

Fig. S1. PACRG co-elutes with endogenous LUBAC components.

HEK293T cells expressing PACRG were lysed, and soluble proteins were separated by size-exclusion chromatography. Fractions were collected and analyzed by Western blotting using antibodies against HOIP, HOIL-1L, SHARPIN, and PACRG. Blot is representative of 3 independent experiments.

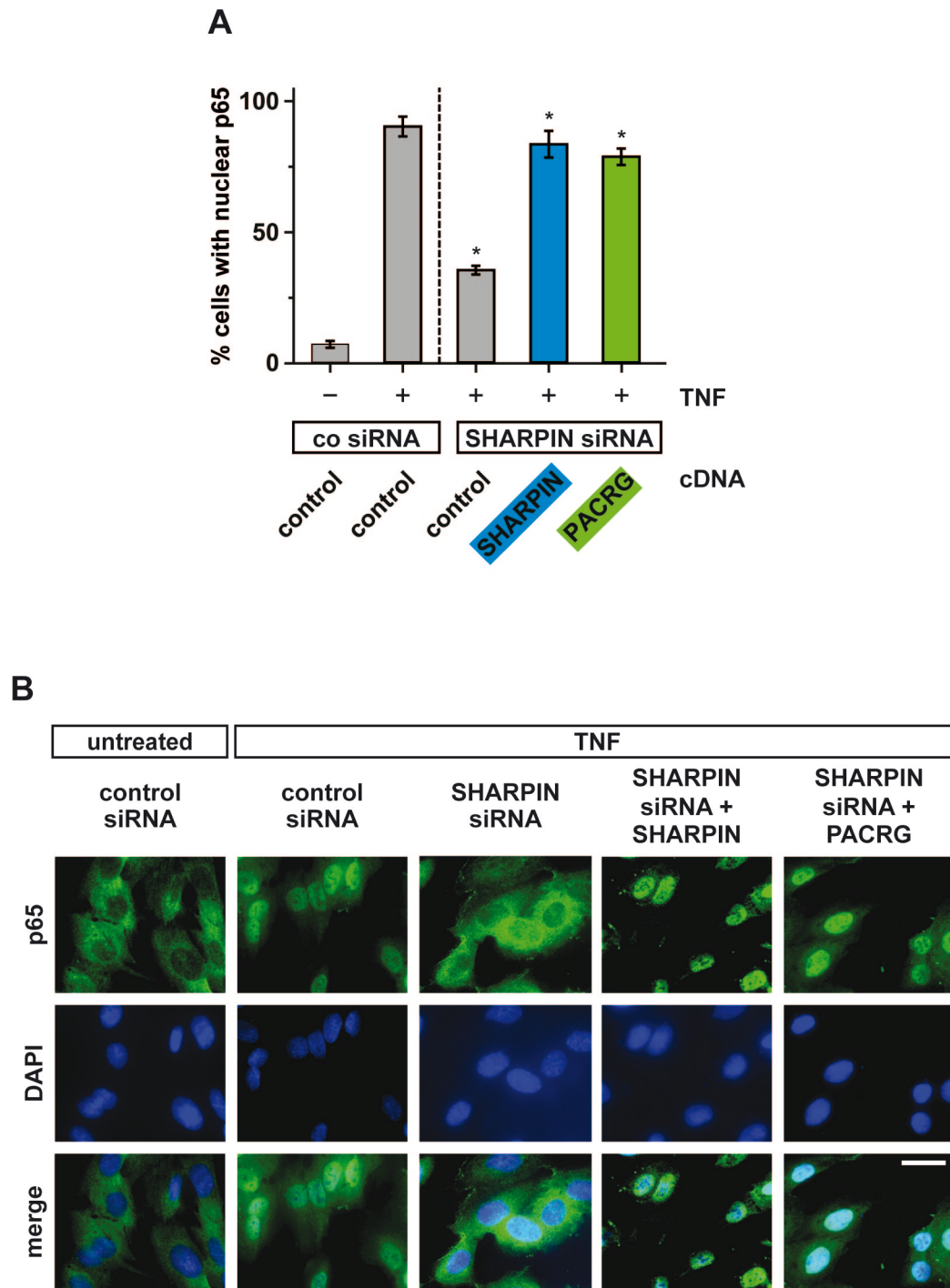


Fig. S2. PACRG restores defective p65 translocation in SHARPIN-deficient cells.

