; 4) Study of other pathogens (not coronavirus); 5) Other studies of coronavirus. Articles containing information in both categories 1 and 2 were identified as such; articles were otherwise categorized according to the lowest numerical category (e.g., an article involving both human host genetic factors to coronavirus as well as immunopathogenesis would be categorized into group 1. Articles that did not involve investigations of specific DNA-based genetic changes (e.g., transcriptomic or proteomic studies) were categorized into group 3, as were studies that only included analyses of sex without other genetic analyses. Other publications, including: 6) Untranslated studies in another language (not English); 7) Not relevant (unrelated to coronavirus or other pathogens); 8) No data available; were removed from further analysis after categorization into these latter four categories.

Data from category 1 publications were manually extracted for relevant information pertaining to: coronavirus studied; general methods and questions analyzed; gene(s), variant(s), or loci analyzed; size of cohorts studied; geographic or ancestral composition of cohorts; statistical results, including (where available) odds ratios, confidence intervals, and p-values.

## **Results**

Our search identified 1,187 articles of potential relevance (Figure 1, Supplementary Table 1). Of these, 45 involved study of human host genetic factors related to coronavirus (Table 1); 35 of the 45 human studies involved analysis of specific genes or loci (only one was a non-candidate study), while 10 involved biological, computational, or case report studies of human host genetic factors. Twenty-eight involved both human and non-human host genetic factors related to coronavirus (these largely investigated inter-species differences in disease susceptibility and pathogenesis, such as related to differences in *ACE2*); 174 involved study of non-human (animal) host genetic factors related to coronavirus; 584 involved study of non-genetic host factors related to coronavirus, including involving immunopathogenesis; 16 involved study of other pathogens (not coronavirus); 321 involved other studies of coronavirus. 18 studies were assigned to the other categories and removed.

We organized our analysis and findings into the schema presented below.

### ***Animal studies***

Coronaviruses affect many species, from Beluga whales to spotted hyenas to turkeys, and sequelae of disease can range from apparently asymptomatic infections to severe or lethal effects on different organ systems, potentially manifesting as diarrheal, encephalitic, nephritic, respiratory, and other types of disease.<sup>26,27</sup> There are numerous non-observational animal studies of coronaviruses, such as involving hamsters,<sup>28-30</sup> guinea pigs,<sup>31</sup> rats,<sup>10,32-35</sup> and non-human primates.<sup>36-38</sup> However, formal host genetic

studies have been described for some but not all species. Many studies have involved examination of differences in species susceptibility and pathogenesis to human and non-human coronaviruses.<sup>14,39-41</sup>

Among the host genetic studies in animals, the objectives and methods used differ significantly depending on the species studied. For example, in chickens and other livestock, the types of published studies predictably differ from those conducted on experimental mice. That is, while MHV represents a problem for mouse colonies, the rationale of the livestock studies may focus more purely on economic repercussions versus attempts to use a model organism to understand immunopathogenesis of infectious disease.<sup>42</sup> The degree to which results may be reported through the scientific literature (versus other routes) is also anticipated to differ depending on the species studied and the reason for the study. See Figure 2 for a summary of interrogated loci in animal studies.

We describe representative studies and key findings below, but the descriptions should not be considered as truly comprehensive; additionally, as noted above, many studies compared susceptibility across species, both through cell-based assays and experimental animals. Many investigations using other methods (e.g., transcriptomics or proteomics) have identified key molecules involved in coronavirus susceptibility and pathogenesis. Though beyond the scope of this article, these molecules should also be considered in future SARS-CoV-2 host genetic studies.

### ***Model animal strains, experimental animals, and domesticated animals***

#### ***Chicken***

In chickens, the infectious bronchitis virus (IBV) coronavirus can cause disease affecting different organ systems and tissues, such as IBV-associated nephritis. As with other species, inbred status and specific chicken lines impact host susceptibility, immune response, and outcomes, and virus/host genetic interactions have been described.<sup>43-47</sup> Breeding experiments have suggested different inheritance patterns related to susceptibility and outcomes, and have implicated both MHC and non-MHC loci.<sup>48,49</sup>



Multiple GWAS investigating the immune response to IBV have identified significantly-associated polymorphisms in the breeds studied;<sup>50,51</sup> the implicated or nearest genes include: *AKT1*, *AvBD12*, *CEP170B*, *CRYL1*, *CWF19L2*, *DHRX*, *FAM19A2*, *GABRB3*, *INTS9*, *NMNAT3*, *PINX1*, *RAB39A*, *VRK1*, *YEATS2*; and *SETBP1*.<sup>50,51</sup>

#### *Domestic cat*

Felines can be infected by feline coronavirus (FCoV), which include feline infectious peritonitis (FIPV) and feline enteric coronavirus (FECV).<sup>52</sup> As with other species, cats demonstrate a range of potential effects. In addition to association with traits such as age, sex, and reproductive status, purebred status and loss of heterozygosity has been shown to be associated with the effects of disease. Susceptibility and outcomes also appear to vary between different breeds.<sup>52-60</sup> A small study of feline leukocyte antigen (FLA)-DRB alleles did not show a statistically significant association between the number of FLA-DRB alleles and FCoV infection outcome.<sup>61</sup> Polymorphisms in *IFNG* (investigated as FIP can result in decreased interferon-gamma levels) were shown to correlate with plasma interferon-gamma levels and outcomes.<sup>62</sup> Polymorphisms in *TNFA* and *CD209* were also shown to be associated with outcomes in one inbred line.<sup>63</sup>

In addition to candidate studies, several GWAS have been performed in cats. One small study on outcomes in experimentally-induced infections in random-bred cats identified one associated genomic region (which did not harbor any obvious candidate genes).<sup>52</sup> Another small study on an inbred breed identified multiple candidate genes (*ELMO1*, *ERAP1*, *ERAP2*, *RRAGA*, *TNSF10*) but none was fully concordant with the FIP disease phenotype.<sup>64</sup> Recent studies on SARS-CoV-1 and SARS-CoV-2 have investigated the susceptibility of cats as well as other animals;<sup>65</sup> see further details below (under Ferrets).

#### *Dromedary camel*

Camels are an important reservoir of coronaviruses that can infect humans; this became especially relevant in the context of MERS.<sup>66-68</sup> Many studies have analyzed factors that contribute to spread,<sup>69</sup> though the searches employed in this analysis identified relatively few host genetic studies separate from analyses of DPP4 receptor characteristics and tropism, including comparisons between camels, humans, and other species.<sup>70-74</sup>

### *Ferret*

Several studies have investigated the susceptibility of various species to coronaviruses. One objective relates to identifying useful animal models of disease, in which non-human species show similar infection and disease outcomes to humans upon exposure to coronaviruses.<sup>65,75,76</sup> For example bat, camel, and humans can be infected by MERS, unlike mouse, ferret, hamster, and guinea pig. SARS-CoV-2 replicates better in ferrets and cats than in dogs, pigs, chickens, and ducks. One explanation involves genetic characteristics of the host receptor for the relevant virus.<sup>76,77</sup> Additionally, within an infected animal, the site of viral replication appears to vary according to the species and coronavirus, and is additionally potentially related to tissue-specific receptor expression.<sup>78</sup> This line of reasoning may also be relevant to age-specific differences observed with SARS-CoV-2 and human infections.<sup>79</sup>

### *Hamster*

As noted above, hamsters have been used as model organisms to study coronaviruses, including studies of host receptors. This includes studies using standard hamster cell lines as well as other approaches involving hamster models.<sup>80-85</sup> For example, hamster models have been used to study species susceptibility to MHV (related to *Ceacam1*),<sup>86</sup> how alterations of specific Dpp4 amino acids in hamster affect susceptibility to MERS,<sup>71,87</sup> and the roles of ACE2 and CD209L in SARS-CoV-1 susceptibility.<sup>82</sup>

### *Mouse*

MHV has represented a challenge for the health of mouse colonies, though relatively recent improvements in animal care practices have been beneficial.<sup>88</sup> Differences in the susceptibility of different mouse strains to MHV has been noted for seven decades.<sup>89-91</sup> Studies have examined a number of different MHV strains. These strains demonstrate different tissue tropism and have different effects on various mouse lines.<sup>92</sup> One distinct example is the JHM strain of MHV, which causes encephalitis in susceptible animals.<sup>93,94</sup> In the discussion below, though susceptibility and outcome findings will be summarized, it is important to note that studies generally focus on the interactions between certain MHV strains and mouse lines, and it is not always clear how well these findings extrapolate to other strains and lines.

Many studies have investigated biological explanations for differences in MHV susceptibility and pathogenesis.<sup>95-97</sup> Studies examining different laboratory mouse strains have suggested that multiple loci are involved.<sup>98-111</sup> Early studies suggested various models, including potential monogenic/Mendelian explanations as well as more complex explanations involving interacting loci.<sup>92,112-115</sup>

Among many studies aiming to understand the underlying pathophysiology, mouse studies originally focused on strains believed to be involved in host susceptibility and reaction to infection. Importantly, these studies have identified interactions of host genetic factors with other factors, such as the cellular environment,<sup>116,117</sup> cell and tissue-specific effects related to viral as well as host genetics,<sup>118-123</sup> and host age.<sup>124-126</sup> Unsurprisingly, some aspects of the disease process appear to be independent of observed strain differences.<sup>127</sup> These studies also showed that host genetic factors influence different parts of the disease process, from initial virus-receptor binding,<sup>117</sup> to cellular viral spreading<sup>128,129</sup> and multiple aspects of the immune response.<sup>101,130-134</sup> These studies enabled the cloning of *Ceacam1*, the MHV receptor gene,<sup>81</sup> as well as related work regarding how genetic changes affecting this receptor confers MHV resistance in SJL mouse lines via inhibition of viral integration into host cells.<sup>103,106,135,136</sup>

In addition to the above studies, MHV-based mouse studies have used transgenic models to directly test the role of implicated pathways (summarized in Table 2). Not surprisingly, the majority of work in mouse models have focused on pathways already implicated in viral infection susceptibility including adaptive immune responses including both humoral and cellular, specific cytokine and immune receptor pathways, viral receptors, complement pathway, apoptosis, autophagy, and tissue repair. These studies have prominently implicated Type I ( $\alpha/\beta$ ) and II ( $\gamma$ ) interferon responses in host response and predominantly protection against MHV infection. However, not all pro-inflammatory pathways are protective. For example, complement activation promotes tissue damage caused by MHV infection, highlighting the complex interplay between the host and virus. In addition to targeted gene disruptions described above, a GWAS using a recombinant inbred mouse panel implicated *Trim55*, which is involved in vascular cuffing and inflammation in response to SARS-CoV-1.<sup>137</sup>

Additional transgenic studies have investigated multiple biologic effects as well as returning to questions regarding susceptibility of different strains.<sup>138</sup> Other mouse models (including knockouts, specific knock-in mutations, humanized mice, and other models involving genetic manipulation) have been used to study human pathogens such as SARS-CoV and MERS; revealing similar properties for viral receptors, *Dpp4* for MERS, *Ace2* for SARS-CoV, cytokine and immune receptor pathways, and complement pathway as with mouse models of MHV. Intriguingly, there are differences between the importance of interferon pathways in host response to SARS-CoV1, where these pathways are dispensable as compared to MHV, where they are protective. Together, these different pathogen models have shown overlapping and unique pathways of host response between coronaviruses and highlight the potential relevance for SARS-CoV-2. See also the *Additional papers on humans and other species* section regarding further examples of studies involving mice and humans, as well as other species.

*Pigs*

Pigs can be infected by transmissible gastroenteritis virus (TGEV) and porcine epidemic diarrhea virus (PEDV), as well as the more recently-identified porcine deltacoronavirus (PDCoV). Like coronavirus disease in chickens, these diseases can have economic effects on the food industry,<sup>139</sup> and analyses aim to address ways to ameliorate disease, such as the development of vaccines. Importantly, variants (both natural and experimentally-induced) may have different effects on different coronaviruses. For example, aminopeptidase N, encoded by *ANPEP* (also called *APN*) was reported as a functional receptor for TGEV and PEDV (as well as HCoV-229E), but multiple models, including CRISPR/Cas9- generated knock-outs, show differences in cellular susceptibility to TGEV and PEDV.<sup>139,140</sup> In another study, infection by PEDV and TGEV correlated positively with *ANPEP* expression, but PEDV and TGEV could infect *ANPEP*-positive and negative enterocytes, with differences observed between viral strains. Overall, the results suggest the presence of an additional receptor.<sup>141</sup> Building on this type of work, site-specific editing of *ANPEP* has been suggested as a potential means to breed resistant animals.<sup>142</sup> Studies focusing on PEDV have shown that knock-out of *CMAH* (hypothesized to affect cellular binding) does not result in immunity, but may improve outcomes.<sup>143</sup>

### *Rats*

Rats can be affected by rat coronaviruses, and can be hosts to a number of different coronaviruses that affect other species.<sup>144,145</sup> Rats have been used as model systems to investigate MHV, including through cellular-based assays.<sup>146,147</sup> Several studies have examined rat susceptibility to various coronaviruses. As with many other studies, these have implicated key interactions between viral and host genetics that affect species and tissue tropism [17151094].<sup>33</sup> In addition to computational approaches examining receptor characteristics, such as involving ACE2 in the context of SARS-CoV-1,<sup>10</sup> experimental studies suggest that rats are not susceptible to MERS based on *Dpp4* characteristics.<sup>148</sup>

### ***Non-domesticated animals***

As described, many species can be infected by coronaviruses. These species include wild as well as domesticated animals. The below section provides select examples of genetic studies on wild animals. Others studies been conducted on coronaviruses (as well as other pathogens),<sup>149</sup> especially related to host ranges or reservoirs and involving host/pathogen co-evolution.<sup>150,151</sup> Related to host genetic studies that are particularly relevant to the current SARS-CoV-2 pandemic (e.g., pangolin), our searches did not identify relevant articles.

