Periostin and tenascin-C interaction promotes

angiogenesis in ischemic proliferative retinopathy

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Supplementary Information



Supplementary Figure S1. Full-length gel images of Figure 2B and 2C

A: Full-length gel images of western blot analysis of PN, TNC and FN in the cell lysates of HRECs after stimulation by IL-13 at different time points. B: Full-length gel images of western blot analysis of co-immunoprecipitation assay of cell lysates from HRECs stimulated by IL-13. The cell lysates were immunoprecipitated with IgG, anti-PN antibodies, anti-TNC antibodies and anti-FN antibodies. The precipitates were analyzed by western blotting with anti-PN, anti-TNC and anti-FN antibodies. Cropped areas in the Figure 2B and 2C are lanes pointed by red arrows.



Supplementary Figure S2. Quantification of signal intensity of co-IP western blot samples. Signal intensity of co-IP samples (Figure 2C) were quantified by ImageJ software and relative intensity was calculated. Error bars represent standard deviations. *; p<0.05, Wilcoxon rank sum test, n=4/group.



Supplementary Figure S3. Normal retinal angiogenesis in developmental stage.

This image shows normal retinal angiogenesis of WT, PN-/-, TNC-/- and dKO mice at post natal 5. There seems to be no obvious difference in normal angiogenesis between these four strain of mice.



Supplementary Figure S4. Alteration of mRNA expression of HIF pathway and related genes. A: Alteration of mRNA expression of HRECs by IL-13 stimulation. HIF2 α and VEGFA was significantly up-regulated. *; p<0.05, Wilcoxon rank sum test. B: Effect of PN or TNC inhibition on IL-13 induced mRNA expression of HIF2 α and VEGFA. PN or TNC inhibition did not effect on the mRNA expression of HIF2 α and VEGFA.



Supplementary Figure S5. TNC mRNA expression was not affected by siRNA of PN

TNC mRNA expression after IL-13 stimulation with control RNA or PN siRNA transfection determined by qRT-PCR. mRNA levels were normalized to GAPDH mRNA expression. Relative mRNA levels are shown as fold changes.

Error bars represent standard deviations. Wilcoxon rank sum test, n=4/group



Supplementary Figure S6. Suppressive effect on PN and TNC expression by siRNA treatment. mRNA and protein expression after IL-13 stimulation with control or PN/TNC siRNA transfection. mRNA levels were determined by qRT-PCR, and were normalized to GAPDH mRNA expression. Relative mRNA levels are shown as fold changes. Error bars represent standard deviations. *; p<0.05, Wilcoxon rank sum test, n=4/group. Protein levels were determined by western blot.

Supplementary Table

ELISA kits

PN: #326070442; SHINO-TEST

TNC : #27767; IBL

FN : BMS2028; Invitrogen

Primers for SYBR green

PN : forward 5'-TGCCCAGCAGTTTTGCCCAT-3', reverse 5'-CGTTGCTCTCCAAACCTCTA-3' FN : forward 5'-GAGAATAAGCTGTACCATCGCAA-3', reverse 5'-CGACCACATAGGAAGTCCCAG-3' GAPDH : forward 5'-GAGTCAACGGATTTGGTCGT-3', reverse 5'-CTTGATTTTGGAGGGATCTCGC-3' HIF1α : forward 5'-TATGAGCCAGAAGAACTTTTAGGC-3' reverse 5'-CACCTCTTTTGGCAAGCATCCTG-3' HIF1β : forward 5'-CTGCCAACCCCGAAATGACAT-3' reverse 5'-CGCCGCTTAATAGCCCTCTG-3' HIF2α : forward 5'-TTGCTCTGAAAACGAGTCCGA-3' reverse 5'-GGTCACCACGGCAATGAAAC-3' pVHL : forward 5'-GGAGCCTAGTCAAGCCTGAGA-3' reverse 5'-CATCCGTTGATGTGCAATGCG-3' VEGFA : forward 5'-CTTGCCTTGCTGCTCTACC-3' reverse 5'-CACACAGGATGGCTTGAAG-3' VEGER2 : forward 5'-GGCCCAATAATCAGAGTGGCA-3' reverse 5'-TGTCATTTCCGATCACTTTTGGA-3' ANGPT2 : forward 5'-AACTTTCGGAAGAGCATGGAC-3' reverse 5'-CGAGTCATCGTATTCGAGCGG-3'

TIE2 : forward 5'-TTAGCCAGCTTAGTTCTCTGTGG-3' reverse 5'-AGCATCAGATACAAGAGGTAGGG-3'

Supplementary Table

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TaqMan probe for qRT-PCR

TNC : Hs01115665_m1; Applied Biosystems GAPDH : Hs02758991_g1; Applied Biosystems

Primary antibodies for immunohistochemistry and immunofluorescence

PN : AF3548; R&D systems, 10 µg/ml (for immunohistochemistry of FVMs) 201466-T10; Sino Biological, 1/200 (for immunofluorescence of FVM sections) 50450-RP02; Sino Biological, 1/500 (for immunohistochemistry and immunofluorescence of mice OIR sections) TNC: NB110-68136; Novus Biologicals, 1/30 FN : ab23750; abcam, 1/40 (for immunohistochemistry of FVMs and mice OIR sections and immunofluorescence of FVM sections) MAB88916; Millipore, 1/50 (for immunofluorescence of mice OIR sections) CD34 : ab81289; abcam, 1/100 (for immunohistochemistry of FVMs and mice OIR sections) RAM34; Thermo Fisher, 1/100 (for immunofluorescence of mice OIR sections) Secondary antibodies for immunofluorescence

Anti-goat : A11056; Invitrogen, 1/1000 Anti-mouse : A21245; Invitrogen, 1/1000 Anti-rabbit : A21235; Invitrogen, 1/1000

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Primary antibodies for western blotting

PN : AF3548, 1 μg/ml TNC : NB110-68136, 5 μg/ml for lysate #12221; CST, 1/500 for blotting of immunoprecipitation FN : ab23750, 1 μg/ml α-Tublin : #2125; CST, 1/1000

HRP conjugated secondary antibody for western blotting

Anti-goat : P0449; DAKO, 1/500 Anti-mouse : P0447; DAKO, 1/500 Anti-rabbit : P0448; DAKO, 1/500

Antibodies for immunoprecipitation

PN : AF3548, 25 μg/ml TNC : #12221, 1/25 FN : ab2413; abcam, 25 μg/ml Goat IgG : NI02; Millipore, 25 μg/ml Rabbit IgG : NI01; Millipore, 25 μg/ml

HRP conjugated antibody for immunoprecipitation

Anti-goat : 18-8814-31; ROCKLAND, 1/1000 Anti-mouse : 18-8817-33; ROCKLAND, 1/1000 Anti-rabbit : 18-8816-33; ROCKLAND, 1/1000

siRNAs

PN : sc-61324; Santa Cruz Biotechnology TNC : sense 5'-CUGAAAUUGGAAACUUAAATT-3', antisense 5'- UUUAAGUUUCCAAUUUCAGTT-3' FN : SASI_Hs01_00207242 ; Sigma-Aldrich

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