(A) SH-SY5Y cells were transfected with control or SHARPIN-specific siRNA with or without plasmids coding for SHARPIN or PACRG as indicated. One day after transfection, cells were stimulated with TNF, and nuclear translocation of p65 was determined by indirect immunofluorescence using an antibody against p65. Nuclei were stained with DAPI. Data represent the mean \pm SEM of 4 independent experiments. For statistical analysis one-tailed

Mann-Whitney U-test was performed. At least 100 cells were analyzed per condition. $*p \leq 0.05$.
(B) Representative examples of the indirect immunofluorescence experiment quantified in (A).
 Scale bar, 100 μm .

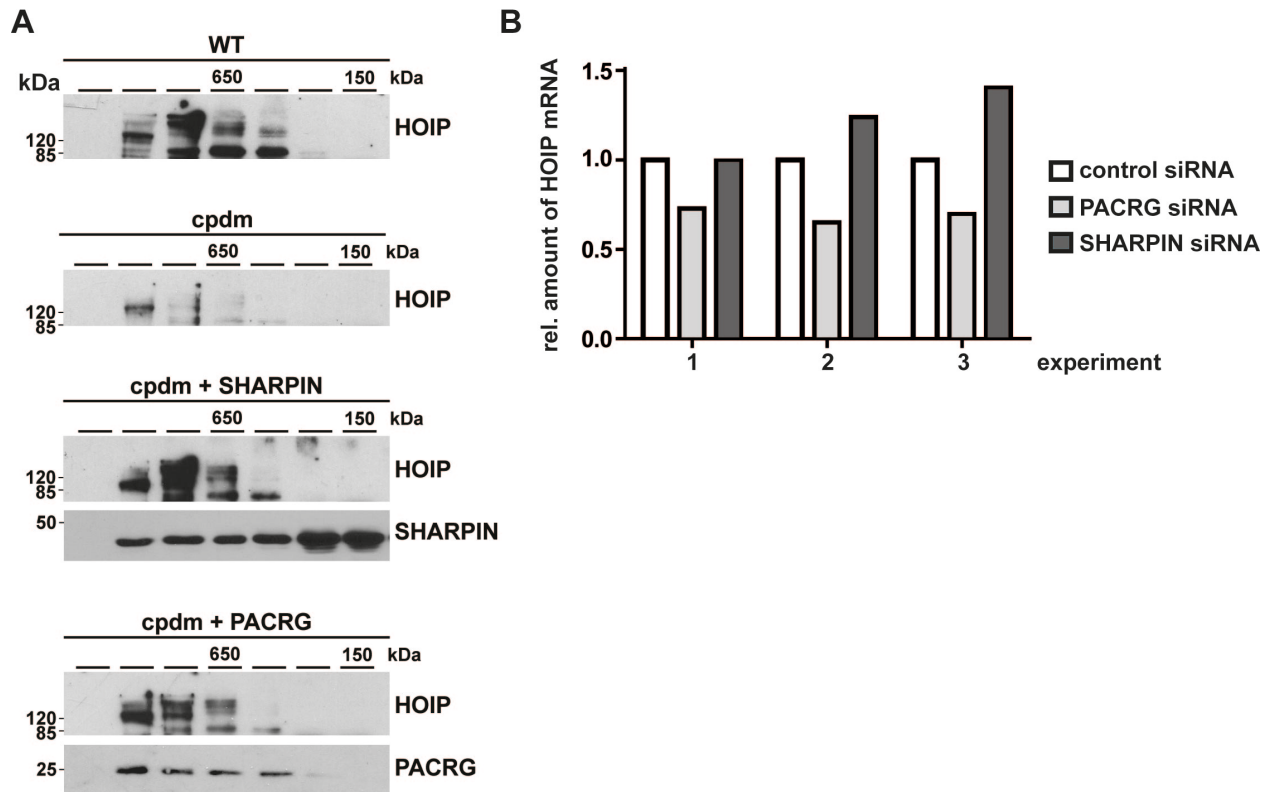


Fig. S3. PACRG stabilizes LUBAC.