### *Cheetah*

Among wild animals, severe population bottlenecks (resulting in reduced genetic diversity) in cheetahs has been used to explain their increased susceptibility to infection by FIPV as well as other infectious diseases. Several such bottlenecks appear to have occurred in cheetah, due to a combination of factors.<sup>152-154</sup> Among possible explanations for this susceptibility, genetic uniformity of the major histocompatibility complex (MHC) has been suspected to be involved.<sup>155</sup>

### *Civet*

Studies have focused on palm civets (as well as other species) related to zoonotic implications as this species has been implicated as the reservoir associated with introduction of SARS-CoV-1 into humans. Specifically, questions about host receptor characteristics (*ACE2*) have been described in the context of SARS-CoV-1.<sup>156-158</sup> As with other coronaviruses and species, the interactions of viral and host genetics have been shown to be important.<sup>159,160</sup>

### *Bat*

As a natural reservoir for many coronaviruses, bats have been studied more extensively than other species outside of laboratory-based animals and livestock. Studies have included co-evolutionary studies between coronaviruses and the genomes of bat hosts (e.g., by correlating phylogenetic analyses of bat coronaviruses with *CYTB* in multiple bat species)<sup>161</sup> as well as genetic/biologic studies related to

host genetic factors. These have involved well-studied genes such as *ACE2* with SARS-CoV-1<sup>162,163</sup> and *DPP4* with MERS.<sup>164,165</sup> In addition to allowing analyses of host susceptibility, these and similar studies help provide estimates for the time-frame of coronavirus circulation in species and populations, and explore cross-species transmission.<sup>150</sup>

## Human

Details of the human studies are presented in Table 1, Figure 3, and Supplementary Table 2. Forty-five studies were initially identified by the methods described. Of these, 35 involved association or other studies related to human host genetic factors (see summary in the next paragraph). Ten others involved biological, computational, or other non-genetic association studies. Many other studies were identified that used a combination of human and animal models, but were categorized separately; additionally, many studies that might be considered genetic studies - if the definition were applied less stringently - were grouped in category 3. For example, studies have examined how specific genes are involved in aspects of viral disease but did not strictly study how DNA-based host genetic variants affect this process. In summary, these ten included mapping of a susceptibility locus to HCoV-229E to chromosome 15.<sup>166</sup> Multiple studies examined the biological effects of mutant genes. Studying the effects of mutant *ACE2* on SARS-CoV-1 entry provided evidence that the cytoplasmic tail of *ACE2* is not required for SARS-CoV-1 penetration.<sup>167</sup> Studies of mutant *TRIM56* on antiviral activity against HCoV-OC43 and other viruses showed that anti-HCoV-OC43 activity relies solely upon TRIM56 E3 ligase activity; this appears different from the mechanisms related to other viral pathogens.<sup>168</sup> Knockout culture cells and nonsynonymous variant *PPIA* models result in limitation of HCoV-229E replication.<sup>169</sup> (Please note that we did not separately or exhaustively investigate human genetic experiments involving cell culture systems.) Specific variants in *IFITM* genes (*IFITM1* and *IFITM3* were studied) modulate the entry of multiple human coronaviruses (HCoV-229E; HCoV-NL63; HCoV OC43; MERS-CoV; and SARS-CoV-1 were studied).<sup>170</sup> Computational models suggest that, while most *ACE2* variants have similar binding

affinity for SARS-CoV-2 spike protein, certain variants (rs73635825 and rs143936283) demonstrate different intermolecular interactions with the spike protein.<sup>171</sup> An in silico analysis of viral peptide-MHC class I binding affinity related to HLA genotypes for SARS-CoV-2 peptides, as well as potential cross-protective immunity related to four common human coronaviruses, provides evidence that HLA-B\*46:01 may be associated COVID-19 vulnerability, while HLA-B\*15:03 may enable cross-protective T-cell based immunity.<sup>172</sup> A recent study on viral cell entry showed that SARS-CoV-2 uses ACE2 for cell entry and TMPRSS2 for S protein priming; potential interventions based on these results include TMPRSS2 inhibition and convalescent sera.<sup>173</sup> In addition to these examples, there are undoubtedly other biological, computational, and other studies examining how changes in and affecting key proteins may modulate disease.

Of the 35 human studies meeting the host genetic study criteria described above, 32 (91%) involved SARS-CoV-1, while 3 (9%) involved SARS-CoV-2. Two of the three SARS-CoV-2 studies were case reports (one on a single family, the other on two patients with a rare immunodeficiency) without specific studies related to host factors; it is anticipated that many more studies on SARS-CoV-2 will be published soon. All of the association studies except one were candidate-gene analyses based on genes hypothesized to be important in disease susceptibility or clinical variables/outcome. The exception was a meta-analysis of 386 studies on susceptibility to tuberculosis, influenza, respiratory syncytial virus, SARS-CoV-1, and pneumonia.<sup>174</sup>

Candidate studies ranged from studies of single variants to studies of over 50 genes selected due to biological plausibility; seven of these studies focused on HLA alleles. Sixteen significant loci related to susceptibility to coronavirus were reported (of which 7 identified protective alleles). Sixteen significant loci related to outcomes or clinical variables were reported (of which 3 identified protective alleles). The types of cases and controls used varied. Only four studies used separate cohorts for replication/validation. However, the studies used many different types of cases and controls, including



within the same study. For example, some studies compared healthcare workers with SARS-CoV-1 infection with healthcare workers who tested negative. Others compared data from individuals with documented infection with data from control samples taken from blood donors. Four studies conducted laboratory-based biological studies in addition to association analyses. These studies are summarized in Table 1 and Figure 3; more details are available in Supplementary Table 2.

#### *Additional papers on humans and other species*

As described, human and animal studies have examined various host factors related to coronavirus infection. For example, human<sup>175</sup> and animal<sup>33,52,125</sup> studies have implicated age as having significant associations with outcomes; age appears to be strongly correlated with COVID-19 outcomes.<sup>176</sup> The overall explanations remain unclear, but could at least partially involve age-related gene expression. Sex also appears to have a role. Human studies of SARS-CoV-1 and SARS-CoV2 suggest a correlation between sex and certain clinical parameters, perhaps rooted in sex-based or related immunologic differences.<sup>175,177,178</sup> However, separating biological differences from sex-related cultural practices (e.g., different rates of social distancing) may be difficult.

Animal studies also suggest sex effects in multiple species, such as related to disease severity.<sup>57,179</sup>

Multiple studies examined different genes/proteins to determine disease susceptibility, transmissibility, and pathogenesis in various species. In addition to humanized genes, such as used in mouse models, studies have involved a combination of computational and biological approaches, and have investigated the viral entry receptors *ACE2* in SARS-CoV-1<sup>10,35,157,158,180-183</sup> and SARS-CoV-2<sup>184</sup> (for which there already exists a large body of unpublished and preprint work) and *DPP4* in MERS.<sup>71,148,185-188</sup> Among other findings, these studies examined specific protein residues that are critical in viral-host interactions [18448527].<sup>157</sup> Other studies examined manipulations of various genes/proteins to study the functional biological effects, including of *ANPEP*,<sup>189</sup> *GLTSCR2*,<sup>190</sup> *IFITM1*, *IFITM2*, and *IFITM3*,<sup>191</sup> and *MAVS*.<sup>192</sup>

## Discussion: human studies

Traditional genome-wide methods have been applied to human viral infections generally,<sup>174,193</sup> but results have not been specific to coronaviruses, and it is unclear to what extent the observations are relevant to the current pandemic. Several dozen studies have investigated human genetic factors related to coronavirus infection. However, these studies have been limited by several potential factors. For endemic human coronaviruses, the mildness of disease may have deprioritized these studies; similar observations may explain the relative dearth of serologic knowledge related to these pathogens.<sup>6</sup> For coronaviruses associated with more severe human disease, such as MERS and SARS-CoV-1, the fact that these epidemics were limited more than the current pandemic crisis may have fortunately led to a lack of cases with which one might conduct traditional association studies (unlike some other respiratory infections leading to more widespread disease).<sup>194,195</sup> Additionally, these two severe conditions primarily affected human populations prior to the technological developments that led to wide availability of much cheaper and faster genomic sequencing.<sup>196</sup>

As shown (Supplementary Table 2), the small sample sizes of previous studies may have led to the preponderance of candidate gene studies. The sample sizes may also have precluded significant findings due to limitations of statistical power and the ability to replicate or validate findings. As previous human studies occurred in areas of the world affected by the coronavirus studied, it is possible that results from these studies would not extrapolate to other populations. Finally, different genes and loci are involved than those previously hypothesized. That is, hypothesis-free approaches may identify significant loci that were not identified by candidate approaches.

Based on announcements about multiple large-scale projects on host genetic factors and SARS-CoV-2, as well as the existence of larger genomic datasets that can be mined quickly and new methods that can be

used to address biological questions,<sup>197</sup> it is anticipated that considerable efforts - and an unfortunately large pool of research subjects - and may yield significant new results quickly.

### *Limitations*

There are multiple limitations to our summaries and analyses. First, it is likely that relevant articles were missed by our search process, and that key findings - including the study of certain genes - were therefore omitted. Along these lines, important findings within identified articles may also have been missed. Second, this analysis focused on DNA-based variants. These DNA-based genetic changes include those studied and identified through association studies as well as genes that were manipulated in experimental approaches, such as via knockout models to understand disease pathogenesis. Related 'omic approaches, such as targeted or broad transcriptomic or proteomic studies, are frequently used to understand important aspects of disease. These approaches can lead to knowledge regarding specific genetic changes. For example, observed transcriptomic changes may enable the identification of important DNA-based variants that explain disease by correlating transcriptomic data with results of DNA sequencing.<sup>198</sup> However, we categorized non-DNA based 'omic approaches separately from DNA-based studies, and did not attempt to comprehensively recapitulate what is known about host reaction to disease. Finally, as the studies varied in many aspects, such as how cases and controls were defined, and which loci were interrogated, we were careful about comparing or combining data between different studies.

**Table 1.** Summary of human studies (related to specific genes or loci) on host genetic factors related to coronaviruses. More details are available in Supplementary Table 2.

<b>Human</b>	<b>Method(s) or approach(es)</b>	<b>Key findings</b>	<b>PMID</b>
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coronavirus studied (other coronaviruses or pathogens)			
SARS-CoV-1	Analysis of association of <i>HLA</i> gene polymorphisms with susceptibility to SARS-CoV-1 infection or clinical parameters	Association of HLA-B* 4601 with severity of SARS-CoV-1 infection	12969506 <sup>199</sup>
SARS-CoV-1	Analysis of association of <i>HLA</i> gene polymorphisms with susceptibility to SARS-CoV-1 infection	HLA-B*0703, HLA-DRB1*0301 and co-inheritance of HLA-B*0703 and HLA-B60) were associated with susceptibility to SARS-CoV-1 infection	15243926 <sup>200</sup>
SARS-CoV-1	Analysis of association of <i>ACE2</i> polymorphisms with SARS-CoV-1 clinical parameters	No association of <i>ACE2</i> polymorphisms with SARS-CoV-1 outcomes	15331509 <sup>201</sup>
SARS-CoV-1	Analysis of association of <i>ACE</i> insertion/deletion (I/D) polymorphism with susceptibility to SARS-CoV-1 or	<i>ACE</i> D allele was associated with hypoxemia in SARS-CoV-1 infections	15381116 <sup>202</sup>

	clinical parameters		
SARS-CoV-1	Analysis of association of <i>OAS1</i> , <i>PKR</i> , <i>MX1</i> polymorphisms with susceptibility to SARS-CoV-1 or clinical parameters	<i>OAS1</i> rs3741981 and rs2660 were associated with SARS-CoV-1 susceptibility; <i>MX1</i> rs2071430 was associated in hypoxemia in SARS-CoV-1 infections	15766558 <sup>203</sup>
SARS-CoV-1	Analysis of association of <i>ACE</i> insertion/deletion (I/D) polymorphism with susceptibility to SARS-CoV-1 or clinical parameters	No association was found with <i>ACE</i> insertion/deletion (I/D) polymorphism and susceptibility to SARS-CoV-1 or clinical parameters	15819995 <sup>175</sup>
SARS-CoV-1	Analysis of association of <i>MBL</i> polymorphisms susceptibility to SARS-CoV-1 or clinical parameters and biological study of MBL	Serum MBL was lower in patients with SARS-CoV-1 infections than controls, and haplotypes associated with lower serum MBL were more frequent in patients with SARS-CoV-1 infections than in control subjects, but there was not association with mortality	15838797 <sup>204</sup>
SARS-CoV-1	Analysis of association of <i>ACE2</i> polymorphisms and	No association was found with <i>ACE2</i> polymorphisms and	15937940 <sup>205</sup>

	susceptibility to SARS-CoV-1 infection	susceptibility to SARS-CoV-1 infection	
SARS-CoV-1	Analysis of association of <i>MBL</i> polymorphisms and susceptibility to SARS-CoV-1 infection	<i>MBL</i> rs1800450 was associated with susceptibility to SARS-CoV-1 infection	16170752 <sup>206</sup>
SARS-CoV-1	Analysis of association of <i>FCGR2A</i> and <i>MBL</i> polymorphisms and susceptibility to SARS-CoV-1 infection or clinical parameters	Homozygosity for <i>FCGR2A</i> rs1801274, as well as a linear trend of <i>FCGR2A</i> genotypes, was associated with severe SARS-CoV-1 infection	16185324 <sup>207</sup>
SARS-CoV-1	Analysis of association of <i>CLEC4M</i> VNTR polymorphism with susceptibility to SARS-CoV-1 and biological studies of cells with these polymorphisms	Homozygosity for the <i>CLEC4M</i> VNTR polymorphism was associated with susceptibility to SARS-CoV-1, and homozygous cells had higher binding capacity for SARS-CoV-1, higher proteasome-dependent viral degradation, and lower capacity for trans infection.	16369534 <sup>208</sup>

SARS-CoV-1	Analysis of association of <i>HLA</i> polymorphisms with SARS-CoV-1 susceptibility	HLA-Cw*0801 was associated with susceptibility to SARS-CoV-1 infection	16455884 <sup>209</sup>
SARS-CoV-1	Analysis of association of polymorphisms in 65 genes with SARS-CoV-1 viral shedding	SARS-CoV-1 shedding was associated with alleles of <i>IL18</i> , <i>IL1A</i> , <i>RELB</i> , and <i>FLG2</i>	16652313 <sup>210</sup>
SARS-CoV-1	Analysis of association of <i>OAS1</i> and <i>MX1</i> polymorphisms with susceptibility to SARS-CoV-1	<i>OAS1</i> 3'-UTR rs2660 and <i>MX1</i> promoter rs2071430 were associated with susceptibility to SARS-CoV-1	16824203 <sup>211</sup>
SARS-CoV-1	Analysis of association of <i>CLEC4M</i> VNTR polymorphism with susceptibility to SARS-CoV-1 infection	No association was found with homozygosity for the <i>CLEC4M</i> VNTR polymorphism and susceptibility to SARS-CoV-1	17534354 <sup>212</sup>
SARS-CoV-1	Analysis of association of <i>CLEC4M</i> VNTR polymorphism with susceptibility to SARS-	No association was found with homozygosity for the <i>CLEC4M</i> VNTR polymorphism and	17534355 <sup>213</sup>

	CoV-1 infection	susceptibility to SARS-CoV-1	
SARS-CoV-1	Analysis of association of <i>CCL5</i> , <i>CXCL9</i> , and <i>CXCL10</i> polymorphisms with susceptibility to SARS-CoV-1 infection or clinical parameters	<i>CCL5</i> rs2107538 was associated with susceptibility to SARS-CoV-1 in one cohort and severe outcomes of SARS-CoV-1 infection in another cohort	17540042 <sup>214</sup>
SARS-CoV-1	Analysis of association of <i>FCER2</i> and <i>ICAM3</i> polymorphisms with susceptibility to SARS-CoV-1 or clinical parameters	Homozygosity for <i>ICAM</i> rs2304237 was associated with higher LDH levels and lower total WBC counts	17570115 <sup>215</sup>
SARS-CoV-1	Analysis of association of <i>CD14</i> , <i>TLR2</i> , and <i>TLR4</i> polymorphisms with susceptibility to SARS-CoV-1 or clinical parameters	<i>CD14</i> rs2569190 was associated with severe SARS-CoV-1 infection (this data was also combined with previous data, suggesting that this and <i>FCGR2A</i> -RR131 are risk genotypes for severe SARS-CoV-1 infection)	17913858 <sup>216</sup>
SARS-CoV-1	Analysis of association of <i>TNF</i> polymorphisms with	<i>TNF</i> polymorphisms were associated with susceptibility to	18312678 <sup>217</sup>



	interstitial lung fibrosis and femoral head osteonecrosis in discharged SARS-CoV-1 patients	SARS-CoV-1 and with femoral head necrosis in discharged SARS-CoV-1 patients	
SARS-CoV-1	Analysis of association of polymorphisms in <i>IL12RB1</i> with susceptibility to SARS-CoV-1 or clinical outcomes	<i>IL12RB1</i> (+1664) polymorphism was associated with susceptibility to SARS-CoV-1 infection	18478121 <sup>218</sup>
SARS-CoV-1	Analysis of association of polymorphisms in 4 C-type lectin genes with susceptibility to SARS-CoV-1 infection	No association of polymorphisms in C-type lectin genes with SARS-CoV-1 susceptibility	18697825 <sup>219</sup>
SARS-CoV-1	Analysis of association of polymorphisms in 9 inflammatory response genes with susceptibility to SARS-CoV-1 or clinical outcomes	No association of polymorphisms in inflammatory response genes with SARS-CoV-1 susceptibility or clinical outcomes	18708672 <sup>220</sup>
SARS-CoV-1	Analysis of association of polymorphisms in <i>MASP2</i> with susceptibility to SARS-CoV-1 infection	No association of <i>MASP2</i> polymorphisms with SARS-CoV-1 susceptibility	19405982 <sup>221</sup>

SARS-CoV-1	Analysis of association of <i>HLA</i> polymorphisms with SARS-CoV-1 susceptibility	HLA-DRB1*12 was more frequent in SARS-CoV-1 patients versus controls; HLA-DRB1*1202 showed the strongest association with SARS-CoV-1 infection in a dominant model	19445991 <sup>222</sup>
SARS-CoV-1	Analysis of association of polymorphisms in 64 genes with susceptibility to SARS-CoV-1 infection	CXCL10(-938AA) is protective (but appears jointly with other variants); <i>FGL2</i> (+158T/*) is associated with higher susceptibility unless combined with CXCL10/(-938AA), when jointly is associated with lower susceptibility	19590927 <sup>223</sup>
SARS-CoV-1	Analysis of association of <i>CD209</i> rs4804803 with SARS-CoV-1 outcomes	<i>CD209</i> polymorphism rs4804803 is associated with lower LDH levels (and therefore, worse prognosis)	20359516 <sup>224</sup>

SARS-CoV-1	Biological study and analysis of <i>MX1</i> promoter polymorphisms with suppressed interferon beta induction and association of <i>MX1</i> promoter polymorphisms with susceptibility to SARS-CoV-1 infection	Differences were observed in binding affinity to nuclear proteins related to IFN-beta stimulation; <i>MX1</i> rs2071430 was associated with lower risk of SARS-CoV-1 infection	20462354 <sup>225</sup>
SARS-CoV-1	Analysis of association of <i>HLA</i> gene polymorphisms with SARS-CoV-1 susceptibility	No significant associations (after correction) <i>HLA</i> gene polymorphisms with SARS-CoV-1 susceptibility were identified	20864745 <sup>226</sup>
SARS-CoV-1	Biological study of in vitro functional effects of rs4804803 and analysis of association of <i>CD209</i> rs4804803 with SARS-CoV-1 outcomes	<i>CD209</i> polymorphism rs4804803 was associated with lower risk of high admission LDH levels, and may contribute to a reduced immune response/reduced lung injury during disease progression	20864747 <sup>227</sup>
SARS-CoV-1	Analysis of association of <i>AHSG</i> and <i>CYP4F3A</i> polymorphisms with SARS-CoV-1 susceptibility	<i>AHSG</i> polymorphism rs2248690 was associated with SARS-CoV-1 susceptibility (as well as higher	21904596 <sup>228</sup>

		AHSG serum concentration)	
SARS-CoV-1	Analysis of association of <i>HLA</i> polymorphisms with SARS-CoV-1 susceptibility	<i>HLA-Cw*1502</i> conferred resistance against SARS infection is associated with resistance to SARS-CoV-1 infection	21958371 <sup>229</sup>
SARS-CoV-1	Analysis of association of <i>HLA</i> polymorphisms with SARS-CoV-1 susceptibility and outcome	No association of <i>HLA</i> polymorphisms with SARS-CoV-1 susceptibility and outcome were identified	24643938 <sup>230</sup>
SARS-CoV-1	Analysis of association of <i>CCL2</i> and <i>MBL</i> polymorphisms with susceptibility to SARS-CoV-1 infection	Variants in <i>MBL</i> (rs1800450) and <i>CCL2</i> (rs1024611) ( <i>CCL2</i> ) were cumulatively associated with SARS-CoV-1 susceptibility	25818534 <sup>231</sup>
SARS-CoV-1 (and other respiratory pathogens)	Meta-analysis of 386 studies on susceptibility to tuberculosis, influenza, respiratory syncytial virus, SARS-CoV-1, and pneumonia	In a pooled model, variants in <i>IL4</i> were positively associated with susceptibility after multiple testing correction	26524966 <sup>174</sup>
SARS-CoV-2	Case report of death due to	Suggestion of genetic	32277694 <sup>232</sup>

	COVID-19 in three previously healthy adult brothers	predisposition due to apparent familial clustering	
SARS-CoV-2	Case reports of two patients with X-linked agammaglobulinemia (and documented pathogenic variants in <i>BTK</i> )	Patients recovered, suggesting that B cell response might not be required to overcome the SARS-CoV-2 infection	32319118 <sup>233</sup>
SARS-CoV-2	Analysis of association of <i>IFITM3</i> rs12252 with clinical outcomes of SARS-CoV-2 infection	Significant association of homozygosity <i>IFITM3</i> rs12252 with disease severity	32348495 <sup>234</sup>

Abbreviations: CCoV: canine coronavirus; FCoV: feline coronavirus; human coronavirus 229E: HCoV-229E; human coronavirus NL63: HCoV NL63; human coronavirus OC43: HCoV OC43; LDH: lactate-dehydrogenase; MBL: Mannose-binding lectin; MERS-CoV: middle east respiratory syndrome coronavirus; SARS-CoV-1: severe acute respiratory syndrome coronavirus 1; SARS-CoV-2: severe acute respiratory syndrome coronavirus 2; SL-CoV: SARS-Cov-1-like coronaviruses; TGEV: porcine transmissible gastroenteritis coronavirus; WBC: white blood cell; WT: wild-type

**Table 2.** Summary of relevant mouse studies related to coronavirus. Note that the different studies have disparate objectives, many of which more directly involve aspects of immunopathogenesis versus standard host genetic questions regarding why specific genetic variants may affect disease susceptibility and outcomes.

<b>Mouse (Human gene)</b>	<b>Method(s) or approach(es)</b>	<b>Pathway: Key findings</b>	<b>PMID</b>
<i>Ace2 (ACE2)</i>	Humanized mice, SARS-CoV1	Viral receptor: humanized <i>Ace2</i> mice, increased infection, permissive gene	18495771 <sup>235</sup>
<i>Atg5 (ATG5)</i>	KO, MHV infection	Autophagy: required for MHV replication, permissive gene	14699140 <sup>236</sup>
<i>Atp1a1 (ATP1A1)</i>	knockdown and chemical inhibition across many coronaviruses	Ion channel: chemical inhibition or gene silencing, results in blocking viral entry, permissive gene	25653449 <sup>237</sup>
<i>B2m (B2M)</i>	KO, MHV infection	Adaptive immunity: MHC Class I/CD8 T-cells required for host immune response, protective gene	8799201 <sup>238</sup> ; 10023135 <sup>239</sup>
<i>Bnip3 (BNIP3)</i>	Cull culture model, MHV infection	Apoptosis: pro-apoptotic gene is suppressed upon viral entry, likely protective	14599795 <sup>240</sup>
<i>C3 (C3)</i>	KO, SARS-CoV1	Complement pathway: decreased complement activation leads to less	30301856 <sup>241</sup>

		severe disease, implicated immune driven component of disease, gene is permissive	
<i>C5ar1(C5AR1)</i>	KO, MHV infection	Complement pathway: Complement pathway exacerbates hepatitis, KO decreases manifestations, decreased susceptibility, permissive gene	24604562 <sup>242</sup>
<i>Ccr1 (CCR1)</i>	KO, MHV infection	Cytokine pathways: Loss of <i>Ccr1</i> increased mortality, protective gene	18158733 <sup>243</sup>
<i>Ccr2 (CCR2)</i>	KO, MHV infection	Cytokine pathways: <i>Ccr2</i> required for clearance of the virus from CNS, KO increased susceptibility, protective gene	15518805 <sup>244</sup>
<i>Ccr5 (CCR5)</i>	KO, MHV infection	Cytokine pathways: KO decreased severity of demyelination disease, permissive gene	11543653 <sup>245</sup>
<i>Cd200r1(CD200R1)</i>	KO, MHV infection	Immune receptor: Cd200 KO increases clearance of MHV, decreases susceptibility, permissive gene	22615569 <sup>179</sup>
<i>Ceacam1</i>	Isoform specific	Viral receptor: KO are fully resistant to	11483763 <sup>246</sup> ;

<i>(CEACAM1)</i>	transgenic and KO, MHV infection	infection, liver, and CNS manifestations, permissive gene	15331748 <sup>247</sup>
<i>Cxcl10 (CXCL10)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T2), KO leads to increased mortality, protective gene	17142734 <sup>248</sup> ; 17617609 <sup>249</sup>
<i>Cxcl9 (CXCL9)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T2), KO had increased MHV associated mortality, protective gene	18973912 <sup>250</sup>
<i>Dpp4 (DPP4)</i>	Various transgenic and humanized models, MERS infection	Viral receptor: humanized <i>Dpp4</i> or mutations, deletions in mouse <i>Dpp4</i> leads to MERS induced ARDS, permissive gene	24574399 <sup>251</sup> ; 25653445 <sup>252</sup> ; 29691378 <sup>253</sup> ; 30142928 <sup>254</sup> ; 31883094 <sup>255</sup>
<i>Ebi3 (EBI3)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T2), KO leads to increased mortality, protective gene	23102608 <sup>256</sup>
<i>Foxn1 (FOXN1)</i>	KO, MHV infection	Adaptive immunity: Athymic mice lacking T-cells unable to clear infection cause severe disseminated disease, protective gene	8799201 <sup>238</sup> ; 15070459 <sup>257</sup>



<i>H2-Ab1 (H2AB1)</i>	KO, MHV infection	Adaptive immunity: MHC Class I/CD4 T-cells required for host immune response, protective gene	8799201 <sup>238</sup>
<i>Ifih1 (IFIH1)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T1), KO more severe, disseminated MHV infection, decreased survival, protective gene	26423942 <sup>258</sup>
<i>Ifnar, (IFNAR)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T1), KO leads to increased mortality and higher viral titers, protective gene	18667505 <sup>259</sup> ; 19215224 <sup>260</sup> ; 19650917 <sup>261</sup>
<i>Ifnar1 (IFNAR1)</i>	KO, SARS-CoV1	Interferon pathway: Type 1, II and III interferon does not alter infection for SARS-CoV-1, in contrast to MHV	20386712 <sup>262</sup>
<i>Ifng (IFNG)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T2), KO has increased mortality, decreased viral clearance, protective gene	9973424 <sup>263</sup> ; 11864749 <sup>138</sup>
<i>Ifngr1 (IFNGR1)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T2), KO has increased mortality, decreased viral clearance, protective	8752933 <sup>264</sup> ; 15039522 <sup>265</sup> ;

		gene	20042510 <sup>266</sup>
<i>Ifngr1 (IFNGR1)</i>	KO, SARS-CoV1	Interferon pathway: Type 1, II and III interferon does not alter infection for SARS-CoV-1, in contrast to MHV	20386712 <sup>262</sup>
<i>Ighm (IGHM)</i>	KO, MHV infection	Adaptive immunity: B-cell deficient develop subclinical infection and transmit virus for increased time span, protective gene	15027615 <sup>267</sup>
<i>Il1r1 (IL1R1)</i>	KO, MHV infection	Cytokine pathways: KO shows reduced viral replication, mortality, and disease progression, permissive gene	26367131 <sup>268</sup>
<i>Mavs (MAVS)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T1), viral sensor, studied in the presence of attenuated virus, protective gene	29717007 <sup>269</sup>
<i>Myd88 (MYD88)</i>	KO, rMA15 infection	Cytokine pathways: downstream of multiple pathways, KO increased susceptibility to MHV infection and mortality, protective gene	19079579 <sup>270</sup>

<i>Prkdc (PRKDC)</i>	KO, MHV infection	Adaptive immunity: Loss of T- and B-cells cause severe disseminated infection, protective gene	8799201 <sup>238</sup>
<i>Rag1 (RAG1)</i>	KO, MHV infection	Adaptive immunity: Loss of mature T- and B-cells leads to failure to clear infection, protective gene	17142734 <sup>248</sup> ; 18973912 <sup>250</sup> ; 25428866 <sup>271</sup> ; 27604627 <sup>272</sup>
<i>Serpine1 (SERPINE1)</i>	KO, SARS-CoV1 infection	Tissue remodeling: KO mice are more susceptible to infection and inflammation, protective gene	23919993 <sup>273</sup>
<i>Stat1 (STAT1)</i>	KO/KI, HCoV-229E infection	Cytokine pathways: Interferon related (T1), KO increased susceptibility HCoV in transgenic APN model, protective	15919828 <sup>274</sup>
<i>Stat1 (STAT1)</i>	KO, SARS-CoV-1	Cytokine pathways, KO worsens disease, increases susceptibility, protective gene	20386712 <sup>262</sup> ; 23142821 <sup>275</sup>
<i>Stat6 (STAT6)</i>	Conditional KO, <i>LysM and FoxJ1, Stat1/Stat6 -/- double knockout,</i>	Cytokine pathways: conditional KO of Stat1 in macrophages but not ciliated epithelial cells showed pulmonary disease, double knockout of <i>Stat1</i> and	23015710 <sup>276</sup>