(A) Wild-type (WT) MEFs, cpdm MEFs and cpdm MEFs expressing either SHARPIN or PACRG were lysed, and soluble proteins were separated by size-exclusion chromatography. Fractions were collected and analyzed by Western blotting using antibodies against HOIP, SHARPIN or PACRG. Blot is representative of 3 independent experiments. **(B)** Quantification of HOIP mRNA by real-time RT-PCR in MEFs silenced for PACRG or SHARPIN expression (corresponding to Fig. 5E).

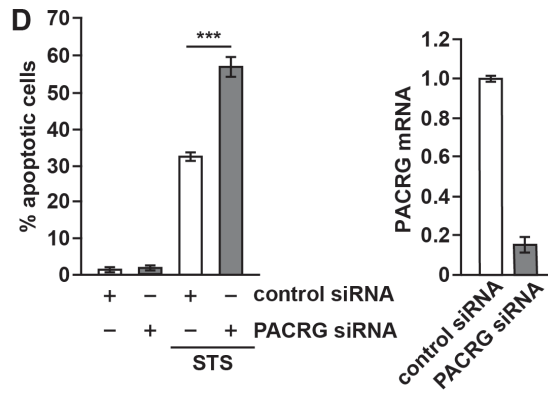
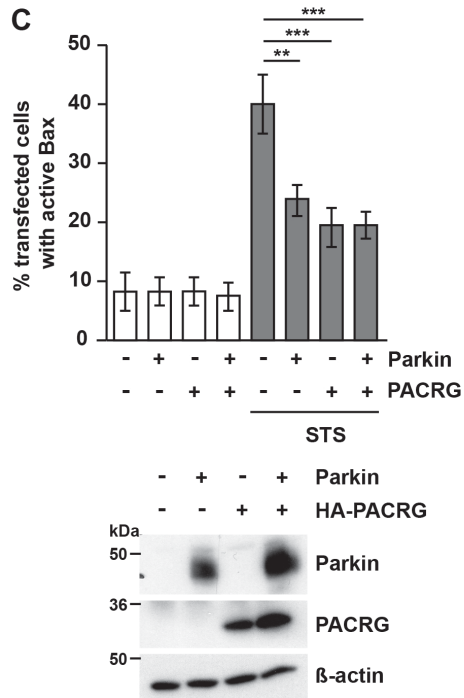
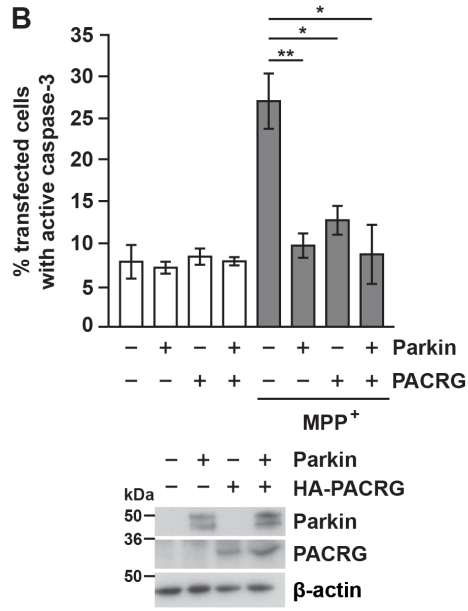
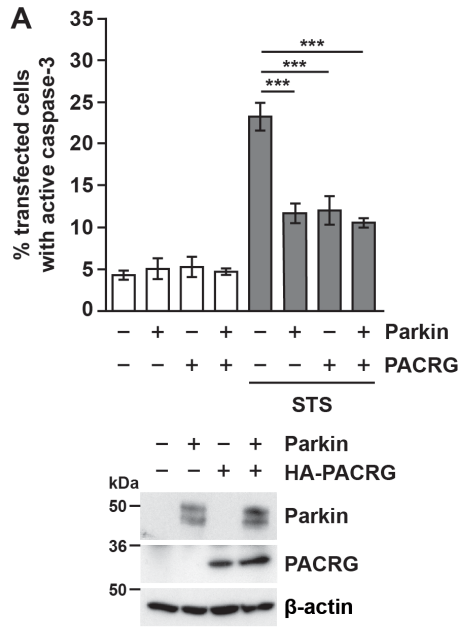