	SARS-CoV-1	<i>Stat6</i> relieves pulmonary disease, implicates alternatively activated macrophages, permissive gene	
<i>Ticam2 (TICAM2)</i>	KO, SARS-CoV1	Immune receptor: TLR-mediated, KO developed more severe infection, increased viral titer, and increased weight loss, protective gene	28592648 <sup>277</sup>
<i>Tlr2 (TLR2)</i>	KO, MHV infection	Immune receptor: KO decreases inflammatory response, protective gene	19740307 <sup>278</sup>
<i>Tlr3 (TLR3)</i>	KO, SARS-CoV1	Immune receptor: TLR mediated, KO more susceptible for SARS-CoV-1 infection, although no increased mortality, protective gene	26015500 <sup>279</sup>
<i>Tlr4 (TLR4)</i>	KO, SARS-CoV1	Immune receptor: TLR mediated, KO more susceptible for SARS-CoV-1 infection, although no increased mortality, protective gene	26015500 <sup>279</sup>
<i>Tlr7 (TLR7)</i>	KO, MHV infection	Immune receptor: viral sensor, KO prolonged infection, protective gene	29717007 <sup>269</sup>

<i>Tram1 (TRAM1)</i>	KO, SARS-CoV1	Immune receptor: TLR mediated, KO more susceptible for SARS-CoV-1 infection, although no increased mortality, protective gene	26015500 <sup>279</sup>
<i>Trif (TRIF)</i>	KO, SARS-CoV1	Immune receptor: TLR mediated, KO more susceptible to SARS-CoV-1 infection, more severe infection with increased interferon signaling, protective gene	26015500 <sup>279</sup>
<i>Trim55 (TRIM55)</i>	KO, SARS-CoV1	Uncharacterized pathway: contributed to lung pathology, KO decreased severity, permissive gene	26452100 <sup>137</sup>
<i>Usp18 (USP18)</i>	KO, MHV infection	Cytokine pathways: Interferon related (T1), KO leads to increased survival, decreased pathology and viral titer, gene is permissive	24648452 <sup>280</sup>

Abbreviations: ARDS: acute respiratory distress syndrome; CNS: central nervous system; KI: knock-in;

KO: knockout; MERS: middle east respiratory syndrome; MHC: major histocompatibility complex; MHV:

mouse hepatitis virus; SARS-CoV-1: SARS-CoV-1: severe acute respiratory syndrome coronavirus 1; T1:

type 1; T2: type 2; TLR: Toll-like receptor

### Figure Legends

**Figure 1.** Description of articles identified through PubMed searches described in Methods.

**Figure 2.** Genes investigated in animal studies related to coronavirus disease. Human genes are shown only for those studies in multiple species analyses; other human gene details are presented elsewhere.

**Figure 3.** 3A: Significant genetic associations with human susceptibility to coronavirus disease. Both protective and permissive genes are shown. Only studies reporting odds ratios (OR) and confidence intervals are shown. 3B: Significant genetic associations with human clinical variables and outcomes related to coronavirus disease. Both protective and permissive genes are shown. Only studies reporting odds ratios (OR) and confidence intervals (CI) are shown (PMID 32348495 did not include CI).

### **Acknowledgments**

This research was supported [in part] by the Intramural Research Program of the National Human Genome Research Institute, National Institutes of Health. PD was supported by the US National Institutes of Health award number 2R01AI148049-21A1. DATC was supported by the US National Institutes of Health award number R01-AI114703-01.

### **References**

1. Zhang X, Tan Y, Ling Y, et al. Viral and host factors related to the clinical outcome of COVID-19. *Nature* 2020.
2. Letko M, Miazgowiec K, McMinn R, et al. Adaptive Evolution of MERS-CoV to Species Variation in DPP4. *Cell Rep* 2018;24:1730-7.
3. Initiative C-HG. The COVID-19 Host Genetics Initiative, a global initiative to elucidate the role of host genetic factors in susceptibility and severity of the SARS-CoV-2 virus pandemic. *Eur J Hum Genet* 2020.

4. Gaunt ER, Hardie A, Claas EC, Simmonds P, Templeton KE. Epidemiology and clinical presentations of the four human coronaviruses 229E, HKU1, NL63, and OC43 detected over 3 years using a novel multiplex real-time PCR method. *J Clin Microbiol* 2010;48:2940-7.
5. Perlman S, Dandekar AA. Immunopathogenesis of coronavirus infections: implications for SARS. *Nat Rev Immunol* 2005;5:917-27.
6. Huang AT, Garcia-Carrera B, Hitchings MDT, et al. A systematic review of antibody immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease. *medRxiv* 2020.
7. Bakkers MJ, Zeng Q, Feitsma LJ, et al. Coronavirus receptor switch explained from the stereochemistry of protein-carbohydrate interactions and a single mutation. *Proc Natl Acad Sci U S A* 2016;113:E3111-9.
8. Hofmann H, Pyrc K, van der Hoek L, Geier M, Berkhout B, Pohlmann S. Human coronavirus NL63 employs the severe acute respiratory syndrome coronavirus receptor for cellular entry. *Proc Natl Acad Sci U S A* 2005;102:7988-93.
9. Li W, Moore MJ, Vasilieva N, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature* 2003;426:450-4.
10. Li KK, Yip CW, Hon CC, Lam CY, Zeng F, Leung FC. Characterisation of animal angiotensin-converting enzyme 2 receptors and use of pseudotyped virus to correlate receptor binding with susceptibility of SARS-CoV infection. *Hong Kong Med J* 2012;18 Suppl 3:35-8.
11. Yan R, Zhang Y, Li Y, Xia L, Guo Y, Zhou Q. Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2. *Science* 2020;367:1444-8.
12. Yeager CL, Ashmun RA, Williams RK, et al. Human aminopeptidase N is a receptor for human coronavirus 229E. *Nature* 1992;357:420-2.

13. Li Z, Tomlinson AC, Wong AH, et al. The human coronavirus HCoV-229E S-protein structure and receptor binding. *Elife* 2019;8.
14. Tresnan DB, Levis R, Holmes KV. Feline aminopeptidase N serves as a receptor for feline, canine, porcine, and human coronaviruses in serogroup I. *J Virol* 1996;70:8669-74.
15. Benbacer L, Kut E, Besnardeau L, Laude H, Delmas B. Interspecies aminopeptidase-N chimeras reveal species-specific receptor recognition by canine coronavirus, feline infectious peritonitis virus, and transmissible gastroenteritis virus. *J Virol* 1997;71:734-7.
16. Delmas B, Gelfi J, L'Haridon R, et al. Aminopeptidase N is a major receptor for the enteropathogenic coronavirus TGEV. *Nature* 1992;357:417-20.
17. Wang N, Shi X, Jiang L, et al. Structure of MERS-CoV spike receptor-binding domain complexed with human receptor DPP4. *Cell Res* 2013;23:986-93.
18. Raj VS, Mou H, Smits SL, et al. Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature* 2013;495:251-4.
19. Peck KM, Scobey T, Swanstrom J, et al. Permissivity of Dipeptidyl Peptidase 4 Orthologs to Middle East Respiratory Syndrome Coronavirus Is Governed by Glycosylation and Other Complex Determinants. *J Virol* 2017;91.
20. Williams RK, Jiang GS, Holmes KV. Receptor for mouse hepatitis virus is a member of the carcinoembryonic antigen family of glycoproteins. *Proc Natl Acad Sci U S A* 1991;88:5533-6.
21. Rockx B, Baas T, Zornetzer GA, et al. Early upregulation of acute respiratory distress syndrome-associated cytokines promotes lethal disease in an aged-mouse model of severe acute respiratory syndrome coronavirus infection. *J Virol* 2009;83:7062-74.
22. Ostaszewski M, Mazein A, Gillespie ME, et al. COVID-19 Disease Map, building a computational repository of SARS-CoV-2 virus-host interaction mechanisms. *Sci Data* 2020;7:136.



23. de Lang A, Baas T, Teal T, et al. Functional genomics highlights differential induction of antiviral pathways in the lungs of SARS-CoV-infected macaques. *PLoS Pathog* 2007;3:e112.
24. Murray MF, Kenny EE, Ritchie MD, et al. COVID-19 outcomes and the human genome. *Genet Med* 2020.
25. Zhou W, Zhao Z, Nielsen JB, et al. Scalable generalized linear mixed model for region-based association tests in large biobanks and cohorts. *Nat Genet* 2020.
26. East ML, Moestl K, Benetka V, et al. Coronavirus infection of spotted hyenas in the Serengeti ecosystem. *Vet Microbiol* 2004;102:1-9.
27. Le Poder S. Feline and canine coronaviruses: common genetic and pathobiological features. *Adv Virol* 2011;2011:609465.
28. Roberts A, Vogel L, Guarner J, et al. Severe acute respiratory syndrome coronavirus infection of golden Syrian hamsters. *J Virol* 2005;79:503-11.
29. Chan JF, Zhang AJ, Yuan S, et al. Simulation of the clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in golden Syrian hamster model: implications for disease pathogenesis and transmissibility. *Clin Infect Dis* 2020.
30. Lau SY, Wang P, Mok BW, et al. Attenuated SARS-CoV-2 variants with deletions at the S1/S2 junction. *Emerg Microbes Infect* 2020;9:837-42.
31. Liang L, He C, Lei M, et al. Pathology of guinea pigs experimentally infected with a novel reovirus and coronavirus isolated from SARS patients. *DNA Cell Biol* 2005;24:485-90.
32. Liu RY, Wu LZ, Huang BJ, et al. Adenoviral expression of a truncated S1 subunit of SARS-CoV spike protein results in specific humoral immune responses against SARS-CoV in rats. *Virus Res* 2005;112:24-31.

33. Nagata N, Iwata N, Hasegawa H, et al. Participation of both host and virus factors in induction of severe acute respiratory syndrome (SARS) in F344 rats infected with SARS coronavirus. *J Virol* 2007;81:1848-57.
34. Fukushi S, Mizutani T, Sakai K, et al. Amino acid substitutions in the s2 region enhance severe acute respiratory syndrome coronavirus infectivity in rat angiotensin-converting enzyme 2-expressing cells. *J Virol* 2007;81:10831-4.
35. Ren W, Qu X, Li W, et al. Difference in receptor usage between severe acute respiratory syndrome (SARS) coronavirus and SARS-like coronavirus of bat origin. *J Virol* 2008;82:1899-907.
36. Fouchier RA, Kuiken T, Schutten M, et al. Aetiology: Koch's postulates fulfilled for SARS virus. *Nature* 2003;423:240.
37. Rowe T, Gao G, Hogan RJ, et al. Macaque model for severe acute respiratory syndrome. *J Virol* 2004;78:11401-4.
38. McAuliffe J, Vogel L, Roberts A, et al. Replication of SARS coronavirus administered into the respiratory tract of African Green, rhesus and cynomolgus monkeys. *Virology* 2004;330:8-15.
39. Compton SR, Stephensen CB, Snyder SW, Weismiller DG, Holmes KV. Coronavirus species specificity: murine coronavirus binds to a mouse-specific epitope on its carcinoembryonic antigen-related receptor glycoprotein. *J Virol* 1992;66:7420-8.
40. Gagneten S, Scanga CA, Dveksler GS, Beauchemin N, Percy D, Holmes KV. Attachment glycoproteins and receptor specificity of rat coronaviruses. *Lab Anim Sci* 1996;46:159-66.
41. Baric RS, Yount B, Hensley L, Peel SA, Chen W. Episodic evolution mediates interspecies transfer of a murine coronavirus. *J Virol* 1997;71:1946-55.
42. Bumstead N. Genetic resistance to avian viruses. *Rev Sci Tech* 1998;17:249-55.
43. Cook J, Otsuki K, Huggins M, Bumstead N. Investigations into resistance of chicken lines to infection with infectious bronchitis virus. *Adv Exp Med Biol* 1990;276:491-6.

44. Ignjatovic J, Reece R, Ashton F. Susceptibility of three genetic lines of chicks to infection with a nephropathogenic T strain of avian infectious bronchitis virus. *J Comp Pathol* 2003;128:92-8.
45. Asif M, Lowenthal JW, Ford ME, Schat KA, Kimpton WG, Bean AG. Interleukin-6 expression after infectious bronchitis virus infection in chickens. *Viral Immunol* 2007;20:479-86.
46. Dawes ME, Griggs LM, Collisson EW, Briles WE, Drechsler Y. Dramatic differences in the response of macrophages from B2 and B19 MHC-defined haplotypes to interferon gamma and polyinosinic:polycytidylic acid stimulation. *Poult Sci* 2014;93:830-8.
47. da Silva AP, Hauck R, Zhou H, Gallardo RA. Understanding Immune Resistance to Infectious Bronchitis Using Major Histocompatibility Complex Chicken Lines. *Avian Dis* 2017;61:358-65.
48. Bumstead N, Huggins MB, Cook JK. Genetic differences in susceptibility to a mixture of avian infectious bronchitis virus and *Escherichia coli*. *Br Poult Sci* 1989;30:39-48.
49. Bacon LD, Hunter DB, Zhang HM, Brand K, Etches R. Retrospective evidence that the MHC (B haplotype) of chickens influences genetic resistance to attenuated infectious bronchitis vaccine strains in chickens. *Avian Pathol* 2004;33:605-9.
50. Luo C, Qu H, Ma J, et al. A genome-wide association study identifies major loci affecting the immune response against infectious bronchitis virus in chicken. *Infect Genet Evol* 2014;21:351-8.
51. Wang W, Zhang T, Zhang G, et al. Genome-wide association study of antibody level response to NDV and IBV in Jinghai yellow chicken based on SLAF-seq technology. *J Appl Genet* 2015;56:365-73.
52. Pedersen NC, Liu H, Gandolfi B, Lyons LA. The influence of age and genetics on natural resistance to experimentally induced feline infectious peritonitis. *Vet Immunol Immunopathol* 2014;162:33-40.
53. Robison RL, Holzworth J, Gilmore CE. Naturally occurring feline infectious peritonitis: signs and clinical diagnosis. *J Am Vet Med Assoc* 1971;158:Suppl 2:981-6.

54. Foley JE, Poland A, Carlson J, Pedersen NC. Risk factors for feline infectious peritonitis among cats in multiple-cat environments with endemic feline enteric coronavirus. *J Am Vet Med Assoc* 1997;210:1313-8.
55. Rohrbach BW, Legendre AM, Baldwin CA, Lein DH, Reed WM, Wilson RB. Epidemiology of feline infectious peritonitis among cats examined at veterinary medical teaching hospitals. *J Am Vet Med Assoc* 2001;218:1111-5.
56. Pesteanu-Somogyi LD, Radzai C, Pressler BM. Prevalence of feline infectious peritonitis in specific cat breeds. *J Feline Med Surg* 2006;8:1-5.
57. Norris JM, Bosward KL, White JD, Baral RM, Catt MJ, Malik R. Clinicopathological findings associated with feline infectious peritonitis in Sydney, Australia: 42 cases (1990-2002). *Aust Vet J* 2005;83:666-73.
58. Worthing KA, Wigney DI, Dhand NK, et al. Risk factors for feline infectious peritonitis in Australian cats. *J Feline Med Surg* 2012;14:405-12.
59. Judd F, Newman LK, Komiti AA. Time for a new zeitgeist in perinatal mental health. *Aust N Z J Psychiatry* 2018;52:112-6.
60. Bell ET, Malik R, Norris JM. The relationship between the feline coronavirus antibody titre and the age, breed, gender and health status of Australian cats. *Aust Vet J* 2006;84:2-7.
61. Addie DD, Kennedy LJ, Ryvar R, et al. Feline leucocyte antigen class II polymorphism and susceptibility to feline infectious peritonitis. *J Feline Med Surg* 2004;6:59-62.
62. Hsieh LE, Chueh LL. Identification and genotyping of feline infectious peritonitis-associated single nucleotide polymorphisms in the feline interferon-gamma gene. *Vet Res* 2014;45:57.
63. Wang YT, Hsieh LE, Dai YR, Chueh LL. Polymorphisms in the feline TNFA and CD209 genes are associated with the outcome of feline coronavirus infection. *Vet Res* 2014;45:123.

64. Golovko L, Lyons LA, Liu H, Sorensen A, Wehnert S, Pedersen NC. Genetic susceptibility to feline infectious peritonitis in Birman cats. *Virus Res* 2013;175:58-63.
65. Martina BE, Haagmans BL, Kuiken T, et al. Virology: SARS virus infection of cats and ferrets. *Nature* 2003;425:915.
66. Sabir JS, Lam TT, Ahmed MM, et al. Co-circulation of three camel coronavirus species and recombination of MERS-CoVs in Saudi Arabia. *Science* 2016;351:81-4.
67. Dudas G, Carvalho LM, Rambaut A, Bedford T. MERS-CoV spillover at the camel-human interface. *Elife* 2018;7.
68. Chu DKW, Hui KPY, Perera R, et al. MERS coronaviruses from camels in Africa exhibit region-dependent genetic diversity. *Proc Natl Acad Sci U S A* 2018;115:3144-9.
69. Miguel E, Chevalier V, Ayelet G, et al. Risk factors for MERS coronavirus infection in dromedary camels in Burkina Faso, Ethiopia, and Morocco, 2015. *Euro Surveill* 2017;22.
70. Eckerle I, Corman VM, Muller MA, Lenk M, Ulrich RG, Drosten C. Replicative Capacity of MERS Coronavirus in Livestock Cell Lines. *Emerg Infect Dis* 2014;20:276-9.
71. van Doremalen N, Miazgowicz KL, Milne-Price S, et al. Host species restriction of Middle East respiratory syndrome coronavirus through its receptor, dipeptidyl peptidase 4. *J Virol* 2014;88:9220-32.
72. Widagdo W, Raj VS, Schipper D, et al. Differential Expression of the Middle East Respiratory Syndrome Coronavirus Receptor in the Upper Respiratory Tracts of Humans and Dromedary Camels. *J Virol* 2016;90:4838-42.
73. Zhang Z, Shen L, Gu X. Evolutionary Dynamics of MERS-CoV: Potential Recombination, Positive Selection and Transmission. *Sci Rep* 2016;6:25049.
74. Corman VM, Eckerle I, Memish ZA, et al. Link of a ubiquitous human coronavirus to dromedary camels. *Proc Natl Acad Sci U S A* 2016;113:9864-9.

75. Chu YK, Ali GD, Jia F, et al. The SARS-CoV ferret model in an infection-challenge study. *Virology* 2008;374:151-63.
76. Kim YI, Kim SG, Kim SM, et al. Infection and Rapid Transmission of SARS-CoV-2 in Ferrets. *Cell Host Microbe* 2020;27:704-9 e2.
77. Shi J, Wen Z, Zhong G, et al. Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS-coronavirus 2. *Science* 2020.
78. van den Brand JM, Haagmans BL, Leijten L, et al. Pathology of experimental SARS coronavirus infection in cats and ferrets. *Vet Pathol* 2008;45:551-62.
79. Bunyavanich S, Do A, Vicencio A. Nasal Gene Expression of Angiotensin-Converting Enzyme 2 in Children and Adults. *JAMA* 2020.
80. Evans MR, Simpson RW. The coronavirus avian infectious bronchitis virus requires the cell nucleus and host transcriptional factors. *Virology* 1980;105:582-91.
81. Dveksler GS, Pensiero MN, Cardellichio CB, et al. Cloning of the mouse hepatitis virus (MHV) receptor: expression in human and hamster cell lines confers susceptibility to MHV. *J Virol* 1991;65:6881-91.
82. Jeffers SA, Tusell SM, Gillim-Ross L, et al. CD209L (L-SIGN) is a receptor for severe acute respiratory syndrome coronavirus. *Proc Natl Acad Sci U S A* 2004;101:15748-53.
83. Lai ZW, Lew RA, Yarski MA, Mu FT, Andrews RK, Smith AI. The identification of a calmodulin-binding domain within the cytoplasmic tail of angiotensin-converting enzyme-2. *Endocrinology* 2009;150:2376-81.
84. Lin SC, Leng CH, Wu SC. Generating stable Chinese hamster ovary cell clones to produce a truncated SARS-CoV spike protein for vaccine development. *Biotechnol Prog* 2010;26:1733-40.

85. Chan CM, Lau SK, Woo PC, et al. Identification of major histocompatibility complex class I C molecule as an attachment factor that facilitates coronavirus HKU1 spike-mediated infection. *J Virol* 2009;83:1026-35.
86. Schickli JH, Thackray LB, Sawicki SG, Holmes KV. The N-terminal region of the murine coronavirus spike glycoprotein is associated with the extended host range of viruses from persistently infected murine cells. *J Virol* 2004;78:9073-83.
87. van Doremalen N, Miazgowicz KL, Munster VJ. Mapping the Specific Amino Acid Residues That Make Hamster DPP4 Functional as a Receptor for Middle East Respiratory Syndrome Coronavirus. *J Virol* 2016;90:5499-502.
88. Guenet JL. Assessing the genetic component of the susceptibility of mice to viral infections. *Brief Funct Genomic Proteomic* 2005;4:225-40.
89. Gledhill AW, Andrewes CH. A hepatitis virus of mice. *Br J Exp Pathol* 1951;32:559-68.
90. Gledhill AW, Andrewes CH, Dick GW. Production of hepatitis in mice by the combined action of two filterable agents. *Lancet* 1952;2:509-11.
91. Gallily R, Warwick A, Bang FB. Effect of Cortisone of Genetic Resistance to Mouse Hepatitis Virus in Vivo and in Vitro. *Proc Natl Acad Sci U S A* 1964;51:1158-64.
92. Kantoch M, Warwick A, Bang FB. The cellular nature of genetic susceptibility to a virus. *J Exp Med* 1963;117:781-98.
93. Wilson GA, Dales S. In vivo and in vitro models of demyelinating disease: efficiency of virus spread and formation of infectious centers among glial cells is genetically determined by the murine host. *J Virol* 1988;62:3371-7.
94. Tardieu M, Boespflug O, Barbe T. Selective tropism of a neurotropic coronavirus for ependymal cells, neurons, and meningeal cells. *J Virol* 1986;60:574-82.

95. Weiser W, Bang FB. Macrophages genetically resistant to mouse hepatitis virus converted in vitro to susceptible macrophages. *J Exp Med* 1976;143:690-5.
96. Bang FB. The use of a genetically incompatible combination of host and virus (MHV) for the study of mechanisms of host resistance. *Adv Exp Med Biol* 1981;142:359-73.
97. Yokomori K, Lai MM. The receptor for mouse hepatitis virus in the resistant mouse strain SJL is functional: implications for the requirement of a second factor for viral infection. *J Virol* 1992;66:6931-8.
98. Dindzans VJ, Skamene E, Levy GA. Susceptibility/resistance to mouse hepatitis virus strain 3 and macrophage procoagulant activity are genetically linked and controlled by two non-H-2-linked genes. *J Immunol* 1986;137:2355-60.
99. Shif I, Bang FB. In vitro interaction of mouse hepatitis virus and macrophages from genetically resistant mice. I. Adsorption of virus and growth curves. *J Exp Med* 1970;131:843-50.
100. Levy-Leblond E, Oth D, Dupuy JM. Genetic study of mouse sensitivity to MHV3 infection: influence of the H-2 complex. *J Immunol* 1979;122:1359-62.
101. Knobler RL, Tunison LA, Oldstone MB. Host genetic control of mouse hepatitis virus type 4 (JHM strain) replication. I. Restriction of virus amplification and spread in macrophages from resistant mice. *J Gen Virol* 1984;65 ( Pt 9):1543-8.
102. Knobler RL, Taylor BA, Wooddell MK, Beamer WG, Oldstone MB. Host genetic control of mouse hepatitis virus type-4 (JHM strain) replication. II. The gene locus for susceptibility is linked to the Svp-2 locus on mouse chromosome 7. *Exp Clin Immunogenet* 1984;1:217-22.
103. Stohlman SA, Frelinger JA, Weiner LP. Resistance to fatal central nervous system disease by mouse hepatitis virus, strain JHM. II. Adherent cell-mediated protection. *J Immunol* 1980;124:1733-9.
104. Smith MS, Click RE, Plagemann PG. Control of mouse hepatitis virus replication in macrophages by a recessive gene on chromosome 7. *J Immunol* 1984;133:428-32.



105. Pereira CA, Lucchiari MA, Modolell M, Kuhn L, Lefkovits I. An attempt to identify gene products related to the induction of an antiviral state in macrophages resistant and sensitive to IFN-gamma. *Res Virol* 1993;144:479-86.
106. Dveksler G, Nedellec P, Lu JH, et al. Characterization of a new gene that encodes a functional MHV receptor and progress in the identification of the virus-binding site(s). *Adv Exp Med Biol* 1995;380:345-50.
107. Weiser WY, Bang FB. Blocking of in vitro and in vivo susceptibility to mouse hepatitis virus. *J Exp Med* 1977;146:1467-72.
108. Kyuwa S, Yamaguchi K, Toyoda Y, Fujiwara K, Hilgers J. Acute and late disease induced by murine coronavirus, strain JHM, in a series of recombinant inbred strains between BALB/cHeA and STS/A mice. *Microb Pathog* 1992;12:95-104.
109. Robbins J, Robbins PF, Kozak CA, Callahan R. The mouse biliary glycoprotein gene (Bgp): partial nucleotide sequence, expression, and chromosomal assignment. *Genomics* 1991;10:583-7.
110. Sussman MA, Shubin RA, Kyuwa S, Stohlman SA. T-cell-mediated clearance of mouse hepatitis virus strain JHM from the central nervous system. *J Virol* 1989;63:3051-6.
111. Castro RF, Evans GD, Jaszewski A, Perlman S. Coronavirus-induced demyelination occurs in the presence of virus-specific cytotoxic T cells. *Virology* 1994;200:733-43.
112. Daya M, Wong F, Cervin M, et al. Mouse fibroblast mutants selected for survival against mouse hepatitis virus infection show increased resistance to infection and virus-induced cell fusion. *Adv Exp Med Biol* 1990;276:59-66.
113. Damy SB, Vassao RC, Lucchiari MA, Pereira CA, Sant'Anna OA. A comparative study of resistance to MHV3 infection in genetically homogeneous and heterogeneous mouse populations. *Braz J Med Biol Res* 1992;25:1025-7.

114. Daya M, Wong F, Cervin M, et al. Mutation of host cell determinants which discriminate between lytic and persistent mouse hepatitis virus infection results in a fusion-resistant phenotype. *J Gen Virol* 1989;70 ( Pt 12):3335-46.
115. Knobler RL, Linthicum DS, Cohn M. Host genetic regulation of acute MHV-4 viral encephalomyelitis and acute experimental autoimmune encephalomyelitis in (BALB/cKe x SJL/J) recombinant-inbred mice. *J Neuroimmunol* 1985;8:15-28.
116. Lavelle GC, Bang FB. Influence of type and concentration of sera in vitro on susceptibility of genetically resistant cells to mouse hepatitis virus. *J Gen Virol* 1971;12:233-8.
117. Boyle JF, Weismiller DG, Holmes KV. Genetic resistance to mouse hepatitis virus correlates with absence of virus-binding activity on target tissues. *J Virol* 1987;61:185-9.
118. Arnheiter H, Haller O. Inborn resistance of mice to mouse hepatitis virus type 3 (MHV3): liver parenchymal cells express phenotype in culture. *Adv Exp Med Biol* 1981;142:409-17.
119. Arnheiter H, Baechi T, Haller O. Adult mouse hepatocytes in primary monolayer culture express genetic resistance to mouse hepatitis virus type 3. *J Immunol* 1982;129:1275-81.
120. Decimo D, Boespflug O, Meunier-Rotival M, Hadchouel M, Tardieu M. Genetic restriction of murine hepatitis virus type 3 expression in liver and brain: comparative study in BALB/c and C3H mice by immunochemistry and hybridization in situ. *Arch Virol* 1993;130:269-77.
121. Wang Y, Burnier M, Detrick B, Hooks JJ. Genetic predisposition to coronavirus-induced retinal disease. *Invest Ophthalmol Vis Sci* 1996;37:250-4.
122. Wang Y, Detrick B, Yu ZX, Zhang J, Chesky L, Hooks JJ. The role of apoptosis within the retina of coronavirus-infected mice. *Invest Ophthalmol Vis Sci* 2000;41:3011-8.
123. Lassnig C, Kolb A, Strobl B, Enjuanes L, Muller M. Studying human pathogens in animal models: fine tuning the humanized mouse. *Transgenic Res* 2005;14:803-6.