Fig. S4. PACRG protects against STS- and MPP⁺-induced apoptotic cell death.

(A, B) SH-SY5Y cells transiently expressing EYFP (control), HA-PACRG, Parkin, or HA-PACRG plus Parkin were treated with STS or MPP⁺. Apoptotic cell death was visualized by indirect immunofluorescence using an antibody against active caspase-3 and quantified by counting transfected cells positive for active caspase-3. Lysates were immunoblotted using antibodies specific for Parkin and HA. β -actin is a loading control. **(C)** SH-SY5Y cells transiently expressing EYFP, HA-PACRG, Parkin, or HA-PACRG plus Parkin were treated with STS. Apoptotic cell death was visualized by indirect immunofluorescence using an antibody against active Bax and quantified by counting transfected cells positive for active Bax. Lysates were immunoblotted using antibodies specific for Parkin, HA, and β -actin. **(D)** SY5Y cells transfected with control or PACRG siRNA were treated with STS. Apoptotic cell death was visualized by indirect immunofluorescence using an antibody against active caspase-3 and quantified by counting transfected cells positive for active caspase-3. PACRG knockdown efficiencies were determined by real-time RT-PCR. For all panels, data represent the mean \pm SEM of at least 3 independent experiments. At least 900 transfected SH-SY5Y cells were assessed per condition. Data represent the mean \pm SEM. Statistical analysis was carried out using the unpaired Student's t-test. * $p \leq 0.05$; ** $p \leq 0.01$; *** $p \leq 0.001$.