124. Sorensen O, Dugre R, Percy D, Dales S. In vivo and in vitro models of demyelinating disease: endogenous factors influencing demyelinating disease caused by mouse hepatitis virus in rats and mice. *Infect Immun* 1982;37:1248-60.
125. Barthold SW, Smith AL. Role of host age and genotype in murine enterotropic coronavirus infection. *Adv Exp Med Biol* 1993;342:371-6.
126. MacNamara KC, Chua MM, Phillips JJ, Weiss SR. Contributions of the viral genetic background and a single amino acid substitution in an immunodominant CD8+ T-cell epitope to murine coronavirus neurovirulence. *J Virol* 2005;79:9108-18.
127. Mansour S, Mercier G, Oth D. Lymphokine release as measurement of anti-mouse hepatitis virus type 3 (MHV3) cellular reactions in various mouse lines exhibiting differential susceptibilities to MHV3-induced paralysis. *Acta Virol* 1990;34:423-32.
128. Barthold SW, Smith AL. Viremic dissemination of mouse hepatitis virus-JHM following intranasal inoculation of mice. *Arch Virol* 1992;122:35-44.
129. Lamontagne L, Decarie D, Dupuy JM. Host cell resistance to mouse hepatitis virus type 3 is expressed in vitro in macrophages and lymphocytes. *Viral Immunol* 1989;2:37-45.
130. Knobler RL, Haspel MV, Oldstone MB. Mouse hepatitis virus type 4 (JHM strains). induced fatal central nervous system disease. I. genetic control and murine neuron as the susceptible site of disease. *J Exp Med* 1981;153:832-43.
131. Vassao RC, Cabrera WH, Ibanez OC, Pereira CA. Specific T-cell response correlates with resistance of genetic heterogeneous mouse populations to mouse hepatitis virus 3 infection. *Arch Virol* 1995;140:1235-45.
132. Vassao RC, Sant' Anna OA, Pereira CA. A genetic analysis of macrophage activation and specific antibodies in relation to the resistance of heterogeneous mouse populations to MHV3 infection. *Arch Virol* 1994;139:417-25.

133. Vassao RC, Mello IG, Pereira CA. Role of macrophages, interferon gamma and procoagulant activity in the resistance of genetic heterogeneous mouse populations to mouse hepatitis virus infection. *Arch Virol* 1994;137:277-88.
134. Chung S, Sinclair S, Leibowitz J, Skamene E, Fung LS, Levy G. Cellular and metabolic requirements for induction of macrophage procoagulant activity by murine hepatitis virus strain 3 in vitro. *J Immunol* 1991;146:271-8.
135. Chen W, Madden VJ, Bagnell CR, Jr., Baric RS. Host-derived intracellular immunization against mouse hepatitis virus infection. *Virology* 1997;228:318-32.
136. Williams RK, Snyder SW, Holmes KV. MHV-resistant SJL/J mice express a non-functional homolog to the MHV receptor glycoprotein. *Adv Exp Med Biol* 1990;276:45-50.
137. Gralinski LE, Ferris MT, Aylor DL, et al. Genome Wide Identification of SARS-CoV Susceptibility Loci Using the Collaborative Cross. *PLoS Genet* 2015;11:e1005504.
138. Kyuwa S, Shibata S, Tagawa Y, Iwakura Y, Machii K, Urano T. Acute hepatic failure in IFN-gamma-deficient BALB/c mice after murine coronavirus infection. *Virus Res* 2002;83:169-77.
139. Li W, Luo R, He Q, van Kuppeveld FJM, Rottier PJM, Bosch BJ. Aminopeptidase N is not required for porcine epidemic diarrhea virus cell entry. *Virus Res* 2017;235:6-13.
140. Whitworth KM, Rowland RRR, Petrovan V, et al. Resistance to coronavirus infection in amino peptidase N-deficient pigs. *Transgenic Res* 2019;28:21-32.
141. Cui T, Theuns S, Xie J, Van den Broeck W, Nauwynck HJ. Role of Porcine Aminopeptidase N and Sialic Acids in Porcine Coronavirus Infections in Primary Porcine Enterocytes. *Viruses* 2020;12.
142. Chen J, Pan K, Chen Z, et al. Production of porcine aminopeptidase N (pAPN) site-specific edited pigs. *Anim Sci J* 2019;90:366-71.

143. Tu CF, Chuang CK, Hsiao KH, et al. Lessening of porcine epidemic diarrhoea virus susceptibility in piglets after editing of the CMP-N-glycolylneuraminic acid hydroxylase gene with CRISPR/Cas9 to nullify N-glycolylneuraminic acid expression. *PLoS One* 2019;14:e0217236.
144. Compton SR, Barthold SW, Smith AL. The cellular and molecular pathogenesis of coronaviruses. *Lab Anim Sci* 1993;43:15-28.
145. Lau SK, Woo PC, Li KS, et al. Discovery of a novel coronavirus, China Rattus coronavirus HKU24, from Norway rats supports the murine origin of Betacoronavirus 1 and has implications for the ancestor of Betacoronavirus lineage A. *J Virol* 2015;89:3076-92.
146. Liu Y, Cai Y, Zhang X. Induction of caspase-dependent apoptosis in cultured rat oligodendrocytes by murine coronavirus is mediated during cell entry and does not require virus replication. *J Virol* 2003;77:11952-63.
147. Liu Y, Zhang X. Expression of cellular oncogene Bcl-xL prevents coronavirus-induced cell death and converts acute infection to persistent infection in progenitor rat oligodendrocytes. *J Virol* 2005;79:47-56.
148. Fukuma A, Tani H, Taniguchi S, Shimojima M, Saijo M, Fukushi S. Inability of rat DPP4 to allow MERS-CoV infection revealed by using a VSV pseudotype bearing truncated MERS-CoV spike protein. *Arch Virol* 2015;160:2293-300.
149. Fish I, Boissinot S. Contrasted patterns of variation and evolutionary convergence at the antiviral OAS1 gene in old world primates. *Immunogenetics* 2015;67:487-99.
150. Leopardi S, Holmes EC, Gastaldelli M, et al. Interplay between co-divergence and cross-species transmission in the evolutionary history of bat coronaviruses. *Infect Genet Evol* 2018;58:279-89.
151. Ar Gouilh M, Puechmaille SJ, Diancourt L, et al. SARS-CoV related Betacoronavirus and diverse Alphacoronavirus members found in western old-world. *Virology* 2018;517:88-97.

152. Evermann JF, Heeney JL, Roelke ME, McKeirnan AJ, O'Brien SJ. Biological and pathological consequences of feline infectious peritonitis virus infection in the cheetah. *Arch Virol* 1988;102:155-71.
153. O'Brien SJ, Troyer JL, Roelke M, Marker L, Pecon-Slattery J. Plagues and adaptation: Lessons from the Felidae models for SARS and AIDS. *Biol Conserv* 2006;131:255-67.
154. Dobrynin P, Liu S, Tamazian G, et al. Genomic legacy of the African cheetah, *Acinonyx jubatus*. *Genome Biol* 2015;16:277.
155. O'Brien SJ, Roelke ME, Marker L, et al. Genetic basis for species vulnerability in the cheetah. *Science* 1985;227:1428-34.
156. Li W, Zhang C, Sui J, et al. Receptor and viral determinants of SARS-coronavirus adaptation to human ACE2. *EMBO J* 2005;24:1634-43.
157. Li F. Structural analysis of major species barriers between humans and palm civets for severe acute respiratory syndrome coronavirus infections. *J Virol* 2008;82:6984-91.
158. Sheahan T, Rockx B, Donaldson E, Corti D, Baric R. Pathways of cross-species transmission of synthetically reconstructed zoonotic severe acute respiratory syndrome coronavirus. *J Virol* 2008;82:8721-32.
159. Song HD, Tu CC, Zhang GW, et al. Cross-host evolution of severe acute respiratory syndrome coronavirus in palm civet and human. *Proc Natl Acad Sci U S A* 2005;102:2430-5.
160. Haagmans BL, Andeweg AC, Osterhaus AD. The application of genomics to emerging zoonotic viral diseases. *PLoS Pathog* 2009;5:e1000557.
161. Cui J, Han N, Streicker D, et al. Evolutionary relationships between bat coronaviruses and their hosts. *Emerg Infect Dis* 2007;13:1526-32.
162. Yu M, Tachedjian M, Crameri G, Shi Z, Wang LF. Identification of key amino acid residues required for horseshoe bat angiotensin-I converting enzyme 2 to function as a receptor for severe acute respiratory syndrome coronavirus. *J Gen Virol* 2010;91:1708-12.

163. Hou Y, Peng C, Yu M, et al. Angiotensin-converting enzyme 2 (ACE2) proteins of different bat species confer variable susceptibility to SARS-CoV entry. *Arch Virol* 2010;155:1563-9.
164. Cui J, Eden JS, Holmes EC, Wang LF. Adaptive evolution of bat dipeptidyl peptidase 4 (dpp4): implications for the origin and emergence of Middle East respiratory syndrome coronavirus. *Virology* 2013;10:304.
165. Lau SKP, Fan RYY, Luk HKH, et al. Replication of MERS and SARS coronaviruses in bat cells offers insights to their ancestral origins. *Emerg Microbes Infect* 2018;7:209.
166. Sakaguchi AY, Shows TB. Coronavirus 229E susceptibility in man-mouse hybrids is located on human chromosome 15. *Somatic Cell Genet* 1982;8:83-94.
167. Inoue Y, Tanaka N, Tanaka Y, et al. Clathrin-dependent entry of severe acute respiratory syndrome coronavirus into target cells expressing ACE2 with the cytoplasmic tail deleted. *J Virol* 2007;81:8722-9.
168. Liu B, Li NL, Wang J, et al. Overlapping and distinct molecular determinants dictating the antiviral activities of TRIM56 against flaviviruses and coronavirus. *J Virol* 2014;88:13821-35.
169. von Brunn A, Ciesek S, von Brunn B, Carbajo-Lozoya J. Genetic deficiency and polymorphisms of cyclophilin A reveal its essential role for Human Coronavirus 229E replication. *Curr Opin Virol* 2015;14:56-61.
170. Zhao X, Sehgal M, Hou Z, et al. Identification of Residues Controlling Restriction versus Enhancing Activities of IFITM Proteins on Entry of Human Coronaviruses. *J Virol* 2018;92.
171. Hussain M, Jabeen N, Raza F, et al. Structural variations in human ACE2 may influence its binding with SARS-CoV-2 spike protein. *J Med Virol* 2020.
172. Nguyen A, David JK, Maden SK, et al. Human leukocyte antigen susceptibility map for SARS-CoV-2. *J Virol* 2020.

173. Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* 2020;181:271-80 e8.
174. Patarcic I, Gelemanovic A, Kirin M, et al. The role of host genetic factors in respiratory tract infectious diseases: systematic review, meta-analyses and field synopsis. *Sci Rep* 2015;5:16119.
175. Chan KC, Tang NL, Hui DS, et al. Absence of association between angiotensin converting enzyme polymorphism and development of adult respiratory distress syndrome in patients with severe acute respiratory syndrome: a case control study. *BMC Infect Dis* 2005;5:26.
176. Verity R, Okell LC, Dorigatti I, et al. Estimates of the severity of coronavirus disease 2019: a model-based analysis. *Lancet Infect Dis* 2020.
177. La Vignera S, Cannarella R, Condorelli RA, Torre F, Aversa A, Calogero AE. Sex-Specific SARS-CoV-2 Mortality: Among Hormone-Modulated ACE2 Expression, Risk of Venous Thromboembolism and Hypovitaminosis D. *Int J Mol Sci* 2020;21.
178. Li MY, Li L, Zhang Y, Wang XS. Expression of the SARS-CoV-2 cell receptor gene ACE2 in a wide variety of human tissues. *Infect Dis Poverty* 2020;9:45.
179. Karnam G, Rygiel TP, Raaben M, et al. CD200 receptor controls sex-specific TLR7 responses to viral infection. *PLoS Pathog* 2012;8:e1002710.
180. Zhang Y, Zheng N, Nan P, Cao Y, Hasegawa M, Zhong Y. Computational simulation of interactions between SARS coronavirus spike mutants and host species-specific receptors. *Comput Biol Chem* 2007;31:134-7.
181. Yang XH, Deng W, Tong Z, et al. Mice transgenic for human angiotensin-converting enzyme 2 provide a model for SARS coronavirus infection. *Comp Med* 2007;57:450-9.
182. Yoshikawa N, Yoshikawa T, Hill T, et al. Differential virological and immunological outcome of severe acute respiratory syndrome coronavirus infection in susceptible and resistant transgenic mice expressing human angiotensin-converting enzyme 2. *J Virol* 2009;83:5451-65.



183. Miyoshi-Akiyama T, Ishida I, Fukushi M, et al. Fully human monoclonal antibody directed to proteolytic cleavage site in severe acute respiratory syndrome (SARS) coronavirus S protein neutralizes the virus in a rhesus macaque SARS model. *J Infect Dis* 2011;203:1574-81.
184. Bao L, Deng W, Huang B, et al. The pathogenicity of SARS-CoV-2 in hACE2 transgenic mice. *Nature* 2020.
185. Raj VS, Smits SL, Provacia LB, et al. Adenosine deaminase acts as a natural antagonist for dipeptidyl peptidase 4-mediated entry of the Middle East respiratory syndrome coronavirus. *J Virol* 2014;88:1834-8.
186. Kandeel M, Elaiziz MA, Kandeel A, Altaher AA, Kitade Y. Association of host tropism of Middle East syndrome coronavirus with the amino acid structure of host cell receptor dipeptidyl peptidase 4. *Acta Virol* 2014;58:359-63.
187. Agrawal AS, Garron T, Tao X, et al. Generation of a transgenic mouse model of Middle East respiratory syndrome coronavirus infection and disease. *J Virol* 2015;89:3659-70.
188. Algaissi A, Agrawal AS, Han S, et al. Elevated Human Dipeptidyl Peptidase 4 Expression Reduces the Susceptibility of hDPP4 Transgenic Mice to Middle East Respiratory Syndrome Coronavirus Infection and Disease. *J Infect Dis* 2019;219:829-35.
189. Tusell SM, Schittone SA, Holmes KV. Mutational analysis of aminopeptidase N, a receptor for several group 1 coronaviruses, identifies key determinants of viral host range. *J Virol* 2007;81:1261-73.
190. Wang P, Meng W, Han SC, Li CC, Wang XJ, Wang XJ. The nucleolar protein GLTSCR2 is required for efficient viral replication. *Sci Rep* 2016;6:36226.
191. Zhao X, Guo F, Liu F, et al. Interferon induction of IFITM proteins promotes infection by human coronavirus OC43. *Proc Natl Acad Sci U S A* 2014;111:6756-61.
192. Lei Y, Moore CB, Liesman RM, et al. MAVS-mediated apoptosis and its inhibition by viral proteins. *PLoS One* 2009;4:e5466.

193. Fumagalli M, Pozzoli U, Cagliani R, et al. Genome-wide identification of susceptibility alleles for viral infections through a population genetics approach. *PLoS Genet* 2010;6:e1000849.
194. Curtis J, Luo Y, Zenner HL, et al. Susceptibility to tuberculosis is associated with variants in the *ASAP1* gene encoding a regulator of dendritic cell migration. *Nat Genet* 2015;47:523-7.
195. Sveinbjornsson G, Gudbjartsson DF, Halldorsson BV, et al. HLA class II sequence variants influence tuberculosis risk in populations of European ancestry. *Nat Genet* 2016;48:318-22.
196. Wheeler DA, Srinivasan M, Egholm M, et al. The complete genome of an individual by massively parallel DNA sequencing. *Nature* 2008;452:872-6.
197. Li B, Clohisey SM, Chia BS, et al. Genome-wide CRISPR screen identifies host dependency factors for influenza A virus infection. *Nat Commun* 2020;11:164.
198. Fresard L, Smail C, Ferraro NM, et al. Identification of rare-disease genes using blood transcriptome sequencing and large control cohorts. *Nat Med* 2019;25:911-9.
199. Lin M, Tseng HK, Trejaut JA, et al. Association of HLA class I with severe acute respiratory syndrome coronavirus infection. *BMC Med Genet* 2003;4:9.
200. Ng MH, Lau KM, Li L, et al. Association of human-leukocyte-antigen class I (B\*0703) and class II (DRB1\*0301) genotypes with susceptibility and resistance to the development of severe acute respiratory syndrome. *J Infect Dis* 2004;190:515-8.
201. Chiu RW, Tang NL, Hui DS, et al. ACE2 gene polymorphisms do not affect outcome of severe acute respiratory syndrome. *Clin Chem* 2004;50:1683-6.
202. Itoyama S, Keicho N, Quy T, et al. ACE1 polymorphism and progression of SARS. *Biochem Biophys Res Commun* 2004;323:1124-9.
203. Hamano E, Hijikata M, Itoyama S, et al. Polymorphisms of interferon-inducible genes OAS-1 and MxA associated with SARS in the Vietnamese population. *Biochem Biophys Res Commun* 2005;329:1234-9.

204. Ip WK, Chan KH, Law HK, et al. Mannose-binding lectin in severe acute respiratory syndrome coronavirus infection. *J Infect Dis* 2005;191:1697-704.
205. Itoyama S, Keicho N, Hijikata M, et al. Identification of an alternative 5'-untranslated exon and new polymorphisms of angiotensin-converting enzyme 2 gene: lack of association with SARS in the Vietnamese population. *Am J Med Genet A* 2005;136:52-7.
206. Zhang H, Zhou G, Zhi L, et al. Association between mannose-binding lectin gene polymorphisms and susceptibility to severe acute respiratory syndrome coronavirus infection. *J Infect Dis* 2005;192:1355-61.
207. Yuan FF, Tanner J, Chan PK, et al. Influence of FcγRIIA and MBL polymorphisms on severe acute respiratory syndrome. *Tissue Antigens* 2005;66:291-6.
208. Chan VS, Chan KY, Chen Y, et al. Homozygous L-SIGN (CLEC4M) plays a protective role in SARS coronavirus infection. *Nat Genet* 2006;38:38-46.
209. Chen YM, Liang SY, Shih YP, et al. Epidemiological and genetic correlates of severe acute respiratory syndrome coronavirus infection in the hospital with the highest nosocomial infection rate in Taiwan in 2003. *J Clin Microbiol* 2006;44:359-65.
210. Chen WJ, Yang JY, Lin JH, et al. Nasopharyngeal shedding of severe acute respiratory syndrome-associated coronavirus is associated with genetic polymorphisms. *Clin Infect Dis* 2006;42:1561-9.
211. He J, Feng D, de Vlas SJ, et al. Association of SARS susceptibility with single nucleic acid polymorphisms of OAS1 and MxA genes: a case-control study. *BMC Infect Dis* 2006;6:106.
212. Tang NL, Chan PK, Hui DS, et al. Lack of support for an association between CLEC4M homozygosity and protection against SARS coronavirus infection. *Nat Genet* 2007;39:691-2; author reply 4-6.
213. Zhi L, Zhou G, Zhang H, et al. Lack of support for an association between CLEC4M homozygosity and protection against SARS coronavirus infection. *Nat Genet* 2007;39:692-4; author reply 4-6.

214. Ng MW, Zhou G, Chong WP, et al. The association of RANTES polymorphism with severe acute respiratory syndrome in Hong Kong and Beijing Chinese. *BMC Infect Dis* 2007;7:50.
215. Chan KY, Ching JC, Xu MS, et al. Association of ICAM3 genetic variant with severe acute respiratory syndrome. *J Infect Dis* 2007;196:271-80.
216. Yuan FF, Boehm I, Chan PK, et al. High prevalence of the CD14-159CC genotype in patients infected with severe acute respiratory syndrome-associated coronavirus. *Clin Vaccine Immunol* 2007;14:1644-5.
217. Wang S, Wei M, Han Y, et al. Roles of TNF-alpha gene polymorphisms in the occurrence and progress of SARS-Cov infection: a case-control study. *BMC Infect Dis* 2008;8:27.
218. Tang F, Liu W, Zhang F, et al. IL-12 RB1 genetic variants contribute to human susceptibility to severe acute respiratory syndrome infection among Chinese. *PLoS One* 2008;3:e2183.
219. Li H, Tang NL, Chan PK, et al. Polymorphisms in the C-type lectin genes cluster in chromosome 19 and predisposition to severe acute respiratory syndrome coronavirus (SARS-CoV) infection. *J Med Genet* 2008;45:752-8.
220. Khoo US, Chan KY, Chan VS, et al. Role of polymorphisms of the inflammatory response genes and DC-SIGNR in genetic susceptibility to SARS and other infections. *Hong Kong Med J* 2008;14 Suppl 4:31-5.
221. Wang Y, Yan J, Shi Y, et al. Lack of association between polymorphisms of MASP2 and susceptibility to SARS coronavirus infection. *BMC Infect Dis* 2009;9:51.
222. Keicho N, Itoyama S, Kashiwase K, et al. Association of human leukocyte antigen class II alleles with severe acute respiratory syndrome in the Vietnamese population. *Hum Immunol* 2009;70:527-31.
223. Hsieh YH, Chen CW, Schmitz SF, et al. Candidate genes associated with susceptibility for SARS-coronavirus. *Bull Math Biol* 2010;72:122-32.

224. Chan KY, Xu MS, Ching JC, et al. CD209 (DC-SIGN) -336A>G promoter polymorphism and severe acute respiratory syndrome in Hong Kong Chinese. *Hum Immunol* 2010;71:702-7.
225. Ching JC, Chan KY, Lee EH, et al. Significance of the myxovirus resistance A (MxA) gene -123C>a single-nucleotide polymorphism in suppressed interferon beta induction of severe acute respiratory syndrome coronavirus infection. *J Infect Dis* 2010;201:1899-908.
226. Ng MH, Cheng SH, Lau KM, et al. Immunogenetics in SARS: a case-control study. *Hong Kong Med J* 2010;16:29-33.
227. Chan KY, Xu MS, Ching JC, et al. Association of a single nucleotide polymorphism in the CD209 (DC-SIGN) promoter with SARS severity. *Hong Kong Med J* 2010;16:37-42.
228. Zhu X, Wang Y, Zhang H, et al. Genetic variation of the human alpha-2-Heremans-Schmid glycoprotein (AHSG) gene associated with the risk of SARS-CoV infection. *PLoS One* 2011;6:e23730.
229. Wang SF, Chen KH, Chen M, et al. Human-leukocyte antigen class I Cw 1502 and class II DR 0301 genotypes are associated with resistance to severe acute respiratory syndrome (SARS) infection. *Viral Immunol* 2011;24:421-6.
230. Yuan FF, Velickovic Z, Ashton LJ, et al. Influence of HLA gene polymorphisms on susceptibility and outcome post infection with the SARS-CoV virus. *Virology* 2014;29:128-30.
231. Tu X, Chong WP, Zhai Y, et al. Functional polymorphisms of the CCL2 and MBL genes cumulatively increase susceptibility to severe acute respiratory syndrome coronavirus infection. *J Infect* 2015;71:101-9.
232. Yousefzadegan S, Rezaei N. Case Report: Death Due to Novel Coronavirus Disease (COVID-19) in Three Brothers. *Am J Trop Med Hyg* 2020.
233. Soresina A, Moratto D, Chiarini M, et al. Two X-linked agammaglobulinemia patients develop pneumonia as COVID-19 manifestation but recover. *Pediatr Allergy Immunol* 2020.

234. Zhang Y, Qin L, Zhao Y, et al. Interferon-induced transmembrane protein-3 genetic variant rs12252-C is associated with disease severity in COVID-19. *J Infect Dis* 2020.
235. Netland J, Meyerholz DK, Moore S, Cassell M, Perlman S. Severe acute respiratory syndrome coronavirus infection causes neuronal death in the absence of encephalitis in mice transgenic for human ACE2. *J Virol* 2008;82:7264-75.
236. Prentice E, Jerome WG, Yoshimori T, Mizushima N, Denison MR. Coronavirus replication complex formation utilizes components of cellular autophagy. *J Biol Chem* 2004;279:10136-41.
237. Burkard C, Verheije MH, Haagmans BL, et al. ATP1A1-mediated Src signaling inhibits coronavirus entry into host cells. *J Virol* 2015;89:4434-48.
238. Houtman JJ, Fleming JO. Dissociation of demyelination and viral clearance in congenitally immunodeficient mice infected with murine coronavirus JHM. *J Neurovirol* 1996;2:101-10.
239. Lavi E, Das Sarma J, Weiss SR. Cellular reservoirs for coronavirus infection of the brain in beta2-microglobulin knockout mice. *Pathobiology* 1999;67:75-83.
240. Cai Y, Liu Y, Yu D, Zhang X. Down-regulation of transcription of the proapoptotic gene BNip3 in cultured astrocytes by murine coronavirus infection. *Virology* 2003;316:104-15.
241. Gralinski LE, Sheahan TP, Morrison TE, et al. Complement Activation Contributes to Severe Acute Respiratory Syndrome Coronavirus Pathogenesis. *mBio* 2018;9.
242. Xu GL, Chen J, Yang F, Li GQ, Zheng LX, Wu YZ. C5a/C5aR pathway is essential for the pathogenesis of murine viral fulminant hepatitis by way of potentiating Fgl2/fibroleukin expression. *Hepatology* 2014;60:114-24.
243. Hickey MJ, Held KS, Baum E, Gao JL, Murphy PM, Lane TE. CCR1 deficiency increases susceptibility to fatal coronavirus infection of the central nervous system. *Viral Immunol* 2007;20:599-608.

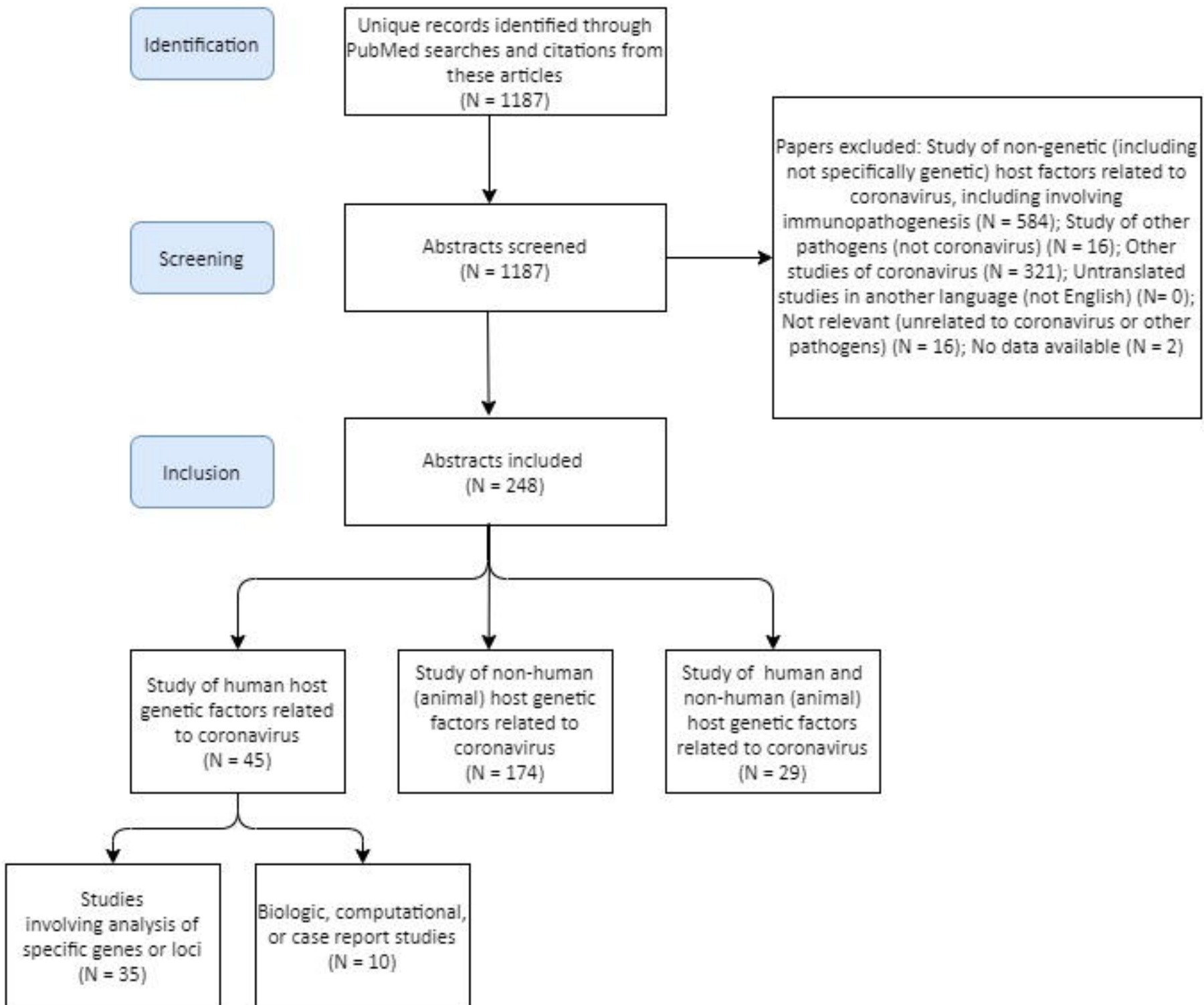
244. Held KS, Chen BP, Kuziel WA, Rollins BJ, Lane TE. Differential roles of CCL2 and CCR2 in host defense to coronavirus infection. *Virology* 2004;329:251-60.
245. Glass WG, Liu MT, Kuziel WA, Lane TE. Reduced macrophage infiltration and demyelination in mice lacking the chemokine receptor CCR5 following infection with a neurotropic coronavirus. *Virology* 2001;288:8-17.
246. Blau DM, Turbide C, Tremblay M, et al. Targeted disruption of the Ceacam1 (MHVR) gene leads to reduced susceptibility of mice to mouse hepatitis virus infection. *J Virol* 2001;75:8173-86.
247. Hemmila E, Turbide C, Olson M, Jothy S, Holmes KV, Beauchemin N. Ceacam1a<sup>-/-</sup> mice are completely resistant to infection by murine coronavirus mouse hepatitis virus A59. *J Virol* 2004;78:10156-65.
248. Stiles LN, Hardison JL, Schaumburg CS, Whitman LM, Lane TE. T cell antiviral effector function is not dependent on CXCL10 following murine coronavirus infection. *J Immunol* 2006;177:8372-80.
249. Walsh KB, Edwards RA, Romero KM, Kotlajich MV, Stohlman SA, Lane TE. Expression of CXC chemokine ligand 10 from the mouse hepatitis virus genome results in protection from viral-induced neurological and liver disease. *J Immunol* 2007;179:1155-65.
250. Muse M, Kane JA, Carr DJ, Farber JM, Lane TE. Insertion of the CXC chemokine ligand 9 (CXCL9) into the mouse hepatitis virus genome results in protection from viral-induced encephalitis and hepatitis. *Virology* 2008;382:132-44.
251. Cockrell AS, Peck KM, Yount BL, et al. Mouse dipeptidyl peptidase 4 is not a functional receptor for Middle East respiratory syndrome coronavirus infection. *J Virol* 2014;88:5195-9.
252. Peck KM, Cockrell AS, Yount BL, Scobey T, Baric RS, Heise MT. Glycosylation of mouse DPP4 plays a role in inhibiting Middle East respiratory syndrome coronavirus infection. *J Virol* 2015;89:4696-9.
253. Jiang Y, Zhao G, Song N, et al. Blockade of the C5a-C5aR axis alleviates lung damage in hDPP4-transgenic mice infected with MERS-CoV. *Emerg Microbes Infect* 2018;7:77.