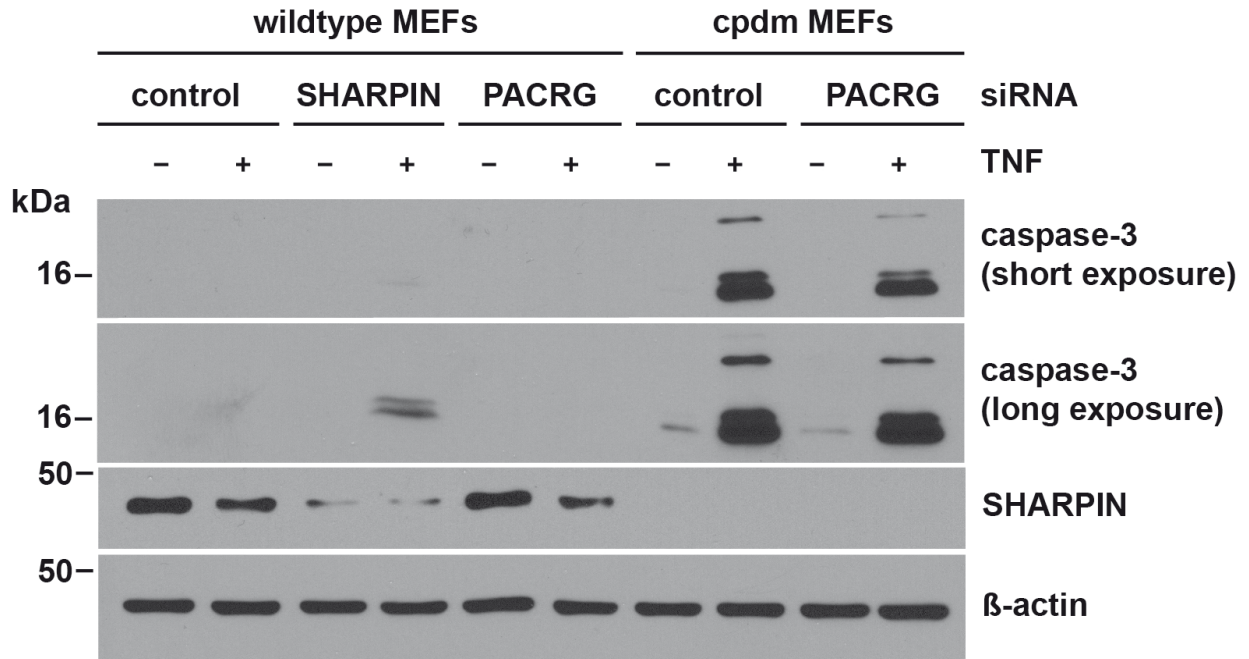


Fig. S5. TNF treatment of PACRG-silenced MEFs does not activate caspase-3.

Wild-type MEFs and cpdm MEFs were transfected with the indicated siRNAs then treated with TNF. Cell lysates were analyzed by Western blotting using an antibody against active caspase-3. Blot is representative of 3 independent experiments.

Table S1. Antibodies

| Antibody Target | Clone Name | Supplier | Dilution Western Blot | Dilution Immunocytochemistry |
|--|--------------------|------------------------|------------------------------|-------------------------------------|
| PACRG | C-8 | Santa Cruz | 1:1000 | |
| Parkin | PRK8 | Santa Cruz | 1:1000 | 1:500 |
| Hsp60 | N-20 | Santa Cruz | 1:1000 | 1:500 |
| p65 | A | Santa Cruz | 1:1000 | 1:500 |
| M1 ubiquitin | 1E3 | Millipore | 1:1000 | 1:200 |
| M1 ubiquitin | 1F11/3F5/Y10 2L | Genentech | 1:10000 | 1:4000 |
| HOIL-1L | AB38540 | Abcam | 1:1000 | |
| Cleaved Caspase-3 | 5A1E | Cell Signalling | 1:1000 | 1:400 |
| SHARPIN | 4444 | Cell Signalling | 1:1000 | |
| SHARPIN | 14626-1-AP | Proteintech | 1:2000 | |
| I κ B α | 9242 | Cell Signalling | 1:1000 | |
| HA tag | 16B12 | Covance | 1:1000 | 1:1000 |
| β -actin | AC-74 | Sigma Aldrich | 1:5000 | 1:1000 |
| HOIP | SAB2102031 | Sigma Aldrich | 1:1000 | |
| HOIP | A303-560A | Bethyl Laboratories | 1:1000- 1:2000 | 1:500 |
| NEMO | HPA000426 | Sigma Aldrich | 1:1000 | |
| BAX | 6A7 | eBioscience | 1:500 | 1:500 |
| anti-rabbit IgG (H+L) | Alexa488 | Thermo Scientific | | 1:1000 |
| anti-mouse IgG (H+L) | Alexa488 | Thermo Scientific | | 1:1000 |
| anti-rabbit IgG (H+L) | Alexa555 | Thermo Scientific | | 1:1000 |
| anti-goat IgG (H+L) | Alexa555 | Thermo Scientific | | 1:1000 |
| anti-mouse IgG (H+L) | HRP-tagged | Promega | 1:10000 | |
| anti-rabbit IgG (H+L) | HRP-tagged | Promega | 1:10000 | |
| anti-goat IgG (H+L) | HRP-tagged | Promega | 1:10000 | |
| anti-HA agarose (mouse) | A2095 | Sigma Aldrich | | |
| anti-cMyc agarose (mouse or rabbit) | #20168 | Thermo Scientific | | |
| anti-V5 agarose (mouse) | A7345 | Sigma Aldrich | | |