254. Fan C, Wu X, Liu Q, et al. A Human DPP4-Knockin Mouse's Susceptibility to Infection by Authentic and Pseudotyped MERS-CoV. *Viruses* 2018;10.
255. Leist SR, Cockrell AS. Genetically Engineering a Susceptible Mouse Model for MERS-CoV-Induced Acute Respiratory Distress Syndrome. *Methods Mol Biol* 2020;2099:137-59.
256. Tirotta E, Duncker P, Oak J, et al. Epstein-Barr virus-induced gene 3 negatively regulates neuroinflammation and T cell activation following coronavirus-induced encephalomyelitis. *J Neuroimmunol* 2013;254:110-6.
257. Scavizzi F, Raspa M. Tissue distribution and duration of mouse hepatitis virus in naturally infected immunocompetent ICR (CD-1) and immunodeficient athymic nude-nu mouse strains used for ovarian transplantation and in vitro fertilization. *Lab Anim* 2004;38:189-99.
258. Zalinger ZB, Elliott R, Rose KM, Weiss SR. MDA5 Is Critical to Host Defense during Infection with Murine Coronavirus. *J Virol* 2015;89:12330-40.
259. Roth-Cross JK, Bender SJ, Weiss SR. Murine coronavirus mouse hepatitis virus is recognized by MDA5 and induces type I interferon in brain macrophages/microglia. *J Virol* 2008;82:9829-38.
260. Raaben M, Prins HJ, Martens AC, Rottier PJ, De Haan CA. Non-invasive imaging of mouse hepatitis coronavirus infection reveals determinants of viral replication and spread in vivo. *Cell Microbiol* 2009;11:825-41.
261. Raaben M, Groot Koerkamp MJ, Rottier PJ, de Haan CA. Type I interferon receptor-independent and -dependent host transcriptional responses to mouse hepatitis coronavirus infection in vivo. *BMC Genomics* 2009;10:350.
262. Frieman MB, Chen J, Morrison TE, et al. SARS-CoV pathogenesis is regulated by a STAT1 dependent but a type I, II and III interferon receptor independent mechanism. *PLoS Pathog* 2010;6:e1000849.



263. Parra B, Hinton DR, Marten NW, et al. IFN-gamma is required for viral clearance from central nervous system oligodendroglia. *J Immunol* 1999;162:1641-7.
264. Schijns VE, Wierda CM, van Hoeij M, Horzinek MC. Exacerbated viral hepatitis in IFN-gamma receptor-deficient mice is not suppressed by IL-12. *J Immunol* 1996;157:815-21.
265. de Wit MC, Horzinek MC, Haagmans BL, Schijns V. Host-dependent type 1 cytokine responses driven by inactivated viruses may fail to default in the absence of IL-12 or IFN-alpha/beta. *J Gen Virol* 2004;85:795-803.
266. Parra GI, Bergmann CC, Phares TW, Hinton DR, Atkinson R, Stohlman SA. Gamma interferon signaling in oligodendrocytes is critical for protection from neurotropic coronavirus infection. *J Virol* 2010;84:3111-5.
267. Compton SR, Ball-Goodrich LJ, Paturzo FX, Macy JD. Transmission of enterotropic mouse hepatitis virus from immunocompetent and immunodeficient mice. *Comp Med* 2004;54:29-35.
268. Guo S, Yang C, Diao B, et al. The NLRP3 Inflammasome and IL-1beta Accelerate Immunologically Mediated Pathology in Experimental Viral Fulminant Hepatitis. *PLoS Pathog* 2015;11:e1005155.
269. Athmer J, Fehr AR, Grunewald ME, et al. Selective Packaging in Murine Coronavirus Promotes Virulence by Limiting Type I Interferon Responses. *mBio* 2018;9.
270. Sheahan T, Morrison TE, Funkhouser W, et al. MyD88 is required for protection from lethal infection with a mouse-adapted SARS-CoV. *PLoS Pathog* 2008;4:e1000240.
271. Fehr AR, Athmer J, Channappanavar R, Phillips JM, Meyerholz DK, Perlman S. The nsp3 macrodomain promotes virulence in mice with coronavirus-induced encephalitis. *J Virol* 2015;89:1523-36.
272. Dickey LL, Worne CL, Glover JL, Lane TE, O'Connell RM. MicroRNA-155 enhances T cell trafficking and antiviral effector function in a model of coronavirus-induced neurologic disease. *J Neuroinflammation* 2016;13:240.

273. Gralinski LE, Bankhead A, 3rd, Jeng S, et al. Mechanisms of severe acute respiratory syndrome coronavirus-induced acute lung injury. *mBio* 2013;4.
274. Lassnig C, Sanchez CM, Egerbacher M, et al. Development of a transgenic mouse model susceptible to human coronavirus 229E. *Proc Natl Acad Sci U S A* 2005;102:8275-80.
275. Graham RL, Becker MM, Eckerle LD, Bolles M, Denison MR, Baric RS. A live, impaired-fidelity coronavirus vaccine protects in an aged, immunocompromised mouse model of lethal disease. *Nat Med* 2012;18:1820-6.
276. Page C, Goicochea L, Matthews K, et al. Induction of alternatively activated macrophages enhances pathogenesis during severe acute respiratory syndrome coronavirus infection. *J Virol* 2012;86:13334-49.
277. Gralinski LE, Menachery VD, Morgan AP, et al. Allelic Variation in the Toll-Like Receptor Adaptor Protein Ticam2 Contributes to SARS-Coronavirus Pathogenesis in Mice. *G3 (Bethesda)* 2017;7:1653-63.
278. Jacques A, Bleau C, Turbide C, Beauchemin N, Lamontagne L. Macrophage interleukin-6 and tumour necrosis factor-alpha are induced by coronavirus fixation to Toll-like receptor 2/heparan sulphate receptors but not carcinoembryonic cell adhesion antigen 1a. *Immunology* 2009;128:e181-92.
279. Totura AL, Whitmore A, Agnihothram S, et al. Toll-Like Receptor 3 Signaling via TRIF Contributes to a Protective Innate Immune Response to Severe Acute Respiratory Syndrome Coronavirus Infection. *mBio* 2015;6:e00638-15.
280. Ma XZ, Bartczak A, Zhang J, et al. Protein interferon-stimulated gene 15 conjugation delays but does not overcome coronavirus proliferation in a model of fulminant hepatitis. *J Virol* 2014;88:6195-204.





#### Chicken:

(Natural coronavirus: Infectious bronchitis virus [IBV])

*AKT1*  
*AvBD12*  
*CEP170B*  
*CRYL1*  
*CWF19L2*  
*DHRX*  
*FAM19A2*  
*GABRB3*  
*INTS9*  
*NMNAT3*  
*PINX1*  
*RAB39A*  
*VRK1*  
*YEATS2*  
*SETBP1*



#### Domestic cat:

(Natural coronaviruses: feline infectious peritonitis [FIPV] and feline enteric coronavirus [FECV])

*CD209*  
*ELMO1*  
*ERAP1*  
*ERAP2*  
*IFNG*  
*RRAGA*  
*TNFA*  
*TNSF10*



#### Pig:

(Natural coronaviruses: feline infectious peritonitis (transmissible gastroenteritis coronavirus [TGEV] and porcine transmissible gastroenteritis coronavirus [TGEV])

*ANPEP*  
*CMAH*



#### Mouse:

(Natural coronaviruses: mouse hepatitis virus [MHV]; mice have been extensively used to study human coronaviruses)

*Ace2*  
*Atg5*  
*Atp1a1*  
*B2m*  
*Bnip3*  
*C3*  
*C5ar1*  
*Ccr1*  
*Ccr2*  
*Ccr5*  
*Cd200r1*  
*Cxcl10*  
*Cxcl9*  
*Dpp4*

*Ebi3*  
*Foxn1*  
*H2-Ab1*  
*Ifih1*  
*Ifnar*  
*Ifng*  
*Ifngr1*  
*Ighm*  
*Il1r1*  
*Mavs*  
*Myd88*  
*Prkdc*  
*Rag1*  
*Serpine1*  
*Stat1*  
*Stat6*  
*Ticam2*  
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*Tlr7*  
*Trif*  
*Trim55*  
*Usp18*

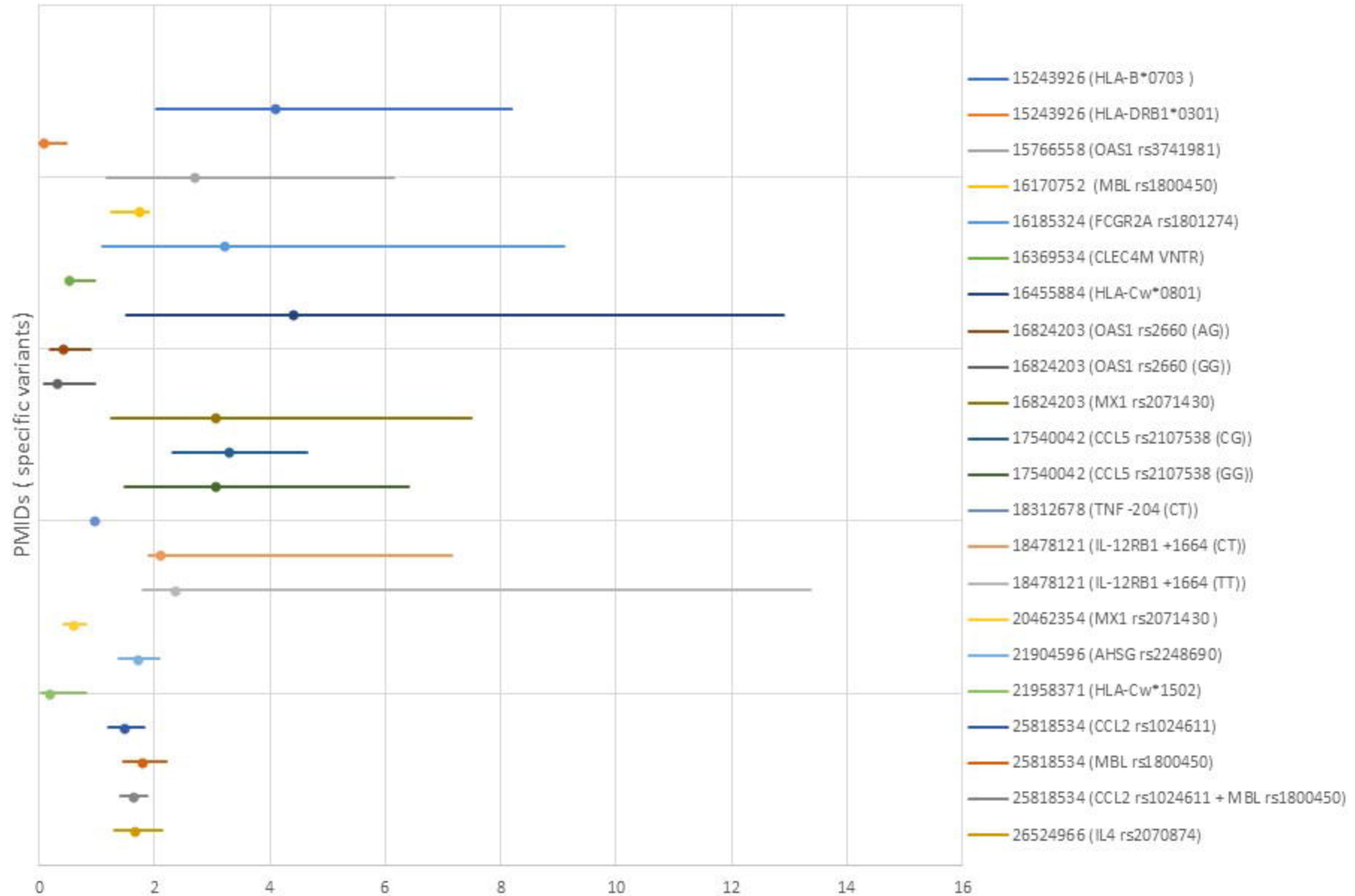


#### Multiple species (e.g., bat, camel, guinea pig, hamster, ferret, rat):

(Studies have been conducted on natural coronaviruses as well as human coronaviruses; co-evolution studies are not included)

*ACE2*  
*CD209L*  
*CEACAM1*  
*DPP4*  
*MHC*

Significant associations with susceptibility  
(includes both protective alleles and risk factors)



Significant associations with clinical variables/outcomes  
(includes both protective alleles and risk factors)

